

Supplementation of *Acacia dealbata* versus *Acacia mearnsii* leaf-meal has potential to maintain growth performance of lambs grazing low-quality communal rangelands in South Africa

Lovelyn Hellen Mushunje

Stellenbosch University

Tawanda Marandure

Swedish University of Agricultural Sciences: Sveriges lantbruksuniversitet

Obert Chenjerai Chikwanha

Stellenbosch University

James Bennett

Coventry University

Heidi Hawkins

Conservation International

Antony Palmer

Rhodes University

Lianhai Wu

Chinese Academy of Sciences

Munyaradzi Marufu

University of Pretoria

Cletos Mapiye

`cmapiye@sun.ac.za`

Stellenbosch University <https://orcid.org/0000-0002-1474-8648>

Research Article

Keywords: *Acacia* foliage, faecal egg count, growth performance, polyphenols, South Africa

Posted Date: March 7th, 2024

DOI: <https://doi.org/10.21203/rs.3.rs-3967742/v1>

License:  This work is licensed under a Creative Commons Attribution 4.0 International License. [Read Full License](#)

Version of Record: A version of this preprint was published at Tropical Animal Health and Production on May 1st, 2024. See the published version at <https://doi.org/10.1007/s11250-024-04004-z>.

Abstract

Supplementing livestock grazing communal rangelands with leaf-meals from *Acacia* trees, which are currently considered as problematic invasive alien plants globally, may be a sustainable way of exploiting their desirable nutritional and anthelmintic properties. The current study evaluated worm burdens and growth performance of lambs grazing low-quality communal rangelands supplemented with leaf-meals prepared from the invasive alien plant species; *Acacia mearnsii* or *A. dealbata*. Forty, three-month-old ewe lambs weighing an average of 18.9 ± 0.60 kg were randomly allocated to four supplementary diets: (1) rangeland hay only (control), (2) commercial protein supplement plus rangeland hay, (3) *A. mearnsii* leaf-meal plus rangeland hay and (4) *A. dealbata* leaf-meal plus rangeland hay. All the supplementary diets were formulated to meet the lambs' minimum maintenance requirements for protein. All the lambs were grazed on communal rangelands daily from 0800 to 1400 after which they were penned to allow them access to their respective supplementary diets until 08:00 the following morning. The respective supplementary diets were offered at the rate of $400 \text{ g ewe}^{-1} \text{ day}^{-1}$ for 60 days. Lambs fed the commercial protein supplement had the highest dry matter intake followed by those fed the *Acacia* leaf-meals and the control diet, respectively ($P \leq 0.05$). Relative to the other supplementary diets, lambs fed the commercial protein supplement and *A. dealbata* leaf-meal had higher ($P \leq 0.05$) final body weight and average daily gains. Dietary supplementation did not affect lamb faecal worm egg counts over the study period ($P > 0.05$). There was no association between supplementary diets and lamb FAMACHA© scores ($P > 0.05$). It was concluded that the *Acacia dealbata* relative to *Acacia mearnsii* has potential to maintain growth performance of lambs grazing communal rangelands in the dry season.

1. Introduction

Seasonal fluctuations in quality and quantity of natural forage are largely responsible for the limitations for inferior performance in livestock grazing communal rangelands (Mphinyane et al., 2015; Mthi and Nyangiwe, 2018). A significant drop in nutritive value, particularly crude protein (CP) from about 12% in the wet season to as low as < 2% in the dry season (Mapiye et al., 2009; Moyo et al., 2012) result in drastic shortfalls in meeting the protein requirements for both maintenance and production of sheep (NRC, 2007). In addition, the proliferation of invasive alien plants (IAPs), especially Australian Acacias (i.e., *Acacia dealbata* and *A. mearnsii*) has reduced rangeland production (Yapi et al., 2018; Gouws and Shackleton, 2019) and impacted negatively on livestock health, meat production and quality (Dezah et al., 2021; Uushona et al., 2021). Invasive Acacias are currently harvested by local communities for timber and fuelwood, and this often yields by-products such as leaves and bark, which currently have limited local value. Strategies are being developed in the tropics to increase the economic value of invasive Acacias' leaves and bark to local communities, including their use as livestock feed and meat preservatives because they contain moderate to high levels of nutrients and polyphenols (Dezah et al., 2021; Uushona et al., 2021; 2022).

Effective exploitation of leaves from *Acacia* species could be sustainable way of managing invasions by these alien plants and may provide a lucrative alternative to more expensive processed protein sources for resource-poor farmers, whose animals mostly rely on communal rangelands throughout the year (Mapiliyao, 2012; Cheema et al., 2011; Maguraushe, 2015). Communal farmers have previously attested to observing increased browsing of *Acacia* leaves by ruminants in the dry season to compensate for the decline in forage quality (Mphinyane et al., 2015). However, high condensed tannins (CT) content in *Acacia* leaves, affects their intake and digestibility by ruminants (Payne et al., 2013; Chikwanha et al., 2023; Uushona et al., 2023). Co-feeding these browse species with low condensed tannin-containing ingredients such as rangeland hay could be a cost-effective way of reducing the negative effects of these secondary metabolites (Salem and Smith, 2008).

Dezah et al. (2021) and Uushona et al. (2021) reported consistently high levels of CP (16.2%), and NDF (36.1%) in Australian *Acacia* foliage. This is consonant with Acacias being N-fixing and staying green during the dry season due to

their long tap roots that can reach deep, ground water sources (Mapiliyao, 2010; Cheema et al., 2011; Maguraushe, 2015). In addition to being limited by low quantity and quality forages, particularly in the dry season, ruminant livestock production in communal areas is compromised by high loads of gastrointestinal parasites (Mapiliyao, 2010). Given the predominance of sheep over other ruminant livestock among communal farmers in the Eastern Cape Province of South Africa (StatsSA, 2017), it is important to focus on the species for testing the complementary anthelmintic and feed supplement potential of acacia leaf-meals. Previous studies that investigated the feed and anthelmintic potential of *Acacia* species on livestock growth performance and parasitic worm burden largely focused on bark tannin extracts. For example, Cenci et al. (2007), observed better growth performance and lower nematode loads in sheep fed *A. mearnsii* extracts containing up to $3.2 \text{ gCT kg}^{-1} \text{ DM}$. On the contrary, low average daily gain (ADG) and carcass weights were reported for lambs fed *A. mearnsii* extracts containing $\geq 60 \text{ g CT kg}^{-1} \text{ DM}$ (Costa et al., 2021). Recently, feeding *A. mearnsii* leaf-meal up to $100 \text{ gkg}^{-1} \text{ DM}$ replacing *Triticum aestivum* (wheat) bran in lamb finisher diets had no effect on growth performance and carcass weights (Uushona et al., 2023b). To the authors' knowledge, there have been no studies on the use of *Acacia* leaf-meals to supplement sheep grazing communal rangelands in South Africa in the dry season. The objective of the current study was, therefore, to evaluate the effects of supplementing *A. mearnsii* and *A. dealbata* leaf-meals on worm burdens and growth performance of lambs grazing South African communal rangelands in the dry season.

2. Materials and methods

2.1 Study area

The study was conducted in the Magxeni community in the Mvenyane area (Matatiele, Eastern Cape, South Africa; (30° 32' 42" S 29° 3' 21"E). The community was purposively selected because of its active involvement with a non-profit organization (NPO) called Conservation South Africa (CSA). The NPO assisted farmers in the Magxeni community to form a grazing association to help them practice sustainable rangeland and livestock management in return for various incentives, including livestock vaccinations, organising sheep auctions and offering farmer training programs. Magxeni community lies within the East Griqualand Grassland vegetation unit of the sub-escarpment Grassland bioregion (Mucina and Rutherford, 2006). It is characterised by well-drained, erosion-prone soils and hilly terrain with an altitude range of 920–1740 m above sea level. The region receives summer rainfall and winter snowfall (at high altitudes), with about 30 days of annual frost, mean annual precipitation of approximately 780 mm and a mean annual temperature of 14.7°C (Gouws and Shackleton, 2019). The area is heavily invaded by *Acacia* species up to 7000 stems/ ha with an annual increase of the *Acacia* biomass invasion estimated to be 26% (Gouws and Shackleton, 2019). In Mvenyani district about 50% of the area is invaded by *A. mearnsii* and *A. dealbata* (Gouws and Shackleton, 2019).

2.2 Sourcing of feed ingredients and preparations of experimental diets

Acacia leaf-meals were harvested within a 10 km radius of the study site between September and October 2020 by cutting small *A. mearnsii* and *A. dealbata* trees (1.5-2.0 m tall) with a hand-held petrol chainsaw. Trees were cut into small branches, which were then stacked on plastic sheets and sun-dried for four days. On the fifth day, dry leaves were shaken off branches, collected from the plastic sheet, bagged and stored in a well-ventilated, dry shaded area. Rangeland hay (~ 70 g CP/ kg DM) harvested in the same season was sourced from a nearby commercial farm and ground using a hammer mill fitted with a 30 mm sieve to allow mixing with the *Acacia* leaf meals. A commercial protein supplement (Protein Lick 40, Molatek, South Africa) was purchased from an animal feed store in Matatiele (Eastern Cape Province, South Africa).

Supplementary diets were formulated using a Pearson square method to meet the lambs' daily minimum maintenance requirements for protein (i.e., 70 g CP/ kg DM; NRC, 2007). The diets were prepared by mixing rangeland hay with either commercial protein or *Acacia* leaf-meals supplements. The four diets formulated contained (1) rangeland hay only, (2) 94% rangeland hay plus 6% commercial protein supplement, (3) 78% rangeland hay plus 22% of either *A. mearnsii* leaf-meal and (4) or *A. dealbata* leaf-meal. The supplementary diets were bagged, labelled, and stored in a well-ventilated cool-dry shed before feeding. All the diets were premixed at the beginning of the trial and safely stored in a dry, well-ventilated place for the duration of the trial.

2.3 Nutritional composition of experimental diets

Samples from rangeland biomass and each diet and were collected weekly and separately pooled for analyses of nutritional composition. Feed ingredients and diets were analysed for dry matter (DM), ash and ether extract (EE) using AOAC International (2002) procedures. A macro-Nitrogen analyser (LECO FP528, LECO Corporation, Miami, USA) was used to determine the total nitrogen content which was multiplied by a factor of 6.25 to calculate crude protein (CP) content. Neutral detergent fibre (NDFom), acid detergent fibre (ADFom) and acid detergent lignin (ADL) of the feed ingredients and diets were analysed using an ANKOM 200 Fibre Analyzer (Ankom Technology Corp., Macedon, NY, USA). Total phenol and tannins were determined using Makkar's (2003) method. Condensed tannins were determined following the procedure by Porter et al. (1985). Analyses of all the proximate, fibre and polyphenolic compositions were done in triplicate. The nutritional composition of the experimental diets and rangeland biomass are shown in Table 1.

Table 1
Nutritional composition of the experimental diets and rangeland biomass (%)

Parameter	Diets		Rangeland biomass					
	Rangeland hay (control)	Protein supplement	<i>A. mearnsii</i>	<i>A. dealbata</i>	August	September	October	SD
Dry matter (%)	79.9	79.9	82.7	72.7	85.7	75.3	83.9	4.30
Crude protein (%)	6.9	9.8	8.1	8.6	7.2	6.4	8.6	0.14
Ether extract (%)	0.8	0.9	0.8	0.6	0.7	0.4	0.4	0.07
Ash (%)	4.2	10.1	4.2	4.6	5.2	4.8	5.1	0.11
Neutral detergent fibre	67.6	65.0	62.3	61.5	66.0	68.2	66.1	1.87
Acid detergent fibre	37.4	36.4	35.6	34.9	37.2	36.8	33.7	0.40
Acid detergent lignin	37.9	36.6	35.9	35.2	37.2	37.2	34.2	0.41
Metabolisable energy (MJ/kg DM)	7.7	7.9	8.1	8.3	7.7	7.8	8.6	0.34
Total phenolics (g /kg DM)	NA	NA	23.0	20.1	NA	NA	NA	0.10
Total tannins (g/kg DM)	NA	NA	2.2	2.6	NA	NA	NA	02.8
Condensed tannins (g/kg DM)	NA	NA	10.3	14.3	NA	NA	NA	1.16
SD – standard deviation, DM – dry matter, NA – not applicable								

2.4 Management of experimental animals

Ethical approval was obtained from the Stellenbosch University Research Ethics Committee for Animal Care and Use (ACU-2020-11758). Forty, 3-months-old Dohne Merino ewe lambs (18.9 ± 0.60 kg) were purchased from a commercial farmer in Matatiele. The animals were ear-tagged for identification but not drenched for internal parasites. Lambs were randomly allocated to the four treatment diets. Each day the lambs grazed communal rangelands and drank water from the river between 0800 and 1400. They were expected to consume 60% of their daily feed intake within the 6 h grazing period (Dias-Silva and Filho, 2021). For the remainder of each day, the animals were individually housed in pens measuring 1.5×1.5 m from 1400 and 0800 and provided with supplementary diets at the rate of $400 \text{ g ewe}^{-1} \text{ day}^{-1}$ (~2% of body weight) and offered clean freshwater *ad libitum*. The animals were adapted to the supplementary diets for 21 days followed by 60 days of data collection (August to October 2021).

2.5 Feed intake and body weights of lambs

During the trial, all the feed offered, and refusals were weighed and recorded daily before feeding in the morning. Feed was given at 10% extra from the previous day's consumption. Lambs were weighed at the beginning of the experiment and thereafter biweekly to give a total of seven measurements until the end of the experimental period using a commercial scale (Rudd scale, Durbanville, South Africa). The animals were weighed in the morning (0600) before being released for grazing. The average daily weight gain was calculated by subtracting the initial body weight from the final body weight and divided by the number of days on feed excluding the adaptation period.

2.6 Faecal worm egg counts and FAMACHA scores

Faecal samples were collected directly from the rectum using the faecal grab technique (Da Costa et al., 2019). Samples were placed in labelled zip lock bags, temporarily stored in a cooler box at 4°C and transported to Queenstown Provincial Veterinary laboratories for determination of faecal nematode egg counts using the McMaster Technique (Miller, 1997). The number of nematode eggs of the McMaster chamber was multiplied by 50 to obtain eggs per gram. FAMACHA scores were determined every two weeks during the trial. Each animal was visually examined by a trained person and classified into one of five categories according to the FAMACHA® eye colour chart (Van Wyk, 2008).

2.7 Statistical analyses

All the data were analysed with SAS v. 9.4 (SAS Institute Inc. Cary, NC, USA). Before analysis, the faecal worm egg count (FEC) values were checked for normality using the PROC UNIVARIATE procedure and transformed to conform to normality by a box-cox transformation using the following formula; $y\lambda = \frac{y^\lambda - 1}{\lambda}$, ($\lambda = 0.25$), where y is the response variable and λ is estimated using the maximum likelihood estimate adapted from the algorithm proposed by Hyde (1999) using the TRANSREG procedure SAS v. 9.4 (SAS Institute Inc. Cary, NC, USA).

The effect of diet on growth performance and FEC was analysed using PROC MIXED procedures SAS v. 9.4 (SAS Institute Inc. Cary, NC, USA). Diet, time on feed and their interaction were fitted as fixed effects, while the animal within a diet was fitted as a random variable, time on feed was a repeated measure and initial weight was a covariate. The following model was used: $Y_{ijk} = \mu + D_i + T_j + (DT)_{ij} + \epsilon_{ijk}$; Where: Y_{ijk} = final weight, ADG, faecal egg count; μ = overall mean; D_i = effect of diet ($i = A. mearnsii$, $A. dealbata$, protein supplement and rangeland hay only); T_j = Time on feed (j = week 1, 3, 5, 7 and 9); $(DT)_{ij}$ = interaction i^{th} diet and j^{th} time on feed and ϵ_{ijk} = residual error. Pair-wise comparisons of the least square means were performed using the PDIFF test with a TUKEY adjustment in SAS version 9.4. The significance threshold for all statistical analyses was set at $P \leq 0.05$. The association between diet and FAMACHA® scores was performed using the chi-square test.

3. Results

3.1 Nutritional composition of the experimental diets and rangeland biomass

The nutritional composition of the experimental diets and the rangelands are shown in Table 1. The protein supplement diet had greatest CP content followed by the *Acacia* leaf-meal diets with the rangeland hay diet (control) having the least. Relative to the other diets, ash content was highest for the protein supplement diet. The fibre components (NDF, ADF and ADL) decreased in the order of rangeland hay > protein supplement > *A. mearnsii* > *A. dealbata*. The *Acacia*

dealbata had higher CT than *A. mearnsii*. Rangeland grazing hay for October had higher CP content and lower ADF and ADL contents than the other months.

3.2 Effect of diet on growth performance of lambs

The commercial protein supplement diet had the highest DMI with intermediate values observed for the *Acacia* diets and the control diet had the least intake ($P \leq 0.05$; Table 2). Diet and time on feed interaction influenced weight of lambs ($P \leq 0.05$; Fig. 1). Body weight increased for lambs on the *A. dealbata* diet until week 3 thereafter decline until week 7 and then increased to week 9. Body weight of lambs on *A. mearnsii* gradually declined until week 7 and then increased up to week 9. Lambs on the protein supplement consistently increased weights throughout the trial period. Body weights for control lambs was not consistent, it increased to week 3 and thereafter declined week 5, increased to week 7 and then declined to week 9. Lambs fed the protein supplement and *A. dealbata* diets had similar ($P > 0.05$) final body weight and ADG, which were higher ($P \leq 0.05$) than that the control and *A. mearnsii* dietary treatments, which were not different ($P > 0.05$) from each other.

Table 2
Least square means ($\pm$ standard error of mean) of growth performance of lambs grazing South African communal rangelands supplemented with *Acacia* leaf-meals

Parameter	Diet				P-value
	Rangeland hay	Protein supplement	<i>A. mearnsii</i>	<i>A. dealbata</i>	
Initial body weight (kg)	19.2 $\pm$ 0.06	19.2 $\pm$ 0.06	19.2 $\pm$ 0.05	19.2 $\pm$ 0.06	0.2532
Final body weight (kg)	20.0 $\pm$ 0.01 ^b	20.7 $\pm$ 0.03 ^a	19.8 $\pm$ 0.04 ^b	20.5 $\pm$ 0.02 ^a	0.0499
Dry matter intake (g)	274.4 $\pm$ 5.22 ^c	318.9 $\pm$ 4.92 ^a	288.6 $\pm$ 5.01 ^b	290.4 $\pm$ 5.21 ^b	< 0.0001
Average daily gain (g)	13.7 $\pm$ 2.67 ^b	25.0 $\pm$ 3.07 ^a	10.7 $\pm$ 2.33 ^b	21.9 $\pm$ 3.67 ^a	0.0399
^{a, b} Values with different superscripts within a row are significantly different ($P \leq 0.05$).					

3.3 Effect of diet on faecal worm egg counts and FAMACHA scores

Diet and time on feed interaction influenced coccidia and strongyle egg counts ($P \leq 0.05$; Fig. 2). Faecal egg counts for coccidia increased for all the diets from week 1 to 7 and then declined to week 9 except for the *A. mearnsii* diet FEC which increased throughout the trial ($P \leq 0.05$). At the end of the trial, coccidia FEC were in the order of *A. mearnsii* > *A. dealbata* > rangeland hay > protein supplement ($P \leq 0.05$). For the *A. mearnsii* diet, strongyle FEC declined ($P \leq 0.05$) from week 1 until week 5 and plateaued ($P > 0.05$) to week 9, while the FEC for the rest of the diets increased ($P \leq 0.05$) from week 1 to 3, declined ($P \leq 0.05$) to week 5 and then remained constant until week 9 without significant differences among them ($P > 0.05$; Fig. 3). There was no association between supplementary diet and FAMACHA scores ($P > 0.05$; Table 3). However, at the end of the trial, there were more lambs observed to be mildly anaemic (C3; Table 3) than at the beginning of the experiment.

Table 3

FAMACHA scores (%) of lambs grazing South African communal rangelands supplemented with Acacia leaf-meals

Parameter	Control	Protein supplement	<i>A. mearnsii</i>	<i>A. dealbata</i>	P-values	Cramer's V
Initial scores						
Non-anaemic (red ocular membrane, A1)	7.5	10	10	10	0.8741	0.1208
Non-anaemic (red/pinkish ocular membrane, B2)	17.5	15	15	15		
Final scores						
Non-anaemic (red/pinkish ocular membrane, B2)	2.8	0	5.6	2.8	0.4517	0.2826
Mildly anaemic (pink ocular membrane, C3)	19.4	22.2	16.7	22.2		
Anaemic (pink, white ocular membrane, D4)	2.8	5.6	0	0		

4. Discussion

The lower average DMI recorded for the rangeland hay and *Acacia* diets compared to the protein supplement diet could be attributed to palatability of the respective diets. It has been demonstrated that palatability is positively correlated with solubility of the dietary components, with protein diets being more soluble, whereas structural components, such as NDF, ADL and phenols present in mature hay and *Acacia* diets are less soluble (Rubanza et al., 2007; Gebeyew et al., 2015). A combination of low CP and high fibre (i.e., NDF and ADL) and phenols negatively influenced DMI of the test diets compared to the control. The relatively high levels of CT present in the *Acacia* diets negatively influenced palatability through the formation of a tannin-salivary protein complex that causes an astringent taste in the mouth and producing a feeling of constriction, dryness and roughness in the oral cavity (Waghorn, 2008; Tadele Yilkal., 2015; Jerónimo et al., 2016). Moreover, CT inhibit activities of ruminal microorganisms by binding to proteins and forming stable complexes in the rumen (Priolo et al., 2000). Priolo et al. (2000) reported that a diet containing 2.5% CT greatly reduced feed palatability and digestibility. Although CT levels in the *Acacia* diets used in the current study are well below 2.5%, their combination with fibre and other phenols could have contributed to the reported differences in DMI across the experimental diets.

The higher final weight and ADG for lambs fed the protein supplement diet corresponds with their higher DMI compared to the other diets. Results concur with Obeidat et al. (2020) who reported high final body weights and ADG on protein diets compared to low quality forage diets. The high ADG observed in lambs supplemented with *A. dealbata* may be partly explained by slightly higher dietary CP and faster adaptation of animals to somewhat high dietary CT diet compared to those on the *A. mearnsii* diet. Declining weights recorded for lambs fed *A. mearnsii* up to week 7 may be reflective of strenuous adaptation to the lower CT diet. Overall, animals fed diets high in CT diets are likely to adapt faster than those fed diets contain less CT (Min et al., 2021). The low ADG for the control diet can be explained by the low levels of protein in the diet, which might have failed to meet the 7% CP requirements for the maintenance of the grazing dry ewes (Nicol and Brooks, 2007).

Average daily gain values (13–25 g/day) recorded in the current study fall far short of the 50 g/day minimum range required to maintain grazing lambs (Vipond et al. 2009; Roberts, 2020; 2022). This could be linked to a combination of

low biomass, nutrient content, and digestibility of the rangeland forages in the current study. Average biomass values of 0.6 t/ ha estimated for the communal grazing in the study area (Matshawule, 2020) were lower than 2 t/ ha, the recommended minimum biomass to meet energy requirements for sheep maintenance on dry natural pasture (Roberts, 2020). Kym et al. (2018) revealed that when energy falls below maintenance for grazing sheep, protein is also utilised less efficiently.

The lowest coccidia faecal oocyst counts observed in the protein supplement diet could be attributed to the better nutritional profile resulting in stronger immune system of the lambs (Ingvarlsen and Moyes, 2013; Bobeck, 2020). Inclusion of CT levels below 1.6% previously led to no effect on FEC in sheep (Hoste et al., 2006). Generally, the ability of an animal to respond to parasite infections is highly associated with nutritional status, especially protein levels (Hoste et al., 2006) and the content of CT in the diet (Mahachi et al., 2020). The inclusion of tanniferous-rich feeds with at least 2.5% CT in small ruminants' diets has been reported to suppress parasitic infections through inhibiting nematode egg hatching and larval eclosion (Priolo et al., 2000; Hoste et al., 2006; Mahachi et al., 2020). The FEC trend reported in the current study contrasts with that reported by Cenci et al. (2007), who previously recorded significant effects of a tanniferous diet on FEC after the eighth week and partly related it to the adaptation of the animal to the diet. The difference with the trend presented in the current study could be largely due the low CT content of the diet as well as short duration of the current trial.

The initial low mean FAMACHA scores indicated that lambs were healthy and non-anaemic at the beginning of the trial. High mean scores at the end of the trial were a proxy of mildly anaemic and anaemic ewes which reflected a significant increase in worm burdens. Worm burdens might have presented an additional nutrient demand (Max et al., 2007) and competed for the nutrients with lambs leaving them in a negative nutrient balance. Overall, the low ADG and final body weights recorded in the current study could have been influenced by a combination of low nutrient quality of the communal rangelands and presence of gastrointestinal worms.

5. Conclusions

It was concluded that supplementing low-quality communal rangelands with *A. dealbata* leaf-meal has potential to maintain lamb growth performance compared to *A. mearnsii* leaf-meal, but both do not affect worm burdens. A follow-up study is recommended to determine the optimum inclusion levels of *A. dealbata* leaf-meal so that worm burdens are lowered while maintaining the growth performance of lambs grazing low-quality communal rangelands in the dry season.

Declarations

Funding

This work was supported by a collaborative Biotechnology and Biological Sciences Research Council-Global Challenges Research Fund (BBSRC-GCRF) funded project (Grant 628 number: BB/S014683/1). Author James Bennet has received research support from BBSRC-GCRF.

Competing Interests

All authors declare that there are no actual or potential conflicts of interest including financial, personal, or other relationships with other people or organisations that could inappropriately influence, or be perceived to influence, this work.

Author Contributions

Lovelyn Hellen Mushunje: Was responsible for data collection and writing the first draft of the manuscript.

Tawanda Marandure: Assisted with data collection and analysis as well as editing the manuscript.

Obert Chenjerai Chikwanha: Co-supervised the research process and was responsible for data analysis and editing the manuscript

James Bennett: Is the grant recipient and Principal Investigator of the study. He was responsible for editing the manuscript.

Heidi Jane Hawkins: Grant co-applicant and provided editorial input to the manuscript.

Antony R. Palmer: Grant co-applicant and provided editorial input to the manuscript

Lianhai Wu: provided editorial input to the manuscript.

Munyaradzi C. Marufu: Co-supervised the research process and provided editorial input to the manuscript

Cletos Mapiye: Grant co-applicant, contributed to the study conception and design

All authors commented on previous versions of the manuscript, read and approved the final manuscript.

Data Availability

The datasets generated during and/or analysed during the current study are available from the corresponding author on reasonable request.

Ethics approval

This study was performed in line with the principles of the Declaration of Helsinki. Approval was granted by the Stellenbosch University Research Ethics Committee for Animal Care and Use (ACU-2020-11758).

Consent to participate

Informed consent was obtained from all individual participants included in the study.

Consent to publish

The authors affirm that human research participants provided informed consent for publication of collected data.

Acknowledgments

The authors are thankful to Mvenyani community, particularly farmers in Magxeni village for their cooperation throughout the study.

References

1. AOAC (2002) Official Method of Analysis. 16th Edition, Association of Official Analytical, Washington DC.
2. Bobeck, E. A. 2020. Nutrition and health: Companion animal applications: Functional nutrition in livestock and companion animals to modulate the immune response. J. Anim. Sci. 98:1–8. doi:10.1093/JAS/SKAA035.

3. Brown, D., J. W. Ng'ambi, and D. Norris. 2018. Effect of tanniniferous acacia karroo leaf meal inclusion level on feed intake, digestibility and live weight gain of goats fed a setaria verticillata grass hay-based diet. *J. Appl. Anim. Res.* 46:248–253. doi:10.1080/09712119.2017.1289939.
4. Cenci F.B., Louvandini H., McManus C.M., dell Porto,A., Costa D.M., Araujo S.C., Minho A.P. and Abdalla A.L., 2007. Effects of condensed tannin from *Acacia mearnsii* on sheep infected naturally with gastrointestinal helminths. *Vet Parasitol*, 144, p. 132-137
5. Cheema, U. B., M. Younas, J. I. Sultan, M. R. Virk, M. Tariq, and A. Waheed. 2011. Fodder tree leaves: an alternative source of livestock feeding. *Adv. Agric. Biotechnol.* 2:22–33.
6. Chikwanha, C.C., Katiyatiya, C.L.F., Strydom, P.E., Mapiye, C. 2023. Substitution effects of *Acacia mearnsii* leaf-meal for *Triticum aestivum* bran on nutrient digestibility, rumen fermentation and nitrogen retention in lambs. *Small Ruminant Research*, 221, 106948. <https://doi.org/10.1016/j.smallrumres.2023.106948>.)
7. Costa, E. I. d. S., Ribeiro, C. V. D. M., Silva, T. M., Batista, A. S. M., Vieira, J. F., Barbosa, A. M., da Silva Júnior, J. M., Bezerra, L. R., Pereira, E. S., & Oliveira, R. L. (2021). Effect of dietary condensed tannins inclusion from *Acacia mearnsii* extract on the growth performance, carcass traits and meat quality of lambs. *Livestock Science*, 253. <https://doi.org/10.1016/J.LIVSCI.2021.104717>
8. Da Costa, M. M. C., L. C. V. Ítavo, C. C. B. Ferreira Ítavo, A. M. Dias, H. V. Petit, F. A. Reis, R. C. Gomes, E. S. Leal, M. V. G. Niwa, and G. J. De Moraes. 2019. Evaluation of internal and external markers to estimate faecal output and feed intake in sheep fed fresh forage. *Anim. Prod. Sci.* 59:741–748. doi:10.1071/AN16567.
9. Dezah, T. J., O. C. Chikwanha, K. Dzama, P. E. Strydom, and C. Mapiye. 2021. Comparison of *Acacia mearnsii* and *Medicago sativa* as substitutes for *Glycine max* in cattle finisher diets: Effects on beef production and quality. *Sci. African.* 11:e00673. doi:10.1016/j.sciaf.2020.e00673.
10. Dias-Silva, T. P., and A. L. A. Filho. 2021. Sheep and goat feeding behavior profile in grazing systems. *Acta Sci. - Anim. Sci.* 43:1–10. doi:10.4025/actascianimsci.v43i1.51265.
11. Gebeyew, K., K. Beriso, A. Mohamed, G. G. S. Melaku, and A. Worku. 2015. Review on the Nutritive Value of Some Selected *Acacia* Species for Livestock Production in Dryland Areas *Advances in Dairy Research*. 3. doi:10.4172/2329-888X.1000139.
12. Gouws, A. J., and C. M. Shackleton. 2019. Abundance and correlates of the *Acacia dealbata* invasion in the northern Eastern Cape, South Africa. *For. Ecol. Manage.* 432:455–466. doi:10.1016/j.foreco.2018.09.048. Available from: <https://doi.org/10.1016/j.foreco.2018.09.048>
13. Hoste, H., F. Jackson, S. Athanasiadou, S. M. Thamsborg, and S. O. Hoskin. 2006. The effects of tannin-rich plants on parasitic nematodes in ruminants. *Trends Parasitol.* 22:253–261. doi:10.1016/J.PT.2006.04.004.
14. Hyde, S. 1999. Likelihood Based Inference on the Box-Cox Family of Transformations: Sas and Matlab Programs.
15. Ingvarsen, K. L., and K. Moyes. 2013. Nutrition, immune function and health of dairy cattle. *Animal.* 7:112-122. doi:10.1017/S175173111200170X. Available from: <http://dx.doi.org/10.1017/S175173111200170X>
16. Jerónimo, E., C. Pinheiro, E. Lamy, M. T. Dentinho, E. Sales-Baptista, O. Lopes, and F. Capela e Silva. 2016. Tannins in ruminant nutrition: Impact on animal performance and quality of edible products. *Tann. Biochem. Food Sources Nutr. Prop.* 121–168.
17. Kym, A., Hynd, P., de Graaf, S., Leahy, T. and Larsen. J. 2018. “The Energy and Protein Nutrition of Grazing Sheep.” In *The Practice of Sheep Veterinary Medicine*, 104–42. University of Adelaide Press.
18. Maguraushe, W. 2015. The potential utilization of *Acacia karroo* in improving communal goat nutrition in the False Thornveld of the Eastern Cape Province , South Africa By. University of Fort Hare.
19. Mahachi, L.N., Chikwanha, O.C., Katiyatiya, C.L.F., Marufu, M.C., Aremu, A.O., Mapiye, C. 2020. *Sericea lespedeza* (*Lespedeza juncea* var. *sericea*) for sustainable small ruminant production: Feed, helminth suppressant and meat

- preservation capabilities. *Animal Feed Science and Technology*, 270. doi:10.1016/j.anifeedsci.2020.114688
20. Makkar, H. P. S. 2003. Quantification of Tannins in Tree and Shrub Foliage. *Quantif. Tann. Tree Shrub Foliage*. doi:10.1007/978-94-017-0273-7.
 21. Mapiliyao, L., D. Pepe, U. Marume, and V. Muchenje. 2012. Flock dynamics, body condition and weight variation in sheep in two ecologically different resource-poor communal farming systems. *Small Rumin. Res.* 104:45–54. doi:10.1016/J.SMALLRUMRES.2011.11.006.
 22. Mapiye, C., M. Chimonyo, K. Dzama, P. E. Strydom, V. Muchenje, and M. C. Marufu. 2009. Nutritional status, growth performance and carcass characteristics of Nguni steers supplemented with Acacia karroo leaf-meal. *Livest. Sci.* 126:206–214. doi:10.1016/j.livsci.2009.07.001. Available from: <http://dx.doi.org/10.1016/j.livsci.2009.07.001>
 23. Matshawule, S. 2020. Species Composition and Biomass Production in Two Communal Coastal Rangelands of the Eastern Cape Province, South Africa. *Advance in Biological Research*. 1: 1-10. 10.26855/abr.20200001.
 24. Max, R. & Kimambo, A & Kassuku, Ayub & Mtenga, L & Buttery, Peter. (2007). Effect of tanniniferous browse meal on nematode faecal egg counts and internal parasite burdens in sheep and goats. *South African Journal of Animal Science*. 37. 10.4314/sajas.v37i2.4033.
 25. Miller, J. 1997. The epidemiology, diagnosis and control of helminth parasites of ruminants. *Prev. Vet. Med.* 31:161–162. doi:10.1016/s0167-5877(97)83404-6.
 26. Min BR, Pinchak WE, Hume ME, Anderson RC. 2021. Effects of Condensed Tannins Supplementation on Animal Performance, Phylogenetic Microbial Changes, and In Vitro Methane Emissions in Steers Grazing Winter Wheat. *Animals (Basel)*;11(8):2391. <http://jas.fass.org/content/78/4/810>
 27. Mphinyane, W. N., G. Tacheba, and M. J. 2015. Seasonal diet preference of cattle, sheep and goats grazing on the communal grazing rangeland in the Central District of Botswana. *African J. Agric. Res.* 10:2791–2803. doi:10.5897/ajar2014.9157.
 28. Moyo, B., P. J. Masika, and V. Muchenje. 2012. Effect of supplementing crossbred Xhosa lop-eared goat castrates with Moringa oleifera leaves on growth performance, carcass and non-carcass characteristics. *Trop. Anim. Health Prod.* 44:801–809. s University doi:10.1007/s11250-011-9970-6.
 29. Mthi, S., and N. Nyangiwe. 2018. Farmers ' perception on sheep production constraints in the communal grazing areas of the Eastern Cape Province , South Africa. 9:334–339. doi:10.5897/IJLP2018.0500.
 30. Mucina, L., & Rutherford, M. C. 2006. The vegetation of South Africa, Lesotho and Swaziland. South African National Biodiversity Institute.
 31. National Research Council, NRC (2007) Nutrient Requirements of Small Ruminants: Sheep, Goats, Cervids, and New World Camelids. The National Academies Press, Washington DC.
 32. Ngorima A. 2014. Perceptions and livelihood uses of an invasive alien tree; MSc Thesis; Rhodes University.
 33. Nicol, A.M. and Brookes, I.M. 2007. Pasture and Supplements for Grazing Animals, edited by PV Rattray, IM Brooks & AM Nicol, New Zealand Society of Animal Production. ISBN No. 0908768257
 34. Obeidat, B. S., H. S. Subih, and M. Ata. 2020. Protein supplementation improves performance of lambs fed low-quality forage. *Animals*. 10:1–7. doi:10.3390/ani10010051.
 35. Porter, L. J., L. N. Hrstich, and B. G. Chan. 1985. The conversion of procyanidins and prodelphinidins to cyanidin and delphinidin. *Phytochemistry*. 25:223–230. doi:10.1016/S0031-9422(00)94533-3.
 36. Priolo, A., Waghorn, G. C., Lanza, M., Biondi, L. and Pennisi, P. 2000. Polyethylene glycol as a means for reducing the impact of condensed tannins in carob pulp: effects on lamb growth performance and meat quality. *Journal of Animal Science*: 78:810-816.
 37. Roberts, D. 2022. Supplementary feeding and feed budgeting for sheep. Agriculture and Food Division, Department of Primary Industries and Regional Development, Government of Western Australia. <https://agric.wa.gov.au/n/1927>

38. Roberts, D. 2020. Grazing stubbles and dry pasture. Agriculture and Food Division, Department of Primary Industries and Regional Development, Government of Western Australia. <https://agric.wa.gov.au/n/1744>
39. Rubanza, C. D. K., M. N. Shem, S. S. Bakengesa, T. Ichinohe, and T. Fujihara. 2007. Effects of *Acacia nilotica*, *A. polyacantha* and *Leucaena leucocephala* leaf meal supplementation on performance of Small East African goats fed native pasture hay basal forages. *Small Rumin. Res.* 70:165–173. doi:10.1016/j.smallrumres.2006.02.008.
40. Salem, H.B., & Smith, T.N. (2008). Feeding strategies to increase small ruminant production in dry environments. *Small Ruminant Research*, 77, 174-194.
41. Tadele Yilkal. 2015. Important Anti-Nutritional Substances and Inherent Toxicants of Feeds. *Food Sci. Qual. Manag.* 36:40–48.
42. Uushona, T., Chikwanha, OC., Katiyatiya, C., Strydom, PE., Mapiye, C (in press). Production and meat quality attributes of lambs fed varying levels of *Acacia mearnsii* leaf-meal as replacement for *Triticum aestivum* bran, *Meat Science*, 109042, <https://doi.org/10.1016/j.meatsci.2022.109042>.
43. Uushona, T., Chikwanha, O.C., Katiyatiya, C.L.F., Strydom, P.E., Mapiye, C., 2023. Production and meat quality attributes of lambs fed varying levels of *Acacia mearnsii* leaf-meal as replacement for *Triticum aestivum* bran. *Meat Sci.* 196, 109042. <https://doi.org/10.1016/j.meatsci.2022.109042>
44. Uushona, T., Chikwanha, O., Tayengwa, T., Katiyatiya, C., Strydom, P., & Mapiye, C. (2021). Nutraceutical and preservative potential of *Acacia mearnsii* and *Acacia dealbata* leaves for ruminant production and product quality enhancement. *The Journal of Agricultural Science*, 159(9-10), 743-756. doi:10.1017/S0021859621001015
45. Van Wyk, J. A. 2008. Production trials involving use of the FAMACHA© system for haemonchosis in sheep: preliminary results. *Onderstepoort J. Vet. Res.* 75:331–345. doi:10.4102/ojvr.v75i4.109.
46. Vipond, J., Morgan, C., and McEvoy, T. 2009. Feeding the ewe for lifetime production, taking theory into practice. *Meat Promotion Wales*. https://meatpromotion.wales/images/resources/Feeding_the_ewe_final.pdf
47. Waghorn, G. 2008. Beneficial and detrimental effects of dietary condensed tannins for sustainable sheep and goat production-Progress and challenges. *Anim. Feed Sci. Technol.* 147:116–139. doi:10.1016/j.anifeedsci.2007.09.013.
48. Yapi, T. S., P. J. O'Farrell, L. E. Dziba, and K. J. Esler. 2018. Alien tree invasion into a South African montane grassland ecosystem: Impact of acacia species on rangeland condition and livestock carrying capacity. *Int. J. Biodivers. Sci. Ecosyst. Serv. Manag.* 14:105–116. doi:10.1080/21513732.2018.1450291. Available from: <https://doi.org/10.1080/21513732.2018.1450291>

Figures

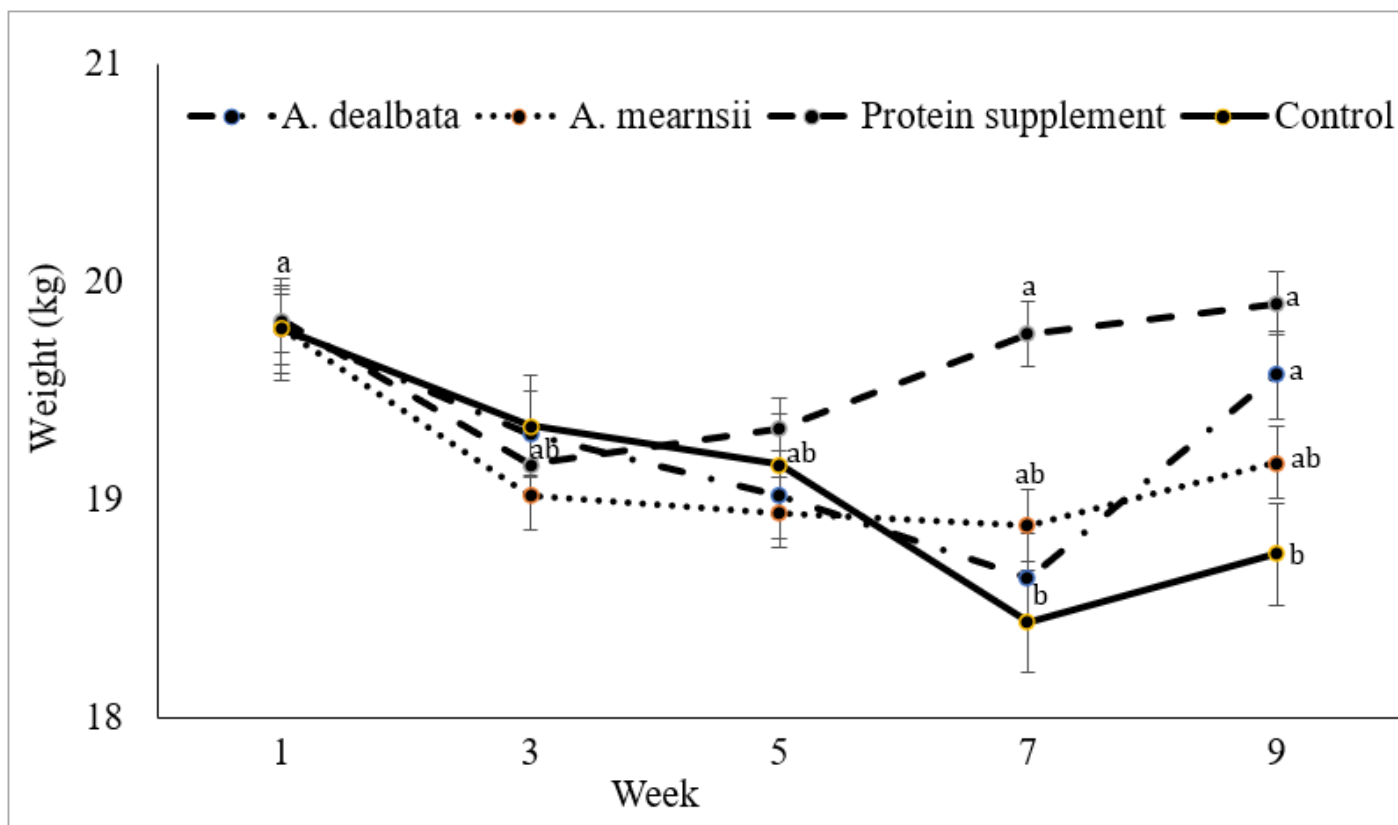


Figure 1

Least square means ($\pm$ standard error of mean) of body weights of lambs grazing South African communal rangelands supplemented with Acacia leaf-meals

^{a-b} Values with different superscripts are significantly different ($P \leq 0.05$).

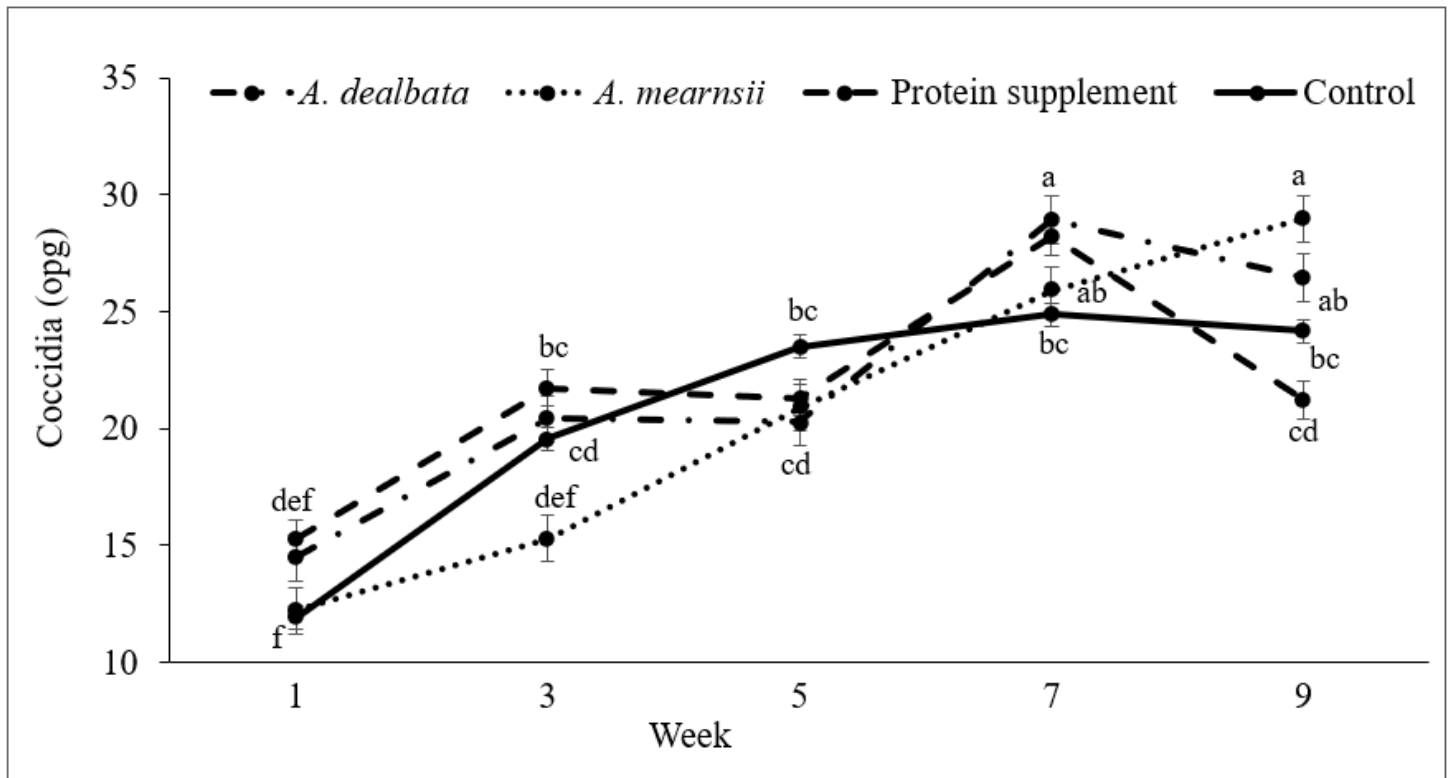


Figure 2

Least square means (± standard error of mean) of coccidia egg counts in lambs grazing South African communal rangelands supplemented with Acacia leaf-meals.

^{a-f} Values with different superscripts are significantly different (P ≤ 0.05)

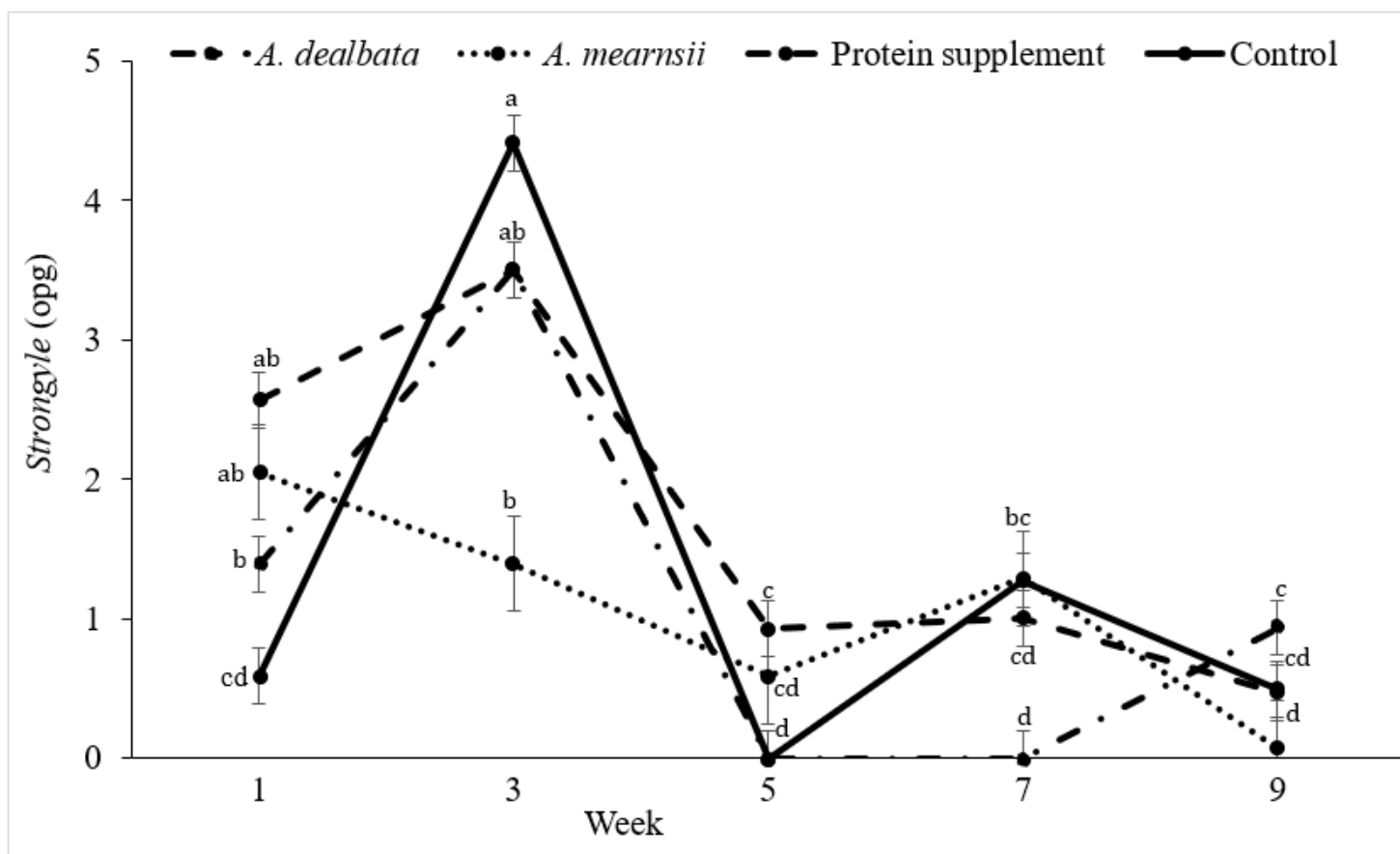


Figure 3

Least square means (± standard error of mean) of strongyles in lambs grazing South African communal rangelands supplemented with Acacia leaf-meals.

^{a-e} Values with different superscripts are significantly different ($P \leq 0.05$).