



Digestate Improves Stinging Nettle (*Urtica dioica*) Growth and Fiber Production at a Chlor-Alkali Site

Chloé Viotti, Coralie Bertheau, Françoise Martz, Loïc Yung, Vincent Placet, Andrea Ferrarini, Flavio Fornassier, Damien Blaudez, Markus Puschenreiter, Michel Chalot

► To cite this version:

Chloé Viotti, Coralie Bertheau, Françoise Martz, Loïc Yung, Vincent Placet, et al.. Digestate Improves Stinging Nettle (*Urtica dioica*) Growth and Fiber Production at a Chlor-Alkali Site. *Plants*, 2024, 13 (17), pp.2425. <10.3390/plants13172425>. <hal-04707519>

HAL Id: hal-04707519

<https://hal.science/hal-04707519v1>

Submitted on 24 Sep 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons CC BY 4.0 - Attribution - International License

Article

Digestate Improves Stinging Nettle (*Urtica dioica*) Growth and Fiber Production at a Chlor-Alkali Site

Chloé Viotti ¹, Coralie Bertheau ¹, Françoise Martz ² , Loïc Yung ³ , Vincent Placet ⁴ , Andrea Ferrarini ⁵ , Flavio Fornassier ⁶, Damien Blaudez ³ , Markus Puschenreiter ⁷ and Michel Chalot ^{1,8,9,*} 

¹ Université de Franche-Comté, CNRS, Chrono-Environnement, F-25200 Montbéliard, France; chloe.viotti@univ-fcomte.fr (C.V.); coralie.bertheau-rossel@univ-fcomte.fr (C.B.)

² Production System Unit, Natural Resources Institute Finland (Luke), Ounasjoentie 6, 96200 Rovaniemi, Finland; francoise.martz@luke.fi

³ Université de Lorraine, CNRS, LIEC, F-54000 Nancy, France; loic.yung@uha.fr (L.Y.); damien.blaudez@univ-lorraine.fr (D.B.)

⁴ Université de Franche-Comté, FEMTO-ST Institute, Department of Applied Mechanics, F-25000 Besançon, France; vincent.placet@univ-fcomte.fr

⁵ Department of Sustainable Crop Production, Università Cattolica del Sacro Cuore, Via Emilia Parmense 84, 29122 Piacenza, Italy; andrea.ferrarini@unicatt.it

⁶ CREA—Centro Viticoltura ed Enologia, Via Trieste 23, 34170 Gorizia, Italy; flavio.fornassier@crea.gov.it

⁷ Institute of Soil Research, University of Natural Resources and Life Sciences, 1180 Vienna, Austria; markus.puschenreiter@boku.ac.at

⁸ Université de Lorraine, Faculté des Sciences et Technologies, F-54000 Nancy, France

⁹ UMR 6249 Laboratoire Chrono-Environnement, Pôle Universitaire du Pays de Montbéliard, 4 Place Tharradin, F-25200 Montbéliard, France

* Correspondence: michel.chalot@univ-fcomte.fr; Tel.: +33-3-81-99-46-76

Abstract: Marginal lands have been proposed to produce non-food crop biomass for energy or green materials. For this purpose, the selection, implementation, and growth optimization of plant species on such lands are key elements to investigate to achieve relevant plant yields. Stinging nettle (*Urtica dioica*) is a herbaceous perennial that grows spontaneously on contaminated lands and was described as suitable to produce fibers for material applications. Two mercury-contaminated soils from industrial wastelands with different properties (grassland soil and sediment landfill) were used in this study to assess the potential growth of stinging nettle in a greenhouse mesocosm experiment. Two organic amendments were studied for their impact on nettle growth. The solid digestate from organic food wastes significantly doubled plant biomass whereas the compost from green wastes had a lower impact. The highest doses of organic amendments significantly increased the number of fibers, which doubled following digestate application, while reducing leaf Hg concentration. Both amendments significantly improved soil respiration and enzymatic activities linked to the microbial biomass in the soil from the sediment landfill by the end of the experiment. In the context of a phytomanagement scenario, solid digestate would be a preferred amendment resource to improve nettle production on industrial wastelands.

Keywords: compost; digestate; enzymatic activities; fiber; marginal lands; N fertilization



Citation: Viotti, C.; Bertheau, C.; Martz, F.; Yung, L.; Placet, V.; Ferrarini, A.; Fornassier, F.; Blaudez, D.; Puschenreiter, M.; Chalot, M. Digestate Improves Stinging Nettle (*Urtica dioica*) Growth and Fiber Production at a Chlor-Alkali Site. *Plants* **2024**, *13*, 2425. <https://doi.org/10.3390/plants13172425>

Academic Editor: Mingsheng Fan

Received: 2 July 2024

Revised: 19 August 2024

Accepted: 23 August 2024

Published: 30 August 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Due to global population growth, agricultural food production has become more important over the years, putting great pressure on the environment [1]. Other needs linked to the bio-based economy (e.g., bioenergy, biomass, fiber) have emerged and have generated new competition for land use between food and non-food crops since the early 2000s [2,3]. Due to low fertility and high environmental stress, marginal lands are unsuitable for food cultivation but have been proposed as relevant land to produce biomass for energy or green material purposes [4,5]. In Europe, marginal lands cover an estimated area of

39 to 111 million hectares [6,7], and their use for cultivation can meet the needs of a circular economy action plan and bioeconomy strategy from the European Green Deal [8,9].

Among marginal lands, trace element (TE)-contaminated soils have been described as a relevant alternative to produce biomass for material applications [10,11]. In addition to the potential economic benefit of biomass production on these lands [12,13], vegetation cover also provides environmental benefits. For example, it can help to prevent the dispersion of contaminated soil particles, reduce the mobility of metals in the rhizosphere [14], and improve soil properties (e.g., organic matter and nutrient content, biological activity) [15]. Although some TE-tolerant plant species growing or implemented in contaminated or polluted soils are known to accumulate TE [16,17], others such as industrial hemp (*Cannabis sativa* L.) or ramie (*Boehmeria Nivea*) exhibit a low bio-concentration factor in the aerial parts. Combined with a high yield crop, the biomass produced can be used for industrial fiber applications [18,19]. However, biomass production should be sufficient and slightly impacted by soil contamination, with yields almost similar to those on uncontaminated soils [20]. Thus, plant tolerance to abiotic stress (e.g., nutrient deficiency) is the most important consideration for establishing a biomass crop on these lands [21,22]. Under such conditions, modifications of the biosynthesis of compounds through specialized metabolism can be observed in plant parts [23,24]. Among these metabolites, phenolic compounds are often produced as a result of the stress response [25]. These metabolites are abundant in plants and involved in different physiological processes such as growth, stress defense, and antioxidant protection [26,27]. Thus, phenolics are considered to be involved in the adaptation of the plant to its environment [28].

Stinging nettle (*Urtica dioica*) is a herbaceous perennial plant that grows spontaneously on contaminated lands, notably in mercury (Hg)-contaminated soils, and that produces fibers that can be used for material applications [29]. Hg contamination has increased over the last few decades and is of concern because of its toxicity to humans and ecosystems and its potential accumulation by plants [30]. Nettle grown under pot-based laboratory-scale experiments exhibited excess levels of metals, above the toxic levels reported by Kabata-Pendias [31], while plant material collected from in situ contaminated sites had TE levels below toxic levels [32]. Although *U. dioica* is native and able to grow in TE-contaminated soils, its cultivation in the context of biomass and fiber production has not yet been investigated. *Urtica dioica* is usually described as a nitrophilous species [33] for which cultivation requires high doses of nitrogen (N) fertilization [32,34]. As a perennial plant, annual re-establishment is not necessary. A duration of four to five years has been described for a stinging nettle crop [35], to a maximum of 10 years with high weed control [32,36]. Although *U. dioica* competes poorly with weeds [37], Müllerová et al. [38] improved stinging nettle expansion and regeneration in cut grasslands using N fertilization (along with phosphorus (P) and potassium (K)). The use of amendments constitutes a factor that assists plant adaptation by improving nutrient availability, soil physicochemical properties, and microbial activity [39–41].

Due to the environmental impacts of inorganic fertilizers [42,43], the use of organic amendments is now encouraged as an alternative to improve both soil fertility and carbon (C) storage [44]. Organic amendments can be produced from agricultural residues or food wastes, and must be rigorously selected as they can have different effects [45,46]. Among these amendments, compost, which results from the stabilization and sanitation of organic waste by aerobic decomposition, is a well-known product [47]. Digestate is another organic amendment, either as solid or liquid material, which is produced under a controlled anaerobic fermentation process from biodegradable materials [48]. The production of such fertilizers from bio-wastes can therefore be part of the bio-based circular economy concept by reincorporating waste materials into the production cycle [49,50], as well as potential substitutes for mineral N fertilizers [51,52].

N cycling in the soil is crucial for plant development and crop yield [53], and N is released from compost and solid digestate slowly over time through mineralization [54,55]. N availability is primarily driven by microbial reactions [56]. Soil microorganisms pro-

duce extracellular hydrolytic and oxidative enzymes involved in C, N, and P cycling and therefore participate in the breakdown of organic matter [57,58]. These enzymes can be stimulated or inhibited by N application [59,60] impacting soil nutrient availability [61]. Moreover, compost and digestate, through adsorption mechanisms and enhancement in microbial activity, may also be involved in soil contaminant immobilization and thus plant uptake limitation [62,63]. Their use therefore appears to be relevant in the context of biomass production from contaminated lands.

The present study aimed to investigate the fitness of *U. dioica* growth on Hg-contaminated soils and to determine the most effective organic amendment (compost: C or digestate: D) and dose (+; ++; +++) for biomass production improvement. We conducted the experiment on two industrial soils with different properties, but both were contaminated with Hg. Stinging nettle plants were grown in pots and amended with compost or solid digestate at different doses. Plant parameters (i.e., biomass, height, stem diameter, chlorophyll content, photosystem II (PSII) activity, phenolic compounds, N and Hg concentrations, fiber characteristics) were measured to evaluate the growth of *U. dioica* on these contaminated soils. Soil parameters (i.e., soil respiration, double-stranded DNA (dsDNA) concentration and enzymatic activities) were assessed to identify the effect of organic amendments on soil activities and microbial biomass that can impact plant growth. We therefore hypothesized that the use of organic amendments would (i) improve nettle fitness in Hg-contaminated soils and (ii) sustain biomass production by improving soil microbial biomass and activities.

2. Results

2.1. Main Impacting Factors and Effect of the Soil on Nettle Growth

All replicates survived during the experiment, with PSII activity that ranged from 0.75 to 0.78 in both soils (Table S1). The application of amendments significantly decreased the chlorophyll content, except for D++, which resulted in the opposite effect (Table S1). Leaves ($<153 \mu\text{g}\cdot\text{kg}^{-1}$ DW) and stems ($\leq 72 \mu\text{g}\cdot\text{kg}^{-1}$ DW) exhibited low Hg concentrations regardless of the soil considered (Table 1). However, D++ significantly divided the Hg concentration in the leaves of nettles from the Tavazzano soil by 2.3 and the Hg concentration in the leaves and stems of nettles from the St-Symphorien-sur-Saône soil by 1.5.

Table 1. Mean Hg concentration ($n = 5$, expressed as $\mu\text{g}\cdot\text{kg}^{-1}$ DW $\pm$ SE) in the leaves and stems of *Urtica dioica* after 83 days of cultivation on contaminated soils (St-Symphorien-sur-Saône and Tavazzano) depending on the nature of amendment (C: compost; D: digestate) and rate applied (+; ++; +++)). Different letters indicate significant differences between treatments and soils (Kruskal–Wallis test, $p < 0.05$).

Soil	Treatment	Leaf Hg Concentration ($\mu\text{g}\cdot\text{kg}^{-1}$ DW)	Steam Hg Concentration ($\mu\text{g}\cdot\text{kg}^{-1}$ DW)
St-Symphorien-sur-Saône	Control	110.8 $\pm$ 15.3 ab	58.8 $\pm$ 4.0 efg
	C++	130.2 $\pm$ 3.5 a	50.0 $\pm$ 6.6 ghi
	C+++	134.5 $\pm$ 14.9 a	72.0 $\pm$ 10.1 cdef
	D+	101.1 $\pm$ 2.7 abc	59.4 $\pm$ 8.1 efg
	D++	74.1 $\pm$ 14.0 defg	36.0 $\pm$ 7.9 hi
Tavazzano	Control	109.3 $\pm$ 23.3 abcd	39.9 $\pm$ 5.5 hi
	C++	73.5 $\pm$ 5.5 bcde	50.2 $\pm$ 7.7 ghi
	C+++	152.5 $\pm$ 37.2 a	50.9 $\pm$ 7.2 fgh
	D+	64.5 $\pm$ 11.3 efg	34.5 $\pm$ 4.4 hi
	D++	47.6 $\pm$ 3.9 ghi	33.2 $\pm$ 2.2 i

When all the measured variables were considered, the applied dose of amendment was the main impacting factor ($R^2 = 0.53$, $p = 0.001$), followed by the treatment (i.e., nature of amendment $\times$ dose $\times$ soil; $R^2 = 0.10$, $p < 0.05$). The two first principal components explained 59.6% and 51.2% of the total variance in the whole dataset for St-Symphorien-sur-

Saône and Tavazzano soils, respectively, with digestate samples that seemed to separate from the control rather than the compost samples (Figure 1a,c). Plant parameters (i.e., aboveground biomass, plant height and leaf area) contributed the most to the variability of the data, along with enzymatic activities involved in N and P cycles (i.e., chit, leu, acP, bisP) for both soils (Figure 1b,d). Except for xilo and uroni, the enzymatic activities involved in the C cycle contributed the least, along with soil respiration and dsDNA for the Tavazzano soil. Finally, the soil had a low significant impact on the variability of the dataset ($R^2 = 0.05$, $p < 0.01$). However, control plants grown on the Tavazzano soil produced significantly twice more aboveground biomass than nettles grown on the St-Symphorien-sur-Saône soil (Figure 2a,d). Nettles resulted in significantly higher plants (Figure 2c,f) with a higher stem diameter (Figure 3a,b).

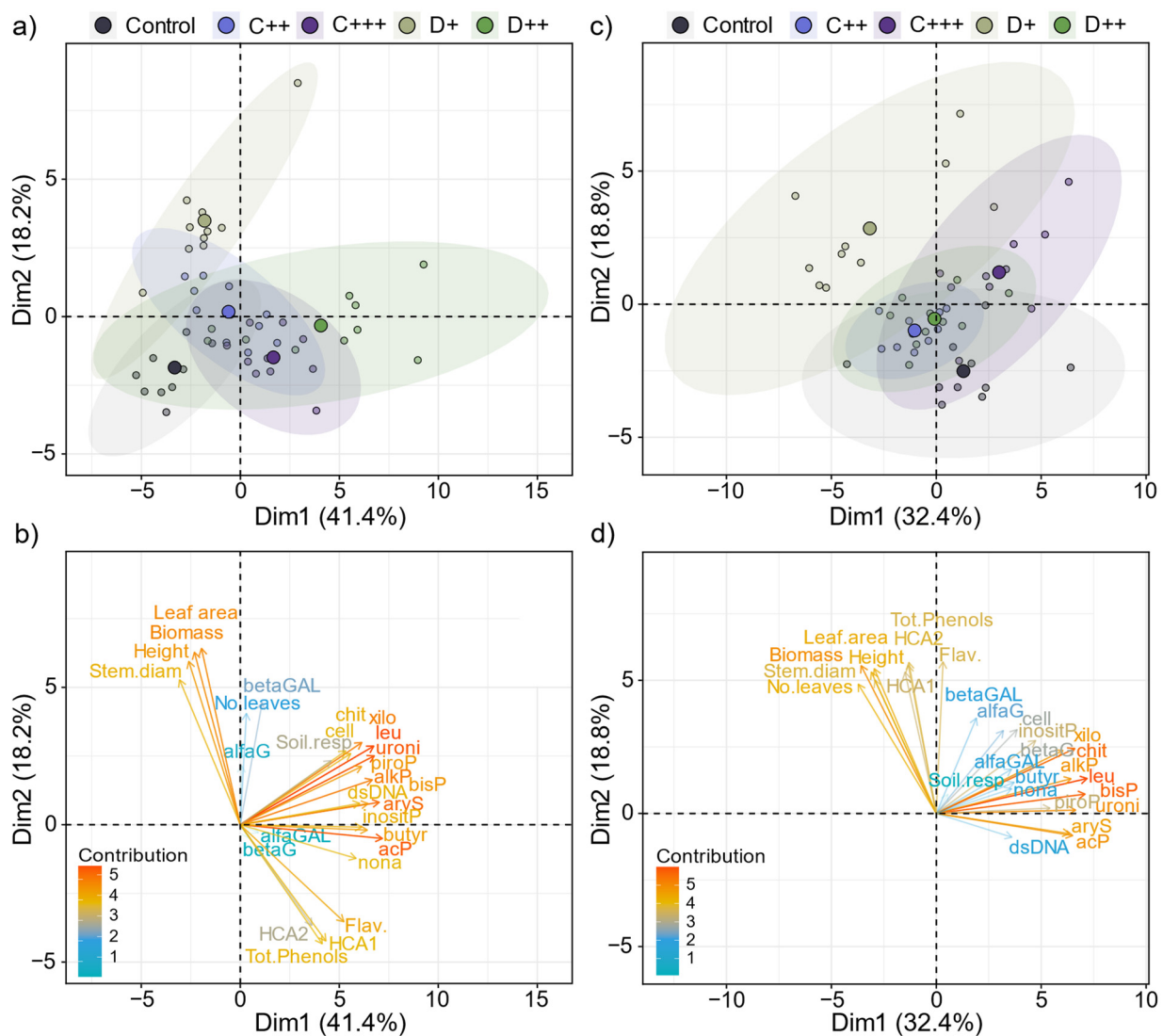


Figure 1. Principal component analysis (PCA) plots showing the ordination of the samples depending on the nature of amendment (C: compost; D: digestate) and rate (+, ++, +++) applied using the whole dataset for (a,b) St-Symphorien-sur-Saône and (c,d) Tavazzano soils. The ellipses represent a 95% confidence interval. Vectors are colored depending on their contribution to the overall distribution and indicate the direction and strength of each environmental variable.

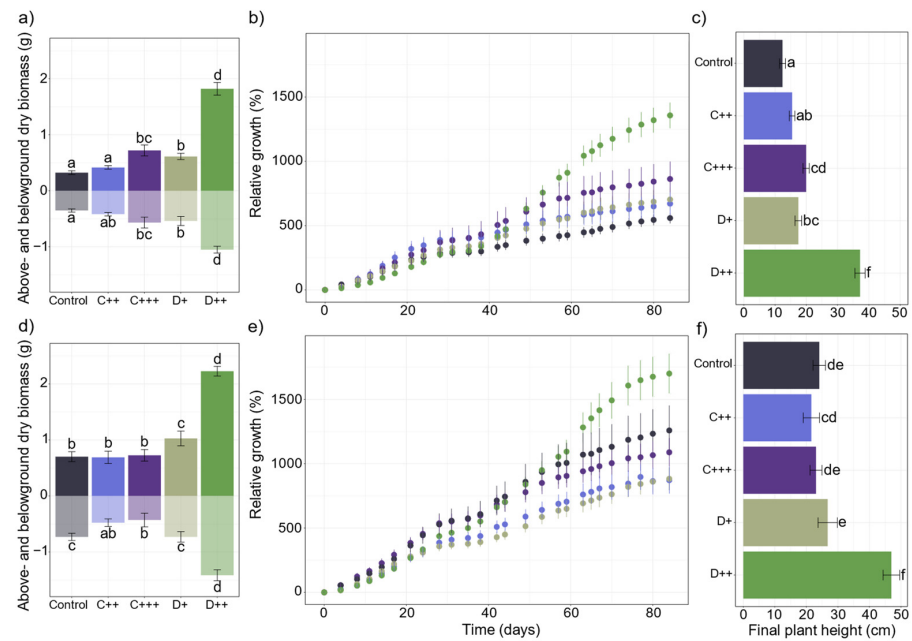


Figure 2. Impact of organic amendments on plant growth parameters ($n = 10$). (a,d) Aboveground and belowground dry biomass ($g \pm SE$); (b,e) relative growth ($\% \pm SE$) of nettle plants during cultivation (days) and (c,f) final plant height ($cm \pm SE$) of *Urtica dioica* after 83 days of cultivation depending on the nature of the amendment (C: compost; D: digestate) and the rate (+; ++; +++) applied to (a–c) St-Symphorien-sur-Saône and (d–f) Tavazzano soils. Different letters indicate significant differences between treatments and soils for each variable (Kruskal–Wallis, $p < 0.05$).

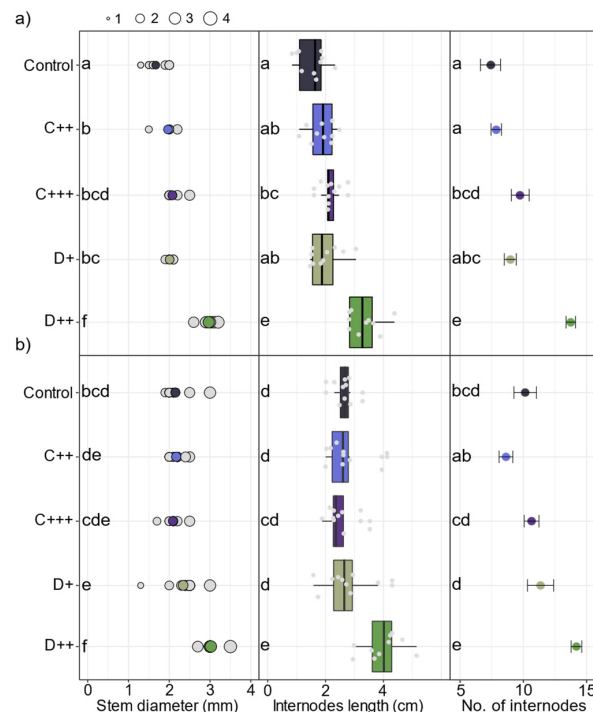


Figure 3. Impact of amendments on nettle stem parameters. Mean stem diameter (mm), internode length (cm), and number of internodes ($n = 10 \pm SE$) per stem after 83 days of cultivation depending on the nature of amendment (C: compost; D: digestate) and rate (+; ++; +++) applied to (a) St-Symphorien-sur-Saône and (b) Tavazzano soils. Different letters indicate significant differences between treatments and soils for each variable (Kruskal–Wallis test, $p < 0.05$).

2.2. Impact of the Nature and Dose of Amendments on Plant Growth

The D+, D++, and C+++ treatments significantly increased the production of below-ground biomass of nettles in the St-Symphorien-sur-Saône soil (Figure 2a). The nature of the amendment and the dose applied mainly affected the final aerial biomass ($F = 147.2$; $p < 0.001$ and $F = 86.3$; $p < 0.001$, respectively) rather than the soil ($F = 24.7$; $p < 0.001$). At harvest, only the plants that grew under the highest dose of digestate reached the flowering stage. In the St-Symphorien-sur-Saône soil, C+++ and D+ significantly doubled the produced aboveground biomass, while D++ more than quintupled it (Figure 2a). During the first four days of cultivation, C+++ and D++ significantly reduced the relative growth compared to control plants, while the positive impact of these treatments started to be significant after 49 days of cultivation in this soil (Figure 2b). From 63 days, D++ significantly improved the relative growth compared to all the other treatments. At harvest, the plant height followed the same increase as above- and belowground biomass, with nettles significantly taller with digestate and C+++ compared to control plants (Figure 2c).

All amendments and doses applied to the St-Symphorien-sur-Saône soil significantly improved the stem diameter, and C+++ and D++ resulted in significantly more and longer internodes (Figure 3a). Concomitantly, the number of fibers increased significantly by 1.3 and 2.4 times under the C+++ and D++ treatments, respectively (Figure 4). Conversely, the fertilization did not significantly impact the fiber diameters, which ranged on average from 37 to 49 μm , or the wall thicknesses (Table S2). Among the plant parameters measured, plant height largely explained the aboveground biomass ($F = 835.1$; $p < 0.001$), followed by the leaf area ($F = 26.7$; $p < 0.001$), the number of leaves ($F = 24.2$; $p < 0.001$), and stem diameter ($F = 9.6$; $p < 0.01$).

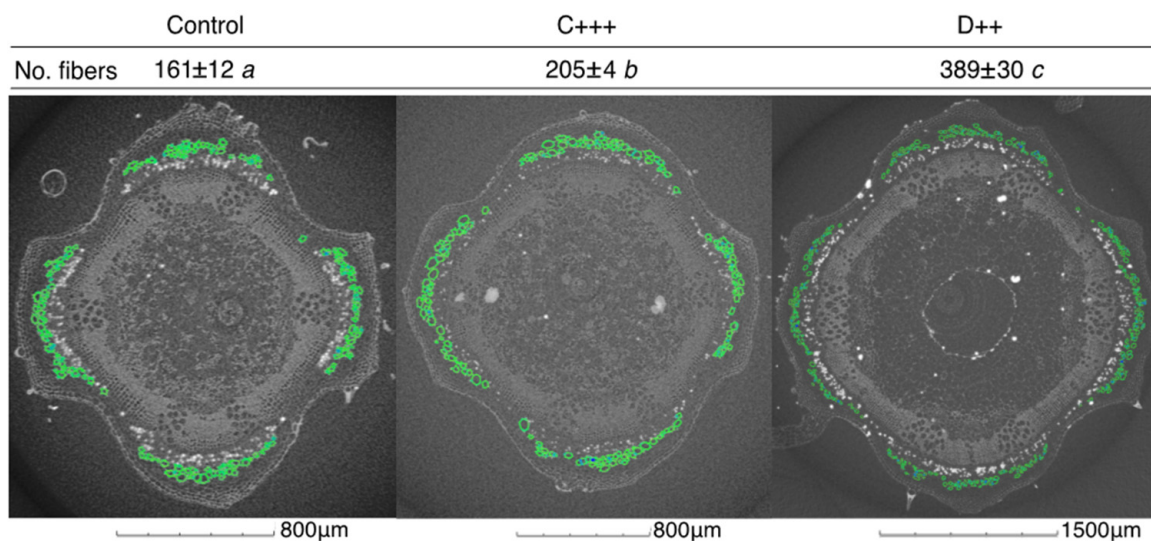


Figure 4. Impact of amendments on the quantity of fibers in nettle stems. Mean number of fibers from *Urtica dioica* stems grown on St-Symphorien-sur-Saône soil ($n = 3 \pm \text{SE}$) depending on the amendment applied (C: compost; D: digestate). Pictures represent the associated micro-computed transverse cross-sections of the stems obtained with VG StudioMax software (version 2023.1). Fibers were identified in green with MATLAB software (version R2023b). Different letters indicate significant differences between treatments (Kruskal–Wallis test, $p < 0.05$).

In the Tavazzano soil, D++ significantly almost doubled the belowground biomass, while the compost application significantly decreased it (Figure 2d). However, only digestate application significantly improved the aerial biomass of nettle (Figure 2d). When applied to the Tavazzano soil, D++ significantly slowed the relative growth during the first eleven days of cultivation before significantly increasing it after 63 days compared to all the other treatments (Figure 2e). At harvest, only D++ significantly doubled the plant height without the impact of the other treatments (Figure 2f). In this soil, only the digestate application had an impact on the stem diameter, and only D++ significantly improved the number and length of internodes (Figure 3b). Among the measured plant parameters, plant height mainly explained the aboveground biomass ($F = 782.1$; $p < 0.001$), followed by the number of leaves ($F = 65.9$; $p < 0.01$), which was the second explanatory factor of aerial biomass before leaf area and stem diameter ($F = 23.3$; $p < 0.01$ and $F = 24.8$; $p < 0.01$, respectively).

2.3. Responses of Soil Activities to Compost and Digestate Application

In the two control rhizospheric soils, alkP, butyr, nona, bisP, leu, and acP enzymatic activities had the highest values (Figure 5a,c; Table S3). Only the compost application slightly but significantly increased the soil pH of the Tavazzano soil at the end of the experiment compared to the control soil (pH = 7.5 and 7.3 in the soil amended with compost; and pH = 6.9 in the control soil), while other treatments did not impact the pH of the soil (Table S4). However, amendment application significantly impacted the enzymatic activities, with the highest impact of the dose compared to the low impact of the treatment (nature of amendment $\times$ dose $\times$ soil) and the soil ($R^2 = 0.39$, $p = 0.001$; $R^2 = 0.07$, $p < 0.01$; $R^2 = 0.04$, $p < 0.05$, respectively). Despite not being the most impacting factor, the Tavazzano control soil exhibited significantly higher enzymatic activities than the St-Symphorien-sur-Saône soil. In both soils, all the amendments, except D+ in the Tavazzano soil, significantly increased butyr (four- and five-fold with C+++ and D++ in the St-Symphorien-sur-Saône soil) and nona (seven-fold increase with the two higher doses of amendments in the St-Symphorien-sur-Saône soil) esterase activities (Figure 5a,c). All the amendments significantly increased the activities of xilo and uroni involved in the C cycle in the St-Symphorien-sur-Saône soil, but the activities remained low (<1 nmol 4-MUF·g⁻¹ soil per hour) (Figure 5a). The digestate significantly increased the enzymatic activities involved in the N cycle, which were, respectively, twice (leu in both concentrations and chit with D+) and three times (chit with D++) higher than in the control (Figure 5a). C+++ and D++ resulted in significantly higher piroP and alkP enzyme activities, and the same was observed for acP, bisP, and aryS activities also with C++. However, amendment application had a lower impact on the enzymatic activities in the Tavazzano soil. C+++ significantly increased the activities of alfaG, xilo, alkP, and bisP (Figure 5b, Table S3). In contrast, the activities of leu, acP, bisP with D+, aryS with C++ and D++, and uroni with digestate and C+ were slightly but significantly decreased compared to the activities in the control soil (Figure 5b).

At the end of the cultivation period, all the amendments except C++ significantly improved the respiration of the St-Symphorien-sur-Saône soil, which doubled with digestate application (Figure 6a). Both amendments significantly increased dsDNA (Figure S1), which was correlated with soil respiration and enzymatic activities involved in C and N cycles (i.e., xilo, uroni, chit, and leu), alkP, and butyr activities (Figure 5b). In contrast, only the digestate application significantly impacted the respiration of the Tavazzano soil, with a two-fold increase with D++ treatment (Figure 6b). No correlation between soil respiration, dsDNA, and enzymatic activity was observed (Figure 5d).

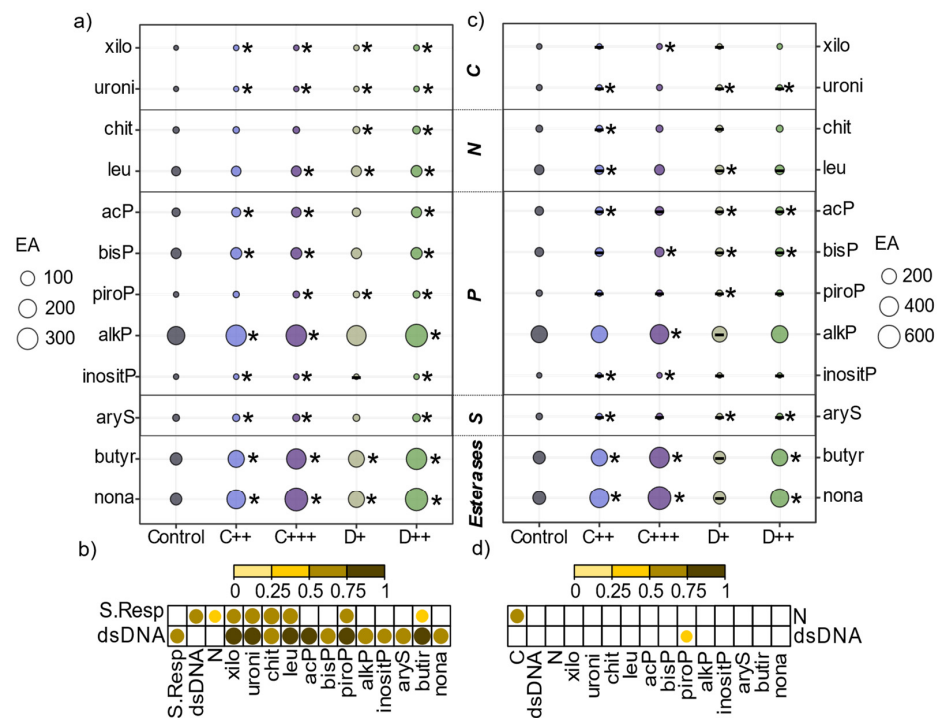


Figure 5. Impact of amendments on rhizospheric soil enzymatic activities. (a,c) Mean ($n = 10$) enzymatic activities (EA) expressed in nanomoles of $4\text{-MUF}\cdot\text{g}^{-1}$ or $\text{AMC}\cdot\text{g}^{-1}$ soil per hour involved in carbon (C), nitrogen (N), phosphate (P), and sulfur (S) cycles and esterases depending on the nature of amendment (C: compost; D: Digestate) and rate (+; ++; +++ applied). * indicates a mean significantly different from that of the control treatment for each soil (Kruskal–Wallis or Tukey's test, $p < 0.05$); - indicates a lower mean than that of the control. (b,d) Spearman's correlation matrix between soil respiration (S.Resp), dsDNA, nitrogen (N) concentration, and enzymatic activities in (a,b) St-Symphorien-sur-Saône and (c,d) Tavazzano rhizospheric soils. Only the significant correlations are represented.

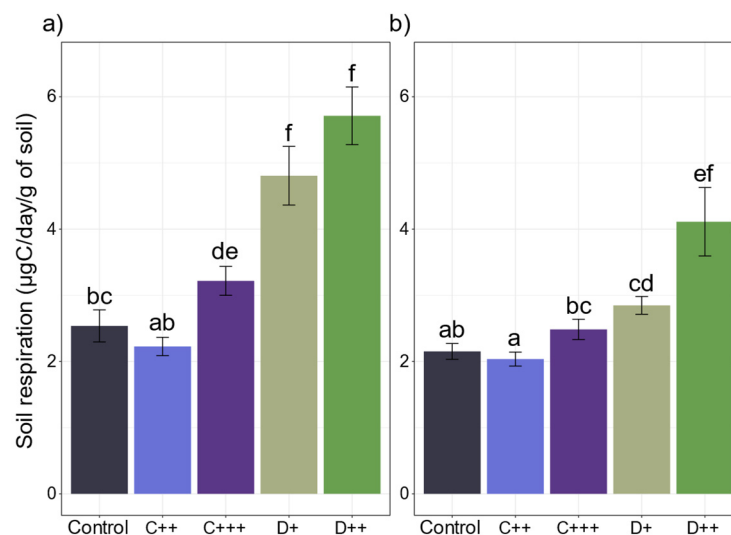


Figure 6. Impact of amendments on soil respiration. Soil respiration ($\mu\text{gC/day/g soil}$; $n = 5 \pm \text{SE}$) after 83 days of cultivation on the (a) St-Symphorien-sur-Saône and (b) Tavazzano soils depending on the nature of amendment (C: compost; D: digestate) and rate (+; ++; +++ applied). Different letters indicate significant differences between treatments for each soil (Kruskal–Wallis test, $p < 0.05$).

2.4. Impact of Organic Amendments on Leaf Phenolic Compounds

The percentage of N in nettle leaves was not significantly impacted using compost and digestate or by soil (i.e., ~2% on average) (Figure S2). However, the soil ($R^2 = 0.33$, $p = 0.001$), the treatment (nature of amendment $\times$ dose $\times$ soil; $R^2 = 0.28$, $p = 0.001$), and, to a lesser extent, the dose of applied amendment ($R^2 = 0.14$, $p < 0.01$) significantly impacted the leaf phenolic compound concentration.

Hydroxycinnamic acids (HCAs) and flavonoids constituted the main soluble phenolics detected in the nettle leaves. Caffeoyl esters (i.e., HCA1) and more specifically chlorogenic acid (i.e., CGA; $33.8 \pm 2.2\%$ and $27.4 \pm 2.0\%$ of total phenolic compounds for the St-Symphorien-sur-Saône and Tavazzano soil, respectively) and caffeoyl malic (i.e., CMA; ~37% of total phenolic compounds for both soils) mainly represented the phenolic compounds (Figure 7a,b). Flavonoids were the second most abundant phenolic compounds ($12.90 \pm 0.81\%$ and $16.43 \pm 1.12\%$ on average for the St-Symphorien-sur-Saône and Tavazzano soils, respectively) followed by p-coumaric esters (i.e., HCA2; $5.8 \pm 0.6\%$ and $7.4 \pm 1.5\%$ on average for the St-Symphorien-sur-Saône and Tavazzano soils, respectively). Overall, amendment application (except with C+++) significantly halved and divided by four the total phenolic compounds and HCA1 concentration in the leaves of nettles from the St-Symphorien-sur-Saône soil (Figure 7a). C++ and D++ significantly decreased the HCA2 concentration, while only C++ significantly reduced the flavonoid concentration by three times. In contrast, applying amendments to the Tavazzano soil significantly increased the leaf total phenolic compounds and HCA1 concentrations by three- to four-fold (Figure 7b). Only digestate significantly increased and almost tripled the HCA2 concentration (Figure 7b), while all amendment treatments at least significantly doubled the flavonoid concentration.

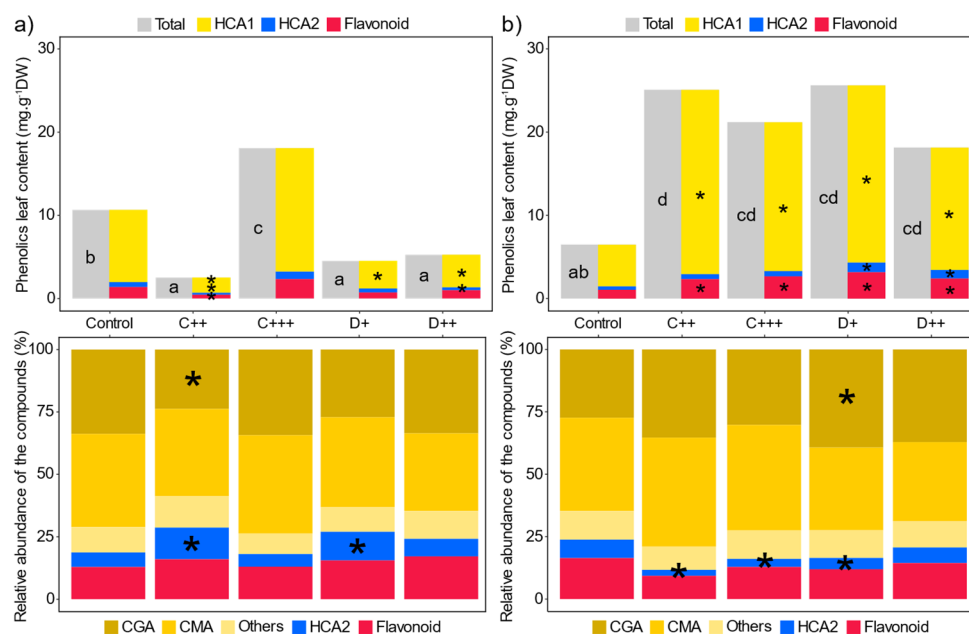


Figure 7. Impact of amendments on leaf phenolic compounds. Mean leaf total phenolic, hydroxycinnamic acid 1 (HCA1), hydroxycinnamic acid 2 (HCA2), and flavonoid concentrations ($n = 5$, $\text{mg}\cdot\text{g}^{-1}\text{DW} \pm \text{SE}$) depending on the nature of amendment (C: compost; D: digestate) and rate (+; ++; +++) applied in (a) St-Symphorien-sur-Saône and (b) Tavazzano soils and their relative quantitative abundances in percentage (chlorogenic acid (CGA), caffeoylmalic acid (CMA) and others are related to HCA1). Different letters indicate significant differences between treatments and soils for total phenolic compounds (Kruskal–Wallis test, $p < 0.05$). * indicates significant differences from the control for each considered compound and for each soil (Tukey's test, $p < 0.05$).

These observed changes in concentrations did not strongly impact the quantitative abundance of the CMA and flavonoid compounds. In the St-Symphorien-sur-Saône soil, C++ significantly reduced the abundance of CGA by ten and concomitantly increased the abundance of HCA2 by six with D+ treatment (Figure 7a). In the Tavazzano soil, D+ significantly increased the abundance of CGA by more than 10% but decreased the percentage of HCA2 by three to four along with compost (Figure 7b).

3. Discussion

3.1. *Urtica dioica* Can Be Grown in Hg-Contaminated Soils, but Its Growth Is Impacted by Soil Properties

Urtica dioica grows spontaneously in contaminated lands but its cultivation in such soils has not been previously studied. However, the contamination by inducing stress can affect plant growth and physiology [64,65]. In this context, assessment and improvement of plant growth and physiological traits are important [66]. In our study, stinging nettle grew well in Hg-contaminated soils, with a photosystem II photochemical efficiency slightly below 0.8, suggesting that the nettles were not stressed by soil conditions [67,68]. The two soils investigated in the present study exhibited different soil properties, particularly in terms of Hg concentration, which was six times higher for the Tavazzano soil. However, the levels of Hg found in the tissues of nettles grown in both soils were similar to those measured in crops (e.g., 6–139 $\mu\text{g}\cdot\text{kg}^{-1}$, [31]), suggesting that Hg was poorly transferred to the aboveground part of the plant, as previously described [29,69,70]. However, the Hg levels were approximately five times greater than those found in our previous study [71], which is probably because the nettle leaves from the present study were much younger, closer to the soil, and probably more active in terms of Hg uptake.

The stinging nettles grown in the Tavazzano soil produced overall more biomass, with higher plants and greater stem diameters than the nettles from the St-Symphorien-sur-Saône soil. Soil is the main driver of nettle growth, and although the Tavazzano soil is slightly lower in N, it contains more phosphorus, more clay, and two times less organic matter than the St-Symphorien-sur-Saône soil. This suggests that, probably more than N, described as the most important element in stinging nettle nutrition [72], P and soil texture are important parameters for the growth and development of *U. dioica*.

3.2. Solid Digestate Has a Greater Impact Than Compost on Stinging Nettle Growth

Compost and solid digestate have been described as effective amendments to improve plant growth [73,74]. Nevertheless, the effects of digestate described in the literature are highly variable due to the dose, fraction (i.e., liquid or solid), the type of feedstock used for digestion, and the type of soil studied [75]. In our study, the application of digestate and compost had different impacts. The two higher doses of amendments (C+++ and D++) increased the biomass, relative growth, plant height, stem diameter and length, and number of internodes of nettle grown in the St-Symphorien-sur-Saône soil. In the Tavazzano soil, only digestate significantly improved plant growth parameters. Overall, the nature of the amendments had a great impact on plant parameters since the effect of compost differed according to the soil type [76]. Notably, D++ significantly increased all measured parameters compared to compost while providing half as much N. In the St-Symphorien-sur-Saône soil, D+ significantly doubled plant biomass, as did C+++ while C++ did not improve biomass. Thus, much more compost than digestate would be needed to improve plant parameters at this site. These results confirm the lower impact of compost on plant growth and biomass in contaminated soils [77,78] when compared to digestate [79]. This may be due to the composition of the digestate, which contains more directly available plant N than the compost [80,81]. In addition, although the compost used contained more organic matter than the solid digestate, this amendment was produced from plant material. Therefore, this OM is partly composed of lignin, which is hardly biodegradable [81]. N is therefore mainly present in organic form and thus more slowly released and available to plants [82]. Mineralization of compost may be a longer process whereas stinging nettle has

a high growth rate. Nitrogen release is therefore not synchronous with plant N uptake, as previously demonstrated for sweet corn [83]. Another possibility for the low impact of compost on plant growth may be the maturity of the amendment [40]. N immobilization can indeed occur quickly in less mature composts [84]. As a consequence, microorganisms scavenge available N from the soil that becomes unavailable to plants [85,86], explaining that nettles were similar to the control in terms of plant parameters, without N accumulation in aerial parts but with higher microbial biomass in all treatments in the St-Symphorien-sur-Saône soil.

Compost application had no impact on nettles grown in the Tavazzano soil, but it can provide a longer-lasting effect [87]. Compost thus does not appear to be suitable for improving the growth of *U. dioica* in soils with low nutrient availability in the short term. The low impact of compost on plant yield following the first application has already been described in the case of fertile soils [55,83], but further investigations are needed to assess its potential effect in the long term. In addition, relative growth was negatively impacted by digestate during the first days of cultivation. Temporary immobilization of soil N has already been described during the use of organic amendments [88] and may be responsible for this slowing in plant growth. It can therefore be recommended to add the amendments two weeks before planting the nettles. Nevertheless, the nettles have started to grow despite the low availability of nutrients, probably thanks to their rhizomes, which constitute an N reserve that can be mobilized primarily for shoot development [89]. Finally, the relative growth of *U. dioica* was enhanced using amendments after several weeks of cultivation (i.e., 63 days).

3.3. The Dose of Amendment Applied Impacts the Growth of the Stinging Nettle and Reduces the Concentration of Hg in the Aerial Parts

Although described as a crop with low requirements for agricultural soils, the use of quite high fertilization doses has been reported for *U. dioica* cultivation [32,34]. In the present study, nettle biomass followed the dose of amendment applied. However, high doses of amendments were used in this study to double the nettle biomass. Growing *U. dioica* on marginal lands seems to require high inputs to enhance its biomass, which should be considered regarding environmental concerns (e.g., gaseous emission, nutrients leaching, eutrophication) [90,91]. Moreover, under the highest fertilization dose of digestate, the nettle plants all reached the flowering stage, while flowers only started to appear in the other treatments except for the control. As the flowering stage induces the cessation of vegetative growth [92], the digestate significantly improved the biomass produced but reduced the growth period.

In addition to impacting the biomass, the production of fibers was also enhanced without impact on the fiber size, which was similar to those previously reported and therefore similar to fibers of hemp (*Cannabis sativa* L.) and flax (*Linum usitatissimum* L.) [37]. As nettle fibers are potentially relevant candidates for composite reinforcement applications [29,93], improving nettle traits without TE accumulation through fertilization can be valuable for biomass production from contaminated lands. Organic amendments can indeed modulate the mobility and bioavailability of TE [94,95]. In our study, D++ significantly reduced the leaf Hg concentration for both soils, and the stem Hg concentration for the St-Symphorien-sur-Saône soil. Digestate efficiently reduced the mobility of Hg in an artificially contaminated chernozem soil [96], suggesting that solid digestate in high doses can reduce Hg transfer from the soil to the plant by reducing its mobility in the soil. Digestate therefore appears to be a relevant candidate to improve the performance of *U. dioica* grown in contaminated soil, but the dose should be adapted for in situ application.

3.4. Organic Amendments Differentially Impact Soil Enzyme Activities Depending on the Soil Type

In addition to its impact on plant parameters, fertilization is also known to induce changes in soil microbial communities and indirectly impact associated enzymatic activities [97,98]. In our study, the application of digestate significantly improved the respiration of the two soils and the microbial biomass of the St-Symphorien-sur-Saône soil, as observed in previous experiments [99–102]. In contrast, compost application in the Tavazzano soil and C++ application in the St-Symphorien-sur-Saône soil did not significantly impact the soil respiration or the root biomass, which was not significantly different or lower compared to the control plants. This result follows previous studies that highlighted a significant correlation between root biomass and soil respiration [103,104]. Nevertheless, soil respiration was also correlated with dsDNA and enzymatic activities in the St-Symphorien-sur-Saône soil, and amendment application significantly increased the esterase activities in both soils. Butyrate esterase (butyr) has been described to be related to living biomass content [105] and was strongly correlated with dsDNA in the St-Symphorien-sur-Saône soil. Therefore, the enzymatic activities measured in our study were likely related to microorganisms rather than to plants [106]. These results follow previous studies that highlighted higher soil microbial biomass and enzymatic activities in response to the application of amendments to contaminated soils [107–110]. However, nettle growth was not affected by the increased microbial biomass and higher enzymatic activities under the C++ treatment. This indicates that compost did not improve plant nutrient uptake [111], or that the released nutrients were immobilized or used by soil microorganisms [112,113]. Heijboer et al. [114] highlighted that soil nitrogen retention or microbial immobilization of nitrogen can occur with organic amendments at the expense of plant growth, and this phenomenon can last for a few months [82]. This may explain the fact that only the high doses of amendments impacted plant growth in our study, and that root biomass followed aerial biomass. We hypothesized that when nutrient release was sufficient for soil microorganisms and plants, the stinging nettle produced more roots to take up nutrients.

However, the amendments differentially impacted the enzymatic activities depending on the studied soil after three months of cultivation. In the St-Symphorien-sur-Saône soil, amendments significantly increased almost all measured activities (except those involved in the C cycle), while they were mainly comparable or slightly reduced in the Tavazzano soil. This may be related to the properties of the soil or to the dose of amendments that may have a negative impact on enzymatic activities [60,115,116]. In our study, the soil pH cannot explain the variations in the enzymatic activities [117], as only compost application slightly increased it in the Tavazzano soil. However, since the mineralization of nutrients from amendments depends on soil properties and may be faster in more fertile soils [118,119], it can be argued that the mineralization rate of amendments was higher in the Tavazzano soil. Thus, all easily degradable organic matter has already been used, resulting in the restoration of soil microbial activity [120]. Previous experiments have shown a return to baseline enzymatic activities 2–3 months after compost application [121]. Additionally, the Tavazzano soil has a higher clay content, which has been described as retaining organic carbon and nitrogen through sorption of organic matter, making them inaccessible to microbes [122,123]. Access to organic matter is therefore easier for microbes in less textured soils such as that of St-Symphorien-sur-Saône, which explains why amendments improved the growth and biomass of *U. dioica* in the St-Symphorien-sur-Saône soil better than in the Tavazzano soil. In addition to improving nettle growth, amendment application also improved soil parameters and conditions that can improve nettle re-establishment and improve re-growth for further harvest.

3.5. Fertilization Impacts the Production of Leaf Phenolic Compounds of the Stinging Nettle

In addition to enzymatic activities, organic amendments differentially affected the leaf phenolic compound concentration of *U. dioica* depending on the soil used. The concentrations of total phenolic compounds, HCA2, and flavonoids did not significantly differ between the control plants grown on the two soils. However, while the amendments significantly increased the total phenolic compound concentrations of nettle leaves grown on the Tavazzano soil, they significantly decreased phenolic compounds in the nettle leaves from the St-Symphorien-sur-Saône soil. The dose of amendments did not differentially affect the concentration of total phenolic compounds following the results from Biesiada et al. [124]. In previous studies, the amount of phenolics decreased upon application of amendment due to an improvement in plant growth status [125,126], as described for *U. dioica* following the growth–differentiation balance hypothesis [127]. Thus, biomass accumulation and specialized metabolism are negatively correlated. As a result, lower production of specialized metabolites is observed when plants are cultivated under sufficient nutrients. A greater improvement in plant growth with the amendments was observed in our study on the St-Symphorien-sur-Saône soil compared to the Tavazzano soil.

In contrast, when growth is reduced due to limited nutrient availability, the production of phenolic compounds is enhanced [128–131]. The accumulation of phenolic compounds is commonly observed as a response to abiotic stress [132] and has been previously described with digestate application [133]. Our results do not follow this rule, as the nettles grown on the Tavazzano soil were not stressed and the application of amendments did not reduce their growth. However, the applied organic amendments contained large amounts of C while N was still limited for the plants. As a consequence, the nettle may have allocated the extra C to the synthesis of phenolic compounds failing to improve its growth [134].

The observed differences in phenolic responses to N fertilization are compound-specific, depending on their biosynthetic pathways [135]. Here, in both soils, hydroxycinnamic acids were the most abundant compounds, as previously described for *U. dioica* [136]. Phenolic acids have been associated with various functions, such as nutrient uptake [137]. The HCA1 concentration was significantly impacted and explained the observed changes in the total phenolic concentration. Flavonoid concentration was impacted, although to a lesser extent, in the nettles from the Tavazzano soil. N limitation has been described as influencing flavonoid biosynthesis [138,139]. When plant nutrient uptake was enhanced, *U. dioica* decreased its phenolic compound production, mainly HCA1, which are the main compounds. In contrast, production is increased when improving growth is not possible, but further investigations on the role of these compounds are still needed.

4. Materials and Methods

4.1. Collection and Characteristics of the Soils

Two Hg-contaminated soils from St-Symphorien-sur-Saône (47°05′03.9″ N, 5°19′48.7″ E, France) and Tavazzano (45°19′17.0″ N, 9°24′04.1″ E, Italy) were used for the experiment that was conducted in pots. The St-Symphorien-sur-Saône site is a chlor-alkali sediment landfill already described in previous studies [140,141]. The Tavazzano site is a grassland adjacent to a chlor-alkali plant using Hg cell technology. Both soils were sampled at a depth of 0–30 cm, air-dried, and sieved to 4 mm. The total N and C contents were analyzed after dry combustion (Vario Macro Cube, Elementar, Langensfeld, Germany). The soil organic matter content (SOM) was determined by heating the soil sample up to 600 °C (soil TOC cube, Elementar, Langensfeld, Germany). The soil total concentrations of major elements and TE were determined by inductively coupled plasma atomic emission spectrometry (ICP–AES, Thermo Fischer Scientific, Inc., Pittsburgh, PA, USA) after acid digestion of 500 mg of sample (Table 2). The pH of the soil and organic amendments was determined after slow shaking for 30 min at room temperature in a 1:25 soil/1 M KCl solution.

Table 2. Properties and elemental composition of St-Symphorien-sur-Saône and Tavazzano soils (mean $\pm$ SE).

	Unit	St-Symphorien-sur-Saône (Fr)	Tavazzano (It)
Sand	%	3.8	10.5
Silt	%	87.4	72.5
Clay	%	8.9	16.9
pH		$8.0 \pm 0.3 \times 10^{-1}$	$6.8 \pm 0.5 \times 10^{-1}$
C/N		24.0 ± 0.7	18.3 ± 0.6
SOM ¹	%	4.0 ± 0.1	1.9 ± 0.2
Total N	%	$0.1 \pm 0.3 \times 10^{-2}$	$0.07 \pm 0.4 \times 10^{-2}$
TOC ²	%	$2.3 \pm 0.5 \times 10^{-1}$	1.1 ± 0.1
Ca	g/kg	236.8 ± 0.2	$3.2 \pm 3.7 \times 10^{-2}$
K	g/kg	$1.0 \pm 1.9 \times 10^{-2}$	$2.5 \pm 0.2 \times 10^{-2}$
Mg	g/kg	1.3 ± 0.2	$4.5 \pm 4.0 \times 10^{-2}$
Na	mg/kg	981.8 ± 16.2	84.2 ± 0.8
P	mg/kg	449.0 ± 7.2	753.5 ± 7.3
Fe	g/kg	$6.0 \pm 7.3 \times 10^{-2}$	14.2 ± 0.1
S	g/kg	13.5 ± 0.1	$0.4 \pm 0.5 \times 10^{-2}$
As	mg/kg	17.1 ± 0.3	9.2 ± 0.1
Cu	mg/kg	12.6 ± 0.1	$16.1 \pm 0.6 \times 10^{-1}$
Hg	mg/kg	6.9 ± 0.3	30.0 ± 0.4
Mn	mg/kg	326.1 ± 6.2	214.1 ± 1.1
Ni	mg/kg	8.7 ± 0.2	26.6 ± 0.2
Pb	mg/kg	17.2 ± 0.8	20.7 ± 0.2
Zn	mg/kg	41.7 ± 0.4	44.5 ± 2.6

¹ SOM: soil organic matter. ² TOC: total organic carbon.

4.2. Greenhouse Experimental Design

The company Agrivalor© (Hirsingue, France) supplied the organic amendments prepared from green wastes (compost: C), or from organic food wastes (i.e., food production byproducts, unsold and expired products; solid digestate: D) (Table S5). These two amendments were mixed with 780 g of sieved soil and placed in one-liter pots. Compost was added to the soil at doses of 0.04% N (i.e., 0.36 g N/kg soil, C++) and 0.07% N (i.e., 0.72 g N/kg soil, C+++), and the solid digestate was added to the soil at doses of 0.02% N (i.e., 0.18 g N·kg⁻¹ soil, D+) and 0.04% N (i.e., 0.36 g N·kg⁻¹ soil, D++). These doses gave the highest stinging nettle (*Urtica dioica*) biomass improvement based on preliminary experiments (Figure S3). One *U. dioica* plantlet harboring a single rhizome was planted per pot. The High School of Horticulture of Roville-aux-Chênes (Roville-aux-Chênes, France) supplied the nettles, which are produced for multiple uses, including textile fiber. Ten replicates were performed per treatment (i.e., control, C++, C+++, D+, and D++) for each soil. Nettles were grown in a greenhouse for 83 days (e.g., until the first nettles reached the flowering stage) from April to June.

4.3. Measured Plant Parameters

Plant height was measured twice a week from Day 0 to Day 83. The leaf chlorophyll content was measured on the second leaf with a chlorophyll content meter (Opti-Sciences, Inc., CCM-200, Tyngsboro, MA, USA) [142]. Photosystem II (PSII) performance was measured on the basis of the fluorescence ratio ($F_m - F_0/F_m$) using a MINI-PAM II (Walz, Germany) after placing the plants in the dark for 2 h [143]. At harvest, aerial parts were separated from roots, washed with tap water, and rinsed in distilled water. The stem diameter was measured with a caliper from the middle part of the stem, and the length of internodes was measured.

One internode from the middle of the stem of nettles amended with the highest doses of compost and digestate on St-Symphorien-sur-Saône soil (n = 3 per treatment) was kept in 70% ethanol to characterize the fiber morphology using X-ray microtomography performed on an EasyTom tomograph (RX Solutions, Chavanod, France). The stem pieces were placed

vertically in an Eppendorf tube in ethanol. The Eppendorf tube was glued on the sample holder to avoid movement during acquisition. The sample holder was mounted on a rotating stage to allow for a rotation of 360°. The X-ray source, a Hamamatsu Open Type Microfocus L10711 (RX Solution, Chavanod, France), was operated with an electron current of 84 μ A and a tube voltage of 60 kV. The X-ray transmission images were acquired using a 2530DX detector of 2176×1792 pixels². The exposure time and the average frame were fixed at 1.5 s and 2 images, respectively, and 1440 images per revolution were acquired. The entire volume was reconstructed at a full resolution with a voxel size of 2 μ m corresponding to a field of view of 3.8×2.8 mm², using filtered back-projection. The data analysis was processed using VG StudioMax software. The analysis of the grey levels was achieved using a threshold to separate the fiber wall from the air and access the geometry of the fiber. These images were then post-processed in MATLAB software. The first step consisted of automatically closing the open contours and removing particles smaller than 5 μ m. Then, each fiber was identified and processed individually. The minimum and maximum Feret diameters of each fiber were determined. The average wall thickness was also determined using the coordinates of the center of the ellipse.

The third nettle leaves from the top were scanned and the leaf area was measured using ImageJ software (version 1.53r) [144]. Samples were then air-dried at room temperature and weighed for dry biomass prior to subsequent analysis.

4.4. Biochemical Analyses on Plant Leaves

Dry leaves were ground using an MM400 Mixer Mill (Retsch, Eragny sur Oise, France) for 1 min at 30 Hz. Composites of two plants were used for subsequent analysis, resulting in $n = 5$ replicates per treatment for each soil. Two milligrams of leaf powder were used for CHN analysis (FlashEA 1112, ThermoFisher Scientific, Waltham, MA, USA). Soluble phenolics were extracted from 15–20 mg of dry powder with 0.6 mL methanol/H₂O (4:1) by 10 min of sonication, followed by shaking for 10 min in the dark. The extraction was repeated twice, and all supernatants were pooled. Trans-cinnamic acid was used as internal standard. Extracts were analyzed by UPLC-DAD-ESI-MS/MS (Nexera2, LCMS-8040, Shimadzu, Kyoto, Japan) using a Luna 5 μ m C18(2) 100 Å, 250×3 mm column (Phenomenex, Torrance, CA, USA) and a C18 guard column with solvent A (10% methanol and 0.2% formic acid) and solvent B (98% methanol and 0.2% formic acid) and the following gradient: 0–1 min of 8% B; 5 min 20% B; 18 min 55% B; 20 min 100% B; 20–26 min of 100% B. The flow rate was 0.35 mL min^{−1}, and the column oven temperature was 40 °C. Quantification was performed using UV detection at 320 nm for hydroxycinnamic acids (chlorogenic acid (CGA) as the standard), 360 nm for flavonols (hyperoside as the standard), and 280 nm for t-cinnamic acid (internal standard). MS detection was used for identification using the following conditions: nebulizing gas (N₂), 3 L min^{−1}; drying gas (N₂), 15 L min^{−1}; desolvation line, 250 °C; heat block temperature, 400 °C; and interface voltage 4.5 kV. Finally, Hg quantification was performed from 25 mg of leaves using an AMA-254 (Altec Co, Chotěboř, Czech Republic) as described in Maillard et al. [140].

4.5. Analyses of Soil Enzymatic Activities, Microbial Biomass, and Respiration

Fresh rhizospheric soil was sampled for enzymatic activities and soil microbial biomass. Seventeen enzymatic activities (EA) involved in C, N, P, and S cycling were measured according to Ferrarini et al. [145]: α -glucosidase (alfaG, EC 3.2.1.20), β -glucosidase (betaG, EC 3.2.1.21), α -galactosidase (alfaGAL, EC 3.2.1.22), β -galactosidase (beta GAL, EC 3.2.1.23), β -D-glucuronidase (uroni, EC 3.2.1.31), β -1,4-glucanase (cell, EC 3.2.1.4), β -1,4-xylanase (xilo, EC 3.2.1.14), N-acetyl- β -D-glucosaminidase (chit, EC 3.1.3.2), leucine amino-peptidase (leu, EC 3.4.11.1), acid (acP, EC 3.1.3.2) and alkaline phosphomonoesterase (alkP, EC 3.1.3.1), phosphodiesterase (bisP, EC 3.1.4.1), pyrophosphodiesterase (piroP, EC 3.6.1.9), inositol-P phosphatase (inositP, EC 3.1.3.25), arylsulfatase (aryS, EC 3.1.6.1), butyrate esterase (butyr, EC 3.1.1.1), and nonanoate esterase (nona, EC 3.1). Briefly, enzymatic activities were measured from 400 mg of soil added to 1.4 mL of 3% lysozyme solution and glass plus

ceramic beads. After shaking and centrifugation, enzymatic activities were quantified fluorometrically from the supernatant and expressed as nanomoles of 4-MUF·g⁻¹ soil per hour. Fluorogenic substrates with 7-amino-4-methyl coumarin (AMC; leu) and 4-methyl-umbelliferyl (MUF; other enzymes) were used as fluorophores.

Soil microbial biomass was measured from the double-stranded DNA (dsDNA) concentration using 400 mg of soil in 0.12 M sodium phosphate solution at pH 7.8 as an extraction buffer [146]. PicoGreen reagent (Life Technologies, Carlsbad, CA, USA) was then used to quantify dsDNA, expressed as nanograms of dsDNA g⁻¹ dry soil after being corrected for soil moisture content.

Soil microbial respiration was monitored using an Xstream gas analyzer (Emerson, Langenfeld, Germany). At the end of the experiment, 18 g of fresh soil samples adjusted at 63% of the field capacity were placed in tight flasks and incubated at 23 °C. The produced CO₂ was measured at the end of the experiment and every 4 days, for a total of 16 days using 5 mL aliquots of the flask atmosphere (sampled using a syringe). An empty sealed flask was used as a negative control. The respiration activity was represented by the produced CO₂ expressed as carbon mass per gram of DW soil per day (μgC·g⁻¹·d⁻¹).

4.6. Statistical Analyses

Statistical analyses and data visualization were performed using R software (version 2022.07.2+576) [147] and were considered significant at $p < 0.05$. All individuals and variables were plotted on a PCA biplot using the FactoMineR package (version 2.4) [148]. Variables were colored according to their contribution to the principal components. The effect of the nature of the amendment, the dose applied, and the soil on plant parameters, enzymatic activities, and phenolic compounds was assessed using a permutational multivariate analysis of variance (PERMANOVA) based on the Euclidean distance matrix using the vegan package (version 2.5-7) [149]. Three-way analyses of variance (ANOVAs) were performed to test the effects of the nature of the amendment, the dose applied, and the soil on the nettle aerial biomass. All the variables were checked for normality distribution (Shapiro–Wilk test) and homoscedasticity (Levene test). For variables that fit to normal distribution, comparisons between soils and treatments were assessed by Tukey's HSD multiple comparison post-hoc analyses. The other variables were analyzed using the Kruskal function from the agricolae package (version 1.3-1) [150]. A correlation matrix was performed using the corrplot package to assess the correlations between soil respiration, dsDNA, the percentage of N in the leaves, and the enzymatic activities significantly impacted by the addition of amendments.

5. Conclusions

Urtica dioica was able to grow in the two Hg-contaminated soils of the present study, without any induced stress. The solid digestate in both doses greatly enhanced the biomass of the nettle in contrast to the compost and seems suitable for improving nettle parameters and fiber production. Moreover, digestate application decreased the leaf Hg concentration at high doses. The digestate, and to a lesser extent the compost, improved soil functions, particularly soil respiration and the enzyme activities involved in C, N, P, and S cycling. The use of organic amendments also impacted the leaf synthesis of hydroxycinnamic acids, but their role still needs to be further studied. The plant response differed depending on the soil characteristics, but the application of an appropriate dose of digestate could be proposed as an efficient strategy for the improvement of nettle cultivation on industrial lands. Field cultivation and *U. dioica* re-establishment need further investigation to assess the long-term efficiency of these organic amendments, and the quantity and quality of fibers for industrial use.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/plants13172425/s1>: Figure S1: Mean soil dsDNA ($n = 10$, $\text{ng}\cdot\text{g}^{-1} \pm \text{SE}$) after 83 days of cultivation on St-Symphorien-sur-Saône soil depending on the nature of amendment (C: compost; D: digestate) and rate (+; ++; ++++) applied. Different letters indicate significant differences between treatments (Kruskal–Wallis test, $p < 0.05$); Figure S2: Mean percentage of nitrogen ($n = 5 \pm \text{SE}$) in *Urtica dioica* leaves after 83 days of cultivation on (a) St-Symphorien-sur-Saône and (b) Tavazzano soils depending on the nature of amendment (C: compost; D: digestate) and rate (+; ++; ++++) applied. Different letters indicate significant differences between treatments and soils (Tukey’s test, $p < 0.05$); Figure S3: Aboveground dry biomass of *Urtica dioica* ($n = 10$) in response to different rates (indicated by + and –) of (a) solid digestate (D) and (b) compost (C) in St-Symphorien-sur-Saône soil. Different letters indicate significant differences between treatments (Kruskal–Wallis test, $p < 0.05$); Table S1: Mean chlorophyll content and photosystem II activity (PSII) ($n = 10 \pm \text{SE}$) of *Urtica dioica* leaves after 83 days of cultivation depending on the nature of amendment (C: compost; D: digestate), rate (+; ++; ++++) applied, and soil (St-Symphorien-sur-Saône or Tavazzano). Different letters indicate significant differences between treatments and soils for each variable (Kruskal–Wallis test, $p < 0.05$); Table S2: Fiber properties from *Urtica dioica* grown on St-Symphorien-sur-Saône soil depending on the amendment (C: compost; D: digestate) and rate (++; ++++) applied ($n = 3 \pm \text{SE}$); Table S3: Mean enzymatic activities ($n = 10 \pm \text{SE}$) expressed in nanomoles of $4\text{-MUF}\cdot\text{g}^{-1}$ or $\text{AMC}\cdot\text{g}^{-1}$ soil in St-Symphorien-sur-Saône and Tavazzano rhizospheric soils after 83 days of cultivation depending on the nature of amendment (C: compost; D: digestate) and rate (+; ++; ++++) applied. * indicates a mean significantly different from the control for each soil; Table S4: Soil pH at T0 and at the end of the experiment (T83) for the two soils depending on the nature of amendment (C: compost; D: digestate) and the rate applied (+; ++; ++++). Different letters indicate significant differences between treatments (Kruskal–Wallis test, $p < 0.05$); Table S5: Properties of the compost and digestate used.

Author Contributions: Conceptualization, C.V., C.B. and M.C.; formal analysis, C.V.; data curation, C.V., F.M., L.Y., V.P., A.F., F.F. and M.P.; writing—original draft preparation, C.V. and M.C.; writing—review and editing, C.B., F.M., L.Y., V.P., A.F. and D.B.; visualization, C.V.; supervision, M.C.; project administration, M.C.; funding acquisition, M.C. All authors have read and agreed to the published version of the manuscript.

Funding: This project received funding from the European Union’s Horizon 2020 research and innovation program under grant agreement No. 771134. The project NETFIB was carried out under the ERA-NET Cofund SusCrop (Grant No. 771134), being part of the Joint Programming Initiative on Agriculture, Food Security, and Climate Change (FACCE-JPI). C.V. received a PhD grant from the Ministère de l’Enseignement supérieur, de la Recherche et de l’Innovation (MESRI). D.B. received funding from the French PIA project “Lorraine Université d’Excellence” Deepsurf, reference ANR-15-IDEX-04-LUE. Part of this work was also supported by the EIPHI Graduate School (contract “ANR-17-EURE-0002”). F.M. received a Grant from; ERDF, grant number A75832 (decision EURA 2014/9165/09 02 01 01/2019/POPELY).

Data Availability Statement: Data are contained within the article and Supplementary Materials.

Acknowledgments: We acknowledge Léa Pouget for her involvement in plant cultivation and harvest. V.P. acknowledges the MIFHySTO technological platform (FEMTO-ST, France) for the use of X-ray nanotomography and Xavier Gabrion and Thomas Jeannin for their support in micro-computed tomography experiments and data processing.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. Niewöhner, J.; Bruns, A.; Haberl, H.; Hostert, P.; Krueger, T.; Lauk, C.; Lutz, J.; Müller, D.; Nielsen, J.Ø. Land Use Competition: Ecological, Economic and Social Perspectives. In *Land Use Competition: Ecological, Economic and Social Perspectives*; Niewöhner, J., Bruns, A., Hostert, P., Krueger, T., Nielsen, J.Ø., Haberl, H., Lauk, C., Lutz, J., Müller, D., Eds.; Human-Environment Interactions; Springer International Publishing: Cham, Switzerland, 2016; pp. 1–17; ISBN 978-3-319-33628-2.
2. Harvey, M.; Pilgrim, S. The New Competition for Land: Food, Energy, and Climate Change. *Food Policy* **2011**, *36*, S40–S51. [CrossRef]
3. Hertel, T.; Steinbuks, J.; Baldos, U. Competition for Land in the Global Bioeconomy. *Agric. Econ.* **2013**, *44*, 129–138. [CrossRef]

4. Dauber, J.; Brown, C.; Fernando, A.; Finnan, J.; Krasuska, E.; Ponitka, J.; Styles, D.; Thrän, D.; van Groenigen, K.J.; Weih, M.; et al. Bioenergy from “Surplus” Land: Environmental and Socio-Economic Implications. *BioRisk* **2012**, *7*, 5–50. [\[CrossRef\]](#)
5. Mehmood, M.A.; Ibrahim, M.; Rashid, U.; Nawaz, M.; Ali, S.; Hussain, A.; Gull, M. Biomass Production for Bioenergy Using Marginal Lands. *Sustain. Prod. Consum.* **2017**, *9*, 3–21. [\[CrossRef\]](#)
6. Cai, X.; Zhang, X.; Wang, D. Land Availability for Biofuel Production. *Environ. Sci. Technol.* **2011**, *45*, 334–339. [\[CrossRef\]](#)
7. Gerwin, W.; Repmann, F.; Galatsidas, S.; Vlachaki, D.; Gounaris, N.; Baumgarten, W.; Volkmann, C.; Keramitzis, D.; Kiourtsis, F.; Freese, D. Assessment and Quantification of Marginal Lands for Biomass Production in Europe Using Soil-Quality Indicators. *SOIL* **2018**, *4*, 267–290. [\[CrossRef\]](#)
8. Muscat, A.; de Olde, E.M.; Candel, J.J.L.; de Boer, I.J.M.; Ripoll-Bosch, R. The Promised Land: Contrasting Frames of Marginal Land in the European Union. *Land. Use Policy* **2022**, *112*, 105860. [\[CrossRef\]](#)
9. Scordia, D.; Papazoglou, E.G.; Kotoula, D.; Sanz, M.; Ciria, C.S.; Pérez, J.; Maliarenko, O.; Prysiazhniuk, O.; von Cossel, M.; Greiner, B.E.; et al. Towards Identifying Industrial Crop Types and Associated Agronomies to Improve Biomass Production from Marginal Lands in Europe. *GCB Bioenergy* **2022**, *14*, 710–734. [\[CrossRef\]](#)
10. Blanco-Canqui, H. Growing Dedicated Energy Crops on Marginal Lands and Ecosystem Services. *Soil. Sci. Soc. Am. J.* **2016**, *80*, 845–858. [\[CrossRef\]](#)
11. Mohanty, A.K.; Vivekanandhan, S.; Pin, J.-M.; Misra, M. Composites from Renewable and Sustainable Resources: Challenges and Innovations. *Science* **2018**, *362*, 536–542. [\[CrossRef\]](#)
12. Bernal, M.P.; Gómez, X.; Chang, R.; Arco-Lázaro, E.; Clemente, R. Strategies for the Use of Plant Biomass Obtained in the Phytostabilisation of Trace-Element-Contaminated Soils. *Biomass Bioenergy* **2019**, *126*, 220–230. [\[CrossRef\]](#)
13. Evangelou, M.W.H.; Robinson, B.H.; Günthardt-Goerg, M.S.; Schulin, R. Metal Uptake and Allocation in Trees Grown on Contaminated Land: Implications for Biomass Production. *Int. J. Phytoremediation* **2013**, *15*, 77–90. [\[CrossRef\]](#) [\[PubMed\]](#)
14. Robinson, B.; Schulin, R.; Nowack, B.; Roulier, S.; Menon, M.; Clothier, B.; Green, S.; Mills, T. Phytoremediation for the Management of Metal Flux in Contaminated Sites. *For. Snow Landsc. Res.* **2006**, *80*, 221–224.
15. Burges, A.; Alkorta, I.; Epelde, L.; Garbisu, C. From Phytoremediation of Soil Contaminants to Phytomanagement of Ecosystem Services in Metal Contaminated Sites. *Int. J. Phytoremediation* **2018**, *20*, 384–397. [\[CrossRef\]](#) [\[PubMed\]](#)
16. Rascio, N.; Navari-Izzo, F. Heavy Metal Hyperaccumulating Plants: How and Why Do They Do It? And What Makes Them so Interesting? *Plant Sci.* **2011**, *180*, 169–181. [\[CrossRef\]](#)
17. Robinson, B.H.; Bañuelos, G.; Conesa, H.M.; Evangelou, M.W.H.; Schulin, R. The Phytomanagement of Trace Elements in Soil. *Crit. Rev. Plant Sci.* **2009**, *28*, 240–266. [\[CrossRef\]](#)
18. De Vos, B.; Souza, M.F.; Michels, E.; Meers, E. Industrial Hemp (*Cannabis sativa* L.) in a Phytoattenuation Strategy: Remediation Potential of a Cd, Pb and Zn Contaminated Soil and Valorization Potential of the Fibers for Textile Production. *Ind. Crops Prod.* **2022**, *178*, 114592. [\[CrossRef\]](#)
19. Yang, B.; Zhou, M.; Shu, W.S.; Lan, C.Y.; Ye, Z.H.; Qiu, R.L.; Jie, Y.C.; Cui, G.X.; Wong, M.H. Constitutional Tolerance to Heavy Metals of a Fiber Crop, Ramie (*Boehmeria Nivea*), and Its Potential Usage. *Environ. Pollut.* **2010**, *158*, 551–558. [\[CrossRef\]](#)
20. Perlein, A.; Bert, V.; Desannaux, O.; Fernandes de Souza, M.; Papin, A.; Gaucher, R.; Zdanevitch, I.; Meers, E. The Use of Sorghum in a Phytoattenuation Strategy: A Field Experiment on a TE-Contaminated Site. *Appl. Sci.* **2021**, *11*, 3471. [\[CrossRef\]](#)
21. Quinn, L.D.; Straker, K.C.; Guo, J.; Kim, S.; Thapa, S.; Kling, G.; Lee, D.K.; Voigt, T.B. Stress-Tolerant Feedstocks for Sustainable Bioenergy Production on Marginal Land. *Bioenerg. Res.* **2015**, *8*, 1081–1100. [\[CrossRef\]](#)
22. Kidd, P.; Mench, M.; Álvarez-López, V.; Bert, V.; Dimitriou, I.; Friesl-Hanl, W.; Herzig, R.; Olga Janssen, J.; Kolbas, A.; Müller, I.; et al. Agronomic Practices for Improving Gentle Remediation of Trace Element-Contaminated Soils. *Int. J. Phytoremediation* **2015**, *17*, 1005–1037. [\[CrossRef\]](#)
23. Michalak, A. Phenolic Compounds and Their Antioxidant Activity in Plants Growing under Heavy Metal Stress. *Pol. J. Environ. Stud.* **2006**, *15*, 523–530.
24. Šamec, D.; Karalija, E.; Šola, I.; Vujčić Bok, V.; Salopek-Sondi, B. The Role of Polyphenols in Abiotic Stress Response: The Influence of Molecular Structure. *Plants* **2021**, *10*, 118. [\[CrossRef\]](#)
25. Kisa, D.; Elmastaş, M.; Öztürk, L.; Kayır, Ö. Responses of the Phenolic Compounds of Zea Mays under Heavy Metal Stress. *Appl. Biol. Chem.* **2016**, *59*, 813–820. [\[CrossRef\]](#)
26. Bautista, I.; Boscaiu, M.; Lidón, A.; Llinares, J.V.; Lull, C.; Donat, M.P.; Mayoral, O.; Vicente, O. Environmentally Induced Changes in Antioxidant Phenolic Compounds Levels in Wild Plants. *Acta Physiol. Plant* **2015**, *38*, 9. [\[CrossRef\]](#)
27. Cheynier, V.; Comte, G.; Davies, K.M.; Lattanzio, V.; Martens, S. Plant Phenolics: Recent Advances on Their Biosynthesis, Genetics, and Ecophysiology. *Plant Physiol. Biochem.* **2013**, *72*, 1–20. [\[CrossRef\]](#)
28. Sharma, A.; Shahzad, B.; Rehman, A.; Bhardwaj, R.; Landi, M.; Zheng, B. Response of Phenylpropanoid Pathway and the Role of Polyphenols in Plants under Abiotic Stress. *Molecules* **2019**, *24*, E2452. [\[CrossRef\]](#)
29. Jeannin, T.; Yung, L.; Evon, P.; Labonne, L.; Ouagne, P.; Lecourt, M.; Cazaux, D.; Chalot, M.; Placet, V. Native Stinging Nettle (*Urtica dioica* L.) Growing Spontaneously under Short Rotation Coppice for Phytomanagement of Trace Element Contaminated Soils: Fibre Yield, Processability and Quality. *Ind. Crops Prod.* **2020**, *145*, 111997. [\[CrossRef\]](#)
30. Xu, J.; Bravo, A.G.; Lagerkvist, A.; Bertilsson, S.; Sjöblom, R.; Kumpiene, J. Sources and Remediation Techniques for Mercury Contaminated Soil. *Environ. Int.* **2015**, *74*, 42–53. [\[CrossRef\]](#)
31. Kabata-Pendias, A. *Trace Elements in Soils and Plants*, 4th ed.; CRC Press: Boca Raton, FL, USA, 2011; ISBN 978-1-4200-9368-1.

32. Viotti, C.; Albrecht, K.; Amaducci, S.; Bardos, P.; Bertheau, C.; Blaudez, D.; Bothe, L.; Cazaux, D.; Ferrarini, A.; Govilas, J.; et al. Nettle, a Long-Known Fiber Plant with New Perspectives. *Materials* **2022**, *15*, 4288. [\[CrossRef\]](#)
33. Taylor, K. Biological Flora of the British Isles: *Urtica dioica* L. *J. Ecol.* **2009**, *97*, 1436–1458. [\[CrossRef\]](#)
34. Di Virgilio, N.; Papazoglou, E.; Jankauskiene, Z.; Lonardo, S.; Praczyk, M.; Wielgusz, K. The Potential of Stinging Nettle (*Urtica dioica* L.) as a Crop with Multiple Uses. *Ind. Crops Prod.* **2015**, *68*, 42–49. [\[CrossRef\]](#)
35. Bredemann, G. *Die Große Brennessel Urtica dioica* L.; Akademie-Verlag: Berlin, Germany, 1959; p. 137.
36. Vogl, C.; Hartl, A. Production and Processing of Organically Grown Fiber Nettle (*Urtica dioica* L.) and Its Potential Use in the Natural Textile Industry: A Review. *Am. J. Altern. Agric.* **2003**, *18*, 119–128. [\[CrossRef\]](#)
37. Bacci, L.; Baronti, S.; Predieri, S.; di Virgilio, N. Fiber Yield and Quality of Fiber Nettle (*Urtica dioica* L.) Cultivated in Italy. *Ind. Crops Prod.* **2009**, *29*, 480–484. [\[CrossRef\]](#)
38. Müllerová, V.; Hejman, M.; Hejmanová, P.; Pavlů, V. Effect of Fertilizer Application on *Urtica dioica* and Its Element Concentrations in a Cut Grassland. *Acta Oecol.* **2014**, *59*, 1–6. [\[CrossRef\]](#)
39. Franco-Otero, V.G.; Soler-Rovira, P.; Hernández, D.; López-de-Sá, E.G.; Plaza, C. Short-Term Effects of Organic Municipal Wastes on Wheat Yield, Microbial Biomass, Microbial Activity, and Chemical Properties of Soil. *Biol. Fertil. Soils* **2012**, *48*, 205–216. [\[CrossRef\]](#)
40. Liu, L.; Wang, S.; Guo, X.; Wang, H. Comparison of the Effects of Different Maturity Composts on Soil Nutrient, Plant Growth and Heavy Metal Mobility in the Contaminated Soil. *J. Environ. Manage.* **2019**, *250*, 109525. [\[CrossRef\]](#)
41. Schröder, P.; Mench, M.; Povilaitis, V.; Rineau, F.; Rutkowska, B.; Schloter, M.; Szulc, W.; Žydelis, R.; Loit, E. Relaunch Cropping on Marginal Soils by Incorporating Amendments and Beneficial Trace Elements in an Interdisciplinary Approach. *Sci. Total Environ.* **2022**, *803*, 149844. [\[CrossRef\]](#)
42. Pahalvi, H.N.; Rafiya, L.; Rashid, S.; Nisar, B.; Kamili, A.N. Chemical Fertilizers and Their Impact on Soil Health. In *Microbiota and Biofertilizers, Vol 2: Ecofriendly Tools for Reclamation of Degraded Soil Environments*; Dar, G.H., Bhat, R.A., Mehmood, M.A., Hakeem, K.R., Eds.; Springer International Publishing: Cham, Switzerland, 2021; pp. 1–20. ISBN 978-3-030-61010-4.
43. Savci, S. An Agricultural Pollutant: Chemical Fertilizer. *IJESD* **2012**, *3*, 73–80. [\[CrossRef\]](#)
44. Lazcano, C.; Zhu-Barker, X.; Decock, C. Effects of Organic Fertilizers on the Soil Microorganisms Responsible for N₂O Emissions: A Review. *Microorganisms* **2021**, *9*, 983. [\[CrossRef\]](#) [\[PubMed\]](#)
45. Fedeli, R.; Celletti, S.; Loppi, S.; Vannini, A. Comparison of the Effect of Solid and Liquid Digestate on the Growth of Lettuce (*Lactuca sativa* L.) Plants. *Agronomy* **2023**, *13*, 782. [\[CrossRef\]](#)
46. Roy, S.; Arunachalam, K.; Dutta, B.K.; Arunachalam, A. Effect of Organic Amendments of Soil on Growth and Productivity of Three Common Crops Viz. Zea Mays, Phaseolus Vulgaris and Abelmoschus Esculentus. *Appl. Soil. Ecol.* **2010**, *45*, 78–84. [\[CrossRef\]](#)
47. Martínez-Blanco, J.; Lazcano, C.; Christensen, T.H.; Muñoz, P.; Rieradevall, J.; Møller, J.; Antón, A.; Boldrin, A. Compost Benefits for Agriculture Evaluated by Life Cycle Assessment. A Review. *Agron. Sustain. Dev.* **2013**, *33*, 721–732. [\[CrossRef\]](#)
48. Guilayn, F.; Rouez, M.; Crest, M.; Patureau, D.; Jimenez, J. Valorization of Digestates from Urban or Centralized Biogas Plants: A Critical Review. *Rev. Environ. Sci. Biotechnol.* **2020**, *19*, 419–462. [\[CrossRef\]](#)
49. Chojnacka, K.; Mikula, K.; Skrzypczak, D.; Izydorczyk, G.; Gorazda, K.; Kulczycka, J.; Kominko, H.; Moustakas, K.; Witek-Krowiak, A. Practical Aspects of Biowastes Conversion to Fertilizers. *Biomass Conv. Bioref.* **2022**, *14*, 1515–1533. [\[CrossRef\]](#)
50. Fernández-Delgado, M.; del Amo-Mateos, E.; Lucas, S.; García-Cubero, M.T.; Coca, M. Liquid Fertilizer Production from Organic Waste by Conventional and Microwave-Assisted Extraction Technologies: Techno-Economic and Environmental Assessment. *Sci. Total Environ.* **2022**, *806*, 150904. [\[CrossRef\]](#) [\[PubMed\]](#)
51. Nevens, F.; Reheul, D. The Application of Vegetable, Fruit and Garden Waste (VFG) Compost in Addition to Cattle Slurry in a Silage Maize Monoculture: Nitrogen Availability and Use. *Eur. J. Agron.* **2003**, *19*, 189–203. [\[CrossRef\]](#)
52. Riva, C.; Orzi, V.; Carozzi, M.; Acutis, M.; Boccasile, G.; Lonati, S.; Tambone, F.; D'Imporzano, G.; Adani, F. Short-Term Experiments in Using Digestate Products as Substitutes for Mineral (N) Fertilizer: Agronomic Performance, Odours, and Ammonia Emission Impacts. *Sci. Total Environ.* **2016**, *547*, 206–214. [\[CrossRef\]](#)
53. Sinclair, T.R.; Ruffy, T.W. Nitrogen and Water Resources Commonly Limit Crop Yield Increases, Not Necessarily Plant Genetics. *Glob. Food Secur.* **2012**, *1*, 94–98. [\[CrossRef\]](#)
54. Al-Bataina, B.B.; Young, T.M.; Ranieri, E. Effects of Compost Age on the Release of Nutrients. *Int. Soil. Water Conserv. Res.* **2016**, *4*, 230–236. [\[CrossRef\]](#)
55. Erhart, E.; Hartl, W.; Putz, B. Biowaste Compost Affects Yield, Nitrogen Supply during the Vegetation Period and Crop Quality of Agricultural Crops. *Eur. J. Agron.* **2005**, *23*, 305–314. [\[CrossRef\]](#)
56. Kuypers, M.M.M.; Marchant, H.K.; Kartal, B. The Microbial Nitrogen-Cycling Network. *Nat. Rev. Microbiol.* **2018**, *16*, 263–276. [\[CrossRef\]](#) [\[PubMed\]](#)
57. Baldrian, P. Distribution of Extracellular Enzymes in Soils: Spatial Heterogeneity and Determining Factors at Various Scales. *Soil. Sci. Soc. Am. J.* **2014**, *78*, 11–18. [\[CrossRef\]](#)
58. Wallenstein, M.D.; Burns, R.G. Ecology of Extracellular Enzyme Activities and Organic Matter Degradation in Soil: A Complex Community-Driven Process. In *Methods of Soil Enzymology*; John Wiley & Sons, Ltd.: Hoboken, NJ, USA, 2011; pp. 35–55; ISBN 978-0-89118-858-2.

59. Piazza, G.; Pellegrino, E.; Moscatelli, M.C.; Ercoli, L. Long-Term Conservation Tillage and Nitrogen Fertilization Effects on Soil Aggregate Distribution, Nutrient Stocks and Enzymatic Activities in Bulk Soil and Occluded Microaggregates. *Soil. Tillage Res.* **2020**, *196*, 104482. [\[CrossRef\]](#)
60. Shen, W.; Lin, X.; Shi, W.; Min, J.; Gao, N.; Zhang, H.; Yin, R.; He, X. Higher Rates of Nitrogen Fertilization Decrease Soil Enzyme Activities, Microbial Functional Diversity and Nitrification Capacity in a Chinese Polyunnel Greenhouse Vegetable Land. *Plant Soil.* **2010**, *337*, 137–150. [\[CrossRef\]](#)
61. Bowles, T.M.; Acosta-Martínez, V.; Calderón, F.; Jackson, L.E. Soil Enzyme Activities, Microbial Communities, and Carbon and Nitrogen Availability in Organic Agroecosystems across an Intensively-Managed Agricultural Landscape. *Soil. Biol. Biochem.* **2014**, *68*, 252–262. [\[CrossRef\]](#)
62. O'Connor, J.; Hoang, S.A.; Bradney, L.; Dutta, S.; Xiong, X.; Tsang, D.C.W.; Ramadass, K.; Vinu, A.; Kirkham, M.B.; Bolan, N.S. A Review on the Valorisation of Food Waste as a Nutrient Source and Soil Amendment. *Environ. Pollut.* **2021**, *272*, 115985. [\[CrossRef\]](#)
63. Ventorino, V.; Pascale, A.; Fagnano, M.; Adamo, P.; Faraco, V.; Rocco, C.; Fiorentino, N.; Pepe, O. Soil Tillage and Compost Amendment Promote Bioremediation and Biofertility of Polluted Area. *J. Clean. Prod.* **2019**, *239*, 118087. [\[CrossRef\]](#)
64. Dimkpa, C.O.; Merten, D.; Svatoš, A.; Büchel, G.; Kothe, E. Metal-Induced Oxidative Stress Impacting Plant Growth in Contaminated Soil Is Alleviated by Microbial Siderophores. *Soil. Biol. Biochem.* **2009**, *41*, 154–162. [\[CrossRef\]](#)
65. Ghorri, N.-H.; Ghorri, T.; Hayat, M.Q.; Imadi, S.R.; Gul, A.; Altay, V.; Ozturk, M. Heavy Metal Stress and Responses in Plants. *Int. J. Environ. Sci. Technol.* **2019**, *16*, 1807–1828. [\[CrossRef\]](#)
66. Raveau, R.; Fontaine, J.; Bert, V.; Perleir, A.; Tisserant, B.; Ferrant, P.; Lounès-Hadj Sahraoui, A. In Situ Cultivation of Aromatic Plant Species for the Phytomanagement of an Aged-Trace Element Polluted Soil: Plant Biomass Improvement Options and Techno-Economic Assessment of the Essential Oil Production Channel. *Sci. Total Environ.* **2021**, *789*, 147944. [\[CrossRef\]](#) [\[PubMed\]](#)
67. Krause, G.H.; Weis, E. Chlorophyll Fluorescence and Photosynthesis: The Basics. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* **1991**, *42*, 313–349. [\[CrossRef\]](#)
68. Parkhill, J.-P.; Maillet, G.; Cullen, J.J. Fluorescence-Based Maximal Quantum Yield for Psii as a Diagnostic of Nutrient Stress. *J. Phycol.* **2001**, *37*, 517–529. [\[CrossRef\]](#)
69. Edwards, S.; MacLeod, C.; Lester, J. The Bioavailability of Copper and Mercury to the Common Nettle (*Urtica dioica*) and the Earthworm *Eisenia Fetida* from Contaminated Dredge Spoil. *Water Air Soil. Pollut.* **1998**, *102*, 75–90. [\[CrossRef\]](#)
70. Hiller, E.; Jurkovič, L.; Majzlan, J.; Kulikova, T.; Faragó, T. Environmental Availability of Trace Metals (Mercury, Chromium and Nickel) in Soils from the Abandoned Mine Area of Merník (Eastern Slovakia). *Pol. J. Environ. Stud.* **2021**, *30*, 5013–5025. [\[CrossRef\]](#)
71. Yung, L.; Bertheau, C.; Cazaux, D.; Regier, N.; Slaveykova, V.; Chalot, M. Insect Life Traits Are Key Factors in Mercury Accumulation and Transfer within the Terrestrial Food Web. *Environ. Sci. Technol.* **2019**, *53*, 11122–11132. [\[CrossRef\]](#)
72. Rutto, L.; Ansari, M.; Brandt, M. Biomass Yield and Dry Matter Partitioning in Greenhouse-Grown Stinging Nettle under Different Fertilization Regimes. *Hort. Technol.* **2012**, *22*, 751–756. [\[CrossRef\]](#)
73. Dutta, S.; He, M.; Xiong, X.; Tsang, D.C.W. Sustainable Management and Recycling of Food Waste Anaerobic Digestate: A Review. *Bioresour. Technol.* **2021**, *341*, 125915. [\[CrossRef\]](#)
74. O'Brien, B.J.; Neher, D.A.; Roy, E.D. Nutrient and Pathogen Suppression Properties of Anaerobic Digestates from Dairy Manure and Food Waste Feedstocks. *Waste Biomass Valor.* **2020**, *11*, 6565–6573. [\[CrossRef\]](#)
75. Karimi, B.; Sadet-Bourgeteau, S.; Cannavacciuolo, M.; Chauvin, C.; Flamin, C.; Haumont, A.; Jean-Baptiste, V.; Reibel, A.; Vrignaud, G.; Ranjard, L. Impact of Biogas Digestates on Soil Microbiota in Agriculture: A Review. *Environ. Chem. Lett.* **2022**, *20*, 3265–3288. [\[CrossRef\]](#)
76. van Herwijnen, R.; Hutchings, T.R.; Al-Tabbaa, A.; Moffat, A.J.; Johns, M.L.; Ouki, S.K. Remediation of Metal Contaminated Soil with Mineral-Amended Composts. *Environ. Pollut.* **2007**, *150*, 347–354. [\[CrossRef\]](#) [\[PubMed\]](#)
77. Alvarenga, P.; Gonçalves, A.P.; Fernandes, R.M.; de Varennes, A.; Vallini, G.; Duarte, E.; Cunha-Queda, A.C. Evaluation of Composts and Liming Materials in the Phytostabilization of a Mine Soil Using Perennial Ryegrass. *Sci. Total Environ.* **2008**, *406*, 43–56. [\[CrossRef\]](#)
78. de la Fuente, C.; Clemente, R.; Martínez-Alcalá, I.; Tortosa, G.; Bernal, M.P. Impact of Fresh and Composted Solid Olive Husk and Their Water-Soluble Fractions on Soil Heavy Metal Fractionation; Microbial Biomass and Plant Uptake. *J. Hazard. Mater.* **2011**, *186*, 1283–1289. [\[CrossRef\]](#) [\[PubMed\]](#)
79. Montemurro, F.; Ferri, D.; Tittarelli, F.; Canali, S.; Vitti, C. Anaerobic Digestate and On-Farm Compost Application: Effects on Lettuce (*Lactuca sativa* L.) Crop Production and Soil Properties. *Compost. Sci. Util.* **2010**, *18*, 184–193. [\[CrossRef\]](#)
80. Möller, K.; Müller, T. Effects of Anaerobic Digestion on Digestate Nutrient Availability and Crop Growth: A Review. *Eng. Life Sci.* **2012**, *12*, 242–257. [\[CrossRef\]](#)
81. Teglia, C.; Tremier, A.; Martel, J.-L. Characterization of Solid Digestates: Part 2, Assessment of the Quality and Suitability for Composting of Six Digested Products. *Waste Biomass Valor.* **2011**, *2*, 113–126. [\[CrossRef\]](#)
82. Chadwick, D.R.; John, F.; Pain, B.F.; Chambers, B.J.; Williams, J. Plant Uptake of Nitrogen from the Organic Nitrogen Fraction of Animal Manures: A Laboratory Experiment. *J. Agric. Sci.* **2000**, *134*, 159–168. [\[CrossRef\]](#)
83. Evanylo, G.; Sherony, C.; Spargo, J.; Starner, D.; Brosius, M.; Haering, K. Soil and Water Environmental Effects of Fertilizer-, Manure-, and Compost-Based Fertility Practices in an Organic Vegetable Cropping System. *Agric. Ecosys. Environ.* **2008**, *127*, 50–58. [\[CrossRef\]](#)

84. Amlinger, F.; Götz, B.; Dreher, P.; Geszti, J.; Weissteiner, C. Nitrogen in Biowaste and Yard Waste Compost: Dynamics of Mobilisation and Availability—A Review. *Eur. J. Soil. Biol.* **2003**, *39*, 107–116. [\[CrossRef\]](#)
85. Juárez, M.F.-D.; Gómez-Brandón, M.; Insam, H. Merging Two Waste Streams, Wood Ash and Biowaste, Results in Improved Composting Process and End Products. *Sci. Total Environ.* **2015**, *511*, 91–100. [\[CrossRef\]](#)
86. Mohanty, M.; Sinha, N.K.; Sammi Reddy, K.; Chaudhary, R.S.; Subba Rao, A.; Dalal, R.C.; Menzies, N.W. How Important Is the Quality of Organic Amendments in Relation to Mineral N Availability in Soils? *Agric. Res.* **2013**, *2*, 99–110. [\[CrossRef\]](#)
87. Larney, F.J.; Angers, D.A. The Role of Organic Amendments in Soil Reclamation: A Review. *Can. J. Soil. Sci.* **2012**, *92*, 19–38. [\[CrossRef\]](#)
88. Goss, M.J.; Tubeileh, A.; Goorahoo, D. Chapter Five—A Review of the Use of Organic Amendments and the Risk to Human Health. In *Advances in Agronomy*; Sparks, D.L., Ed.; Academic Press: Cambridge, MA, USA, 2013; Volume 120, pp. 275–379.
89. Rosnitschek-Schimmel, I. Seasonal Dynamics of Nitrogenous Compounds in a Nitrophilic Weed I. Changes in Inorganic and Organic Nitrogen Fractions of the Different Plant Parts of *Urtica dioica*. *Plant Cell Physiol.* **1985**, *26*, 169–176. [\[CrossRef\]](#)
90. Nkoa, R. Agricultural Benefits and Environmental Risks of Soil Fertilization with Anaerobic Digestates: A Review. *Agron. Sustain. Dev.* **2014**, *34*, 473–492. [\[CrossRef\]](#)
91. Urrea, J.; Alkorta, I.; Garbisu, C. Potential Benefits and Risks for Soil Health Derived From the Use of Organic Amendments in Agriculture. *Agronomy* **2019**, *9*, 542. [\[CrossRef\]](#)
92. Jung, C.; Müller, A.E. Flowering Time Control and Applications in Plant Breeding. *Trends Plant Sci.* **2009**, *14*, 563–573. [\[CrossRef\]](#)
93. Dabi, G.G.; Wakjira, Y.T.; Feysa, H.E.; Abebe, W.M. Development and Characterization of Laminated Fiber Reinforced Bio-Composite From Nettle and Poly Lactic Acid Fiber. *J. Indus. Text.* **2022**, *52*, 15280837221118064. [\[CrossRef\]](#)
94. Pérez-Esteban, J.; Escolástico, C.; Ruiz-Fernández, J.; Masaguer, A.; Moliner, A. Bioavailability and Extraction of Heavy Metals from Contaminated Soil by Atriplex Halimus. *Environ. Exp. Bot.* **2013**, *88*, 53–59. [\[CrossRef\]](#)
95. Singh, B.S.M.; Singh, D.; Dhal, N.K. Enhanced Phytoremediation Strategy for Sustainable Management of Heavy Metals and Radionuclides. *Case Stud. Chem. Environ. Eng.* **2022**, *5*, 100176. [\[CrossRef\]](#)
96. García-Sánchez, M.; Šípková, A.; Száková, J.; Kaplan, L.; Ohecová, P.; Tlustoš, P. Applications of Organic and Inorganic Amendments Induce Changes in the Mobility of Mercury and Macro- and Micronutrients of Soils. *Sci. World J.* **2014**, *2014*, 407049. [\[CrossRef\]](#)
97. Jian, S.; Li, J.; Chen, J.; Wang, G.; Mayes, M.A.; Dzantor, K.E.; Hui, D.; Luo, Y. Soil Extracellular Enzyme Activities, Soil Carbon and Nitrogen Storage under Nitrogen Fertilization: A Meta-Analysis. *Soil. Biol. Biochem.* **2016**, *101*, 32–43. [\[CrossRef\]](#)
98. Sekaran, U.; McCoy, C.; Kumar, S.; Subramanian, S. Soil Microbial Community Structure and Enzymatic Activity Responses to Nitrogen Management and Landscape Positions in Switchgrass (*Panicum virgatum* L.). *GCB Bioenergy* **2019**, *11*, 836–851. [\[CrossRef\]](#)
99. Alburquerque, J.A.; de la Fuente, C.; Campoy, M.; Carrasco, L.; Nájera, I.; Baixauli, C.; Caravaca, F.; Roldán, A.; Cegarra, J.; Bernal, M.P. Agricultural Use of Digestate for Horticultural Crop Production and Improvement of Soil Properties. *Eur. J. Agron.* **2012**, *43*, 119–128. [\[CrossRef\]](#)
100. Mortola, N.; Romaniuk, R.; Cosentino, V.; Eiza, M.; Carfagno, P.; Rizzo, P.; Bres, P.; Riera, N.; Roba, M.; Butti, M.; et al. Potential Use of a Poultry Manure Digestate as a Biofertiliser: Evaluation of Soil Properties and *Lactuca sativa* Growth. *Pedosphere* **2019**, *29*, 60–69. [\[CrossRef\]](#)
101. Nabel, M.; Schrey, S.D.; Poorter, H.; Koller, R.; Jablonowski, N.D. Effects of Digestate Fertilization on *Sida* Hermaphrodita: Boosting Biomass Yields on Marginal Soils by Increasing Soil Fertility. *Biomass Bioenergy* **2017**, *107*, 207–213. [\[CrossRef\]](#)
102. Watson, C.; Schlösser, C.; Vögel, J.; Wichern, F. Hydrochar, Digestate, and Process Water Impacts on a Soil's Microbial Community, Processes, and Metal Bioavailability. *Soil. Sci. Soc. Am. J.* **2021**, *85*, 717–731. [\[CrossRef\]](#)
103. Jia, S.; Wang, Z.; Li, X.; Sun, Y.; Zhang, X.; Liang, A. N Fertilization Affects on Soil Respiration, Microbial Biomass and Root Respiration in *Larix gmelinii* and *Fraxinus mandshurica* Plantations in China. *Plant Soil.* **2010**, *333*, 325–336. [\[CrossRef\]](#)
104. Zhang, H.; Liu, Y.; Zhou, Z.; Zhang, Y. Inorganic Nitrogen Addition Affects Soil Respiration and Belowground Organic Carbon Fraction for a *Pinus tabulaeformis* Forest. *Forests* **2019**, *10*, 369. [\[CrossRef\]](#)
105. Wittmann, C.; Kähkönen, M.A.; Ilvesniemi, H.; Kurola, J.; Salkinoja-Salonen, M.S. Areal Activities and Stratification of Hydrolytic Enzymes Involved in the Biochemical Cycles of Carbon, Nitrogen, Sulphur and Phosphorus in Podsolized Boreal Forest Soils. *Soil. Biol. Biochem.* **2004**, *36*, 425–433. [\[CrossRef\]](#)
106. Kracmarova, M.; Kratochvilova, H.; Uhlik, O.; Strejcek, M.; Szakova, J.; Cerny, J.; Tlustos, P.; Balik, J.; Demnerova, K.; Stiborova, H. Response of Soil Microbes and Soil Enzymatic Activity to 20 Years of Fertilization. *Agronomy* **2020**, *10*, 1542. [\[CrossRef\]](#)
107. de Varennes, A.; Abreu, M.M.; Qu, G.; Cunha-Queda, C. Enzymatic Activity of a Mine Soil Varies According to Vegetation Cover and Level of Compost Applied. *Int. J. Phytoremediation* **2010**, *12*, 371–383. [\[CrossRef\]](#) [\[PubMed\]](#)
108. Malabad, A.M.; Zapata-Carbonell, J.; Maurice, N.; Ciadamidaro, L.; Pfendler, S.; Tatin-Froux, F.; Ferrarini, A.; Fornasier, F.; Toussaint, M.-L.; Parelle, J.; et al. Digestate Improved Birch (*Betula pendula*) Growth and Reduced Leaf Trace Element Contents at a Red Gypsum Landfill. *Ecol. Eng.* **2022**, *185*, 106815. [\[CrossRef\]](#)
109. Pardo, T.; Clemente, R.; Bernal, M.P. Effects of Compost, Pig Slurry and Lime on Trace Element Solubility and Toxicity in Two Soils Differently Affected by Mining Activities. *Chemosphere* **2011**, *84*, 642–650. [\[CrossRef\]](#) [\[PubMed\]](#)
110. Pérez-de-Mora, A.; Burgos, P.; Madejón, E.; Cabrera, F.; Jaekel, P.; Schlöter, M. Microbial Community Structure and Function in a Soil Contaminated by Heavy Metals: Effects of Plant Growth and Different Amendments. *Soil. Biol. Biochem.* **2006**, *38*, 327–341. [\[CrossRef\]](#)

111. Agegnehu, G.; Nelson, P.N.; Bird, M.I. Crop Yield, Plant Nutrient Uptake and Soil Physicochemical Properties under Organic Soil Amendments and Nitrogen Fertilization on Nitisols. *Soil. Tillage Res.* **2016**, *160*, 1–13. [\[CrossRef\]](#)
112. Camberato, J.J.; Gagnon, B.; Angers, D.A.; Chantigny, M.H.; Pan, W.L. Pulp and Paper Mill By-Products as Soil Amendments and Plant Nutrient Sources. *Can. J. Soil. Sci.* **2006**, *86*, 641–653. [\[CrossRef\]](#)
113. Wu, J.; Huang, M.; Xiao, H.-A.; Su, Y.-R.; Tong, C.-L.; Huang, D.-Y.; Syers, J.K. Dynamics in Microbial Immobilization and Transformations of Phosphorus in Highly Weathered Subtropical Soil Following Organic Amendments. *Plant Soil.* **2007**, *290*, 333–342. [\[CrossRef\]](#)
114. Heijboer, A.; ten Berge, H.F.M.; de Ruiter, P.C.; Jørgensen, H.B.; Kowalchuk, G.A.; Bloem, J. Plant Biomass, Soil Microbial Community Structure and Nitrogen Cycling under Different Organic Amendment Regimes; a ¹⁵N Tracer-Based Approach. *Appl. Soil. Ecol.* **2016**, *107*, 251–260. [\[CrossRef\]](#)
115. Liao, L.; Feng, Z.; Fu, J.; Liu, X.; Dong, Z.; Dong, T.; Wang, Z. Increasing the Amount of Nitrogen Fertilizer Decreased the Activity of Soil Enzyme in Cv.Huangguogan. *AIP Conf. Proc.* **2017**, *1890*, 040023. [\[CrossRef\]](#)
116. Sawicka, B.; Krochmal-Marczak, B.; Pszczółkowski, P.; Bielińska, E.J.; Wójcikowska-Kapusta, A.; Barbaś, P.; Skiba, D. Effect of Differentiated Nitrogen Fertilization on the Enzymatic Activity of the Soil for Sweet Potato (*Ipomoea batatas* L. [Lam.]) Cultivation. *Agronomy* **2020**, *10*, 1970. [\[CrossRef\]](#)
117. Naz, M.; Dai, Z.; Hussain, S.; Tariq, M.; Danish, S.; Khan, I.U.; Qi, S.; Du, D. The Soil pH and Heavy Metals Revealed Their Impact on Soil Microbial Community. *J. Environ. Manage.* **2022**, *321*, 115770. [\[CrossRef\]](#) [\[PubMed\]](#)
118. Antil, R.S.; Bar-Tal, A.; Fine, P.; Hadas, A. Predicting Nitrogen and Carbon Mineralization of Composted Manure and Sewage Sludge in Soil. *Compost. Sci. Util.* **2011**, *19*, 33–43. [\[CrossRef\]](#)
119. Busby, R.R.; Allen Torbert, H.; Gebhart, D.L. Carbon and Nitrogen Mineralization of Non-Composted and Composted Municipal Solid Waste in Sandy Soils. *Soil. Biol. Biochem.* **2007**, *39*, 1277–1283. [\[CrossRef\]](#)
120. Serra-Wittling, C.; Houot, S.; Barriuso, E. Soil Enzymatic Response to Addition of Municipal Solid-Waste Compost. *Biol. Fertil. Soils* **1995**, *20*, 226–236. [\[CrossRef\]](#)
121. Albiach, R.; Canet, R.; Pomares, F.; Ingelmo, F. Microbial Biomass Content and Enzymatic Activities after the Application of Organic Amendments to a Horticultural Soil. *Bioresour. Technol.* **2000**, *75*, 43–48. [\[CrossRef\]](#)
122. Bolan, N.S.; Kunhikrishnan, A.; Choppala, G.K.; Thangarajan, R.; Chung, J.W. Stabilization of Carbon in Composts and Biochars in Relation to Carbon Sequestration and Soil Fertility. *Sci. Total Environ.* **2012**, *424*, 264–270. [\[CrossRef\]](#) [\[PubMed\]](#)
123. Kome, G.K.; Enang, R.K.; Tabi, F.O.; Yerima, B.P.K. Influence of Clay Minerals on Some Soil Fertility Attributes: A Review. *Open J. Soil. Sci.* **2019**, *9*, 155–188. [\[CrossRef\]](#)
124. Biesiada, A.; Wołoszczak, E.; Sokol-Letowska, A.; Kucharska, A.; Nawirska-Olszańska, A. The Effect of Nitrogen Form and Dose on Yield, Chemical Composition and Antioxidant Activity of Stinging Nettle (*Urtica dioica* L.). *Herba Pol.* **2009**, *55*, 84–93.
125. Besharati, J.; Shirmardi, M.; Meftahizadeh, H.; Dehestani Ardakani, M.; Ghorbanpour, M. Changes in Growth and Quality Performance of Roselle (*Hibiscus sabdariffa* L.) in Response to Soil Amendments with Hydrogel and Compost under Drought Stress. *S. Afr. J. Bot.* **2022**, *145*, 334–347. [\[CrossRef\]](#)
126. Heimler, D.; Romani, A.; Ieri, F. Plant Polyphenol Content, Soil Fertilization and Agricultural Management: A Review. *Eur. Food Res. Technol.* **2017**, *243*, 1107–1115. [\[CrossRef\]](#)
127. Radman, S.; Zutic, I.; Fabek, S.; Zlabur, J.; Benko, B.; Toth, N.; Coga, L. Influence of Nitrogen Fertilization on Chemical Composition of Cultivated Nettle. *Emir. J. Food Agric.* **2015**, *27*, 889–896. [\[CrossRef\]](#)
128. Bustamante, M.Á.; Michelozzi, M.; Barra Caracciolo, A.; Grenni, P.; Verbokkem, J.; Geerdink, P.; Safi, C.; Nogues, I. Effects of Soil Fertilization on Terpenoids and Other Carbon-Based Secondary Metabolites in Rosmarinus Officinalis Plants: A Comparative Study. *Plants* **2020**, *9*, E830. [\[CrossRef\]](#) [\[PubMed\]](#)
129. Glynn, C.; Herms, D.A.; Oriens, C.M.; Hansen, R.C.; Larsson, S. Testing the Growth-Differentiation Balance Hypothesis: Dynamic Responses of Willows to Nutrient Availability. *New Phytol.* **2007**, *176*, 623–634. [\[CrossRef\]](#) [\[PubMed\]](#)
130. Keski-Saari, S.; Julkunen-Tiitto, R. Resource Allocation in Different Parts of Juvenile Mountain Birch Plants: Effect of Nitrogen Supply on Seedling Phenolics and Growth. *Physiol. Plant.* **2003**, *118*, 114–126. [\[CrossRef\]](#)
131. Radušienė, J.; Marksa, M.; Ivanauskas, L.; Jakštas, V.; Čališkan, Ö.; Kurt, D.; Odabaş, M.S.; Çirak, C. Effect of Nitrogen on Herb Production, Secondary Metabolites and Antioxidant Activities of Hypericum Prunatum under Nitrogen Application. *Indus. Crops Prod.* **2019**, *139*, 111519. [\[CrossRef\]](#)
132. Ramakrishna, A.; Ravishankar, G.A. Influence of Abiotic Stress Signals on Secondary Metabolites in Plants. *Plant Signal Behav.* **2011**, *6*, 1720–1731. [\[CrossRef\]](#)
133. Lee, M.E.; Steiman, M.W.; Angelo, S.K.S. Biogas Digestate as a Renewable Fertilizer: Effects of Digestate Application on Crop Growth and Nutrient Composition. *Renew. Agric. Food Syst.* **2021**, *36*, 173–181. [\[CrossRef\]](#)
134. Ibrahim, M.H.; Jaafar, H.Z.E.; Rahmat, A.; Rahman, Z.A. Effects of Nitrogen Fertilization on Synthesis of Primary and Secondary Metabolites in Three Varieties of Kacip Fatimah (*Labisia pumila* Blume). *Int. J. Mol. Sci.* **2011**, *12*, 5238–5254. [\[CrossRef\]](#)
135. Witzell, J.; Shevtsova, A. Nitrogen-Induced Changes in Phenolics of Vaccinium Myrtillus—Implications for Interaction with a Parasitic Fungus. *J. Chem. Ecol.* **2004**, *30*, 1937–1956. [\[CrossRef\]](#)
136. Pinelli, P.; Ieri, F.; Vignolini, P.; Bacci, L.; Baronti, S.; Romani, A. Extraction and HPLC Analysis of Phenolic Compounds in Leaves, Stalks, and Textile Fibers of *Urtica dioica* L. *J. Agric. Food Chem.* **2008**, *56*, 9127–9132. [\[CrossRef\]](#)

137. Robbins, R.J. Phenolic Acids in Foods: An Overview of Analytical Methodology. *J. Agric. Food Chem.* **2003**, *51*, 2866–2887. [[CrossRef](#)] [[PubMed](#)]
138. Deng, B.; Li, Y.; Lei, G.; Liu, G. Effects of Nitrogen Availability on Mineral Nutrient Balance and Flavonoid Accumulation in *Cyclocarya paliurus*. *Plant Physiol. Biochem.* **2019**, *135*, 111–118. [[CrossRef](#)]
139. Li, Z.; Jiang, H.; Yan, H.; Jiang, X.; Ma, Y.; Qin, Y. Carbon and Nitrogen Metabolism under Nitrogen Variation Affects Flavonoid Accumulation in the Leaves of *Coreopsis tinctoria*. *PeerJ* **2021**, *9*, e12152. [[CrossRef](#)] [[PubMed](#)]
140. Maillard, F.; Girardclos, O.; Assad, M.; Zappelini, C.; Pérez Mena, J.M.; Yung, L.; Guyeux, C.; Chrétien, S.; Bigam, G.; Cosio, C.; et al. Dendrochemical Assessment of Mercury Releases from a Pond and Dredged-Sediment Landfill Impacted by a Chlor-Alkali Plant. *Environ. Res.* **2016**, *148*, 122–126. [[CrossRef](#)]
141. Zappelini, C.; Karimi, B.; Foulon, J.; Lacercat-Didier, L.; Maillard, F.; Valot, B.; Blaudez, D.; Cazaux, D.; Gilbert, D.; Yergeau, E.; et al. Diversity and Complexity of Microbial Communities from a Chlor-Alkali Tailings Dump. *Soil. Biol. Biochem.* **2015**, *90*, 101–110. [[CrossRef](#)]
142. Markwell, J.; Osterman, J.C.; Mitchell, J.L. Calibration of the Minolta SPAD-502 Leaf Chlorophyll Meter. *Photosynth. Res.* **1995**, *46*, 467–472. [[CrossRef](#)]
143. Maxwell, K.; Johnson, G.N. Chlorophyll Fluorescence—A Practical Guide. *J. Exp. Bot.* **2000**, *51*, 659–668. [[CrossRef](#)]
144. Schneider, C.A.; Rasband, W.S.; Eliceiri, K.W. NIH Image to ImageJ: 25 Years of Image Analysis. *Nat. Methods* **2012**, *9*, 671–675. [[CrossRef](#)]
145. Ferrarini, A.; Martani, E.; Fornasier, F.; Amaducci, S. High C Input by Perennial Energy Crops Boosts Belowground Functioning and Increases Soil Organic P Content. *Agric. Ecosyst. Environ.* **2021**, *308*, 107247. [[CrossRef](#)]
146. Fornasier, F.; Ascher, J.; Ceccherini, M.T.; Tomat, E.; Pietramellara, G. A Simplified Rapid, Low-Cost and Versatile DNA-Based Assessment of Soil Microbial Biomass. *Ecol. Indic.* **2014**, *45*, 75–82. [[CrossRef](#)]
147. R Core Team R: A Language and Environment for Statistical Computing; R Foundation for Statistical Computing: Vienna, Austria, 2020.
148. Lê, S.; Josse, J.; Husson, F. FactoMineR: An R package for multivariate analysis. *J. Stat. Softw.* **2008**, *25*. [[CrossRef](#)]
149. Oksanen, J.; Blanchet, F.G.; Friendly, M.; Kindt, R.; Legendre, P.; McGlinn, D.; Minchin, P.R.; O'Hara, R.B.; Simpson, G.L.; Solymos, P.; et al. Vegan Community Ecology Package. 2020, R Package Version 2.5-7. Available online: https://www.researchgate.net/publication/346579465_vegan_community_ecology_package_version_25-7_November_2020 (accessed on 1 July 2024).
150. de Mendiburu, F. Agricolae: Statistical Procedures for Agricultural Research. 2019, R Package Version 1.3-1. Available online: <https://CRAN.R-project.org/package=agricolae> (accessed on 1 July 2024).

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.