

Gas chromatography (GC) fingerprinting and immunomodulatory activity of polysaccharide from the rhizome of *Menispermum dauricum* DC

Pei Yang^{1,*}, Yang Zhai^{2,*}, Yan Ma¹, Beibei Mao¹, Fengshan Wang³, Li Li⁴, Lijuan Luan² and Yuhong Liu¹

¹ School of Pharmaceutical Sciences, Shandong University of Traditional Chinese Medicine, Jinan, China

² Shandong Cancer Hospital and Institute, Shandong First Medical University and Shandong Academy of Medical Sciences, Jinan, China

³ School of Pharmaceutical Sciences, Shandong University, NMPA Key Laboratory for Quality Research and Evaluation of Carbohydrate-based Medicine, National Glycoengineering Research Center, Jinan, China

⁴ Sishui Siheyuan Culture and Tourism Development Company, Ltd, Sishui, China

* These authors contributed equally to this work.

ABSTRACT

This research aimed to establish the gas chromatography (GC) fingerprints and examine the immunomodulatory activity of the rhizome of *Menispermum dauricum* polysaccharides. In this study, the preparation conditions were optimized by the response surface method (RSM). GC is an effective and sensitive technique employed to measure the composition of monosaccharides; the GC fingerprints of total polysaccharides from 10 batches of the rhizome of *M. dauricum* (tMDP) were established, and chemometrics methods were adopted to examine the differences and similarities of tMDP from distinct regions. The similarity evaluation illustrated that the polysaccharides derived from the rhizome of *M. dauricum* from different origins were highly similar. The results of principal components analysis (PCA) illustrated that all the tMDPs may be integrated into one group within the 95% confidence interval, but the rhizome of *M. dauricum* from different origins could also be distinguished in the plot of PCA scores. Then, the major bioactive fraction MDP was purified and obtained by column chromatography. Our previous study showed that MDP exhibited significant immunomodulatory activity, but the mechanism of the *in vitro* immunomodulatory activity of MDP is unclear. The macrophage activation induced by MDP was abolished when Toll-like receptor 4 (TLR4) signaling was knocked down by the TLR4 inhibitor. Furthermore, western blot analysis illustrated that MDP activated RAW264.7 cells through MAPKs and NFκB pathways induced by TLR4. This research offers a theoretical foundation for quality control and additional study as a potential immunomodulator of MDP.

Subjects Biochemistry, Cell Biology, Molecular Biology, Immunology

Keywords *Menispermum dauricum* DC, GC fingerprinting, Immunomodulatory activity, TLR4-MAPKs/NFκB

Submitted 4 July 2022

Accepted 3 August 2022

Published 22 August 2022

Corresponding authors

Lijuan Luan, 15553115573@163.com

Yuhong Liu, yhlui@sducm.edu.cn

Academic editor

Héctor Mora-Montes

Additional Information and
Declarations can be found on
page 18

DOI 10.7717/peerj.13946

© Copyright

2022 Yang et al.

Distributed under

Creative Commons CC-BY 4.0

OPEN ACCESS

INTRODUCTION

The rhizome of *Menispermum dauricum* DC (Menispermaceae), referred to as Bei Dou Gen in Chinese, which is mainly produced in northeastern, northern, and eastern China, is a traditional Chinese medicine herb that has had widespread application in modern medicine to treat diarrhea, sore throats, colitis, and rheumatic arthralgia, as well as other conditions (Wu & Jin, 2007). It contains various chemical components such as alkaloids, polysaccharides, quinones, cardiac glycosides, lactones, saponins, tannins, proteins, and resins (Wu et al., 2018; Zhou et al., 2018). The *M. dauricum* rhizome's main chemical components are alkaloids. These alkaloids exert a variety of biological properties, including those that influence the cardiovascular system, antitumor activity, and antiarrhythmic function. Additionally, the injection of total alkaloids has been used therapeutically for a considerable period for the treatment of chronic tracheitis, as well as arthralgia and throat sores (Chen et al., 2012).

The polysaccharides are also significant bioactive constituents of the rhizome of *M. dauricum*, which exhibited numerous pharmacological properties while having no toxic effects (Li et al., 2006; Liang et al., 2005; Lin et al., 2013a, 2013b). In most cases, the quality control of polysaccharides is primarily dependent on the phenol sulfuric acid technique, which involves establishing the content of sugar present (Zhang et al., 2021).

The phenol sulfuric acid approach, on the other hand, is incapable of providing any structural properties of polysaccharides because of its low level of specificity. Additionally, research suggested that adulterated or low-quality products cannot be identified in an efficient manner (Li et al., 2019). Moreover, the biological properties of polysaccharides are based on their numerous morphological characteristics, such as the composition of their monosaccharides, the kind of glycosidic bonds, and the distribution of their molecular weight (Li et al., 2019). In particular, the examination of the polysaccharide's monosaccharide composition is the first and initial phase in the analysis of polysaccharides, which needs to be taken into consideration to set up a quality control system for polysaccharides and products associated with them.

Recently, the use of fingerprint profiling as a technology and methodology for quality control of compounds generated from plants has gained widespread recognition as a practical and effective technique (Li et al., 2019). In addition, chemometrics is a robust tool for analyzing fingerprints premised on chemical data, such as principal component analysis (PCA) and similarity analysis (SA), thus contributing to the analysis of the raw data gathered from fingerprints (Dong et al., 2020; Liu et al., 2015a). Fingerprinting methods have been utilized effectively to control quality as well as standardize plant polysaccharides, including *Panax*, *Cordyceps*, *Ganoderma*, tea, and *Lycium barbarum* polysaccharides (Sun et al., 2014).

Polysaccharides derived from natural sources have garnered a lot of interest in recent years because they do not have any toxic effects in studies to date and have potent immunomodulatory properties (Pan et al., 2021; Zhong et al., 2021; Zhou et al., 2021). As a result, there is a remarkable demand for the extraction and purification of new immunomodulatory polysaccharides derived from natural products that are both safe and

Table 1 The source, neutral sugar content and principal component analysis composite score of the total polysaccharides from the rhizome of *M. dauricum*.

No.	Source	Neutral sugar content (%)	Principal component analysis composite score
S1	Shandong	20.34	-1.777427
S2	Shandong	19.54	0.0950919
S3	Heilongjiang	20.32	0.8092878
S4	Shandong	20.77	-1.506477
S5	Heilongjiang	19.47	-0.156665
S6	Liaoning	22.77	0.6893036
S7	Jilin	20.12	0.5907167
S8	Shandong	21.91	0.1607214
S9	Liaoning	22.61	-1.386776
S10	Jilin	20.86	2.4822341

efficacious. Macrophages, which are powerful phagocytic cells, are present in nearly all of the body's tissues. Macrophage activation is an integral process in both adaptive and innate immune responses, which may trigger and propagate defense responses against infections (Ma et al., 2019; Wang et al., 2017b). Some botanical polysaccharides could be recognized and combined with specific receptors on macrophages to enhance the viability of macrophages against pathogenic microorganisms and tumorigenesis by the promotion of phagocytic and the cytokines of nitric oxide (NO), interleukin (IL)-6 and tumor necrosis factor (TNF)- α (Yu et al., 2012). The polysaccharides with immunomodulatory action that are derived from plant products have no reports of allergies or adverse effects and have been believed to be potential candidates for use as immunoregulators (Zhao et al., 2020).

In this research, the fingerprint of the rhizome of *M. dauricum* polysaccharide was established and analyzed using gas chromatography (GC) and chemometrics, which offered a technique that was both reliable and effective for controlling the quality of the rhizome of *M. dauricum* and provided the groundwork for the future expansion of its functional products. Then, the major bioactive fraction MDP was obtained by column chromatography. Moreover, the *in vitro* immunomodulatory activities and the mechanism of MDP on macrophages were also evaluated.

MATERIALS AND METHODS

Materials

The rhizome of *M. dauricum* plants for each of the 10 batches, denoted by the letters S1 to S10, was gathered from various regions throughout China (Table 1). The voucher samples of rhizome of *M. dauricum* were deposited in the School of Pharmaceutical Sciences of Shandong University of Traditional Chinese Medicine, Jinan, China.

Chemicals: The DEAE-cellulose-52 was procured from Yuanye Biological Co., Ltd. (Shanghai, China). Sephadex G-50 and Sephacryl S-100 were procured from GE Healthcare Life Sciences (Piscataway, NJ, USA). Guoyao Group Co., Ltd. (Beijing, China) provided glucose and galactose. Arabinose, fucose, mannose, and rhamnose were all

supplied by Macklin Biochemical Technology Co., Ltd. (Shanghai, China). Lipopolysaccharide (LPS) was procured from Sigma-Aldrich (St. Louis, MO, USA). The ELISA kits employed in the IL-6, NO, TNF- α , and IgM tests were retrieved from Enzyme-linked Biotechnology Co., Ltd. (Shanghai, China). Antibodies against MyD88, NF κ B, JNK, ERK, and P38, as well as GAPDH, were procured from ABclonal (Wuhan, China) while other antibodies were procured from Cell Signaling Technology (Beverly, MA, USA).

Experimental design of RSM

The independent parameters chosen were the liquid-solid ratio (A, mL/g), extraction time (B, min) and extraction temperature (C, °C) whereas the dependent parameter was the polysaccharide extraction yield, premised on the findings from the single-factor experiments. The computation for the extraction yield was performed based on the formula below:

$$\text{Extraction yield (\%)} = [\text{crude polysaccharide weight (g)}/\text{rhizome of } M. \text{ dauricum} \text{ weight (g)}] \times 100\%.$$

A three-factor three-level response surface optimization experiment and prediction model were constructed with the aid of the Design-Expert software (version: 8.0.5).

GC fingerprint analysis

Preparation of polysaccharide extracts

Distilled water was used in the process of extracting crude polysaccharides from the rhizome of *M. dauricum* under the optimal extraction parameters, which were determined through response surface optimization. Following the completion of three rounds of extraction, the aqueous extract was subjected to a vacuum and concentrated. After that, four times the volume of ethanol was introduced into the solution to precipitate the polysaccharide, and afterward, the mixture was left to stand at 4 °C for an entire night. The precipitate was collected and deproteinized by means of the Trichloroacetic acid (TCA)-n-butanol method and then subjected to freeze-drying to produce a total polysaccharide fraction (tMDP) (Yang et al., 2022).

Complete acid hydrolysis

Hydrolysis of 10 mg of tMDP was performed using 3 mL of 2 mol/L trifluoroacetic acid (TFA) in a vial that was sealed at 110 °C for 6 h. To get rid of the TFA residue, the mixture was combined with 1 mL of methanol, and then it was rotary-evaporated thrice under a vacuum until it was completely dry. The hydrolyzed polysaccharide samples were prepared for derivatization (Liu et al., 2015b).

Preparation of derivatization and GC analysis

The GC method was employed to measure the composition of the monosaccharides. The hydrolyzed tMDP and monosaccharide standards were added with hydroxylamine hydrochloride, pyridine, and inositol hexaacetate, and stirred at 90 °C for 30 min. Then,

acetic anhydride was introduced and the reaction continued for 30 min at 90 °C for acetylation. A GC examination was performed on the products of the reaction. The temperature program was set to 170 °C for 3 min, 170 °C to 178 °C at a rate of 0.5 °C/min for 3 min, and elevated to 210 °C for 5 min at a rate of 2 °C/min (Yang et al., 2022).

Extraction and purification of polysaccharides

tMDP was extracted from the S10 batch (highest PCA score) of the rhizome of *M. dauricum* by the method above, and the TCA-n-butanol method was used to remove protein. Subsequently, the purification of tMDP was done utilizing the Sephadex G-50 column (1.75 cm × 66 cm), Sephacryl S-100 column (1.75 cm × 66 cm), and DEAE-52 cellulose column (5.5 cm × 30 cm), and A white purified polysaccharide (MDP) was obtained by collecting the major fraction and lyophilizing it (Yang et al., 2022).

Immunomodulatory activity *in vitro*

Cell culture

Beina Chuanglian Biotechnology Co., Ltd. (Beijing, China) provided the RAW264.7 cells utilized in this study. At a temperature of 37 °C, the cells were incubated in DMEM that had been supplemented with streptomycin sulfate (100 g/ml), penicillin (100 units/mL), and 10% fetal bovine serum. The environment of the incubator was humidified and contained 5% carbon dioxide.

Cell viability test

By conducting the Methyl thiazolyl tetrazolium (MTT) test *in vitro*, we examined the effect that varying concentrations of MDP had on the RAW264.7 cells' viability. In general, 96-well microplates were seeded with RAW264.7 cells at a density of 1×10^4 cells per well. After the cells had been cultured at 37 °C for 24 h with 5% carbon dioxide, they underwent a second round of incubation at 37 °C for another 24 h with MDP samples at increasing dosages (0, 10, 50, 100, 200, and 400 µg/mL) or LPS (1 µg/mL, positive control). Following the initial incubation, 20 µL of the MTT solution with a concentration of 5 mg/mL was introduced into each well, and the plates were subsequently re-incubated at 37 °C for a further 4 h in the medium. After that, the medium was carefully aspirated, and each well was then treated using 150 µL of DMSO so that the dissolving crystals could be dissolved. At last, to ascertain the absorbance at 570 nm, we made use of a microplate reader (BioTek Instruments Inc., Winooski, VT, USA) (Wang et al., 2017a).

$$\text{Cell viability \%} = (\text{OD treatment group})/(\text{OD control group}) \times 100\%.$$

TLR4 pathway blocking experiment

RAW264.7 cells were seeded at a density of 1×10^4 cells/well into 96-well microplates for 24 h. Following the adhesion of the cells, various doses of MDP (50, 100, and 200 µg/mL), as well as LPS, were introduced, and TAK-242 was added at the same time to make the final concentration of 20 µg/mL. Next, the concentrations of IL-6, TNF-α, and NO in the supernatant were detected with the aid of ELISA kits (ML BIO Biotechnology, Shanghai, China) that are commercially available. The assays were carried out in compliance with the

guidelines that were supplied by the manufacturer, after which the cytokine levels were calculated using the standard curves.

Western blotting analysis

After cultivating the RAW 264.7 cells for 24 h on a tissue culture plate with six wells having a flat bottom at 5×10^6 cells per well, the MDP was added at a final concentration of 50 $\mu\text{g/mL}$ in order to activate these cells for 24 h. The positive control consisted of the cells that had been treated using LPS at a concentration of 1 $\mu\text{g/mL}$. At the end of the incubation period, all of the conditioned cells were harvested and processed to produce the total proteins. Briefly, samples were prepared by RIPA (radioimmune precipitation assay) with protease and phosphatase inhibitors. The BCA technique was used to determine the protein. After denaturation, 30 μg of protein per well was deposited *via* electrophoresis onto SDS-PAGE and then transferred onto PVDF membranes. 10% nonfat milk (prepared in TBS supplemented with 0.1% Tween 20) was employed in blocking the membranes at ambient temperature for 4 h. Afterward, the membranes were probed with primary antibodies against P38 (1:750), p-P38 (1:1,000), ERK (1:1,500), p-ERK (1:1,500), JNK (1:1,500), p-JNK (1:2,000), NF κ B (1:1,500), p-NF κ B (1:1,000), MyD88 (1:1,500), TLR4 (1:1,000), as per the guidelines stipulated by the manufacturer. After being rinsed in TBST, the membranes were subjected to incubation with the secondary antibody (1:5,000) for 1 h at ambient temperature. ECL chemiluminescence detection kit (Millipore, MA, USA) was utilized to analyze the signals, which were subsequently quantified with an Amersham Imager 600 Chemiluminescence imaging system (GE, Boston, MA, USA).

RESULTS

Extraction conditions optimization

To maximize the extraction rate of polysaccharides and herb utilization, and to lay the foundation for the subsequent fingerprinting study, RSM was adopted to generate optimal conditions for extracting the rhizome of *M. dauricum* polysaccharides. Premised on the findings obtained from the single-factor experimental analysis [Figs. 1A–1C](#), a 3-factor 3-level response surface experiment was carried out for optimizing the extraction conditions of polysaccharides. To perform the variance analysis, we utilized the Design-Expert software ([Table 2](#)) to identify an optimal region for the study by fitting the mathematical model with the experimental data.

To show the type of interaction between variables, 2D contour and 3D response surface plots are presented in [Figs. 1D–1I](#). According to the plots, the optimum extraction conditions for polysaccharides were a ratio of liquid to the raw material of 19.79 mL/g, an extraction duration of 116.47 min, and a temperature for extraction of 86.86 °C. The maximum anticipated polysaccharides yield, under the optimal conditions, was 1.781%. However, for the sake of the operability and convenience of production, there could be modifications to the experimental parameters: the time for extraction was 116 min, the ratio of liquid to the raw material was 20 mL/g, and the temperature used for extraction was 87 °C.

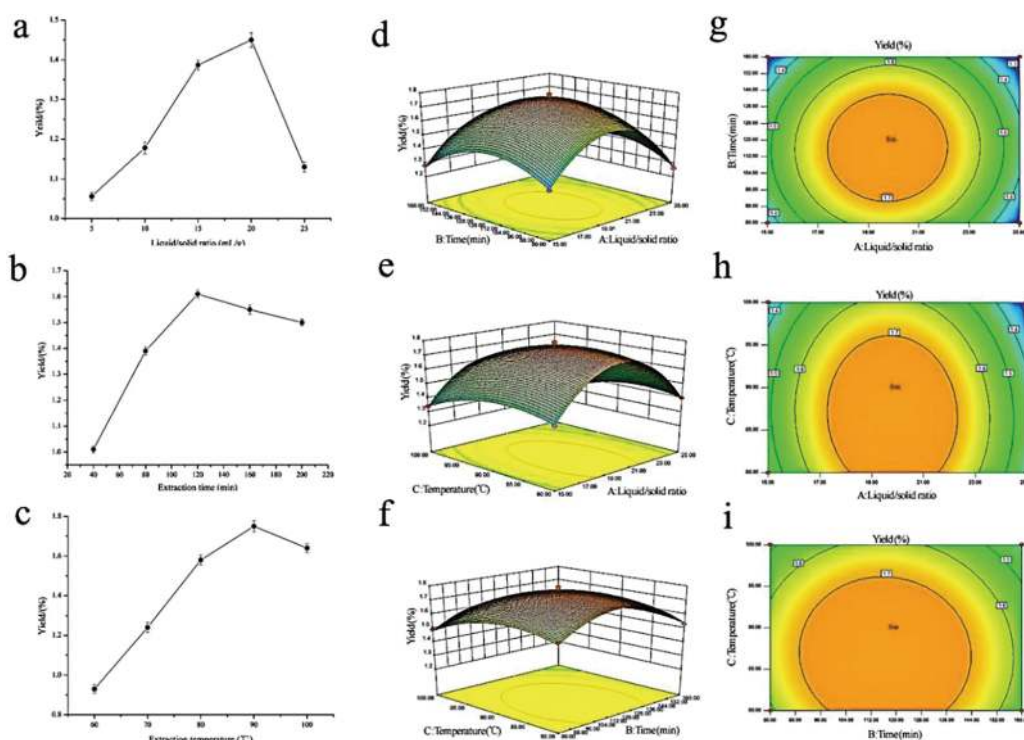


Figure 1 Influences of extraction variables on the polysaccharides yield from the rhizome of *M. dauricum*; response surface plots and contour plots. (A) Effect exerted by the liquid/solid ratio (mL/g) on polysaccharides yield from the rhizome of *M. dauricum*; (B) Effect exerted by the extraction time (min) on polysaccharides yield from the rhizome of *M. dauricum*; (C) Effect exerted by the extraction temperature (°C) on polysaccharides yield from the rhizome of *M. dauricum*; (D) plots of response surface presenting the interactive effects of liquid/solid ratio (a) and extraction time (b); (E) response surface plots presenting the interactive effects of liquid/solid ratio (a) and extraction temperature (c); (F) response surface plots presenting the interactive effects of extraction duration (b) and extraction temperature (c); (G) contour plots presenting the interactive effects of A and B; (H) contour plots presenting the effects of the interaction of a and c; (I) contour plots presenting the interactive effects of b and c. The values are presented as the mean $\pm$ SD, $n = 3$.

Full-size [DOI: 10.7717/peerj.13946/fig-1](https://doi.org/10.7717/peerj.13946/fig-1)

GC fingerprints analysis

GC fingerprints of tMDP

tMDP was prepared by the optimized extraction conditions of RSM and deproteinization using the TCA-n-butanol method, which was used to establish GC fingerprinting, thus laying the foundation for controlling the quality of the rhizome of *M. dauricum*.

To ascertain the structural characteristics of polysaccharides, it is required and essential to take into account the monosaccharide composition. Compositions of monosaccharides and polysaccharide ratios varied remarkably by the source, which was an essential aspect of the polysaccharide's biological activity. In the quality control of active polysaccharides throughout the last several years, the monosaccharide composition-associated fingerprints have seen a widespread application. To determine the standard GC fingerprint, GC was utilized to conduct monosaccharide composition analysis on 10 different batches of tMDP. As shown in Fig. 2A, GC fingerprinting of full acid hydrolysates of 10 different batches of

Table 2 Analysis of variance in the regression model.

Source	Square sum	df	Mean square	F value	P value	Significance
Model	0.69	9	0.077	357.11	<0.0001	**
A-A	0.0072	1	0.0072	33.60	0.0007	**
B-B	0.00845	1	0.00845	39.43	0.0004	**
C-C	0.029	1	0.029	134.4	<0.0001	**
AB	0.000225	1	0.000225	1.05	0.3396	
AC	0.000625	1	0.000625	2.92	0.1314	
BC	0.000225	1	0.000225	1.05	0.3396	
A ²	0.43	1	0.43	1,996.38	<0.0001	**
B ²	0.13	1	0.13	593.19	<0.0001	**
C ²	0.037	1	0.037	172.7	<0.0001	**
Residual	0.00150	7	0.0002143			
Lack of fit	0.0009	3	0.0003	2.0	0.2564	
Pure error	0.0006	4	0.00015			
Cor total	0.69	16				

Note:

** $P < 0.01$ significant different.

tMDP exhibited a high degree of similarity having 11 common peaks. Figure. 2B shows the fingerprint that was used as a reference. The chromatogram of mixed standard monosaccharides depicted in Fig. 2C revealed the presence of seven different components, with peaks 1, 2, 3, 8, 9, 10, and 11 representing rhamnose, arabinose, fucose, mannose, glucose, galactose, and inositol (internal standard), respectively. To sum up the above, tMDP contained six monosaccharides, among which rhamnose, glucose, and galactose were the majority, whereas arabinose, fucose and mannose, were of low content.

The similarity analysis of the GC fingerprints

Based on the professional program “Similarity Evaluation System for Chromatographic Fingerprint of Traditional Chinese Medicine (TCM)” (Version 2012A), similarity values were computed premised on the GC chromatograms of polysaccharide samples as well as the reference fingerprint. Relative to the reference fingerprint, the similarity across 10 batches of tMDP ranged from 0.943% to 0.998 (Table 3). The great similarity revealed that the previously developed GC fingerprint was particularly suited as one of the multi-dimensional fingerprints for quality control of the rhizomes of *M. dauricum*.

PCA of the GC fingerprints

The GC fingerprints of the rhizome of *M. dauricum* samples were then presented quantitatively by utilizing PCA. The relative peak area of seven distinctive chromatographic peaks (peaks 1, 2, 3, 4, 8, 9, and 10) in Fig. 2 was chosen in PCA. As depicted by the scree plot (Fig. 3A), the first three PCs had eigenvalues of 4.302, 1.088, and 1.015, explaining 61.45%, 15.55%, and 14.50% of variance correspondingly,

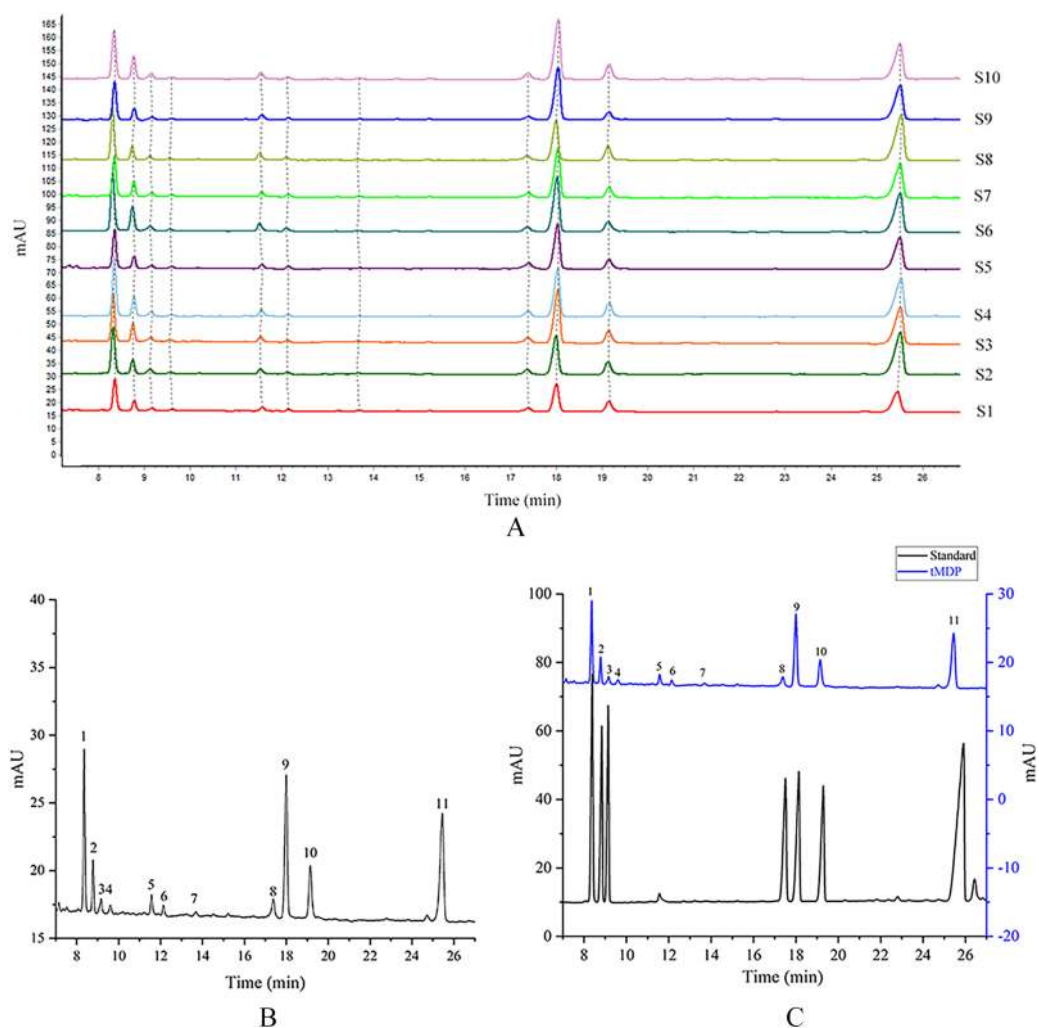


Figure 2 GC chromatograms of 10 different batches of tMDP's complete acid hydrolysates (A), the reference GC fingerprint of tMDP (B), Monosaccharide standards, and tMDP sample (1, Rhamnose; 2, Arabinose; 3, Fucose; 8, Mannose; 9, Glucose; 10, Galactose; 11, Internal standard) (C).

Full-size [DOI: 10.7717/peerj.13946/fig-2](https://doi.org/10.7717/peerj.13946/fig-2)

representing 91.50% of the total variance. The following equations can be used to explain the principal components:

$$PC1 = 0.436 * X1 + 0.437 * X2 + 0.451 * X3 + 0.255 * X4 + 0.325 * X8 + 0.387 * X9 + 0.308 * X10$$

$$PC2 = -0.318 * X1 - 0.156 * X2 - 0.181 * X3 - 0.735 * X4 + 0.481 * X8 + 0.061 * X9 - 0.256 * X10$$

$$PC3 = 0.114 * X1 - 0.237 * X2 - 0.180 * X3 - 0.006 * X4 + 0.362 * X8 - 0.520 * X9 + 0.704 * X10$$

The PCA results were further analyzed by OriginPro 2021. As per the 3D scores plot of PCA depicted in Fig. 3B, all of the rhizomes of *M. dauricum* samples were integrated into a group within the 95% confidence ellipse, which indicated that the quality of the rhizome of *M. dauricum* from Northeast and Shandong of China was similar, which was consistent

Table 3 Similarity analysis of 10 batches of tMDP.

No.	S1	S2	S3	S4	S5	S6	S7	S8	S9	S10	R
S1	1.000	0.984	0.990	0.997	0.990	0.990	0.989	0.973	0.977	0.988	0.995
S2	0.984	1.000	0.977	0.988	0.977	0.976	0.975	0.998	0.964	0.958	0.987
S3	0.990	0.977	1.000	0.991	0.998	0.993	0.998	0.968	0.996	0.994	0.998
S4	0.997	0.988	0.991	1.000	0.990	0.995	0.990	0.977	0.981	0.987	0.997
S5	0.990	0.977	0.998	0.990	1.000	0.990	0.998	0.970	0.993	0.991	0.997
S6	0.990	0.976	0.993	0.995	0.990	1.000	0.992	0.963	0.988	0.993	0.996
S7	0.989	0.975	0.998	0.990	0.998	0.992	1.000	0.967	0.997	0.994	0.997
S8	0.973	0.998	0.968	0.977	0.970	0.963	0.967	1.000	0.955	0.943	0.979
S9	0.977	0.964	0.996	0.981	0.993	0.988	0.997	0.955	1.000	0.990	0.992
S10	0.988	0.958	0.994	0.987	0.991	0.993	0.994	0.943	0.990	1.000	0.991
R	0.995	0.987	0.998	0.997	0.997	0.996	0.997	0.979	0.992	0.991	1.000

with the determination of polysaccharide content (Table 1). The high similarity suggested that the GC fingerprint was very suitable as a quality control indicator for the rhizome of *M. dauricum*. However, the rhizome of *M. dauricum* polysaccharides from different origins also differed slightly and could be clustered into three categories broadly, among which S1, S2, S4, and S8 were all produced in Shandong, S3, S5, S7, and S10 were from Heilongjiang and Jilin, while S6 and S9 were from Liaoning.

Immunomodulatory activity and mechanism of MDP

Effect of MDP on the morphology of RAW264.7 cells

tMDP was further purified by cellulose and gel columns to obtain a homogeneous polysaccharide fraction MDP. To examine the impact of MDP on RAW264.7 cells' morphology, the morphologic changes of RAW264.7 cells were observed under a microscope. As could be observed in Fig. 4A, the cells that belonged to the control group had a round shape and normal structure. Nonetheless, following MDP treatment, there was a remarkable alteration in the structure of the cells. It is quite clear to observe that many of the antennas generated surrounding the cells may enhance the area of contact with exterior substances and were beneficial to the phagocytic uptake. This suggests that concentrations of MDP ranging from 10 to 400 µg/mL are capable of activating macrophages without causing cytotoxic effects.

Effect of MDP on RAW264.7 cell viability

Macrophages are the primary cells implicated in the innate defense mechanism of the immune system. In addition, they are the major cells responsible for inflammation and other immunological processes that occur inside the host. It has been shown that many different polysaccharides stimulate both the activity of cells as well as their proliferation (Wang et al., 2018). As a result, we studied how MDP affected the viability of RAW264.7 cells, and the findings are depicted in Fig. 4B. At concentrations of 10, 50, 100, 200, and 400 µg/mL, the levels of cell viability for the various groups were 120.66%, 123.64%,

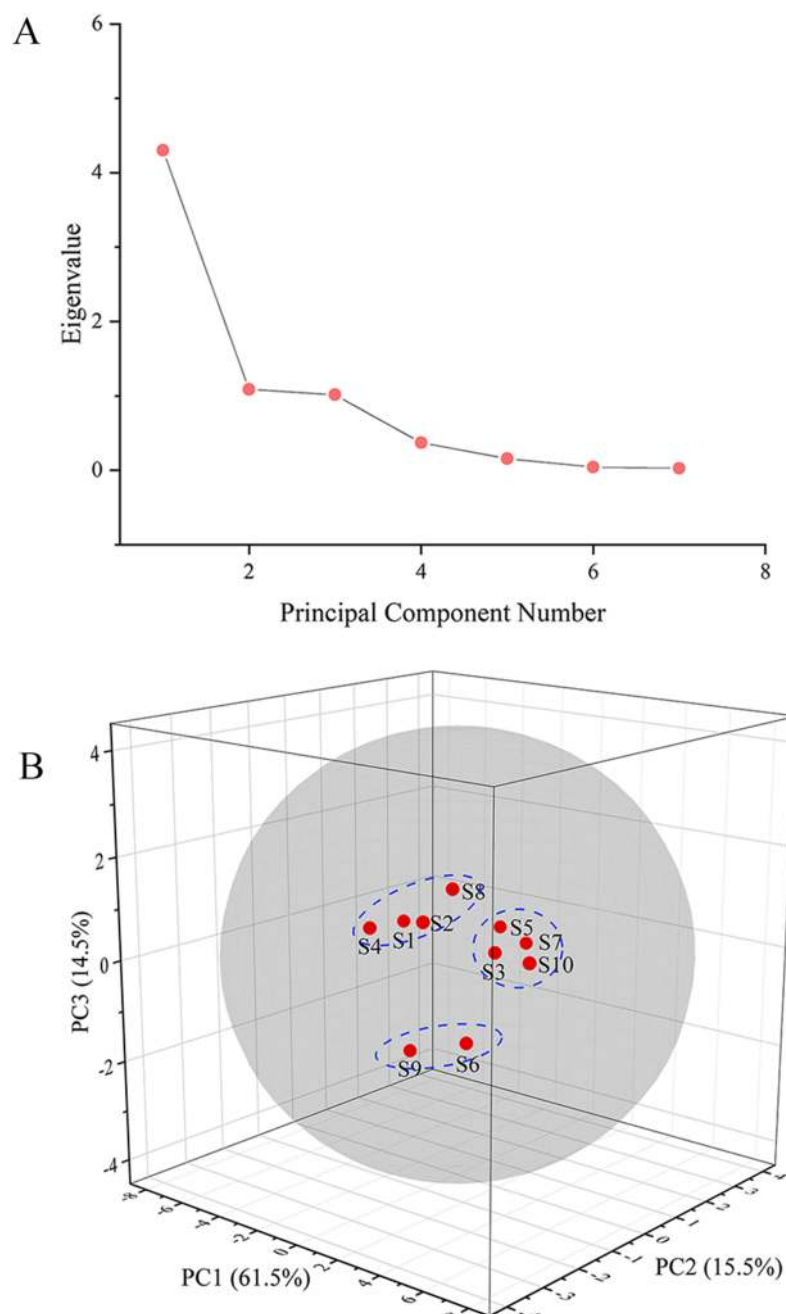


Figure 3 The scree plot (A) and plot of PCA scores (B) of tMDP' GC fingerprints from different regions. [Full-size !\[\]\(e4c51d9db35ee9651ed60d72acdb782c_img.jpg\) DOI: 10.7717/peerj.13946/fig-3](https://doi.org/10.7717/peerj.13946/fig-3)

118.76%, 130.02%, and 114.73%, correspondingly, which illustrate that MDP has the potential to significantly improve RAW264.7 proliferation.

MDP enhanced immunomodulatory effects via TLR4 in vitro

Macrophages are by far the most important body cells that are associated with the immunological defense system (Cheng et al., 2019). After being activated, they are capable of producing a wide variety of chemokines and cytokines. One of the most important

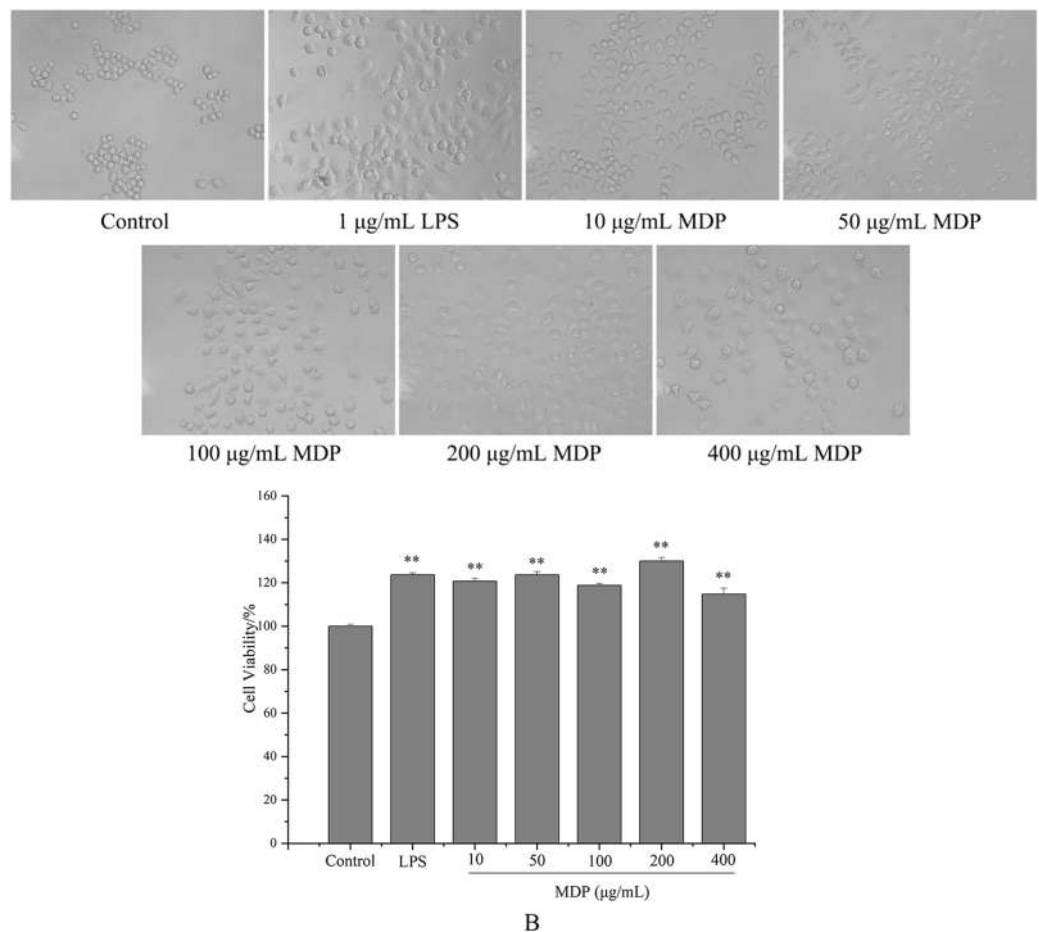


Figure 4 Effect of MDP on the morphology of RAW264.7 cells (A) and the effect of MDP on RAW264.7 cell viability (B). The values are presented as the mean $\pm$ SD, $n = 3$. ** $P < 0.01$ vs. control group. [Full-size !\[\]\(a6fe094a331555a6fd67d72f7f1bf63e_img.jpg\) DOI: 10.7717/peerj.13946/fig-4](https://doi.org/10.7717/peerj.13946/fig-4)

mechanisms that immunoregulators use is to cause the release of several biological components (IL-6, TNF- α , NO, *etc.*) (Tian *et al.*, 2019). NO is a type of vital molecule that is produced by macrophages, and it played an important function in the modulation of apoptosis as well as the host's defense against cancer cells and other pathogens (Ren *et al.*, 2019). In addition, NO may also stimulate phagocytosis and lysis in macrophages, two processes that are essential to the immune system. Because of this, the ability of macrophages to release NO is an indication of the effects that polysaccharides have on the functioning of the immune system (Wang *et al.*, 2017a). Inflammation, cancer, and immunological illnesses may all be traced back to the main active molecules inside organisms, such as TNF- α and IL-6, which perform integral roles in the pathogenesis of these conditions (Wang *et al.*, 2017a). If the host is attacked by external pathogens, activated macrophages produce IL-6 and TNF- α for the purpose of mediating the immune system's response (Sorimachi *et al.*, 1999). Our previous study showed that MDP could promote the production of NO, IL-6 and TNF- α in RAW264.7 cells (Yang *et al.*, 2022). These results suggested that MDP has significant immunomodulatory activities *via* the

mechanism of increasing the release of NO, IL-6, and TNF- α in RAW264.7 cells. However, the mechanism of the *in vitro* immunomodulatory activity of MDP is unclear.

TLRs have been discovered as key membrane receptors that perform an instrumental function in activating macrophages (Fan *et al.*, 2021). Among TLRs, several investigations have demonstrated that TLR4 is implicated in the generation of cytokines in response to polysaccharide stimulation (Liu *et al.*, 2022b). To further investigate whether MDP exerts immune activity through activation of the TLR4 pathway, TAK-242, an inhibitor of TLR4, was utilized so that additional confirmation of the processes that result in MDP-mediated macrophage activation could be achieved. As depicted in Fig. 5, TAK-242 considerably decreased the levels of TNF- α , NO, and IL-6 ($P < 0.05$) as opposed to the MDP treatment group, illustrating that the TLR4 pathway was implicated in the MDP-mediated activation of macrophages.

MDP increased the protein expression of the TLR4-MAPK/NF κ B signaling pathway

By performing a Western blot analysis, we evaluated how MDP affected the protein expression levels of TLR4 to get a deeper insight into its action mechanism. According to the results shown in Fig. 6A, MDP significantly promoted the expression of TLR4 protein, which is consistent with the above experimental results. Once TLR4 is activated, TLR4 is capable of binding with the MyD88 protein to create the TLR4-MyD88 complex, which then stimulates the phosphorylation of the downstream proteins, such as MAPKs (ERK, JNK, and p38) and NF- κ B (Xin *et al.*, 2019). The findings of Western blotting (Fig. 6B) illustrated that the protein expression of MyD88 was remarkably enhanced in RAW264.7 cells following MDP treatment, which indicated that TLR4 protein, after being activated by MDP, combined with a large amount of MyD88 and then activated the downstream pathway.

MAPKs perform crucial functions in immunological and inflammatory responses, including the stimulation of cell proliferation and the regulation of cytokine production *via* the modulation of transcriptional factors (Yang *et al.*, 2019). Phosphorylation is a common mechanism that underlies the functions that are controlled by MAPKs (Yang *et al.*, 2019). To determine if the MAPK signaling pathway was implicated in MDP-related macrophage activation, the phosphorylation levels of the p38, ERK, and JNK MAPK pathways were examined through western blot analysis. As depicted in Figs. 6D–6F, the phosphorylation levels of ERK, JNK, and p38 were substantially elevated in RAW264.7 cells following stimulation with MDP in contrast with the normal control cells. It has been shown that NF κ B is a ubiquitous transcriptional factor that is essential for the activation of macrophages *via* the stimulation of diverse genes implicated in the modulation of immunological as well as inflammatory responses (Yang *et al.*, 2019). As per the findings of Western blot (Fig. 6C), MDP caused an enhancement in the protein expression of p-NF κ B in RAW264.7 cells. In conclusion, following MDP treatment, the key protein of TLR4-MAPK/NF κ B signaling pathways including TLR4, MyD88, p-NF κ B, p-ERK, p-P38, and p-JNK were up-modulated substantially in RAW264.7 cells. As per these findings, it is

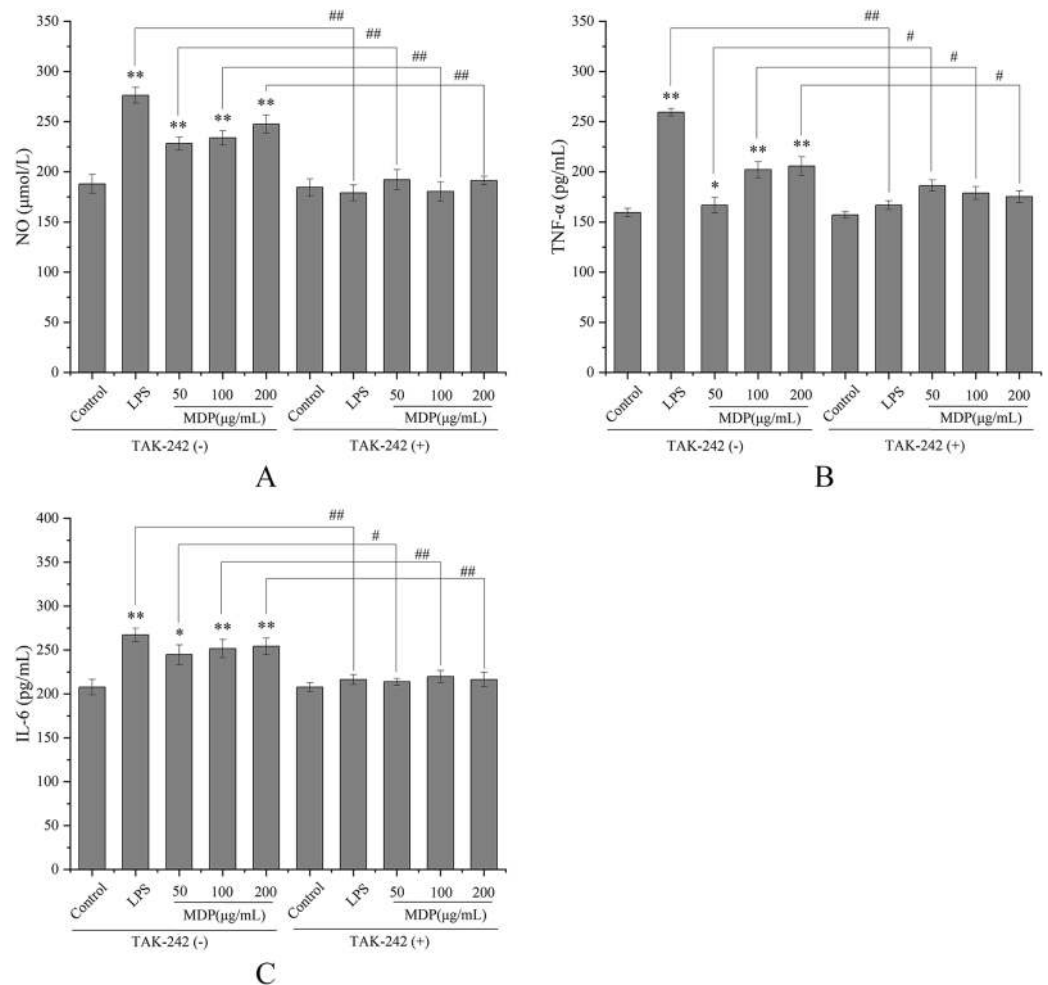


Figure 5 Effects of MDP on the secretion of NO and cytokines with or without inhibitors. (A) NO; (B) TNF- α ; (C) IL-6. Without inhibitors, the secretion of NO and cytokines by macrophages was induced by MDP, however, this impact was hindered when the RAW264.7 cells had been pre-treated with inhibitors. The values are presented as the mean $\pm$ SD, $n = 3$. * $P < 0.05$, ** $P < 0.01$, compared with Control group. # $P < 0.05$, ## $P < 0.01$, compared with the no inhibitor group.

Full-size DOI: [10.7717/peerj.13946/fig-5](https://doi.org/10.7717/peerj.13946/fig-5)

evident that MDP may stimulate the TLR4-MAPK/NF κ B signaling pathways, which in turn exerts immunomodulatory impacts.

DISCUSSION

The detection methods commonly used for monosaccharide composition include high-performance anion-exchange chromatography (HPAEC), gas chromatography (GC), and high-performance liquid chromatography (HPLC) (Liu, 2022a). Among them, the GC is an analytical method that is primarily utilized for characterizing and identifying volatile compounds and is an effective and sensitive technique employed to measure the composition of monosaccharides with the advantages of high sensitivity, good separation, fast analysis, and less sample consumption (Liu, 2022a). Cheong et al. (2016) used GC and HPSEC-RID-MALLS to compare and analyze the monosaccharide composition of three

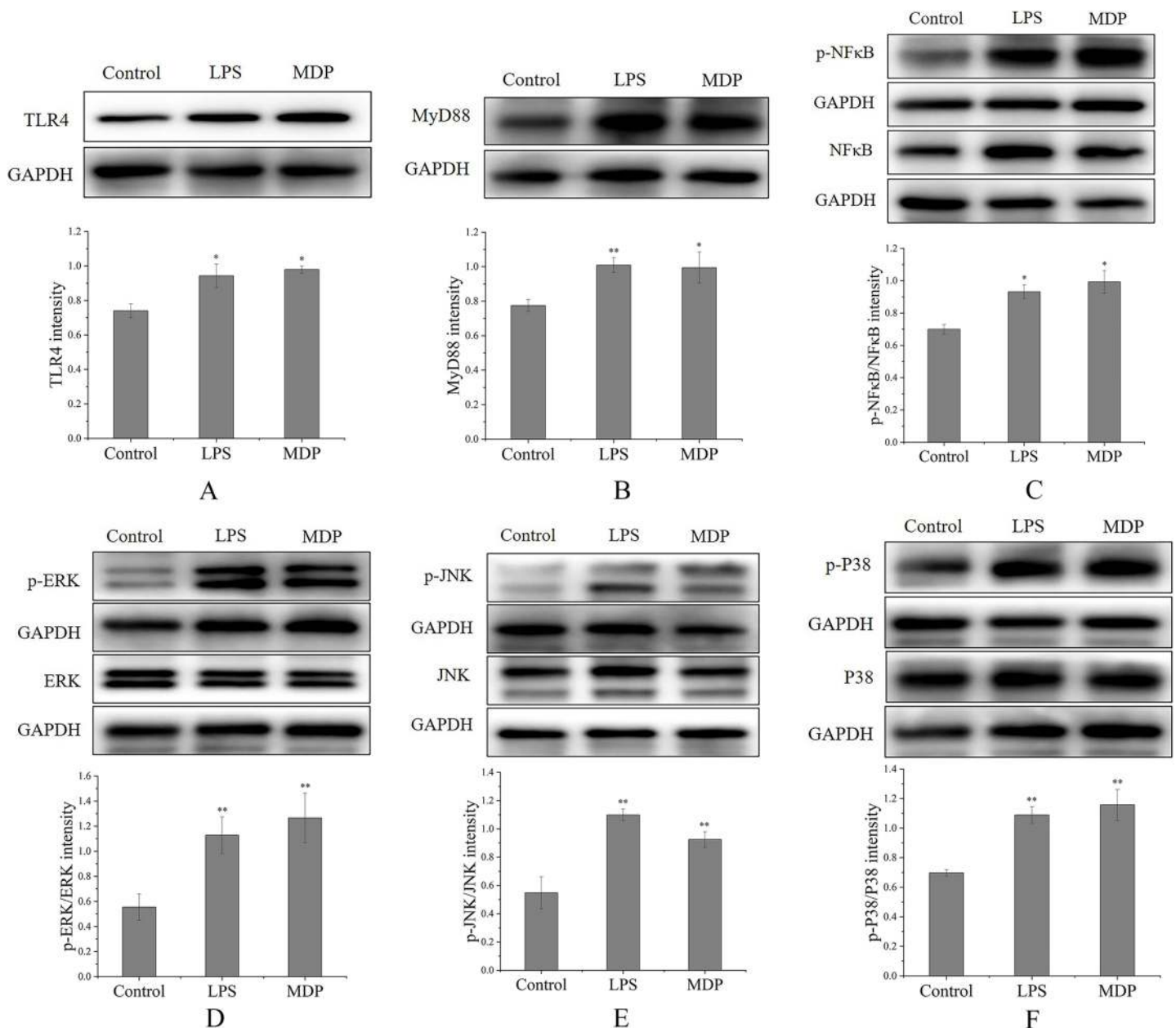


Figure 6 The influence of MDP on the stimulation of TLR4-MAPK/NFκB signaling pathways in RAW 264.7 cells. (A) TLR4, (B) MyD88, (C) p-NFκB/NFκB, (D) p-ERK/ERK, (E) p-JNK/JNK, (F) p-P38/P38. The values are presented as the mean ± SD, $n = 3$. * $P < 0.05$, ** $P < 0.01$, compared with Control group. Full-size [DOI: 10.7717/peerj.13946/fig-6](https://doi.org/10.7717/peerj.13946/fig-6)

ginseng plant polysaccharides, Ginseng, American ginseng, and *Panax notoginseng*, which all consisted of six monosaccharides, but the intensity of each monosaccharide peak differed. This research can both ensure the quality of certain polysaccharides found in *Panax* spp., and perform a fundamental role in controlling the quality of the functional food products associated with these polysaccharides. Ding (2020) established a fingerprint profile of *Lonicera japonica* polysaccharide by GC, and the results of principal component analysis and factor analysis showed that *L. japonica* produced in Shandong had the best

quality and those produced in Jilin were the last, indicating that different climatic environments could lead to differences in quality of *L. japonica* herbs from different origins. It serves as a baseline for determining the overall quality of herbs derived from *L. japonica*.

However, GC is not suitable for the determination of uronic acid fractions. Based on this, we determined the monosaccharide composition of tMDP by thin-layer chromatography. The results (Fig. S1) showed that tMDP did not contain uronic acid components, so GC could be used to detect its monosaccharide composition. Additionally, GC fingerprints may be established to offer a groundwork for the quality control of the rhizome of *M. dauricum* medicinal materials. A total of 11 common peaks were obtained by GC fingerprinting of tMDP, and 6 peaks were identified. Among them, peak 1 was Rha, peak 2 was Ara, peak 3 was Fuc, peak 8 was Man, peak 9 was Glu, peak 10 was Gal, and peak 11 was internal standard. However, peaks 5, 7, and 8 were also present in the mixed standards, which may be by-products of the derivatization process. Through GC fingerprinting and principal component analysis, the better quality herbs were screened out, thus laying the foundation for subsequent experiments.

Polysaccharides have been demonstrated to perform a broad range of biological functions, according to numerous studies, and the immunomodulatory activity of polysaccharides is widely acknowledged to be the most significant biological function of these molecules. Macrophages are immune cells that have a phagocytotic function to eliminate foreign antigens and perform an instrumental function in the case of antimicrobial invasion and host defense (Liu et al., 2022b). By coming into close touch with or eliminating infections or tumor cells, activated macrophages may generate cytokines that trigger other immune cells (Liu et al., 2022b). In this study, MDP was found to activate RAW264.7 cells and enhance the secretion of TNF- α , NO, and IL-6 indicating that MDP has immunomodulatory properties. Furthermore, the influence of MDP in promoting the release of TNF- α , NO, and IL-6 was suppressed after TAK-242 treatment. This suggests the possibility that MDP exerts immunomodulatory activity by activating the TLR4 protein pathway. The connection between the MAPK, TLR4, and TLR4 signaling pathways is known. When TLR4 is expressed in RAW264.7 cells, it activates two distinct signaling networks: the TRIF-dependent pathways and the MyD88-dependent pathways. Once the TLR4 signaling pathway has been activated, it stimulates the activation of the NF- κ B and MAPK signaling pathways in a mechanism that is reliant on MyD88. The findings of western blotting illustrated that MDP could activate the expression of key node proteins on the TLR4-MAPK/NF κ B pathway. In summary, the results of our research showed that MDP does, in fact, remarkably activate macrophages by activating TLR4, which is mediated by MAPK and NF κ B signaling pathways (Fig. 7). This signaling pathway has also been shown to be responsible for the immunomodulatory activities of other polysaccharides, according to multiple research reports (Yang et al., 2019; Liu, 2022a; Wu et al., 2019).

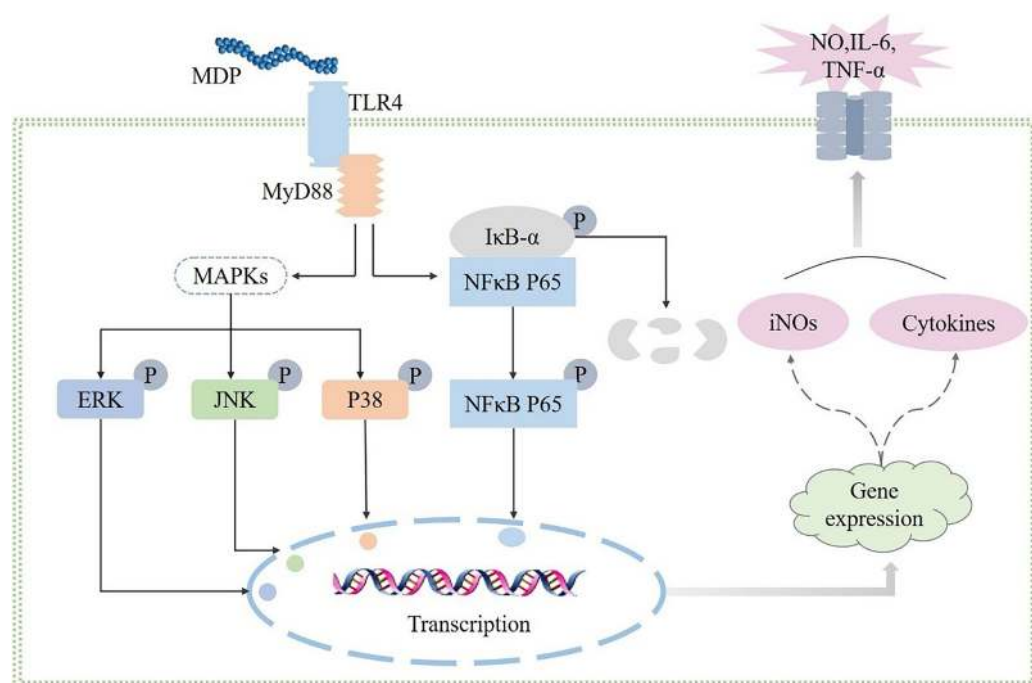


Figure 7 Possible immunomodulatory signaling mechanism of MDP in RAW264.7 macrophages.

Full-size [DOI: 10.7717/peerj.13946/fig-7](https://doi.org/10.7717/peerj.13946/fig-7)

CONCLUSIONS

In this research, the extraction conditions of polysaccharides from the rhizome of *M. dauricum* were successfully optimized by RSM. The GC fingerprints of 10 batches of the rhizome of *M. dauricum* polysaccharides were established, and The findings of SA and PCA of the fingerprints illustrated that the rhizome of *M. dauricum* polysaccharides derived from various sources each had their own distinct chromatographic fingerprint features. Moreover, according to the findings of this research, MDP may substantially activate RAW264.7 cells through the TLR4-MAPK/NFκB dependent signaling pathway so as to generate biological cytokines. In conclusion, our studies have demonstrated an *in vitro* immunomodulatory effect of MDP with a molecular mechanism that is related to the activation of the TLR4-MAPK/NFκB signaling pathway.

ABBREVIATIONS

GC	Gas chromatography
RSM	Response surface method
tMDP	Total polysaccharides from the rhizome of <i>M. dauricum</i>
MDP	Polysaccharides from the rhizome of <i>M. dauricum</i>
PCA	Principal components analysis
TLR4	Toll-like receptor 4
MyD88	Myeloid differentiation factor 88 protein
MAPK	Mitogen-activated protein kinase
NFκB	Nuclear factor kappa-B

TCA	Trichloroacetic acid
TFA	Trifluoroacetic acid
MTT	Methyl thiazolyl tetrazolium
LPS	Lipopolysaccharide
RIPA	Radioimmune precipitation assay
BCA	Bicinchoninic acid

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

We received support in the form of grants from the National Natural Science Foundation of China (81973218), the Taishan Industry Leading Talents Project (tscy20200410), the Technology Development Program of TCM of Shandong Province (2019-0024), the Open Projects Fund of NMPA Key Laboratory for Quality Research and Evaluation of Carbohydrate-based Medicine (No. 2021QRECM02) and the Shandong Key Laboratory of Carbohydrate Chemistry and Glycobiology, Shandong University (2021CCG05). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:

National Natural Science Foundation of China: 81973218.

Taishan Industry Leading Talents Project: tscy20200410.

Technology Development Program of TCM of Shandong Province: 2019-0024.

Open Projects Fund of NMPA Key Laboratory for Quality Research and Evaluation of Carbohydrate-based Medicine: 2021QRECM02.

Shandong Key Laboratory of Carbohydrate Chemistry and Glycobiology, Shandong University: 2021CCG05.

Competing Interests

We hereby declare that there are no conflicting interests involved in the present research. Li Li is employed by Sishui Siheyuan Culture and Tourism Development Company, Ltd.

Author Contributions

- Pei Yang conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, and approved the final draft.
- Yang Zhai conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, and approved the final draft.
- Yan Ma conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, and approved the final draft.
- Beibei Mao performed the experiments, prepared figures and/or tables, and approved the final draft.

- Fengshan Wang analyzed the data, authored or reviewed drafts of the article, and approved the final draft.
- Li Li analyzed the data, authored or reviewed drafts of the article, and approved the final draft.
- Lijuan Luan conceived and designed the experiments, analyzed the data, authored or reviewed drafts of the article, and approved the final draft.
- Yuhong Liu conceived and designed the experiments, analyzed the data, authored or reviewed drafts of the article, and approved the final draft.

Data Availability

The following information was supplied regarding data availability:

Raw data is available in the [Supplemental Files](#).

Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.13946#supplemental-information>.

REFERENCES

- Chen JY, Xie YF, Zhou TX, Qin GW. 2012. Chemical constituents of Menispermum dauricum: chemical constituents of Menispermum dauricum. *Chinese Journal of Natural Medicines* 10(4):292–294 DOI 10.3724/SP.J.1009.2012.00292.
- Cheng XD, Wu QX, Zhao J, Su T, Lu YM, Zhang WN, Wang Y, Chen Y. 2019. Immunomodulatory effect of a polysaccharide fraction on RAW 264.7 macrophages extracted from the wild lactarius deliciosus. *International Journal of Biological Macromolecules* 128:732–739 DOI 10.1016/j.ijbiomac.2019.01.201.
- Cheong KL, Wu DT, Deng Y, Leong F, Zhao J, Zhang WJ, Li SP. 2016. Qualitation and quantification of specific polysaccharides from Panax species using GC-MS, saccharide mapping and HPSEC-RID-MALLS. *Carbohydrate Polymers* 153(3):47–54 DOI 10.1016/j.carbpol.2016.07.077.
- Ding J. 2020. Study on fingerprint and in vitro antiviral activity of Lonicera japonica Polysaccharide. *China Pharmacy* 31:1061–1067.
- Dong Y, Pei F, Su A, Sanidad KZ, Ma G, Zhao L, Hu Q. 2020. Multiple fingerprint and fingerprint-activity relationship for quality assessment of polysaccharides from Flammulina velutipes. *Food and Chemical Toxicology* 135:110944 DOI 10.1016/j.fct.2019.110944.
- Fan S, Wang Y, Zhang Y, Wu Y, Chen X. 2021. Achyranthes bidentata polysaccharide activates nuclear factor-kappa B and promotes cytokine production in J774A.1 Cells through TLR4/MyD88 signaling pathway. *Frontiers in Pharmacology* 12:753599 DOI 10.3389/fphar.2021.753599.
- Li H, Gong X, Wang Z, Pan C, Zhao Y, Gao X, Liu W. 2019. Multiple fingerprint profiles and chemometrics analysis of polysaccharides from Sarcandra glabra. *International Journal of Biological Macromolecules* 123:957–967 DOI 10.1016/j.ijbiomac.2018.11.103.
- Li M, Shan BE, Liang WJ, Ren FZ. 2006. Experimental study on the antimutagenic and mutagenic activity of Rhizoma menispermum extracts. *Chinese Journal of Cancer Prevention and Treatment* 411–413 DOI 10.16073/j.cnki.cjcpt.2006.06.004.

- Liang WJ, Liu DQ, Shan BE, Zhang J, Li QX. 2005. Study of immunoregulatory activity of rhizoma menipermi extracts on mouse and human lymphocytes and macrophages in vitro. *Chinese Journal of Immunology* 56–59.
- Lin M, Xia BR, Yang M, Gao S, Huo YQ, Lou G. 2013a. Anti-ovarian cancer potential of two acidic polysaccharides from the rhizoma of *Menispermum dauricum*. *Carbohydrate Polymers* 92(2):2212–2217 DOI 10.1016/j.carbpol.2012.12.013.
- Lin M, Xia BR, Yang M, Gao S, Huo YQ, Lou G. 2013b. Characterization and antitumor activities of a polysaccharide from the rhizoma of *Menispermum dauricum*. *International Journal of Biological Macromolecules* 53:72–76 DOI 10.1016/j.ijbiomac.2012.11.012.
- Liu J. 2022a. Research progress on fingerprint analysis and profile-effectrelationship of polysaccharides. *Science and Technology of Food Industry* 43:404–412 DOI 10.13386/j.issn1002-0306.2021010244.
- Liu X, Chen X, Xie L, Xie J, Shen M. 2022b. Sulfated Chinese yam polysaccharide enhances the immunomodulatory activity of RAW 264.7 cells via the TLR4-MAPK/NF-κB signaling pathway. *Food & Function* 13(3):1316–1326 DOI 10.1039/D1FO03630K.
- Liu YH, Liu CH, Jiang HQ, Zhou HL, Li PL, Wang FS. 2015b. Isolation, structural characterization and neurotrophic activity of a polysaccharide from *phellinus ribis*. *Carbohydrate Polymers* 133:24–30 DOI 10.1016/j.carbpol.2015.07.013.
- Liu W, Xu J, Zhu R, Zhu Y, Zhao Y, Chen P, Pan C, Yao W, Gao X. 2015a. Fingerprinting profile of polysaccharides from *Lycium barbarum* using multiplex approaches and chemometrics. *International Journal of Biological Macromolecules* 78:230–237 DOI 10.1016/j.ijbiomac.2015.03.062.
- Ma L, Jiao K, Luo L, Xiang J, Fan J, Zhang X, Yi J, Zhu W. 2019. Characterization and macrophage immunomodulatory activity of two polysaccharides from the flowers of *Paeonia suffruticosa* Andr. *International Journal of Biological Macromolecules* 124(4):955–962 DOI 10.1016/j.ijbiomac.2018.12.035.
- Pan Q, Sun Y, Li X, Zeng B, Chen D. 2021. Extraction, structural characterization, and antioxidant and immunomodulatory activities of a polysaccharide from *Notarchus leachii* freeri eggs. *Bioorganic Chemistry* 116(10):105275 DOI 10.1016/j.bioorg.2021.105275.
- Ren D, Zhao Y, Zheng Q, Alim A, Yang X. 2019. Immunomodulatory effects of an acidic polysaccharide fraction from herbal *gynostemma pentaphyllum* tea in RAW264.7 cells. *Food & Function* 10(4):2186–2197 DOI 10.1039/C9FO00219G.
- Sorimachi K, Akimoto K, Hattori Y, Ieiri T, Niwa A. 1999. Secretion of TNF-α, IL-8 and nitric oxide by macrophages activated with polyanions, and involvement of interferon-γ in the regulation of cytokine secretion. *Cytokine* 11(8):571–578 DOI 10.1006/cyto.1998.0472.
- Sun X, Wang H, Han X, Chen S, Zhu S, Dai J. 2014. Fingerprint analysis of polysaccharides from different *Ganoderma* by HPLC combined with chemometrics methods. *Carbohydrate Polymers* 114(2):432–439 DOI 10.1016/j.carbpol.2014.08.048.
- Tian H, Liu ZJ, Pu YW, Bao YX. 2019. Immunomodulatory effects exerted by *poria cocos* polysaccharides via TLR4/TRAE6/NF-κB signaling in vitro and in vivo. *Biomedicine & Pharmacotherapy* 112(1):108709 DOI 10.1016/j.biopha.2019.108709.
- Wang Z, Liu Z, Zhou L, Long T, Zhou X, Bao Y. 2017b. Immunomodulatory effect of APS and PSP is mediated by Ca2(+)-cAMP and TLR4/NF-kappaB signaling pathway in macrophage. *International Journal of Biological Macromolecules* 94(8):283–289 DOI 10.1016/j.ijbiomac.2016.10.018.

- Wang Y, Tian Y, Shao J, Shu X, Jia J, Ren X, Guan Y. 2018. Macrophage immunomodulatory activity of the polysaccharide isolated from Collybia radicata mushroom. *International Journal of Biological Macromolecules* 108(2):300–306 DOI 10.1016/j.ijbiomac.2017.12.025.
- Wang M, Yang XB, Zhao JW, Lu CJ, Zhu W. 2017a. Structural characterization and macrophage immunomodulatory activity of a novel polysaccharide from smilax glabra roxb. *Carbohydrate Polymers* 156(4):390–402 DOI 10.1016/j.carbpol.2016.09.033.
- Wu D, Du JK, Zhang Y, Su Y, Zhang H. 2018. Anti-tumor effects of phenolic alkaloids of menispermum dauricum on gastric cancer in vivo and in vitro. *Journal of Cancer Research and Therapeutics* 14(Supplement):S505–S511 DOI 10.4103/0973-1482.184521.
- Wu MX, Jin LQ. 2007. The research progress of the rhizoma of Menispermum dauricum. 9:35–38 DOI 10.13313/j.issn.1673-4890.2007.09.011.
- Wu Y-G, Wang K-W, Zhao Z-R, Zhang P, Liu H, Zhou G-J, Cheng Y, Wu W-J, Cai Y-H, Wu B-L, Chen F-Y. 2019. A novel polysaccharide from Dendrobium devonianum serves as a TLR4 agonist for activating macrophages. *International Journal of Biological Macromolecules* 133(21):564–574 DOI 10.1016/j.ijbiomac.2019.04.125.
- Xin X, Ma LM, Zhou YR, Shen W, Xu DY, Dou J. 2019. Polysaccharide enhanced NK cell cytotoxicity against pancreatic cancer via TLR4/MAPKs/NF-κB pathway in vitro/vivo. *Carbohydrate Polymers* 225:115223 DOI 10.1016/j.carbpol.2019.115223.
- Yang P, Jin J, Ma Y, Wang FS, Li YY, Duan BG, Zhang Y, Liu Y. 2022. Structure characterization, immunological activity, and mechanism of a polysaccharide from the rhizome of Menispermum dauricum DC. *Frontiers in Nutrition* 9:922569 DOI 10.3389/fnut.2022.922569.
- Yang F, Li XZ, Yang Y, Ayivi-Tosuh SM, Wang F, Li H. 2019. A polysaccharide isolated from the fruits of Physalis alkekengi L. induces RAW264.7 macrophages activation via TLR2 and TLR4-mediated MAPK and NF-κB signaling pathways. *International Journal of Biological Macromolecules* 140(3):895–906 DOI 10.1016/j.ijbiomac.2019.08.174.
- Yu Q, Nie SP, Wang JQ, Yin PF, Li WJ, Xie MY. 2012. Polysaccharide from Ganoderma atrum induces tumor necrosis factor-α secretion via phosphoinositide 3-kinase/Akt, mitogen-activated protein kinase and nuclear factor-κB signaling pathways in RAW264.7 cells. *International Immunopharmacology* 14(4):362–368 DOI 10.1016/j.intimp.2012.09.005.
- Zhang Y, Wu M, Xi J, Pan C, Xu Z, Xia W, Zhang W. 2021. Multiple-fingerprint analysis of Poria cocos polysaccharide by HPLC combined with chemometrics methods. *Journal of Pharmaceutical and Biomedical Analysis* 198(3):114012 DOI 10.1016/j.jpba.2021.114012.
- Zhao Y, Yan BC, Wang ZY, Li MJ, Zhao W. 2020. Natural Polysaccharides with immunomodulatory activities. *Mini-Reviews in Medicinal Chemistry* 20(2):96–106 DOI 10.2174/1389557519666190913151632.
- Zhong RF, Yang JJ, Geng JH, Chen J. 2021. Structural characteristics, anti-proliferative and immunomodulatory activities of a purified polysaccharide from Lactarius volemus Fr. *International Journal of Biological Macromolecules* 192(12):967–977 DOI 10.1016/j.ijbiomac.2021.10.049.
- Zhou Y, Wang S, Feng W, Zhang Z, Li H. 2021. Structural characterization and immunomodulatory activities of two polysaccharides from Rehmanniae Radix Praeparata. *International Journal of Biological Macromolecules* 186:385–395 DOI 10.1016/j.ijbiomac.2021.06.100.
- Zhou YL, Zhao X, Li SX, Lv HM, Wang AL, Zhang Y. 2018. Alkaloids from Rhizome of Menispermum dauricum and Their Anti-inflammatory activity. 20:163–168 DOI 10.13313/j.issn.1673-4890.20170831005.