

PHYTOCHEMICAL CHARACTERIZATION AND TOXICITY ASSESSMENT OF ACUTE AND SUB-ACUTE ORAL ADMINISTRATION OF AQUEOUS EXTRACT OF *JATROPHA TANJORENSIS* LEAF

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ABSTRACT

The phytochemical evaluation of the aqueous extract of *Jatropha tanjorensis* leaf revealed the presence of alkaloids, tannins, flavonoids, phenols, terpenoids, saponins, cardiac glycosides, and steroids. Toxicity studies indicated no mortality up to 6000 mg/kg in mice, with 33.33% mortality at 7000 mg/kg and 100% mortality at 10,000 mg/kg, yielding an LD₅₀ of 7745.97 mg/kg. Biochemical analyses in Wistar rats showed no significant ($p < 0.05$) changes in serum ALT (Alanine Aminotransferase) and AST (Aspartate Aminotransferase) activities but a significant ($p < 0.05$) reduction in ALP (Alkaline Phosphatase) activity, particularly at 400 mg/kg. Serum total protein and albumin concentrations increased significantly ($p < 0.05$), while bilirubin and globulin levels remained unchanged ($p < 0.05$). Urea levels decreased significantly ($p < 0.05$) at 200 mg/kg, with no significant ($p < 0.05$) changes in creatinine, sodium, chloride, and potassium levels. Bicarbonate levels were significantly ($p < 0.05$) reduced at 200 mg/kg. Antioxidant biomarkers—SOD (Superoxide Dismutase), CAT (Catalase), GSH (Reduced glutathione), and GPx (Glutathione peroxidase)—increased ($p < 0.05$) dose-dependently, while MDA (Malondialdehyde) levels decreased ($p < 0.05$). Histological analyses of liver and kidney sections revealed normal ultrastructures, indicating no histopathological damage. These findings suggest that *Jatropha tanjorensis* extract has a favorable safety profile at lower doses, with potential antioxidant benefits and no adverse effects on liver and kidney structures.

Keywords: Antioxidant enzymes, *Jatropha tanjorensis*, Liver enzymes, Phytochemicals, Toxicity

INTRODUCTION

Traditional herbal medicines have long been cherished for their natural origins and perceived minimal side effects, serving as integral components of local and regional healing practices worldwide (WHO, 2019; Rizvi et al., 2022). The World Health Organization defines these medicines as plant-derived substances with minimal industrial processing, used for the prevention and treatment of illnesses. Over millennia, they have found extensive application in both developed and developing countries, evolving from crude formulations like tinctures and teas to contributing directly to the synthesis of modern pharmaceuticals such as aspirin (from willow bark), morphine (from opium poppy), and quinine (from cinchona bark) (Chaachouay & Zidane, 2024). While the historical efficacy of herbal remedies is undeniable, concerns about

their safety persist, prompting rigorous toxicity evaluations. Despite sparse reports of adverse reactions leading to injury or death, the presence of potentially harmful compounds like alkaloids and anthraquinone glycosides necessitates comprehensive safety assessments (Zhang et al., 2015). This underscores the critical importance of evaluating the toxicity of medicinal plants to ensure their therapeutic benefits outweigh any potential risks (Neergheen-Bhujun, 2013).

The assessment of acute, sub-acute, and sub-chronic toxicity in medicinal plants plays a crucial role in ensuring their safety and efficacy for human consumption. Acute toxicity assessments provide valuable insights into the immediate adverse effects of a medicinal plant or its extracts. This helps in determining safe dosage levels and potential risks associated with high doses. Sub-acute toxicity studies, conducted over a longer duration, further

elucidate the cumulative effects and potential toxicity of repeated doses, mimicking more closely the conditions of chronic use. Sub-chronic toxicity studies, on the other hand, evaluate the effects of repeated exposure over a longer duration, mimicking more closely the typical usage patterns of medicinal products. Both assessments contribute significantly to ensuring the safety of herbal medicines (Li et al., 2020; Njinga et al., 2020).

Jatropha tanjorensis (J. L. Ellis & Saroja), a member of the Euphorbiaceae family, stands as a versatile botanical wonder with a plethora of medicinal virtues (Ellis & Saroja, 1961). This gregarious shrub, reaching heights of about 1.8 meters, thrives as a common weed among field crops, primarily flourishing in the rainforest zones of West Africa (Idu et al., 2014). The multifaceted applications of *Jatropha tanjorensis* in traditional medicine and beyond underscore its significance. Abundant scientific evidence supports the medicinal properties of this plant. Every part of the *Jatropha tanjorensis* plant, from its seeds to its leaves and bark, is rich in therapeutic potential. Exploring the medicinal properties of *Jatropha tanjorensis* uncovers a wealth of benefits, including anti-diabetic, cardiovascular, antioxidant, and haematological improvements (Adebajo et al., 2001; Oyewole & Akingbala, 2011; Omoregie & Osagie, 2011; Idu et al., 2014).

The antioxidant and hypoglycaemic properties of *Jatropha tanjorensis* leaf extracts have made them a popular treatment for conditions such as diabetes, malaria, and hypertension in Southern Nigerian regions like Edo State. Here, the leaves are consumed not only as a nutritious vegetable but also as part of daily meals, due to their reputed effectiveness in managing diabetes mellitus, thanks to their anti-hyperglycaemic effects (Olayiwola et al., 2004). These antioxidants enhance the body's defence mechanisms and are crucial in fighting oxidative stress-related conditions like diabetes, malaria, and hypertension, which are especially common in areas where this plant is cultivated. Studies administering *Jatropha tanjorensis* leaf powder to animal models have demonstrated improvements in haematological indices, indicating a potential enhancement in bone marrow function (Orhue et al., 2008). Additionally, the plant's antibacterial attributes have been delineated, affirming its role

in infectious disease management (Viswanathan & Jeyaananthi, 2009). The nutritional profile of *Jatropha tanjorensis* is equally noteworthy, boasting a rich concentration of antioxidant nutrients such as phosphorus, selenium, zinc, vitamin C, and E (Omobuwajo et al., 2011).

Phytochemical investigations revealed the existence of alkaloids, tannins, saponins, flavonoids, terpenoids, cardiac glycosides, and anthraquinones in the aqueous leaf extract of *Jatropha tanjorensis* (Ebenyi et al., 2021). Similarly, alkaloids, tannins, flavonoids, cardiac glycosides, and anthraquinones were detected in the methanol leaf extract of *Jatropha tanjorensis* (Igbinaduwa et al., 2011). Studies have reported that *Jatropha tanjorensis* leaf extract contains phytochemical constituents capable of reducing blood cholesterol levels, which can be beneficial in treating cardiovascular diseases resulting from hyperlipidaemia (Oyewole & Akingbala, 2011). *Jatropha tanjorensis* leaf extract also has antimicrobial properties, inhibiting the growth of *Escherichia coli*, *Staphylococcus aureus*, and *Pseudomonas aeruginosa* (Ewa-Udu et al., 2022). As scientific exploration continues, this botanical gem promises to unveil even more of its healing secrets, potentially revolutionising the landscape of natural medicine. An earlier study examining the toxicity characteristics of *Jatropha tanjorensis* demonstrated the absence of toxicity when administering (2000 and 5000 mg/kg) of aqueous leaf extract to mice and rats within 24 hours after oral ingestion (Ebenyi et al., 2021). Similarly, no observed toxicity was reported at 8000 mg/kg of methanol extracts in the same animal models within the same timeframe (Igbinaduwa et al., 2011).

Insufficient exploration into the toxicity evaluation of the aqueous extract derived from *Jatropha tanjorensis* leaf, especially regarding acute and sub-acute oral administration, limits a thorough understanding of the safety attributes linked to this extract. Assessing toxicity is crucial for identifying potential adverse reactions, setting safe dosage thresholds, and balancing the risks and benefits associated with utilising the extract in therapeutic or medicinal scenarios. The absence of comprehensive toxicity analyses may lead to uncertainties regarding the extract's safety, potentially prompting cautious or restricted usage within clinical settings. The study seeks to identify and quantify the key phytochemical constituents present in the aqueous extract of

Jatropha tanjorensis leaf. This entails gauging the levels of alkaloids, flavonoids, tannins, saponins, terpenoids, cardiac glycosides, anthraquinones, and other bioactive compounds that contribute to the plant's medicinal attributes. Although *Jatropha tanjorensis* offers potential medicinal benefits, the presence of alkaloids and anthraquinones requires caution due to their potential toxic effects such as hepatotoxicity, nephrotoxicity, cardiotoxicity, gastrointestinal distress, and neurotoxicity. Proper dosing, a thorough understanding of the plant's pharmacology, are essential to mitigate these risks. Furthermore, it aims to evaluate the toxicity of the aqueous extract of *Jatropha tanjorensis* leaf through both acute and sub-acute oral administration, with the goal of elucidating its safety profile and potential therapeutic applications. In conclusion, acute and sub-acute toxicity assessment of medicinal plants is indispensable for safety evaluation, risk identification, quality control, optimizing treatment regimens, public health protection, scientific validation, and building consumer confidence. These assessments are essential steps in ensuring the safe and effective use of herbal medicines in healthcare practice.

Materials and Methods

Collection of Plant Materials and Preparation of Aqueous Extract of *Jatropha tanjorensis* Leaves

Fresh leaves of *Jatropha tanjorensis* were collected from the National Crop Research Institute (NCRI) in Umudike, Abia State, Nigeria, and identified by Mr. Pipi Okey from the Department of Plant Science and Biotechnology, Michael Okpara University of Agriculture, Umudike. The specimen was assigned voucher number MOUAU/PSB/18/103592 and deposited in the Department's Herbarium. The leaves were rinsed with distilled water to remove dirt and air-dried at room temperature until completely dry. The dried leaves were ground into a fine powder using an electric blender. A known quantity (188.54 g) of the powdered sample was macerated in 2.5 L of distilled water for 24 hours in a sterile extraction jar. The filtrate was first filtered using muslin cloth and then filtered again using Whatman filter paper No. 1 into a clean, calibrated flask. The filtrate was lyophilized, using a freeze dryer set to a temperature of -40°C. A dark-brown paste with a yield of 50.85% was obtained, transferred into a sterile clean bottle,

and stored in a refrigerator at 4°C until needed for biochemical assays.

Qualitative Phytochemical Analysis

Preliminary analysis of the aqueous extract of *Jatropha tanjorensis* leaf was carried out to identify the presence of various phytoconstituents such as alkaloids, tannins, flavonoids, phenols, terpenoids, saponin, cardiac glycoside, and steroids.

Test for Alkaloid

For qualitative alkaloid determination, a few drops of Meyer's reagent were added to 1 mL of extract. The formation of a creamy white precipitate was considered positive for the alkaloid (Archana et al., 2012).

Test for Tannin

For qualitative tannin analysis, a few drops of lead acetate were added to 1 mL of extract. A large white-brown precipitate formation was considered a positive test for tannin (Archana et al., 2012).

Test for Flavonoids

For the confirmation of flavonoid, 0.5 g of each extract were added in a test tube and 10 mL of distilled water, 5 ml of dilute ammonia solution were added to a portion of the aqueous filtrate of each plant extract followed by addition of 1 mL concentrated H₂SO₄. Indication of yellow colour shows the presence of flavonoid in each extract (Sofowora, 1993).

Test for Phenols

To detect phenols, 1 mL of extract solution was added to 2 mL distilled water and a few drops of 10% ferric chloride (FeCl₃). The appearance of a green or blue colour is the indication of phenols presence. Next, 0.50 g of plant extract was added and allowed to dissolve in distilled water and then 3 mL of 10% lead acetate was added. The presence of phenolic compounds was confirmed by the appearance of white precipitation (Trease et al., 2003).

Test for Saponin

To detect the presence of saponin, 5 mL of distilled water was added to 1 mL of extract and vortexed for 10 minutes. The formation of a foam column that did not disappear with the addition of HCl was evaluated as positive for saponin (Sati and Kumar, 2015; Savithramma et al., 2011).

Test for Cardiac Glycoside

Qualitative analysis of cardiac glycosides in the extracts was performed with the Keller Killiani test. 1 mL of acetic acid and 2 drops of ferric chloride were added to 2 mL of extract, then 2 mL of sulfuric acid (concentrated) was added and the colour change was observed. Reddish-brown colour formation was deemed to be a positive test for cardiac glycosides (Archana et al., 2012; Sati & Kumar, 2015).

Test for Terpenoids and Steroids

The presence of terpenoids and steroids was determined by a previously described method (Kantamreddi & Lakshmi, 2010; Siddiqui & Ali, 1997) with slight modifications. Briefly, 0.5 g of solvent-free extract was added in 2 mL chloroform and then filtered. The filtrate was placed on ice; addition of 2 mL of acetic acid and then a few drops of concentrated sulfuric acid (H_2SO_4) was carefully applied to the inner sides of the test tubes. The emergence of a pink or pinkish brown colour/ring indicates the terpenoids existence, while a blue or bluish green colour for steroids presence and the combination of pink and blue/bluish green colours confirmed the presence of both terpenoids and steroids.

Quantitative Phytochemical Analysis

Quantitative analysis of the phytochemical components, whose presence were tested by qualitative analysis, were also carried out. All the analysis were performed in triplicate.

Estimation of Total Alkaloids

Alkaloid determination was made according to the method reported by Selvakumar et al. (2019). To 1 mL of the test extract 5 mL of phosphate buffer (pH 4.7) and 5 mL of bromocresol green (BCG) solution was added to the solution. The mixture was shaken and the complex extracted with 1-, 2-, 3- and 4-mL of chloroform by vigorous shaking. The extracts were then collected in a 10 mL volumetric flask and then diluted to adjust volume with chloroform. The absorbance of the chloroform complex was measured at 470 nm using a UV spectrophotometer against a blank prepared similarly, but without extract. To quantify the total alkaloid content in the sample, a standard curve was prepared using atropine. Standard solution of atropine was prepared at concentrations of (0.1, 0.2, 0.4, 0.6, 0.8, and 1.0 mg/mL). To generate the standard

curve, the average absorbance values were plotted against the corresponding atropine concentrations. A linear regression analysis was performed to obtain the equation of the line: $y = mx + b$. This linear equation was then used to determine the total alkaloid content in the extract. The alkaloid content was expressed as mg atropine equivalent (AE)/g of sample. All tests were repeated in triplicate.

Estimation of Total Tannins

Tannin content was determined according to the method of Van-Burden & Robinson, 1981. A known weight of sample (500 mg) was weighed into a 100 mL plastic bottle. 50 mL of distilled water was added and shaken for 1 h in a mechanical shaker. This was filtered into a 50 mL volumetric flask and made up to the mark. Then 5 mL of the filtered was pipette out into a test tube and mixed with 2 mL of 0.1 M FeCl_3 in 0.1 N HCl and 0.008 M potassium ferrocyanide. The absorbance of each reaction mixture was thereafter measured at 120 nm against a blank using a spectrophotometer. To quantify the total tannin content in the sample, a standard curve was prepared using gallic acid. Standard solution of gallic acid were prepared at concentrations of (0.1, 0.2, 0.3, 0.4, and 0.5 mg/L). The average absorbance values were plotted against the corresponding concentrations of gallic acid to generate the standard curve. A linear regression analysis was performed to obtain the equation of the line: $y = mx + b$. This linear equation was then used to determine the total tannin content in the extract. Results were quantified in the form of GAE (mg of Gallic Acid Equivalents) per gram of dry plant extract/fraction. Samples were analysed in triplicates.

Estimation of Total Flavonoids

The total flavonoid content of the extract was determined using a slightly modified method reported by Meda et al. (2005). 0.5 mL of the extract samples were mixed with 0.5 mL methanol, 50 L of 10% AlCl_3 , 50 L of 1 mol L^{-1} potassium acetate and 1.4 mL water, and allowed to incubate at room temperature for 30 min. Thereafter, the absorbance of each reaction mixture was subsequently measured at 415 nm against blank. To quantify the total flavonoids content of the extract, a standard curve was prepared using quercetin. Standard solutions of quercetin were prepared at concentrations of 40, 80, 120, 160, 200, and 240 g/mL. The average absorbance values were

plotted against the corresponding concentrations of quercetin to generate the standard curve. A linear regression analysis was performed to obtain the equation of the line: $y = mx + b$. This linear equation was then used to determine the total flavonoid content in the extract. The flavonoid content was expressed as mg quercetin equivalents (QE)/g of sample.

Estimation of Total Phenols

Total phenolic content was determined using the method of Makkar et al. (1993) with some modifications. The reaction mixture contained 100 L of the extract, 2.8 mL distilled water, 2 mL of 2% sodium carbonate and 100 L of 50% Folin–Ciocalteu reagent, and it was incubated at room temperature for 30 min. Thereafter, the absorbance of the mixture was measured at 750 nm using a spectrophotometer against a blank prepared similarly, but without extract. To quantify the total phenolic content of the extract, a standard curve was prepared using tannic acid. Standard solution of tannic acid were prepared at concentrations of 0, 20, 40, 60, 80 and 100 g/mL. The average absorbance values were plotted against the corresponding concentrations of tannic acid to generate the standard curve. A linear regression analysis was performed to obtain the equation of the line: $y = mx + b$. This linear equation was then used to determine the total phenolic content in the extract. The total phenol was expressed as mg tannic acid equivalents (TAE)/g of sample.

Estimation of Total Saponins

Saponin content was determined by the gravimetric method of AOAC, (1990). 20 g of sample was weighed into a 250 mL beaker and 100 mL of 20% aqueous ethanol was added. The suspension was heated over a hot water bath for 4 h with continuous stirring at about 55°C. The mixture was filtered and the residue re-extracted with another 200 mL 20% ethanol. The combined extracts were reduced to 40 mL over water bath at about 90°C. The concentrate was transferred into a 250 mL separatory funnel and 20 mL of diethyl ether was added and shaken vigorously. The aqueous layer was recovered while the ether layer was discarded. The purification process was repeated. 60 mL of n-butanol was added and the combined n-butanol extracts were washed twice with 10 mL of 5% aqueous sodium chloride. The remaining solution was heated in a water bath. After

evaporation the sample was dried in the oven to a constant weight. The saponin content was calculated using the formula:

$$\text{Saponin content (\%)} = \frac{\text{Weight of sample (g)}}{\text{Weight of original sample (g)}} \times 100$$

Estimation of Cardiac Glycosides

The determination of the total cardiac glycosides was performed using the Baljet method as described by (El-Olemy et al., 1994) with slight modifications. Briefly, 1 g of pulverized leaf powder was macerated in 10 mL of 70% alcohol for 30 min. The solution was filtered and the residue re-macerated twice to ensure extraction of the metabolites. The extract was dried at 40°C for 6 h and reconstituted in distilled water. To 1 mL of the reconstituted sample was added 1 mL of freshly prepared Baljet's reagent (95 mL of 1% aqueous picric acid + 5 mL of 10% aqueous NaOH) and the intensity of the colour complex formed was read at 495 nm after 1 h against a blank treated in the same way except that the sample was substituted with distilled water. A blank experiment was also performed using distilled water. To quantify the total cardiac glycosides content of the extract, a standard curve was prepared using digitoxin. Standard solutions of digitoxin were prepared at concentrations of 0.2, 0.4, 0.6, 0.8, 1.0 mg/L. The average absorbance values were plotted against the corresponding concentrations of digitoxin to generate the standard curve. A linear regression analysis was performed to obtain the equation of the line: $y = mx + b$. This linear equation was then used to determine the total cardiac glycosides content in the extract. The total cardiac glycosides were expressed as mg digitoxin equivalents (DE)/g of sample.

Estimation of Terpenoids

The determination of the total terpenoid content was performed using the method of Indumathi and Mohandass, (2014) with slight modifications. 1 mL of the extract was mixed with 3 mL of chloroform and allowed to stand for 1.5 h. Then, 1.5 mL of the mixture was taken and mixed with 3.5 mL concentrated H_2SO_4 . The absorbance of the mixture was read at 538 nm. To quantify the total terpenoids content of the extract, a standard curve was prepared using linalool. Standard solutions of linalool were prepared at concentrations of 40, 80, 120, 160,

200, 240 g/mL. The average absorbance values were plotted against the corresponding concentrations of linalool to generate the standard curve. A linear regression analysis was performed to obtain the equation of the line: $y = mx + b$. This linear equation was then used to determine the total terpenoids content in the extract. The total terpenoids content was expressed as mg linalool equivalents (LE)/g of sample.

Estimation of Steroids

The determination of the total steroid content was performed using the method of Indumathi and Mohandass, (2014) with slight modifications. 0.5 g of each extract was dissolved in 3 mL chloroform. Then, 2 mL of concentrated H_2SO_4 was carefully added to form a lower layer. A reddish brown colour at the interface indicated the presence of steroids. The absorbance of the resulting reddish brown colour was measured at 550 nm against a reagent blank. To quantify the total steroids content of the extract, a standard curve was prepared using linalool. Standard solutions of linalool were prepared at concentrations of 40, 80, 120, 160, 200, 240 g/mL. The average absorbance values were plotted against the corresponding concentrations of linalool to generate the standard curve. A linear regression analysis was performed to obtain the equation of the line: $y = mx + b$. This linear equation was then used to determine the total steroid content in the extract. The total steroids content was expressed as mg linalool equivalents (LE)/g of sample.

Animals

Thirty (30) male albino rats (100-150 g) were procured from the Department of Veterinary Medicine, Michael Okpara University of Agriculture, Umudike, Nigeria. The animals were housed in clean, well-ventilated polypropylene cages, maintained under standard conditions (12 h light and 12 h dark cycle) at an ambient temperature of $25 \pm 2^\circ C$ and were fed with standard rodent pellets and water *ad libitum*. The rats were acclimatized for 14 days before the commencement of the experiment.

Experimental Design

The acclimatized animals were randomly divided into five (5) groups (A, B, C, D, E) of six (6) rats each. They were treated as follows:

1. Group A (normal control): received distilled

water throughout the experimental period.

2. Group B (positive control): received 500 mg/kg b.w. of sodium arsenite only throughout the experimental period.
3. Group C (test group 1): received 500 mg/kg b.w. of sodium arsenite and 100 mg/kg b.w. of *Jatropha tanjorensis* leaves extract throughout the experimental period.
4. Group D (test group 2): received 500 mg/kg b.w. of sodium arsenite and 200 mg/kg b.w. of *Jatropha tanjorensis* leaves extract throughout the experimental period.
5. Group E (test group 3): received 500 mg/kg b.w. of sodium arsenite and 400 mg/kg b.w. of *Jatropha tanjorensis* leaves extract throughout the experimental period.

The treatments were administered orally for 14 days and the animals were weighed every 3 days to monitor weight changes.

Preparation of Tissue Homogenate

After the last administration, the animals were fasted overnight and sacrificed by cervical dislocation. The testes were rapidly excised, rinsed in ice-cold 1.15% KCl solution, blotted dry, and weighed. Each tissue sample was then homogenized in four (4) volumes of ice-cold 0.1 M phosphate buffer (pH 7.4) using a Potter-Elvehjem homogenizer. The homogenate was centrifuged at 10,000 g for 20 minutes at $4^\circ C$ to obtain the post-mitochondrial supernatant, which was used for the biochemical assays.

Biochemical Assays

The following biochemical assays were carried out on the post-mitochondrial supernatant.

Malondialdehyde (MDA) Assay

Lipid peroxidation was estimated by measuring the level of malondialdehyde (MDA), a product of lipid peroxidation, using the method of Buege and Aust (1978). Briefly, 0.1 mL of the post-mitochondrial supernatant was mixed with 1.5 mL of 20% acetic acid (pH 3.5), 0.2 mL of 8.1% SDS, and 1.5 mL of 0.8% TBA. The mixture was made up to 4 mL with distilled water and incubated at $95^\circ C$ for 60 minutes. After cooling, 1 mL of distilled water and 5 mL of n-butanol:pyridine mixture (15:1 v/v) were added.

The mixture was shaken vigorously and centrifuged at 4000 rpm for 10 minutes. The absorbance of the organic layer was measured at 532 nm. The MDA concentration was calculated using the extinction coefficient of $1.56 \times 10^5 \text{ M}^{-1}\text{cm}^{-1}$ and expressed as nmol/mg protein.

Superoxide Dismutase (SOD) Assay

SOD activity was determined using the method of Misra and Fridovich (1972). The reaction mixture consisted of 0.1 mL of the post-mitochondrial supernatant, 1.2 mL of sodium carbonate buffer (50 mM, pH 10.2), and 0.2 mL of epinephrine (30 mM). The reaction was initiated by the addition of epinephrine, and the increase in absorbance was measured at 480 nm for 4 minutes. One unit of SOD activity was defined as the amount of enzyme required to cause 50% inhibition of the oxidation of epinephrine to adrenochrome. The specific activity was expressed as units/mg protein.

Catalase (CAT) Assay

CAT activity was determined according to the method of Sinha (1972). The reaction mixture contained 0.1 mL of the post-mitochondrial supernatant and 1.9 mL of phosphate buffer (50 mM, pH 7.0). The reaction was initiated by the addition of 1 mL of hydrogen peroxide (30 mM), and the decrease in absorbance was measured at 240 nm for 2 minutes. The enzyme activity was calculated using the extinction coefficient of $43.6 \text{ M}^{-1}\text{cm}^{-1}$ and expressed as units/mg protein.

Glutathione Peroxidase (GPx) Assay

GPx activity was measured according to the method of Rotruck et al. (1973). The reaction mixture contained 0.1 mL of the post-mitochondrial supernatant, 1 mL of phosphate buffer (50 mM, pH 7.0), 0.1 mL of GSH (1 mM), 0.1 mL of NADPH (0.12 mM), 0.1 mL of sodium azide (1 mM), and 0.1 mL of hydrogen peroxide (0.15 mM). The decrease in absorbance at 340 nm was recorded for 3 minutes. The enzyme activity was expressed as nmol NADPH oxidized/min/mg protein using the extinction coefficient of $6.22 \times 10^3 \text{ M}^{-1}\text{cm}^{-1}$.

Glutathione-S-Transferase (GST) Assay

GST activity was determined according to the method of Habig et al. (1974). The reaction mixture contained 0.1 mL of the post-mitochondrial supernatant, 1.7 mL of phosphate buffer (50 mM, pH

7.0), 0.2 mL of GSH (1 mM), and 0.1 mL of CDNB (1 mM). The increase in absorbance at 340 nm was recorded for 5 minutes. One unit of GST activity was defined as the amount of enzyme required to conjugate 1 mol of CDNB with GSH per minute. The specific activity was expressed as units/mg protein.

Protein Estimation

The protein content in the post-mitochondrial supernatant was estimated by the method of Lowry et al. (1951) using bovine serum albumin as the standard. The absorbance was read at 750 nm and the protein concentration was expressed as mg/mL of the supernatant.

Statistical Analysis

Data were expressed as mean $\pm$ SEM. Statistical analysis was performed using one-way ANOVA followed by Tukey's post hoc test for multiple comparisons. Values of $P < 0.05$ were considered statistically significant.

1 Results

1.1 Phytochemical Analysis of Aqueous Extract of *Jatropha tanjorensis* Leaf

The result of the phytochemical evaluation of Aqueous Extract of *Jatropha tanjorensis* Leaf is shown in Table 1. It revealed the presence of alkaloids, tannins, flavonoids, phenols, terpenoids, saponins, cardiac glycosides, and steroids.

1.2 Results of Acute Toxicity Study of Aqueous Extract of *Jatropha tanjorensis* Leaf in Wistar Rats

Graded doses up to 6000 mg/kg of the extract produced no mortality in rats as the treated rats remained active and physically stable throughout the 24-hour period of observation. However, 33.33 percent mortality was observed in the third phase and group administered 7000 mg/kg body weight while all the rats in the 10,000 mg/kg treated group died, representing 100 percent mortality. The application of Lorke's formula yielded an LD_{50} value of 7745.97 mg/kg body weight for the aqueous extract. The results are presented in Tables 2, 3, and 4 below.

$$\text{LD}_{50} = \sqrt{D_0 \times D_{100}}$$

Where:

D_0 : Highest dose that gave no mortality.

D_{100} : Lowest dose that produced mortality.

Table 1: Phytochemical Constituents of Aqueous Extract of *Jatropha tanjorensis* Leaf

Phytochemical Parameter	Qualitative	Quantitative (mg/100g)
Alkaloids	++	27.15±0.59
Tannins	+	7.63±0.18
Flavonoids	+++	39.18±1.07
Phenols	+++	46.07±1.14
Terpenoids	+	5.41±0.15
Saponins	+++	33.10±1.25
Cardiac glycosides	+	4.83±0.19
Steroids	+	3.80±0.10

+++ = high amount, ++ = moderate amount, + = low amount

Table 2: Stage 1 Acute Toxicity (LD₅₀) Evaluation of the Extract

Group	Dose (mg/kg)	No. of Deaths	Percentage of mortality	Observations
1	10	0/3	0.00	No mortality observed, instead animals remained active and physically stable.
2	100	0/3	0.00	No mortality observed, instead animals remained active and physically stable.
3	1000	0/3	0.00	No mortality observed, instead animals remained active and physically stable.

$$LD_{50} = \sqrt{6000 \times 10000}$$

$$LD_{50} = 7745.97 \text{ mg/kg body weight}$$

$$LD_{50} = \sqrt{D_0 \times D_{100}}$$

Where:

D_0 : Highest dose that gave no mortality.

D_{100} : Lowest dose that produced mortality

$$LD_{50} = \sqrt{6000 \times 10000}$$

$$LD_{50} = 7745.97 \text{ mg/kg body weight}$$

1.3 Effect of Varying Doses of Aqueous Extract of *Jatropha tanjorensis* Leaf on the Activities of Liver Enzymes and Total Protein Levels in Sera of Male Wistar Rats

The experimental results showed no significant differences ($P > 0.05$) in serum levels of ALT and AST between Wistar rats administered the extract at various doses and the control group (Figures 1 and

2). However, serum ALP levels were significantly reduced ($P < 0.05$) in all experimental groups given the extract compared to the control group, with the lowest ALP level observed in the group administered 400 mg/kg (Figure 3). Additionally, serum total protein and albumin concentrations were significantly increased ($P < 0.05$) in rats given the extract compared to their respective controls (Figures 4 and 5). There were no significant differences ($P > 0.05$) in serum total bilirubin levels between the experimental groups and the control group after 30 days (Figure 6). Similarly, serum globulin concentrations did not show a significant increase ($P > 0.05$) in the experimental groups compared to the control (Figure 7).

Table 3: Stage 2: Acute toxicity (LD50) evaluation of the extract

Group	Dose (mg/kg)	No. of Deaths	Percentage of mortality	Observations
1	1600	0/3	0.00	No mortality observed, instead animals remained active and physically stable.
2	2900	0/3	0.00	No mortality observed, instead animals remained active and physically stable.
3	5000	0/3	0.00	No mortality observed. Animals were initially calm but regained physical activity within one hour of administration.

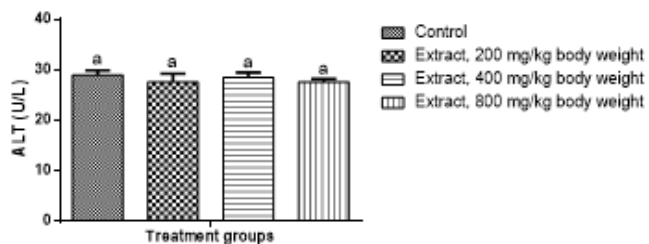


Figure 1: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on serum levels of ALT in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P > 0.05$), whereas data with different lower-case letters are significantly different ($P < 0.05$). Values are presented as Mean $\pm$ SEM.

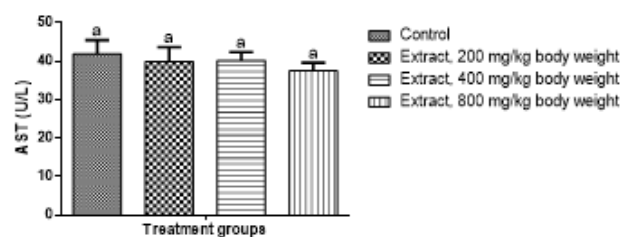


Figure 2: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on serum levels of AST in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P > 0.05$), whereas data with different lower-case letters are significantly different ($P < 0.05$). Values are presented as Mean $\pm$ SEM.

2 Effect of Varying Doses of Aqueous Extract of *Jatropha tanjorensis* Leaf on Kidney Function Indices in Sera of Male Wistar Rats

The serum urea levels in rats given 400 and 800 mg/kg of the extract did not show significant differences ($P > 0.05$) compared to the control group. However, administering 200 mg/kg of the extract resulted in a significant ($P < 0.05$) reduction in urea concentration compared to the control (Figure 8). Similarly, serum creatinine levels in rats given 400 and 800 mg/kg of the extract were not significantly

different ($P > 0.05$) from the control (Figure 9). There were no significant reductions ($P > 0.05$) in serum Na^+ concentration in rats administered 200, 400, and 800 mg/kg of the extract (Figure 10). The serum chloride and potassium ion concentrations in rats given the extract were also not significantly different ($P > 0.05$) from the control (Figures 11 and 12). However, rats administered 200 mg/kg of the extract showed a significant reduction ($P < 0.05$) in serum bicarbonate ion concentration compared to the control and those given 400 mg/kg of the extract. In contrast, rats given 800 mg/kg of the extract did not show significant differences ($P > 0.05$) in serum bicarbonate levels

Table 4: Stage 3 Acute Toxicity (LD₅₀) Evaluation of the Extract

Group	Dose (mg/kg)	No. of Deaths	Percentage of mortality	Observations
1	6000	0/3	0.00	No mortality observed. Animals were initially calm but regained physical activity within 24 hours of administration.
2	7000	1/3	33.33	33.33% mortality was observed. Surviving animals were weak, depressed and calm. They also did not completely regain physical activity within 24 hours of administration.
3	10000	3/3	100.00	Rats were initially calm and depressed and 100% mortality was observed by the end of 24 hours of administration.

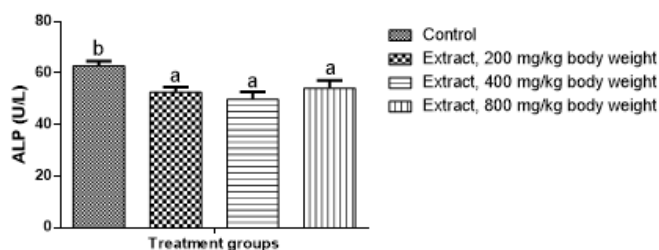


Figure 3: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on serum levels of ALP in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P > 0.05$), whereas data with different lower-case letters are significantly different ($P < 0.05$). Values are presented as Mean $\pm$ SEM.

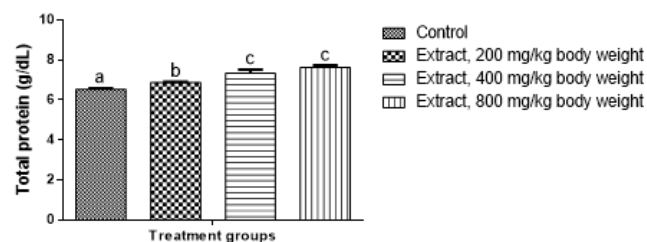


Figure 4: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on serum total protein levels in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P > 0.05$), whereas data with different lower-case letters are significantly different ($P < 0.05$). Values are presented as Mean $\pm$ SEM.

3 Effect of Varying Doses of Aqueous Extract of *Jatropha tanjorensis* Leaf on Sera Antioxidant Status in Male Wistar Rats

In this study, the rats administered varying doses of the extract exhibited significant ($P < 0.05$) increases in

compared to the control (Figure 13).

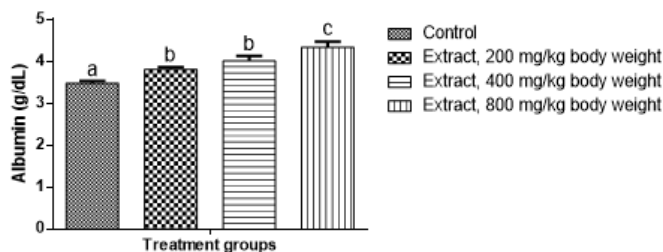


Figure 5: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on serum albumin levels in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P \leq 0.05$). Values are presented as Mean $\pm$ SEM.

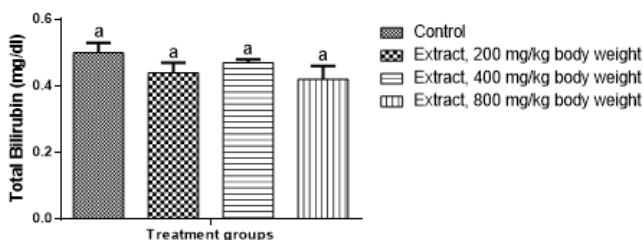


Figure 6: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on serum total bilirubin levels in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P \leq 0.05$). Values are presented as Mean $\pm$ SEM.

serum levels of SOD, CAT, GSH, and GPx, in a dose dependent manner when compared to the control groups. However, at the 800 mg/kg dose, serum levels of SOD and GSH were decreased but not statistically ($P \geq 0.05$) significant when compared to the 400 mg/kg group (Figures 14, 15, 16, 17). Additionally, serum levels of MDA were significantly ($P \leq 0.05$) reduced in a dose dependent manner in all treatment groups when compared to the control (Figure 18).

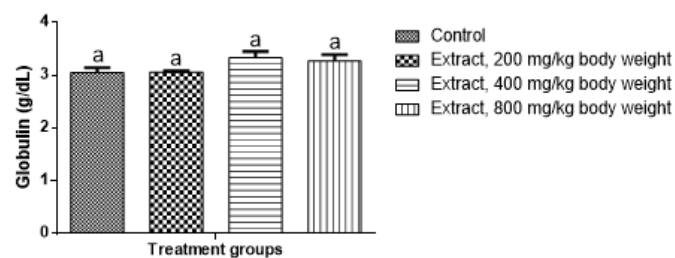


Figure 7: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on serum globulin levels in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P \leq 0.05$). Values are presented as Mean $\pm$ SEM.

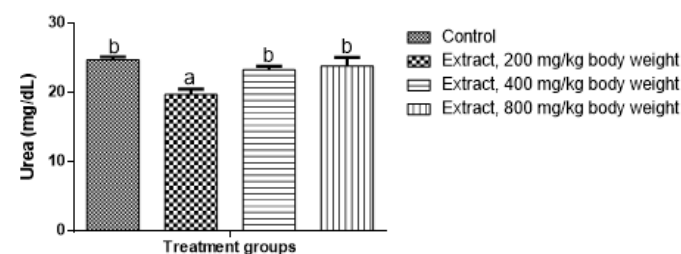


Figure 8: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on serum urea concentrations in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P \leq 0.05$). Values are presented as Mean $\pm$ SEM.

4 Effect of Varying Doses of Aqueous Extract of *Jatropha tanjorensis* Leaf on Kidney Homogenate Supernatant and Antioxidant Status in Male Wistar Rats

In this study, rats administered varying doses of the extract showed significant increases ($P \leq 0.05$) in the antioxidant status of kidney homogenates, as evidenced by elevated levels of SOD, CAT, GSH, and GPx, in a dose-dependent manner compared to the control groups (Figures 19, 20, 21, 22). Additionally, the levels of malondialdehyde (MDA) in the kidney homogenates were significantly reduced ($P \leq 0.05$) in a dose-dependent manner across all treatment groups

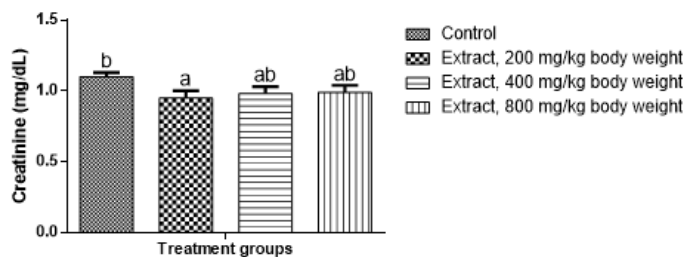


Figure 9: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on serum creatinine concentrations in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P < 0.05$). Values are presented as Mean $\pm$ SEM.

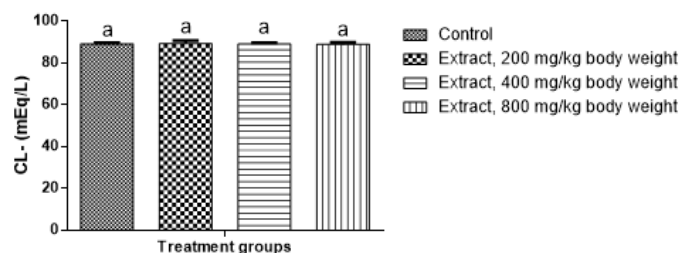


Figure 11: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on chloride ion concentrations in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P < 0.05$). Values are presented as Mean $\pm$ SEM.

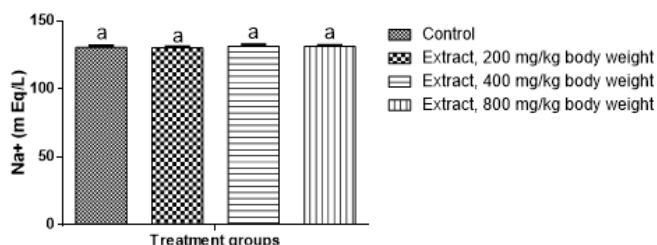


Figure 10: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on sodium ion concentrations in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P < 0.05$). Values are presented as Mean $\pm$ SEM.

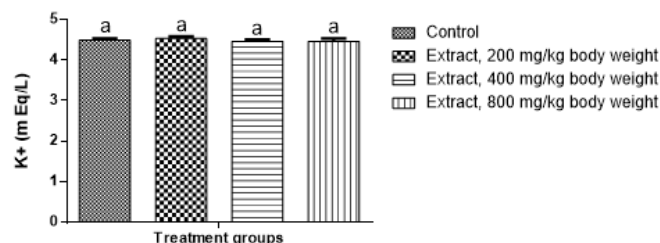


Figure 12: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on potassium ion concentrations in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P < 0.05$). Values are presented as Mean $\pm$ SEM.

compared to the control (Figure 23).

5 Histopathological Assessment of Liver and Kidney Ultrastructures Following Administration of Aqueous Extract of *Jatropha tanjorensis* Leaf

5.1 Liver

Histological analysis of liver sections from rats administered 200, 400, and 800 mg/kg of the extract revealed essentially normal histology, similar to the control group. Sections of the liver in all the groups showed normal hepatic histomorphology, lacking any

signs of acute or chronic damage, and portal tracts without significant inflammation (Plates 1a, b, c, and d). Normal hepatic lobules consisting of hepatocytes arranged in interconnecting cords around the central veins were observed. Likewise, the structures of the portal triads were normal.

Group 1 (Control)

Plate 1a: The photomicrograph of the sectioned liver from the control group revealed normal hepatic histomorphology. The hepatic lobules appeared normal, with hepatocytes arranged in interconnecting cords around the central veins. Additionally, the structures of the portal triads were normal. *H.E x200 magnification / Bar: 50 μ m*. Central veins (V); Portal area (P) consisting of hepatic artery (HA), hepatic

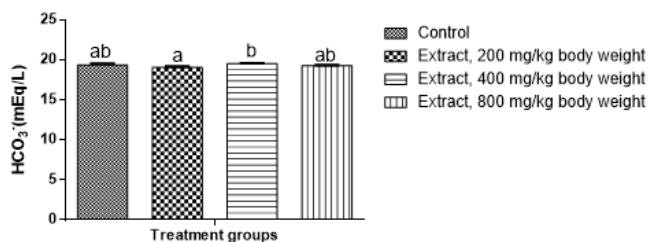


Figure 13: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on serum bicarbonate ion concentrations in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P \leq 0.05$). Values are presented as Mean $\pm$ SEM.

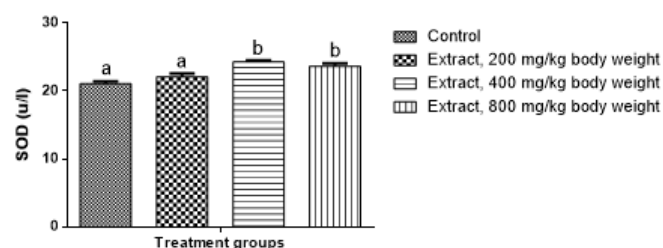


Figure 14: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on serum SOD levels in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P \leq 0.05$). Values are presented as Mean $\pm$ SEM.

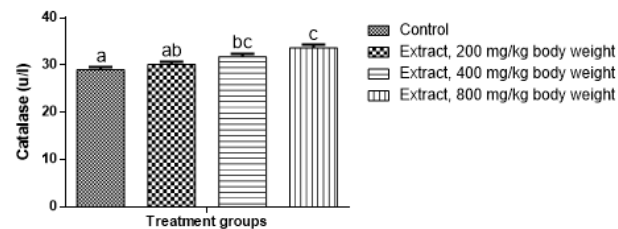


Figure 15: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on serum CAT levels in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P \leq 0.05$). Values are presented as Mean $\pm$ SEM.

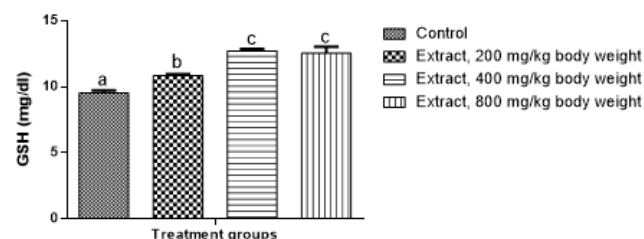


Figure 16: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on serum GSH levels in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P \leq 0.05$). Values are presented as Mean $\pm$ SEM.

vein (HV), and bile duct (BD).

Group 2 (200 mg/kg)

Plate 1b: The photomicrograph of the sectioned liver from the 200 mg/kg treated group revealed normal hepatic histomorphology. The hepatic lobules appeared normal, with hepatocytes arranged in interconnecting cords around the central veins. Additionally, the structures of the portal triads were normal. *H.E x200 magnification / Bar: 50 μ m*. Central veins (V); Portal area (P) consisting of hepatic artery (HA), hepatic vein (HV), and bile duct (BD).

Group 3 (400 mg/kg)

Plate 1c: The photomicrograph of the sectioned liver from the 400 mg/kg treated group revealed normal hepatic histomorphology. The hepatic

lobules appeared normal, with hepatocytes arranged in interconnecting cords around the central veins. Additionally, the structures of the portal triads were normal. *H.E x200 magnification / Bar: 50 μ m*. Central veins (V); Portal area (P) consisting of hepatic artery (HA), hepatic vein (HV), and bile duct (BD).

Group 4 (800 mg/kg)

Plate 1d: The photomicrograph of the sectioned liver from the 800 mg/kg treated group revealed normal hepatic histomorphology. The hepatic lobules appeared normal, with hepatocytes arranged in interconnecting cords around the central veins. Additionally, the structures of the portal triads were normal. *H.E x200 magnification / Bar: 50 μ m*. Central veins (V); Portal area (P) consisting of hepatic artery

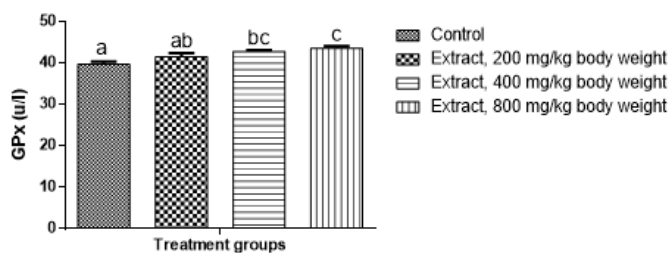


Figure 17: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on serum GPx levels in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P < 0.05$). Values are presented as Mean $\pm$ SEM.

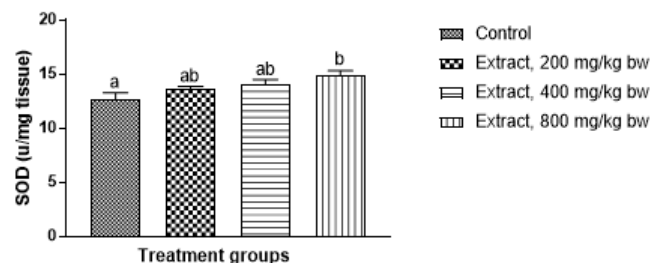


Figure 19: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on SOD of kidney homogenate supernatant in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P < 0.05$). Values are presented as Mean $\pm$ SEM.

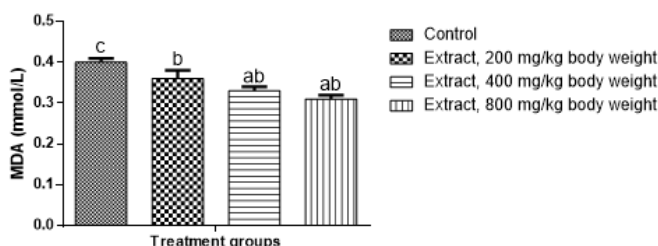


Figure 18: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on serum MDA levels in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P < 0.05$). Values are presented as Mean $\pm$ SEM.

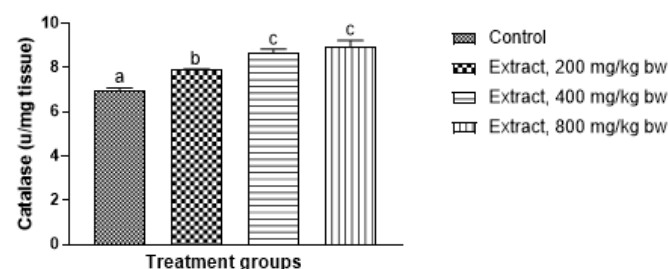


Figure 20: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on CAT of kidney homogenate supernatant in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P < 0.05$). Values are presented as Mean $\pm$ SEM.

(HA), hepatic vein (HV), and bile duct (BD).

5.2 Kidney

Histological analysis of the kidney sections from rats administered the varying doses of the extract revealed essentially normal histology, similar to the control group. Sections of the kidney in the various groups showed normal renal histomorphology, with no indications of glomerular damage or acute or chronic tubular damage (Plates 2a, b, c, and d). Normal glomeruli in their respective Bowman's capsules were observed, surrounded by numerous normal renal tubules (proximal convoluted tubules, pars recta, distal convoluted tubules, and collecting ducts) in a highly vascularized connective tissue matrix.

Group 1 (Control)

Plate 2a: The photomicrograph of the sectioned kidney from the control group displayed normal renal histomorphology. Normal glomeruli within their respective Bowman's capsules were seen, surrounded by numerous normal renal tubules in a highly vascularized connective tissue matrix. *H.E x200 magnification / Bar: 50 μ m*. Glomeruli (G); Renal tubules (arrow); Blood capillary (BC).

Group 2 (200 mg/kg)

Plate 2b: The photomicrograph of the sectioned kidney from the 200 mg/kg treated group displayed normal renal histomorphology. Normal glomeruli

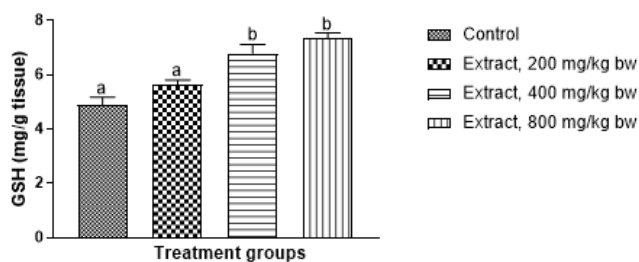


Figure 21: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on GSH of kidney homogenate supernatant in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P \leq 0.05$). Values are presented as Mean $\pm$ SEM.

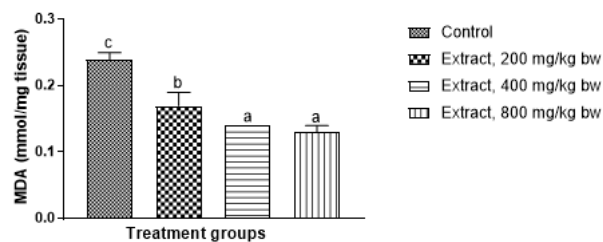


Figure 23: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on MDA of kidney homogenate supernatant in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P \leq 0.05$). Values are presented as Mean $\pm$ SEM.

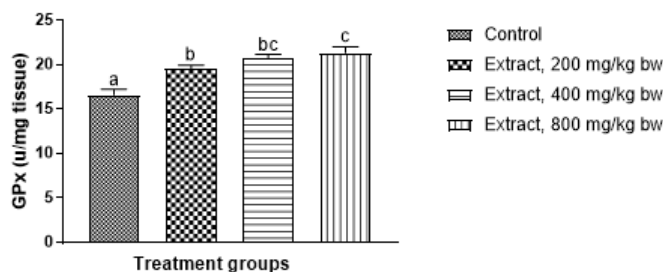


Figure 22: Effect of different doses of aqueous extract of *Jatropha tanjorensis* leaf on GPx of kidney homogenate supernatant in male Wistar rats. Data sharing the same lower-case letters are not significantly different ($P \geq 0.05$), whereas data with different lower-case letters are significantly different ($P \leq 0.05$). Values are presented as Mean $\pm$ SEM.

Group 4 (800 mg/kg)

Plate 2d: The photomicrograph of the sectioned kidney from the 800 mg/kg treated group displayed normal renal histomorphology. Normal glomeruli within their respective Bowman's capsules were seen, surrounded by numerous normal renal tubules in a highly vascularized connective tissue matrix. *H.E x200 magnification / Bar: 50 μ m*. Glomeruli (G); Renal tubules (arrow); Blood capillary (BC).

6 Discussion

The phytochemical evaluation of *Jatropha tanjorensis* extract reveals a rich presence of bioactive compounds with diverse pharmacological properties. The extract contains a diverse array of phytochemicals, including alkaloids, tannins, flavonoids, phenols, terpenoids, saponins, cardiac glycosides, and steroids. This finding aligns with those reported by Ebenyi et al. (2021) regarding the phytochemical analysis of aqueous extract of *Jatropha tanjorensis* leaf, as well as by Igbinaduwa et al. (2011) and Oyewole et al. (2011) concerning the phytochemical analysis of the methanol extract of *Jatropha tanjorensis* leaf.

In this study, alkaloids are present in moderate amounts. Medicinal plants generally contain alkaloids in the range of 10 to 50 mg per 100 grams. This extract, with an alkaloid content of 27.15 mg per 100 grams, falls within the typical range, suggesting a moderate level that could effectively provide therapeutic benefits like pain relief and anticancer activity without being excessively toxic. Alkaloids

within their respective Bowman's capsules were seen, surrounded by numerous normal renal tubules in a highly vascularized connective tissue matrix. *H.E x200 magnification / Bar: 50 μ m*. Glomeruli (G); Renal tubules (arrow); Blood capillary (BC).

Group 3 (400 mg/kg)

Plate 2c: The photomicrograph of the sectioned kidney from the 400 mg/kg treated group displayed normal renal histomorphology. Normal glomeruli within their respective Bowman's capsules were seen, surrounded by numerous normal renal tubules in a highly vascularized connective tissue matrix. *H.E x200 magnification / Bar: 50 μ m*. Glomeruli (G); Renal tubules (arrow); Blood capillary (BC).

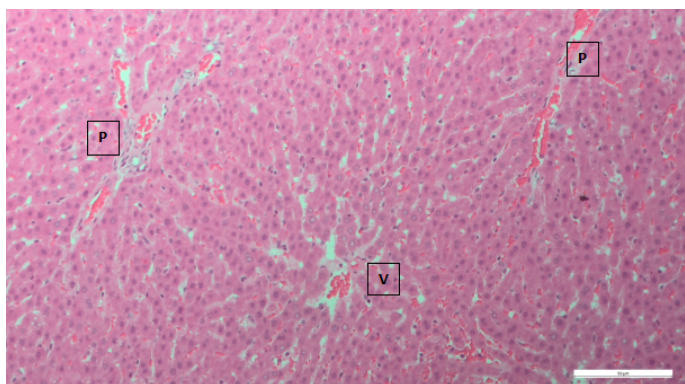


Figure 24: Plate 1a: Photomicrograph of the sectioned liver from the control group, revealing normal hepatic histomorphology. The hepatic lobules and portal triads appear normal. *H.E x200 magnification / Bar: 50 μ m*. Central veins (V); Portal area (P) consisting of hepatic artery (HA), hepatic vein (HV), and bile duct (BD).

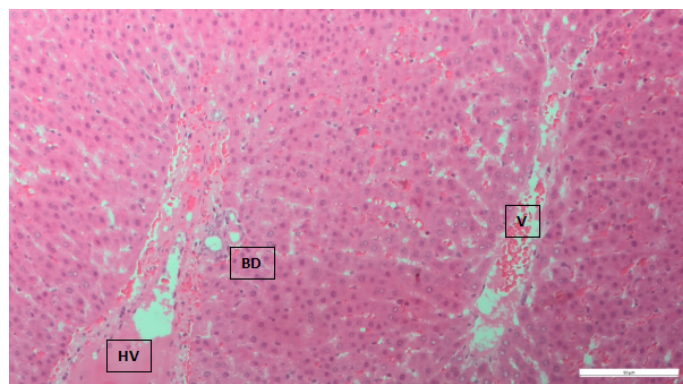


Figure 25: Plate 1b: Photomicrograph of the sectioned liver from the 200 mg/kg treated group, revealing normal hepatic histomorphology. The hepatic lobules and portal triads appear normal. *H.E x200 magnification / Bar: 50 μ m*. Central veins (V); Portal area (P) consisting of hepatic artery (HA), hepatic vein (HV), and bile duct (BD).

are recognized for their analgesic, antimalarial, antimicrobial, anticancer, and anti-inflammatory properties (Heinrich et al., 2021).

Plants known for their astringent and antimicrobial qualities typically contain tannin levels ranging from 5 to 20 mg/100g. The 7.63 mg/100g tannin content in this extract falls towards the lower limit of this range, indicating its potential to deliver advantageous antimicrobial and antioxidant benefits while reducing the risk of excessive astringency. Tannins exhibit astringent, antioxidant, anti-inflammatory, and antimicrobial characteristics (Kováč et al., 2022).

Flavonoids are powerful antioxidants known for their anti-inflammatory, antiviral, and anticancer properties (Hasnat et al., 2024). In medicinal plants, flavonoid content generally ranges from 20 to 50 mg/100g. With a flavonoid content of 39.18 mg/100g, this extract is on the higher end of the spectrum, suggesting it has significant antioxidant and anti-inflammatory potential. This makes the extract beneficial for preventing diseases related to oxidative stress and for managing inflammation.

Plants recognized for their potent antioxidant properties usually have phenol content ranging from 30 to 70 mg/100g. This extract, with a phenol content of 46.07 mg/100g, falls within this typical range, indicating substantial antioxidant properties that can aid in the prevention and management

of diseases linked to oxidative stress, such as cardiovascular diseases and cancer. Phenols are powerful antioxidants that protect against oxidative damage and help reduce the risk of chronic diseases (Kostić et al., 2023).

Typically, medicinal plants have terpenoid levels ranging from 1 to 10 mg/100g. The terpenoid content of 5.41 mg/100g in this extract falls within this anticipated range, suggesting potential benefits in reducing inflammation, fighting infections, and possibly inhibiting cancer cell growth. Terpenoids exhibit anti-inflammatory, anticancer, and antimicrobial properties (Masyita et al., 2022).

Plants known for their health benefits typically contain saponin levels ranging from 20 to 60 mg/100g. With a saponin content of 33.10 mg/100g falling within this usual range, the extract shows promise in managing cholesterol levels, boosting immune function, and potentially preventing cancer. Saponins are recognized for their ability to lower cholesterol, enhance immune response, and exhibit anticancer properties (Juang & Liang, 2020).

Cardiotonic plants typically have cardiac glycoside levels between 1 and 5 mg/100g. This extract contains 4.83 mg/100g of cardiac glycosides, which falls towards the higher end of the usual range. This suggests a robust potential for cardiotonic effects that can be beneficial in the management of heart failure and arrhythmias (Škubník et al., 2021). Cardiac

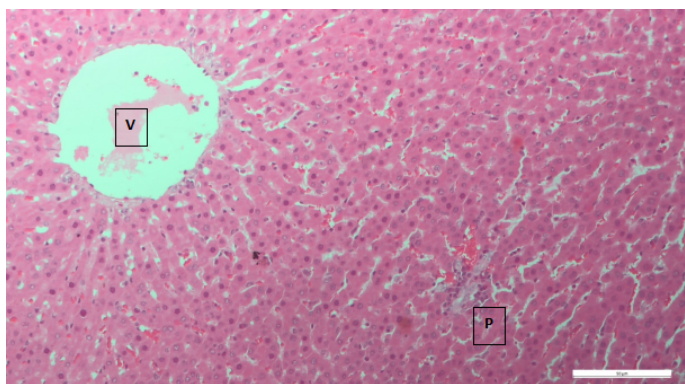


Figure 26: Plate 1c: Photomicrograph of the sectioned liver from the 400 mg/kg treated group, revealing normal hepatic histomorphology. The hepatic lobules and portal triads appear normal. *H.E x200 magnification / Bar: 50 μ m*. Central veins (V); Portal area (P) consisting of hepatic artery (HA), hepatic vein (HV), and bile duct (BD).

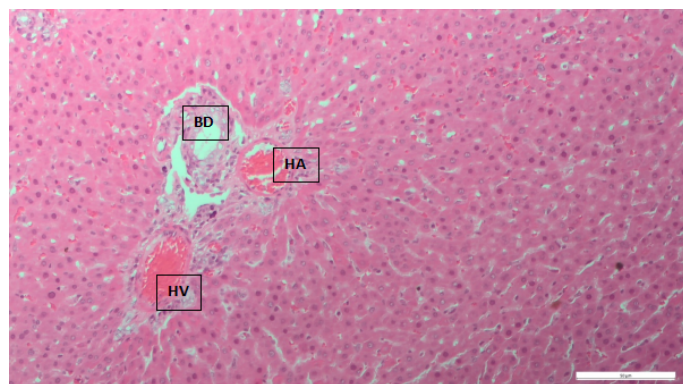


Figure 27: Plate 1d: Photomicrograph of the sectioned liver from the 800 mg/kg treated group, revealing normal hepatic histomorphology. The hepatic lobules and portal triads appear normal. *H.E x200 magnification / Bar: 50 μ m*. Central veins (V); Portal area (P) consisting of hepatic artery (HA), hepatic vein (HV), and bile duct (BD).

glycosides are employed in treating heart conditions by fortifying cardiac contractions and regulating heart rhythm.

Plants with anti-inflammatory and immunomodulatory effects typically have steroid levels ranging from 1 to 10 mg/100g. Steroids are known for their anti-inflammatory and immunomodulatory properties (Dembitsky, 2023). The extract's steroid content of 3.80 mg/100g falls within this anticipated range, indicating potential advantages in reducing inflammation and modulating immune responses. This could be valuable in treating inflammatory and autoimmune conditions like rheumatoid arthritis and type 1 diabetes.

The acute toxicity study of *Jatropha tanjorensis* in Wistar rats provided valuable insights into the safety profile of the plant extract. The absence of mortality and maintained physical stability at doses up to 6000 mg/kg indicates that *Jatropha tanjorensis* extract is relatively non-toxic at lower doses. This suggests a high margin of safety for potential therapeutic use within this dose range. At 7000 mg/kg, a significant increase in mortality rate (33.33%) indicates that toxicity begins to manifest at this dose. While the majority of mice survived, the onset of mortality at this level signals a threshold for adverse effects, highlighting the importance of dose regulation. Complete mortality at 10,000 mg/kg demonstrates that this dose is highly toxic and

lethal. This drastic increase in mortality confirms a critical toxic threshold and emphasizes that doses approaching this level are dangerous and unsafe for use. The LD50 value (the dose at which 50% of the test population is expected to die) of 7745.97 mg/kg indicates the median lethal dose. This relatively high LD50 value suggests that the extract has a broad therapeutic index and is relatively safe at lower doses. The LD50 value is crucial for determining the safety margin and guiding dosage levels for further studies and potential therapeutic applications. This finding is consistent with the report by Ebenyi et al. (2021), which observed no behavioural changes or mortality at oral doses of 2,000 and 5,000 mg/kg within the first 24 hours and during a 14-day study period following administration of an aqueous extract of *Jatropha tanjorensis* leaf. Similarly, Igbinaaduwa et al. (2011) reported no mortality or signs of behavioural changes or toxicity in mice and rats after administering methanol extract of *Jatropha tanjorensis* leaf at doses up to 8,000 mg/kg body weight.

ALT (Alanine aminotransferase) and AST (Aspartate aminotransferase) are enzymes that are commonly used as biomarkers for liver function (McGill, 2016; Chinnappan et al., 2023). The lack of significant ($p < 0.05$) differences in serum ALT and AST levels suggests that the extract, at the administered doses, does not cause hepatocellular damage or stress. This outcome aligns with Ezendiokwere et al.'s

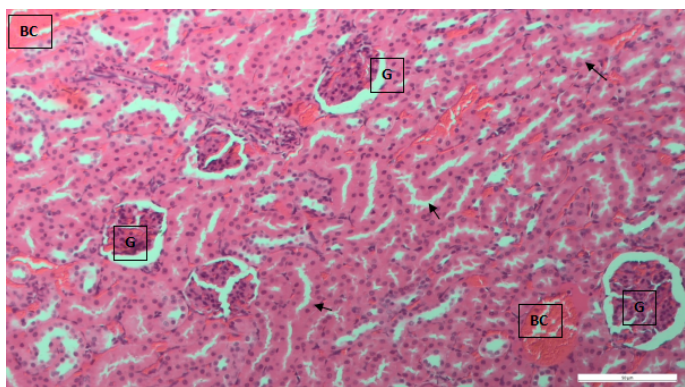


Figure 28: Plate 2a: Photomicrograph of the sectioned kidney from the control group displaying normal renal histomorphology. Normal glomeruli within their respective Bowman's capsules are seen, surrounded by numerous normal renal tubules in a highly vascularized connective tissue matrix. *H.E x200 magnification / Bar: 50 μ m*. Glomeruli (G); Renal tubules (arrow); Blood capillary (BC).

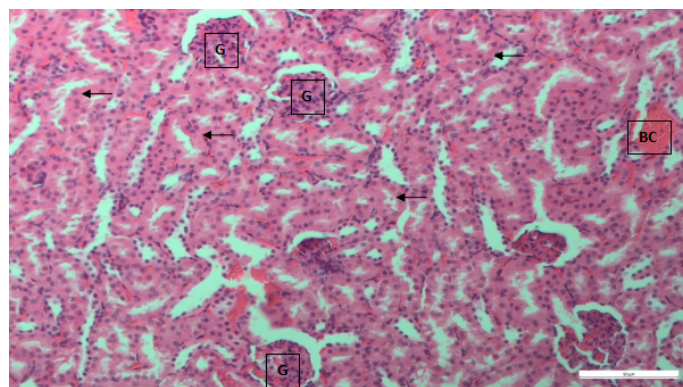


Figure 29: Plate 2b: Photomicrograph of the sectioned kidney from the 200 mg/kg treated group displaying normal renal histomorphology. Normal glomeruli within their respective Bowman's capsules are seen, surrounded by numerous normal renal tubules in a highly vascularized connective tissue matrix. *H.E x200 magnification / Bar: 50 μ m*. Glomeruli (G); Renal tubules (arrow); Blood capillary (BC).

2023 study, which examined the Acute and Subacute Toxicity Profile of Diethyl ether Extract from *Jatropha tanjorensis* leaf in Male Wistar Rats. This indicates that the extract is not hepatotoxic. This implies that the extract's hepatoprotective properties may stem from its tannin content. Tannins are recognized for their antioxidant abilities, capable of scavenging free radicals and mitigating oxidative stress in cells, including those in the liver. By shielding liver cells from oxidative harm, tannins may play a role in preserving the typical levels of ALT and AST (Sobeh et al., 2017; Abu et al., 2022). While the precise mechanisms remain somewhat unclear, it's theorized that tannins might influence enzyme activity in liver cells, including ALT and AST, contributing to the maintenance of serum levels within normal ranges.

ALP (Alkaline phosphatase) is another enzyme associated with liver function (McGill, 2016; Chinnappan et al., 2023), but it can also indicate bone and other tissue-related activity (Fernandez, 2007; Haarhaus et al., 2022). The significant ($p < 0.05$) reduction in ALP levels suggests that the extract might have an inhibitory effect on ALP activity. This result is consistent with the findings of Chibuogwu et al. (2021) and Ezendiokwere et al. (2023). The presence of the phytochemicals (flavonoids, tannins, saponins, and terpenoids), through their combined antioxidant, anti-inflammatory, and

enzyme-modulating properties, could be responsible for the significant reduction in ALP levels observed in the study. This effect can be beneficial in managing conditions where elevated ALP levels are a concern, such as liver diseases or bone disorders (Singab et al., 2005; Sobeh et al., 2017; Abu et al., 2022; Wang et al., 2019). The dose-dependent response, with the most significant reduction at 400 mg/kg, indicates that the extract's effect is potent and potentially dose-related.

Total protein and albumin are indicators of nutritional status and liver function (Keller, 2019). An increase in these parameters suggests that the extract might have a positive effect on protein synthesis or overall nutritional status. This could indicate an anabolic effect or improved liver function in terms of protein production. This result also aligns with the findings of Ezendiokwere et al. (2023).

Bilirubin is a byproduct of the breakdown of red blood cells and is processed by the liver (Guerra Ruiz et al., 2021). Normal levels of bilirubin indicate proper liver function and red blood cell turnover. The lack of significant changes in bilirubin levels further supports the conclusion that the extract does not adversely affect liver function at the administered doses.

The significant reduction in serum cholesterol,

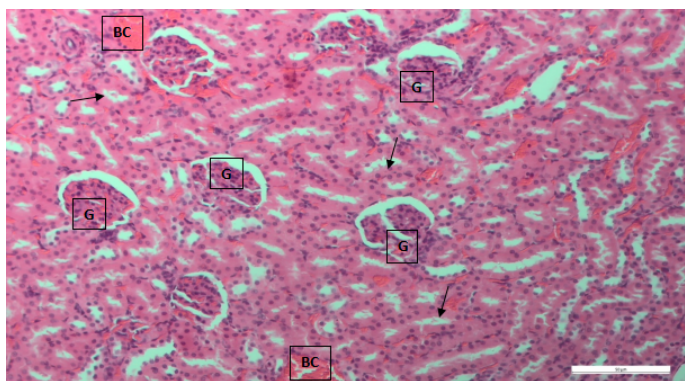


Figure 30: Plate 2c: Photomicrograph of the sectioned kidney from the 400 mg/kg treated group displaying normal renal histomorphology. Normal glomeruli within their respective Bowman's capsules are seen, surrounded by numerous normal renal tubules in a highly vascularized connective tissue matrix. *H.E x200 magnification / Bar: 50 μm*. Glomeruli (G); Renal tubules (arrow); Blood capillary (BC).

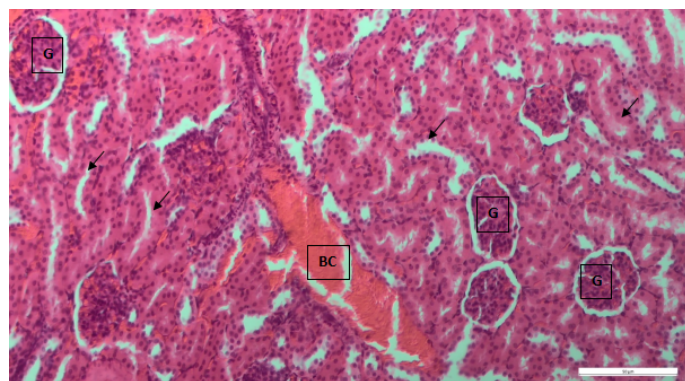


Figure 31: Plate 2d: Photomicrograph of the sectioned kidney from the 800 mg/kg treated group displaying normal renal histomorphology. Normal glomeruli within their respective Bowman's capsules are seen, surrounded by numerous normal renal tubules in a highly vascularized connective tissue matrix. *H.E x200 magnification / Bar: 50 μm*. Glomeruli (G); Renal tubules (arrow); Blood capillary (BC).

triglycerides, and LDL levels in the treated groups suggests that the extract has a lipid-lowering effect. This outcome aligns with the report of Akinloye et al. (2017) which examined the acute toxicity and antioxidant effects of *Jatropha tanjorensis* methanol extract in Wistar rats. This result is consistent with the hypolipidemic activity of several of the phytochemicals present in the extract, such as flavonoids and saponins. These compounds are known to have lipid-lowering properties, possibly through mechanisms involving inhibition of lipid absorption, enhanced lipid metabolism, or modulation of lipid-regulating enzymes (Juang & Liang, 2020; Hasnat et al., 2024). The observed reduction in LDL (Low-Density Lipoprotein) levels is particularly notable, as elevated LDL is a known risk factor for cardiovascular diseases. The decrease in LDL levels indicates a potential cardioprotective effect of the extract. The significant reduction in serum cholesterol, triglycerides, and LDL levels with the highest dose (400 mg/kg) demonstrating the most pronounced effect suggests a dose-dependent response. This implies that higher doses of the extract may lead to more substantial improvements in lipid profiles.

HDL (High-Density Lipoprotein) is often referred to as "good cholesterol" because it helps remove other forms of cholesterol from the bloodstream (FERENCE et al., 2017). The lack of significant changes in

HDL levels suggests that while the extract effectively lowers total cholesterol, triglycerides, and LDL, it does not adversely affect HDL levels. This is beneficial because maintaining or increasing HDL levels is generally desirable for cardiovascular health.

The extract's capacity to lower LDL cholesterol without significantly altering HDL levels highlights its potential as a natural remedy for improving lipid profiles and enhancing cardiovascular health.

7 Conclusion

The comprehensive phytochemical and toxicological evaluation of *Jatropha tanjorensis* leaf extract conducted in this study provides valuable insights into its safety profile and potential health benefits. The phytochemical analysis revealed a rich presence of bioactive compounds, including alkaloids, tannins, flavonoids, phenols, terpenoids, saponins, cardiac glycosides, and steroids, which are known for their diverse pharmacological properties. The acute toxicity study in Wistar rats demonstrated that the extract is relatively non-toxic at lower doses, with an LD50 value of 7745.97 mg/kg, indicating a broad therapeutic index. The biochemical analysis of liver function markers (ALT, AST, and ALP) and lipid profile parameters (cholesterol, triglycerides, LDL, and HDL) suggested that the extract does not cause hepatocellular damage and has a lipid-lowering effect,

particularly at higher doses. These findings align with previous studies and support the potential use of *Jatropha tanjorensis* leaf extract as a safe and effective natural remedy for managing various health conditions. Further research is needed to elucidate the precise mechanisms of action and to explore its potential therapeutic applications in clinical settings.

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