



MOSQUITO LARVICIDAL EFFECT OF *CHROZOPHORA ROTTLERI* AGAINST *ANOPHELES STEPHENSI*, *AEDES AEGYPTI* AND *CULEX QUINQUEFASCIATUS* (DIPTERA: CULICIDAE)

S. Sumithira¹, A. Amsath¹, Marimuthu Govindarajan^{2*} and Udaiyan Muthukumaran³

¹Department of Zoology, Khadir Mohideen College, Adirampattinam-614701, Tamil Nadu, India.

²Department of Zoology, Thiru. Vi. Ka. Government Arts College, Tiruvarur-610003, Tamil Nadu, India

³Unit of Vector Control, Phytochemistry and Nanotechnology, Department of Zoology, Annamalai University, Annamalaiagar-608 002, Tamil Nadu, India

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ABSTRACT

In the present study, the larvicidal efficacy of the crude leaf extracts of *C. rottleri* with five different solvents like methanol, Chloroform, benzene, Ethyl acetate and Hexane was tested against the third instar larvae of *Culex quinquefasciatus*, *Aedes aegypti* and *Anopheles stephensi*. The larval mortality was observed after 24 h of treatment. Among the five solvents the maximum efficacy was observed in methanol. The lethal concentration (LC₅₀) values of *C. rottleri* against third instar larvae of *Culex quinquefasciatus*, *Aedes Aegypti* and *Anopheles stephensi* were 142.90, 133.96, and 122.85 ppm respectively. No mortality was observed in controls. The chi-square value were significant at $p < 0.05$ level. The methanol extract of *C. rottleri* showed good larvicidal activity against three vector mosquitoes.

Keywords: *Chrozophora rottleri*, Larvicidal activity, *Culex quinquefasciatus*, *Aedes aegypti*, *Anopheles stephensi*.

INTRODUCTION

Mosquitoes are the most important single group of insects well-known for their public importance, since they act as vector for many tropical and subtropical disease such as dengue fever, yellow fever, chikungunya, malaria, filariasis and encephalitis of different types including, Japanese encephalitis (Youdeowei, 1983). Larviciding is a successful way of reducing mosquito densities in their breeding places before they emerge into adults. Larviciding largely depends on the use of synthetic chemical insecticides – organophosphates (e.g. temephos and fenthion), insect growth regulators (e.g. diflubenzuron and methoprene), etc. Although effective, their repeated use has disrupted natural biological control systems and sometimes resulting in the widespread development of resistance. These problems have warranted the need for developing alternative strategies using eco-friendly products (Tiwarly *et al.*, 2007). These steadily growing problems demand an intensive search for new products that are environmentally safe, target specific and degradable. The above facts prompted us to undertake investigations of some plant species traditionally used as insecticidal agents, as well as

other endangered plant species, with the aim of identifying lead compounds for the development of new plant based insecticidal agents (Govindarajan *et al.*, 2012).

Chrozophora rottleri belongs to Euphorbiaceae family commonly known as Suryavarti. The plant occurs naturally throughout India, Myanmar, Thailand, Andaman Islands, and Central Java: Malesia. *C. rottleri*, an erect hairy annual common waste lands, blossoms profusely from January to April. It is an erect herb with silvery hairs; lower part of stem is naked, upper part hairy and has slender tap-root. The three-lobed leaves are alternative, thick and rugose. The plants are monoecious, the flowers borne in sessile axillary racemes with staminate flowers in upper and pistillate flowers in the lower part of raceme (Srivastava and Agarwal, 1953). *C. rottleri* is traditionally used by the tribes and native medical practitioners for the treatment of various diseases. In Sudan, powdered stems or whole plants are applied to wounds to improve healing. In Ethiopia, an infusion of the seeds and leaves is taken as a laxative. The plant is also used medicinally in Saudi Arabia, Pakistan and India (e.g. against jaundice and purifying blood). In

*Corresponding author: Dr. M. Govindarajan, Assistant Professor, Department of Zoology, Thiru. Vi. Ka. Government Arts College, Tiruvarur 610003, Tamil Nadu, India, Email: drgovind1979@gmail.com, Mobile: +91 9585265999.

Senegal, the plant is not browsed by most stock, except occasionally by sheep and goats, as it causes vomiting and diarrhoea, where as in Kenya, camels graze it. The fruits yield a purplish blue dye, which is used in East Africa to dye mats (Prota11 (1): Medicinal plants, cited 2010). In Nepal, juice of the fruit is given in cases of cough and colds (Manandhar and Manandhar, 2002), purifying agent (leaf) and laxative (seed), having bioactive components (Singh, 2010). The leaves are very much beneficial in treating skin diseases and also used as a depurative agent (Khare, 2007). The seeds are used as cathartic like Ghodtapde (Sasinath, 2007). Priyanka *et al.* (2010), reported that the aqueous extract of the leaves of this plant have significant anti-helmintic property against *Pheritima posthuma* (Indian Earth worm). Aqueous extract of this plant possessed phytotoxic activity on rice, wheat and mustard. In an experimental study by Suparna and Tapaswi (1999), the leaf extracts exhibited higher inhibition of shoot, root and radial elongation than the stem and root. The aim of the present study was to evaluate the larvicidal activity of different solvent extracts of *Chrozophora rottleri* against *Culex quinquefasciatus*, *Aedes aegypti* and *Anopheles stephensi*.

MATERIALS AND METHODS

Collection of Plants

Fully developed leaves of the *C. rottleri* were collected from Nilgiris, Western Ghats (11° 10'N to 11° 45' N latitude and 76° 14'E to 77° 2' E longitude), Tamil Nadu State, India. The identity was confirmed at the Department of Botany, Annamalai University, Annamalai Nagar, Tamil Nadu. Voucher specimens were numbered and kept in our laboratory and are available upon request.

Extraction

The leaves were washed with tap water, shade dried and finely ground. The finely ground plant material (3.0 kg/solvent) was loaded in Soxhlet apparatus and was extracted with five different solvents namely benzene, hexane, ethyl acetate, chloroform and methanol individually. The solvent from the extract was removed using a rotary vacuum evaporator to collect the crude extract. The crude residue of this plant varies with the solvents used. The *C. rottleri* with five different solvents yielded 84.60, 107.38, 99.37, 114.96 and 136.28 gm of crude residue respectively. Standard stock solutions were prepared at 1% by dissolving the residues with ethanol. From this stock solution, different concentrations were prepared and these solutions were used for ovicidal and repellent activity.

Test Organisms

Laboratory-bred pathogen-free strains of mosquitoes were reared in the vector control laboratory, Department of Zoology, Annamalai University. At the time of adult

feeding, these mosquitoes were 3–4 days old after emergences (maintained on raisins and water) and were starved for 12 h before feeding. Each time, 500 mosquitoes per cage were fed on blood using a feeding unit fitted with Parafilm as membrane for 4 h. *Ae. aegypti* feeding was done from 12:00 p.m. to 4:00 p.m. and *An. stephensi* and *Cx. quinquefasciatus* were fed during 6:00 p.m. to 10:00 p.m. A membrane feeder with the bottom end fitted with Parafilm was placed with 2.0 ml of the blood sample (obtained from a slaughter house by collecting in a heparinized vial and stored at 4 °C) and kept over a netted cage of mosquitoes. The blood was stirred continuously using an automated stirring device, and a constant temperature of 37 °C were maintained using a water jacket circulating system. After feeding, the fully engorged females were separated and maintained on raisins. Mosquitoes were held at 28±2 °C, 70–85 % relative humidity, with a photo period of 12-h light and 12-h dark.

Larvicidal Bioassay

The larvicidal activity of the leaf extract of *C. rottleri* was evaluated as per the method recommended by WHO (1996). Different concentration of the test samples were used. 500 ml glass beaker containing 250 ml of tap water. Early third instar larvae of *Culex quinquefasciatus*, *Aedes aegypti* and *Anopheles stephensi* were introduced to each of the test solution as well as control. Control consists of acetone only. For each experiment six replicates were maintained at a time. The lethal concentration (LC₅₀) values were calculated after 24 h by probit analysis (Finney, 1971).

Statistical Analysis

Statistical evaluation was done using Statistical Package of Social Sciences (SPSS) 13.0 for windows, Significance level was set at p<0.05.

RESULTS

The toxicity of *C. rottleri* was tested against the early third instar larvae of *Culex quinquefasciatus*, *Aedes aegypti* and *Anopheles stephensi*. The data were recorded and statistical data regarding the LC₅₀, LC₉₀, regression equation, Chi-square and 95% confidence limits were calculated (Tables I, II, III.). The methanolic extract of *C. rottleri* showed highest larvicidal activity against *Culex quinquefasciatus*, *Aedes aegypti* and *Anopheles stephensi*. The LC₅₀ values of *C. rottleri* against early third instar larvae of *Culex quinquefasciatus*, *Aedes aegypti* and *Anopheles stephensi* were 142.90, 133.96 and 22.85 ppm, respectively. No mortality was observed in control. The chisquare value were significant at p <0.05 level. In the present study third instar was highly sensitive when compared with larvae of the three vector mosquitoes.

Table 1. Larvicidal activity of different solvent extracts of *Chrozophora rottleri* against *Anopheles stephensi*.

Solvents	Concentration (ppm)	% of mortality $\pm$ SD	LC ₅₀ (ppm) (LCL-UCL)	LC ₉₀ (ppm) (LCL-UCL)	χ^2
Hexane	70	19.3 $\pm$ 1.2	173.01 (157.90-187.13)	315.54 (293.68-344.16)	2.978 n.s.
	140	38.6 $\pm$ 0.6			
	210	61.2 $\pm$ 0.4			
	280	79.5 $\pm$ 1.2			
	350	97.4 $\pm$ 0.8			
Ethyl acetate	70	22.9 $\pm$ 0.6	163.28 (147.23-177.92)	310.48 (288.29-339.69)	1.105 n.s.
	140	40.2 $\pm$ 0.8			
	210	64.8 $\pm$ 1.2			
	280	83.1 $\pm$ 0.4			
	350	96.4 $\pm$ 1.2			
Benzene	70	25.4 $\pm$ 0.4	154.07 (137.52-168.89)	300.68 (278.87-329.39)	1.350 n.s.
	140	43.9 $\pm$ 0.6			
	210	66.2 $\pm$ 1.2			
	280	85.7 $\pm$ 0.8			
	350	97.3 $\pm$ 1.2			
Chloroform	70	28.3 $\pm$ 0.8	143.77 (127.37-158.28)	282.74 (262.31-309.54)	2.840 n.s.
	140	46.2 $\pm$ 1.2			
	210	69.7 $\pm$ 0.6			
	280	88.9 $\pm$ 0.4			
	350	99.1 $\pm$ 1.2			
Methanol	60	29.6 $\pm$ 1.2	122.85 (108.24-135.68)	246.67 (228.44-270.73)	4.404 n.s.
	120	47.8 $\pm$ 0.8			
	180	66.3 $\pm$ 0.4			
	240	88.6 $\pm$ 1.2			
	300	99.1 $\pm$ 0.6			

^a Values are mean $\pm$ SD of five replicates.

No mortality was observed in the control.

SD = standard deviation.

LC₅₀= lethal concentration that kills 50% of the exposed organisms.

LC₉₀= lethal concentration that kills 90% of the exposed organisms.

UCL= 95% upper confidence limit.

LCL= 95% lower confidence limit.

χ^2 = chi square.

n.s. = not significant ($\alpha=0.05$).

Table 2. Larvicidal activity of different solvent extracts of *Chrozophora rottleri* against *Aedes aegypti*.

Solvents	Concentration (ppm)	% of mortality $\pm$ SD	LC ₅₀ (ppm) (LCL-UCL)	LC ₉₀ (ppm) (LCL-UCL)	χ^2
Hexane	70	19.7 $\pm$ 0.8	176.33 (159.95-191.56)	333.92 (309.28-366.83)	1.512 n.s.
	140	40.5 $\pm$ 0.4			
	210	58.2 $\pm$ 1.2			
	280	77.6 $\pm$ 0.6			
	350	94.3 $\pm$ 1.2			
Ethyl acetate	70	21.6 $\pm$ 0.6	172.49 (156.39-187.41)	325.37 (301.78-356.66)	1.425 n.s.
	140	38.2 $\pm$ 1.2			
	210	60.4 $\pm$ 0.4			
	280	79.8 $\pm$ 0.8			
	350	95.3 $\pm$ 1.2			
Benzene	70	24.8 $\pm$ 0.4	160.78 (143.95-175.98)	314.29 (291.11-345.08)	1.410 n.s.
	140	41.6 $\pm$ 1.2			
	210	63.9 $\pm$ 0.6			
	280	82.7 $\pm$ 0.8			
	350	96.2 $\pm$ 1.2			
Chloroform	70	26.7 $\pm$ 1.2	151.77 (134.91-166.78)	299.95 (277.96-329.02)	1.889 n.s.
	140	44.5 $\pm$ 0.6			
	210	66.2 $\pm$ 0.8			
	280	85.8 $\pm$ 0.4			
	350	97.6 $\pm$ 1.2			
Methanol	60	26.8 $\pm$ 0.8	133.96 (119.39-147.00)	264.70 (245.05-290.81)	3.067 n.s.
	120	43.2 $\pm$ 0.6			
	180	62.9 $\pm$ 1.2			
	240	84.1 $\pm$ 0.4			
	300	97.4 $\pm$ 0.8			

^a Values are mean $\pm$ SD of five replicates.

No mortality was observed in the control.

SD = standard deviation.

LC₅₀= lethal concentration that kills 50% of the exposed organisms.

LC₉₀= lethal concentration that kills 90% of the exposed organisms.

UCL= 95% upper confidence limit.

LCL= 95% lower confidence limit.

χ^2 = chi square.

n.s. = not significant ($\alpha=0.05$).

Table 3. Larvicidal activity of different solvent extracts of *Chrozophora rotleri* against *Culex quinquefasciatus*.

Solvents	Concentration (ppm)	% of mortality $\pm$ SD	LC ₅₀ (ppm) (LCL-UCL)	LC ₉₀ (ppm) (LCL-UCL)	χ^2
Hexane	70	21.8 $\pm$ 1.2	178.91 (162.79-194.03)	335.63 (310.89-368.68)	4.803 n.s.
	140	36.3 $\pm$ 0.8			
	210	57.5 $\pm$ 0.6			
	280	74.9 $\pm$ 0.4			
	350	96.2 $\pm$ 1.2			
Ethyl acetate	70	20.4 $\pm$ 0.8	174.58 (159.29-188.91)	320.17 (297.68-349.74)	3.690 n.s.
	140	37.6 $\pm$ 0.4			
	210	59.2 $\pm$ 1.2			
	280	78.9 $\pm$ 0.6			
	350	97.3 $\pm$ 1.2			
Benzene	70	23.6 $\pm$ 1.2	164.01 (148.16-178.54)	309.98 (287.90-339.01)	2.125 n.s.
	140	39.8 $\pm$ 0.8			
	210	61.4 $\pm$ 0.6			
	280	85.1 $\pm$ 0.4			
	350	96.5 $\pm$ 1.2			
Chloroform	70	24.9 $\pm$ 0.6	157.94 (141.61-172.68)	304.97 (282.91-334.08)	3.302 n.s.
	140	42.5 $\pm$ 0.4			
	210	65.3 $\pm$ 1.2			
	280	82.7 $\pm$ 0.8			
	350	98.2 $\pm$ 1.2			
Methanol	60	24.6 $\pm$ 0.4	142.90 (128.39-156.13)	278.57 (257.71-306.47)	2.179 n.s.
	120	40.4 $\pm$ 0.8			
	180	59.2 $\pm$ 1.2			
	240	81.7 $\pm$ 0.6			
	300	95.3 $\pm$ 0.4			

^a Values are mean $\pm$ SD of five replicates.

No mortality was observed in the control.

SD = standard deviation.

LC₅₀= lethal concentration that kills 50% of the exposed organisms.

LC₉₀= lethal concentration that kills 90% of the exposed organisms.

UCL= 95% upper confidence limit.

LCL= 95% lower confidence limit.

χ^2 = chi square.

n.s. = not significant ($\alpha=0.05$).

DISCUSSION

The present findings corporate earlier findings of Macedo *et al.* (1997) who showed that ethanol extract of *Tagetes patula* was less active and only 50% larvae were killed at higher concentration (100 ppm). Methanolic leaf extract of *Cassia fistula* was tested for larvicidal activity against *Culex quinquefasciatus* and *Anopheles stephensi* with LC₅₀ values of 17.97 and 20.57 mg/l, respectively (Govindarajan *et al.*, 2008a). The petroleum ether fraction of *Acacia*

nolotica and *Citrullus colocynthis* showed 100 per cent mortality in 100, 250 and 500 ppm and 60 and 50 percent mortality at 125 and 62.5 ppm respectively against *Culex quinquefasciatus*.

The leaf extract of *Acalypha in dica* with different solvents viz, benzene, chloroform, ethyl acetate and methanol were tested for larvicidal, ovicidal activity and oviposition attractancy against *Anopheles stephensi*. The larval mortality was observed after 24 h exposure. The

LC₅₀ values are 19.25, 27.76, 23.26 and 15.03 ppm, respectively (Govindarajan et al., 2008b). The leaf extract of *Cassia fistula* with different solvents viz, methanol, benzene and acetone were studied for the larvicidal, ovicidal and repellent activity against *Aedes aegypti*. The 24 h LC₅₀ concentration of the extract against *Aedes aegypti* were observed at 10.69, 18.27 and 23.95 mg/l respectively (Govindarajan, 2009).

Cheng et al. (2003) reported that the leaf and bark essential oil of *Cryptomeria japonica* showed larvicidal activity against *Aedes aegypti*. Methanolic fraction of leaves of *Mentha piperita*, *Phyllanthus niruri* and *Letiota aspera* exhibited the LC₅₀ values of 43.65, 1819.70 and 2818.38 respectively against the larvae of *Culex quinquefasciatus* (Pandian et al., 1994). Singh et al. (2003) reported the mosquito larvicidal properties of the leaf extract of a herbaceous plant *Ocinum canum* against *Aedes aegypti*. The LC₅₀ values for 2nd, 3rd and 4th larvae were 177.82, 229.08 and 331.13 ppm respectively. Gusmao et al. (2002) reported that the extract of *Derris urucu* showed larvicidal activity against *Aedes aegypti* with LC₅₀ values of 17.6 µg/ml. Muthukrishnan et al. (1997) reported that ethyl acetate fractions of *Solanum trilobatum* and *Letiota aspera* showed the LC₅₀ values of 23.5 and 138.6 ppm against 2nd and 3rd larvae of *Culex quinquefasciatus*.

Yang et al. (2002) reported that *Piper longum* fruit-isolated pipernonaline had strong larvicidal effects against the 4th stage larvae of *Aedes aegypti*. The LC₅₀ value of pipernonaline was 0.25 mg/L against *Aedes aegypti*. Kabaru and Gichia, (2001) reported that the mangrove plant *Rhizophora mucronata* bark and pith extract showed high toxicity with LC₅₀ values of 157.4 and 168.3 ppm respectively against *Aedes aegypti* larvae.

CONCLUSION

In conclusion, the present study showed that extracts from *C. rotterli* can be effectively used in the control of *Cx. quinquefasciatus*, *Ae. aegypti* and *An. stephensi* mosquito larvae. However, further studies on the identification of the active principles involved and their mode of action and field trials need presently other investigations.

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