

The identification of a cannabinoid-like compound (adrenoyl-EA), in the flowers of a South African medicinal plant (*Leonotis leonurus*)

Ethan Hunter

Stellenbosch University

Marietjie Stander

Stellenbosch University

Jens Kossmann

Stellenbosch University

Suparna Chakraborty

University of Cape Town Department of Biological Sciences

Sharon Prince

University of Cape Town Department of Biological Sciences

Shaun Peters

Stellenbosch University

Bianke Loedolff (✉ bianke@sun.ac.za)

Stellenbosch University Faculty of AgriSciences <https://orcid.org/0000-0001-7519-0257>

Research note

Keywords: Adrenoyl-EA, Traditional medicine, Phytochemistry, Flowers, Cannabis, Phytocannabinoids, *Leonotis leonurus*

Posted Date: September 11th, 2020

DOI: <https://doi.org/10.21203/rs.3.rs-61760/v1>

License: © ⓘ This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

Version of Record: A version of this preprint was published on November 10th, 2020. See the published version at <https://doi.org/10.1186/s13104-020-05372-z>.

Abstract

Objective Current global trends on natural therapeutics suggest an increasing market interest toward the use and discovery of new plant-derived therapeutic compounds, often referred to as traditional medicine (TM). The Cannabis industry is currently one such focal area receiving attention, owing to the occurrence of phytocannabinoids (pCBs) which have shown promise in health-promotion and disease prevention. However, the occurrence of pCBs in other plant species are often overlooked and rarely studied.

Leonotis leonurus (L.) R. Br. is endemic to South Africa with a rich history of use in TM practices amongst indigenous people and, has been recorded to induce mild psychoactive effects akin to Cannabis. While the leaves have been well-reported to contain therapeutic phytochemicals, little information exists on the flowers. Consequently, as part of a larger research venture, we targeted the flowers of *L. leonurus* for the identification of potential pCB or pCB-like compounds.

Results Flower extracts were separated and analyzed using high performance thin layer chromatography (HPTLC). A single pCB candidate was isolated from HPTLC plates and, using liquid chromatography coupled to tandem mass spectrometry (LC-MS/MS), identified as 7,10,13,16-Docosatetraenoylethanolamine (adrenoyl-EA). Adrenoyl-EA is a known bioactive pCB-like compound reported to occur in only one other plant species, *Tropaeolum tuberosum*.

Introduction

Increasing interest from developed economies in the use of TMs has created the second largest global therapeutics market [1, 2]. The medicinal Cannabis industry is currently one such focal area, largely because of the occurrence of phytocannabinoids (pCBs). As part of a unique class of phytochemicals that interact with the human endocannabinoid system (ECS), the use of pCBs as therapeutics holds promise in the treatment of numerous chronic diseases [3–8]. However, many countries still abide by strict regulatory laws pertaining to the commercial growth of Cannabis and, consequently research into pharmacologically relevant products has until recently, been relatively protracted. Since the discovery of pCBs, similar compounds (pCB-like) have been reported in several plant species, beyond Cannabis, that are commonly used in TM practices [9]. The growing interest in the use of TMs enabled an opportunity for the discovery and evaluation of new compounds from medicinal and nonmedicinal plants for the development of natural therapeutics.

The use of the ‘wild cannabis’ plant, *L. leonurus* (L.) R. Br., endemic to South Africa, is commonly reported in TM to treat numerous ailments including eczema, headaches, hypertension, and chest infections [10, 11]. The leaves (when smoked) have been described to elicit similar effects to Cannabis, including mild psychoactive symptoms, as well as the capacity to alleviate anxiety and induce calming effects [10, 12]. Despite the amount of research available on the leaves, little to no information exists on the medicinal properties of the flowers of this plant. Furthermore, the occurrence of pCBs or pCB-like compounds has

never been reported. In this research note, we report on the identification of adrenoyl-EA, a pCB-like compound, in the flowers of the white flower variety of *L. leonurus* (L.) R. Br. var. *albiflora*.

Materials And Methods

Plant material

L. leonurus (L.) R. Br. var. *albiflora* seeds (white flower variety) were obtained from a commercial seed supplier (Seeds for Africa, South Africa) and grown on a residential property at the Bottom Road Sanctuary, Zeekoevlei, Western Cape Province, South Africa (GPS coordinates: -34.057951, 18.499391). Flower samples were harvested at mature stage (Fig. 1, photo courtesy of Dr B Loedolff) and immediately stored at -20°C.

Whole flower metabolite extraction

Flowers were lyophilized (20 h; 100 mTorr, -60°C) and ground to a fine powder using a pestle and mortar. Metabolites were extracted from flower material (50 mg) as previously described [13, 14], evaporated, and the resin reconstituted in 200 µL ddH₂O, calibrating the extracts to 0.50 mg/µL.

Phytocannabinoid screening and isolation using HPTLC

Phytocannabinoid screening was performed on 10 × 20 cm glass-back plates pre-coated with 0.25 mm silicagel 60 (REF 811213, Macherey-Nagel, Germany). Flower derived-extracts (0.50 mg/µL) were applied to the plates in 2 µL increments and allowed to dry completely before placement into a glass chromatography chamber (Camag, Switzerland), pre-conditioned with the mobile phase, chloroform:methanol (9:1; 30 min). Subsequently, the chromatogram was developed in the dark using fast blue B reagent (0.1% w/v; dissolved in 1 mM NaOH). Based on R_f ranges (comparative to the upper R_f values representing the less polar range of pCBs from Cannabis, under the same chromatographic conditions [15]) and change in colour (in the range of orange-red [16]), a single compound of interest was scraped from the preparative HPTLC plate and resuspended in 50% (v/v) methanol, prior to tandem mass spectrometry (LC-MS/MS) analyses.

Tandem mass spectrometry (LC-MS/MS) analyses

LC-MS/MS analyses were performed, as previously described [14], with a Waters Synapt G2 quadrupole time-of-flight mass spectrometer (Waters Corporation, Milford, MA, USA) equipped with a Waters Acquity UPLC. Samples were separated on a Waters UPLC BEH C18 column (2.1 × 100 mm; 1.7 µm) at a flow rate of 0.25 ml/min at 55°C. Solvent A consisted of 0.1% (v/v) formic acid in water and solvent B was 0.1% (v/v) formic acid in acetonitrile. The mobile phase gradient was initiated at 100% solvent A for 1 min and linearly reduced to 28% solvent A over 22 min. Subsequently, the mobile phase was changed to 40% solvent B over 8 min followed by a 1 min wash step in 100% solvent B before the column was re-equilibrated to the initial conditions for 4 min. Electrospray ionization was applied, and samples were

analysed in a negative mode run and a positive mode run. Data was acquired in MS^E mode, which consists of a high collision energy scan range of m/z 125–1500 and a low collision energy scan from m/z 40–1500. The photo diode array detector was set to scan from 220–600 nm. The capillary voltage was set at 3.5 kV and the collision energy either 6 V (low collision energy scan from) or 30–60 V (high collision energy scan), the cone voltage was 15 V, the source temperature 120°C and the desolvation temperature was 275°C. The desolvation and cone gas (nitrogen) flows were 650 L/h and 50 L/h, respectively. Sodium formate was used for calibration and leucine enkephalin was infused in the background as lock mass for accurate mass determinations. Metabolites were monitored using their deprotonated quasi-molecular ions.

Compounds were tentatively identified using the Metabolomics workbench [17]. The database was searched, using the m/z mass obtained from total ion chromatograms, with parameters set to the negative $[M-H]^-$ and positive ion $[M+H]^+$ mode, respectively, and a mass tolerance of ± 0.2 m/z . Tentative identification was based on the accurate mass and fragment ions of the specific peaks compared to literature and was carried out in conjunction with independent metabolite repositories, Metabolomics Workbench, PubChem, and METLIN Metabolite and Chemical Entity Database.

Results And Discussion

L. leonurus (L.) R. Br. is a drought resistant medicinal shrub endemic to South Africa with a bi-annual flowering season (high-yielding medicinal crop). Anecdotally it is known for eliciting mild psychoactive effects akin to the smoking of Cannabis and, has a long-standing history in traditional healing practices in South Africa. Some studies have alluded to its medicinal activities however, these have largely focused on the leaves and only a few reports have dealt with the flowers [10, 11]. Furthermore, there are no reports that describe the occurrence of medicinal pCBs from the leaves or the flowers. Since pCBs have been proposed as effective TMs for the prevention and/or treatment of chronic diseases [18–20], identification of pCBs (or pCB-like compounds) in plants other than Cannabis *spp.* could present an attractive value proposition to complement the emergent Cannabis industry.

Based on its mild Cannabis-like effect when smoked, we conducted analyses into the flower-derived phytochemicals and the potential presence of pCB-like compounds in *L. leonurus* (L.) R. Br. var. *albiflora* (Fig. 1A). Since extensive phytochemical profiling of the leaves have never yielded any pCB-like compounds [10, 11], we suspected that such compounds may be present in flowers (akin to Cannabis). Using HPTLC, coupled to fast blue BB salt for a selective colorimetric detection of pCB compounds [14], a distinct orange spot could be observed (R_f 0.55, Fig. 1B). The fast blue method detects major neutral cannabinoids such as tetrahydrocannabinol (THC), cannabidiol (CBD), and their cannabinoid acid derivatives THCA and CBDA, among other cannabinoids with high sensitivity and selectivity [15, 16].

A single, putative pCB-like compound was isolated, and its identity determined using LC-MS/MS in both positive and negative ionization modes (Fig. 2A, B). Mass spectra from whole flower extracts and the isolated HPTLC compound were compared (Fig. 2C, D) resulting in equivalent retention times and peak

masses in both negative (RT: 30.78 min; $[M-H]^- = 374.26$) and positive (RT: 30.77 min; $[M+H]^+ = 376.26$) ionization mode. Further investigation using the international metabolite repositories, Metabolomics Workbench, PubChem, and METLIN Metabolite and Chemical Entity Database, consistently identified this compound as 7,10,13,16-Docosatetraenylethanolamine ($[M-H]^- = 374.31$; $[M+H]^+ = 376.32$; neutral $m/z = 375.31$), also known as Adrenoyl-ethanolamide (EA). Adrenoyl-EA is a bioactive endocannabinoid previously thought to be unique to mammals [21, 22]. However, it has recently (and for the first time) been reported to occur in methanol extracts of the Mashua plant (*Tropaeolum tuberosum*), extensively used in Andean folk medicine [23]. Our findings similarly demonstrate the occurrence of adrenoyl-EA in acetonitrile extracts from the flowers of *L. leonurus* (L.) R. Br. var. *albiflora*.

Adrenoyl-EA is structurally similar to the major endocannabinoid anandamide and functions as an agonist of the CB_1 and $TRPV_1$ receptor proteins, two major receptors of the human ECS [21, 24]. Agonists of these receptors are ideal candidates in a range of therapeutic targets, specifically exhibiting anti-inflammatory, neuroprotective, and anticancer activities, among others [23, 25, 26]. Within the growing market of TMs, we consider it beneficial to identify sustainable, climate resilient plant resources (high yielding medicinal crops) that accumulate pCBs or PCB-like compounds with the potential for therapeutic purposes.

A South African perspective on the development of pCB products

It is estimated that the domestic South Africa Cannabis market will reach USD \$1.8 billion by 2023 however, this projection is based on export value only, excluding Cannabis-derived pCB products. The value proposition lies within the latter, given the therapeutic potential of pCBs in chronic disease treatment [23–26]. South African agriculture is typified by both large scale mechanized, and smallholder practices and Cannabis cultivation is considered viable. Although the agricultural experience and arable land in South Africa is adequate for the growth and export of Cannabis, the irrigation infrastructure and required daylight might not be sufficient to sustain the production of high quality Cannabis-derived products. One of the major predicted hurdles in establishing a sustainable Cannabis industry is the water-intensive measures required for an efficient pCB yield from Cannabis. If one could exploit a water-efficient medicinal crop capable of producing pCB-like compounds, this would serve as an ideal industry alternative into a realm of “smart-pharming” practices.

As part of a pCB-driven strategy for the development of high value TMs, we suggest that the discovery of pCBs in endemic South African plants could be highly complementary to the Cannabis industry, given that these plants are adapted to the climatic and water-scarce conditions. To our knowledge, we provide the first evidence on the presence of the pCB-like compound, adrenoyl-EA, in the flowers of *L. leonurus* (L.) R. Br. var. *albiflora*. Globally, the production of pCBs from these alternative resources could benefit the future TM market, particularly in countries where agriculture is typified by water scarcity.

Limitations

HPTLC and LC-MS/MS profiles provided secure evidence for the occurrence of the pCB-like compound adrenoyl-EA within the whole flower extract of white *L. leonurus* (L.) R. Br. var. *albiflora*. Further investigations would entail (i) quantification of the identified compound/s with LC-MS/MS using authentic standard compounds and, (ii) determining structural similarities and differences when compared to other plant-, animal- or human-derived adrenoyl-EA, with NMR. However, to support the idea of an alternative and sustainable crop to produce pCB products, it would be beneficial to isolate and identify the other compounds from the HPTLC to uncover the existence of a metabolite pathway capable of producing several different pCB or pCB-like compounds.

Abbreviations

pCB:phytocannabinoid; HPTLC:high performance thin layer chromatography; LC-MS/MS:liquid chromatography linked to tandem mass spectrometry; CM:conventional medicine; TM:traditional medicine; CAGR:compound annual growth rate; CBG:cannabigerol; THC:tetrahydrocannabinol; CBD:cannabidiol; NMR:Nuclear Magnetic Resonance.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Availability of data and materials

Any data, including the full LC-MS/MS datasets, used and/or analysed during the current study are available from the corresponding author (bianke@sun.ac.za) on reasonable request.

Competing interests

Not applicable.

Funding

Not applicable.

Author's contributions

BL and SP¹ conceived the original idea for this research project and dually serve as principal investigators. EH conducted metabolite extractions, HPTLC and LC-MS/MS analyses. MS designed the LC-MS/MS approach as part of the central analytical facility services (CAF) at Stellenbosch University. JK provided funding and intellectual input. SP³ provided intellectual input as a human cancer specialist. All authors (EH, MS, JK, SC, SP¹, SP³, BL) contributed equally to the writing of this research note and approved the final version.

Acknowledgments

Not applicable.

References

1. World Health Organization. (2013). WHO traditional medicine strategy: 2014–2023. WHO. 2013. https://www.who.int/traditional-complementary-integrative-medicine/publications/trm_strategy14_23/en/. Accessed 5 April 2019.
2. Parasuraman S. Herbal drug discovery: Challenges and perspectives. *Curr Pharmacogenomics Person Med*. 2018;63–68.
3. Hazekamp A. The trouble with CBD oil. *Med Cannabis Cannabinoids*. 2018;1:65–72.
4. Bielawiec P, Harasim-Symbor E, Chabowski A. Phytocannabinoids. Useful Drugs for the Treatment of Obesity? Special Focus on Cannabidiol. *Front Endocrinol*. 2020;11:114.
5. Casey S, Vaughan C. (2018). Plant-Based Cannabinoids for the Treatment of Chronic Neuropathic Pain. *Medicines*. 2018;5,3,67.
6. Di Marzo V, Bifulco M, De Petrocellis L. The endocannabinoid system and its therapeutic exploitation. *Nat Rev Drug Discov*. 2004;3:9,771–84.
7. Dinu A, Rogobete A, Bratu T, Popovici S, Bedreag O, Papurica M, Bratu L, Sandesc D. Cannabis sativa revisited – crosstalk between microRNA expression, inflammation, oxidative stress, and endocannabinoid response system in critically ill patients with sepsis. *Cells*. 2020;9:307.
8. Maione S, Costa B, Di Marzo V, Endocannabinoids. A unique opportunity to develop multitarget analgesics. *Pain*. 2013;154:87–93.
9. Hanuš L, Meyer S, Muñoz E, Taglialatela-Scafati O, Appendino G. Phytocannabinoids: a unified critical inventory. *Nat Prod Rep*. 2016;33:1357–92.
10. Mazimba O. Leonotis leonurus: A herbal medicine review. *J Pharmacogn Phytochem*. 2015;3:74–82.
11. Nsuala B, Enslin G, Viljoen A. “Wild cannabis”: A review of the traditional use and phytochemistry of Leonotis leonurus. *J Ethnopharmacol*. 2015;174:520–39.
12. Nsuala B, Kamatou G, Sandasi M, Enslin G, Viljoen A. Variation in essential oil composition of Leonotis leonurus, an important medicinal plant in South Africa. *Biochem Syst Ecol*. 2017;70:155–61.

13. Routaboul JM, Kerhoas L, Debeaujon I, Pourcel L, Caboche M, Einhorn J, Lepiniec L. Flavonoid diversity and biosynthesis in seed of *Arabidopsis thaliana*. *Planta*. 2006;224:96–107.
14. Loedolff B, Brooks J, Stander M, Peters S, Kossmann J. High light bio-fortification stimulates de novo synthesis of resveratrol in *Diplotaxis tenuifolia* (wild rocket) micro-greens. *Funct Food Health Dis*. 2017;7:859–72.
15. Galand N, Ernouf D, Montigny F, Dollet J, Pothier J. Separation and Identification of Cannabis Components by Different Planar Chromatography Techniques (TLC, AMD, OPLC). *J Chromatogr Sci*. 2004;42:130–4.
16. Brenneisen R. Chemistry and Analysis of Phytocannabinoids and Other Cannabis Constituents. *Forensic Sci Med*. 2007; 17–49.
17. Sud M, Fahy E, Cotter D, Azam K, Vadivelu I, Burant C, Edison A, Fiehn O, Higashi R, Nair KS, Sumner S, Subramaniam S. Metabolomics Workbench: An international repository for metabolomics data and metadata, metabolite standards, protocols, tutorials and training, and analysis tools. *Nucleic Acids Res*. 2016;44:D463-D70.
18. Singh R. Medicinal plants: A review. *J Plant Sci*. 2015;3:50–5.
19. Ahn K. Worldwide trend of botanical drug and strategies for developing global drugs. *BMB Rep*. 2017;50:111–6.
20. Katchan V, David P, Shoenfeld Y. Cannabinoids and autoimmune diseases: A systematic review. *Autoimmun Rev*. 2016;15:513–28.
21. Movahed P, Jönsson B, Birnir B, Wingstrand J, Jørgensen T, Ermund A, Högestätt E. Endogenous Unsaturated C18N-Acylethanolamines Are Vanilloid Receptor (TRPV1) Agonists. *J Biol Chem*. 2005;280:46384–96.
22. Zendulkar O, Dovrtelova G, Noskova K, Turjap M, Sulcova A, Hanus L, Jurica J. Cannabinoids and cytochrome P450 interactions. *Curr Drug Metab*. 2016;17:206–26.
23. Luis A, Perez T, Madalina S, Matias A, Angel R. Alkamides from *Tropaeolum tuberosum* inhibit inflammatory response induced by TNF- α and NF- κ B. *J Ethnopharmacol*. 2019;235:199–205.
24. Watson J, Kim J, Das A. Emerging Class of Omega-3 Fatty Acid Endocannabinoids & Their Derivatives. *Prostaglandins Other Lipid Mediat*. 2019;143:106337.
25. Cristino L, Bisogno T, Di Marzo V. Cannabinoids and the expanded endocannabinoid system in neurological disorders. *Nat Rev Neurol*. 2020;16:9–29.
26. Massi P, Solinas M, Cinquina V, Parolaro D. Cannabidiol as potential anticancer drug. *Br J Clin Pharmacol*. 2013;75(2):303–12.

Figures

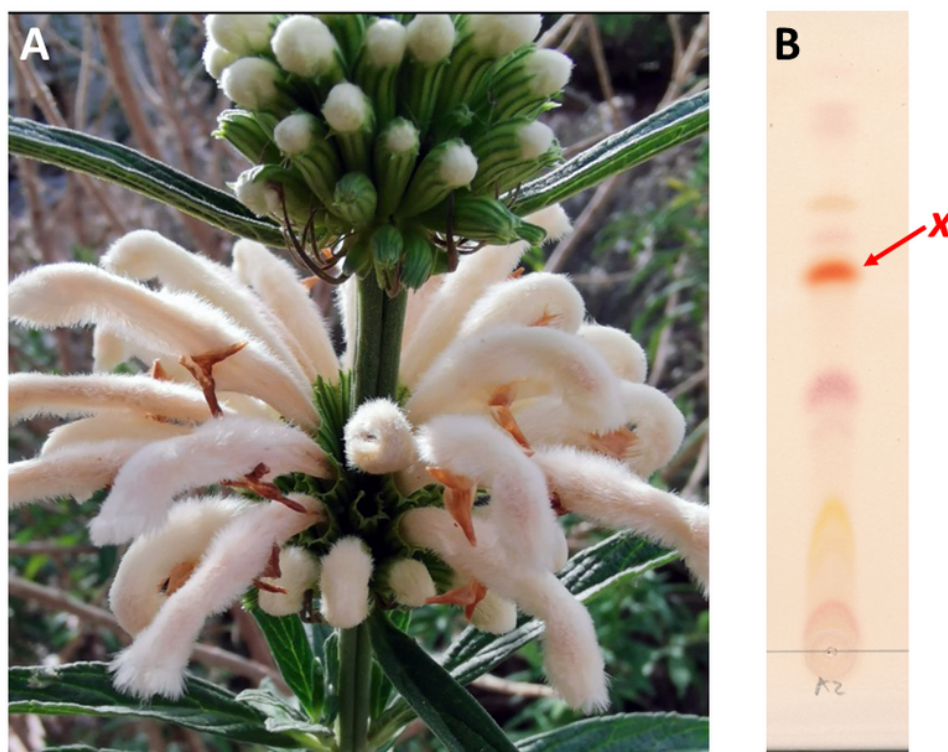


Figure 1

HPTLC phytochemical profile of *Leonotis leonurus* (L.) R. Br. var. *albiflora* whole flower extract. (A) *L. leonurus* (L.) R. Br. var. *albiflora* white flower phenotypic appearance (Photo courtesy of Dr B Loedolff) and (B) its corresponding acetonitrile (75%) extract, HPTLC phytochemical profile. Whole flower metabolite extractions were prepared using various extraction solvents (full HPTLC plate visual is available in Supplementary Figure, acetonitrile cropped area is indicated with red dotted block) and, the acetonitrile (75%) extracts were selected for downstream analyses. The phytochemical profile was developed fully (mobile phase; chloroform:methanol 9:1, stationary phase; silica $60\text{\AA}$) before reagent visualization (0.1% Fast blue B salt dissolved in 1 mM NaOH) was applied to reveal potential pCB candidates, based solely on an orange-red colour development. Compound x (R_f value: 0.55) represents a colorimetric-specific pCB-like candidate compound and, was selected for further LC-MS/MS analyses.

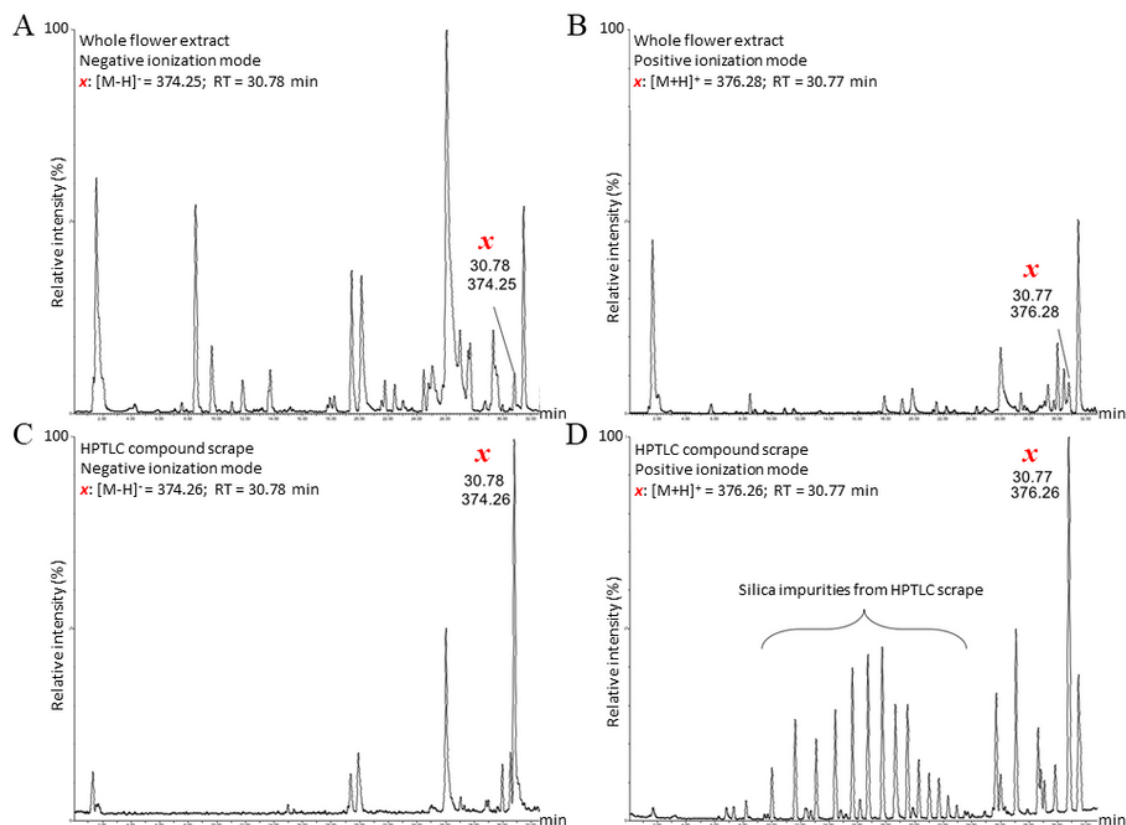


Figure 2

LC-MS/MS spectra of *Leonotis leonurus* (L.) R. Br. var. *albiflora* whole flower extract, targeting the cannabinoid occurrence in flowers. LC-MS/MS spectra of white *L. leonurus* (L.) R. Br. var. *albiflora* whole flower extract in negative (A) and positive (B) ionization modes. Spectra of the preparative HPTLC product, compound x, in negative (C) and positive (D) ionization modes. In accordance with literature, matching retention times and adduct masses for compound x, representing 7,10,13,16-Docosatetraenylethanolamine, are shown in both ionization modes ($[M-H]^- = 374.26$, RT: 30.78 min; $[M+H]^+ = 376.26$, RT: 30.77 min), where chromatogram axis represent relative mass abundance (%) and retention time (RT, minutes), respectively.

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [HunterSuppFigure1.tif](#)