

Shikonin reactivates TSGs GADD45B and PPP3CC to block NSCLC cell proliferation and migration through JNK/P38/MAPK signaling pathways

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Abstract

Background

Shikonin, a natural naphthoquinone compound extracted from the Chinese traditional herbal medicine "Lithospermum erythrorhizon", possesses antitumor activity against various cancer types. Tumor-suppressor genes (TSGs) negatively regulate cell growth, proliferation, and differentiation, thereby inhibiting tumor formation. However, the molecular mechanism of action of shikonin on TSGs in non-small-cell lung cancer (NSCLC) remains unclear.

Methods

The inhibitory effect of shikonin on the proliferation and invasion abilities of lung cancer cells were measured by Cell Counting Kit 8 (CCK8) and wound healing assays. The alteration of genes by shikonin treatment was detected by mRNA high-throughput sequencing and further confirmed by qPCR and western blotting experiments. The dominant functions of the upregulated genes were analyzed by GO and KEGG profiling.

Results

Shikonin inhibited the proliferation and invasion of A549 and H1299 NSCLC cells in a dose-dependent manner. mRNA high-throughput sequencing revealed a total of 1794 upregulated genes in shikonin-treated NSCLC cells. Moreover, bioinformatic analysis of GO and KEGG profiling revealed that the up-regulated genes were mostly involved in the JNK/P38/MAPK signaling pathway, among which the expression of *GADD45B* and *PPP3CC* was significantly enhanced. Finally, we confirmed that *GADD45B* and *PPP3CC* were indeed upregulated in JNK/P38/MAPK pathway.

Conclusions

Taken together, these results suggested that shikonin might affect the expression of *GADD45B* and *PPP3CC* through the JNK/P38/MAPK pathway, therefore exerting an inhibitory effect on the proliferation and migration of cancer cells. To our knowledge, this is the first study reporting the role of shikonin in upregulating TSGs to activate the JNK/P38/MAPK signaling pathways in NSCLC.

1. Introduction

Lung cancer is one of the most diagnosed cancers, with the highest mortality rate among malignancies worldwide. Globally, approximately 2 million new cases and more than 1.76 million deaths are recorded each year (1, 2). Traditionally, lung cancer is histologically classified into small-cell lung cancer and non-small-cell lung cancer (NSCLC), with NSCLC consisting predominantly of lung adenocarcinoma (LUAD)

and lung squamous cell carcinoma. NSCLC is the main histological subtype accounting for 85% of lung cancers (3). According to statistics, approximately two-thirds of patients with NSCLC are in the middle and late stages at the time of diagnosis, losing the best time for treatment. Current clinical treatments for NSCLC mainly contain surgery, chemotherapy, radiation therapy, targeted therapy, and immunotherapy; however, the patients remain to experience an unsatisfactory progression-free survival (PFS)(3–5). As current treatment strategies for NSCLC face issues, such as drug resistance and body tolerance, it is vital to identify additional treatment strategies for the prevention and treatment of NSCLC. Traditional Chinese medicine (TCM) is characterized by its novel pharmacological mechanisms, low toxicity, and limited side-effects. An increasing number of researchers have recently focused their efforts on discovering active TCM-derived anticancer ingredients (6–8).

Shikonin, often referred to as "Zicao" in TCM, is the major bioactive ingredient extracted from the roots of *Lithospermum erythrorhizon*. The chemical structure of shikonin is 5,8-dihydroxy-2-((1R)-1-hydroxy-4-methyl-3-pentenyl)-1,4-naphthoquinone(9). Shikonin has been reported to have various pharmacologic roles, such as antifungal, antibacterial, antitumor, and wound-healing properties(10–12). Recently, a myriad of studies has documented that shikonin exhibits solid antitumor functions against various types of cancer, such as lung, breast, gastrointestinal and urogenital cancers. The underlying mechanisms mostly involve suppression of cell proliferation, induction of apoptosis, and blockade of cell migration and invasion through various molecular pathways(9). For instance, Wang et al. found that shikonin suppress the growth of H1299 lung adenocarcinoma cells through targeting survivin and modulating the expression of apoptosis-related and CDK/cyclin family members, inducing apoptotic cell death and arresting the cell cycle(10). Zang et al. reported that shikonin can induce cell apoptosis to inhibit the growth of paclitaxel-resistant NSCLC through blocking NEAT1 and Akt signaling(13). Given the important role of shikonin in lung cancer treatment, it is particularly important to further explore its mechanism of action.

Tumor-suppressor genes (TSGs) can inhibit the occurrence and development of tumors by participating in DNA damage repair, inhibiting cell division, inducing apoptosis, and inhibiting metastasis(14, 15). Inactivation of TSGs is a common mechanism leading to the occurrence of cancer. Abnormal expression or inactivation of TSGs, because of mutations or other reasons, abolishes their inhibitory effect, resulting in unlimited cell proliferation and leading to tumorigenesis(15). According to the literature, TSGs extensively participate in inhibiting the development of lung cancer(16). Foster et al. demonstrated that *ATMIN* acts as a tumor suppressor in LUAD, and deletion of *Atmin* increasing tumor burden in a LUAD mouse model(17). Han et al. confirmed that as a tumor suppressor, MIR99AHG interacts with *ANXA2* to accelerate the generation of AnxA2-induced ATG16L + vesicles, thereby promoting the assembly of phagocytes and inhibiting the proliferation and metastasis of LUAD cells in vitro and in vivo(18). Wang et al. demonstrated that *SH3GL3* acts as a TSG by inhibiting the cell cycle at the G0/G1 phase and inducing apoptosis in lung cancer cells; as such, it was indicated as a biomarker for the diagnosis and prognosis of lung cancer(19). Given the important role of TSGs in lung cancer, there are relatively few studies on whether shikonin acts on TSGs to further affect the development of lung cancer. Therefore, studying the

mechanism of shikonin and TSGs in lung cancer might be significant for the targeted clinical therapy of lung cancer.

In the present study, we confirmed that shikonin suppressed the proliferation and migration of A549 and H1299 LUAD cells in a dose-dependent manner. Through high-throughput sequencing, we found that the inhibitory effect of shikonin on NSCLC might be related to TSGs. KEGG analysis showed that shikonin-regulated TSGs mainly inhibited lung cancer cells through the MAPK signaling pathway. Furthermore, qPCR and western blot experiments confirmed that TSGs, such as *DUSP5*, *GADD45B*, and *PPP3CC*, were upregulated in shikonin-treated lung cancer cells. Finally, we further validated the expression of *GADD45B* and *PPP3CC* in NSCLC using the TCGA database and KM survival curve analysis, and found that upregulated *GADD45B* and *PPP3CC* improved prognosis. Therefore, we hypothesized that shikonin inhibits the proliferation and migration of A549 and H1299 cells by acting on *GADD45B* and *PPP3CC* and by activating the MAPK pathway. Our study suggested that shikonin is an attractive candidate for further consideration in the development of anti-lung cancer drugs.

2. Materials And Methods

2.1 Reagents and Antibodies

Shikonin (S0921AS; purity > 98%; Meilunbio, China) was dissolved in DMSO to prepare a 1000 μ M stock solution that was stored at -20°C . The concentration of DMSO in the solution did not exceed 0.2%. The RPMI-1640 cell culture medium and RPMI-1640 complete medium (1% HEPES 1 M buffer solution, 1% sodium pyruvate, and 1% glutamine) were purchased from CYTIVA (SH30809.01). Antibodies against GADD45B, PPP3CC and β -actin were purchased from Affinity Biosciences (DF2375), Proteintech (19653-1-AP) and ABclonal (WH256886), respectively, whereas antibodies against p38 MAPK (CST9212S), JNK (CST9252S) and ERK1/2 (CST4695S) were all purchased from Cell Signaling Technology.

2.2 Cell culture

The A549 and NCI-H1299 human lung adenocarcinoma cell lines were purchased from the Cell Bank of the Chinese Academy of Science (Shanghai, China). Cells were cultured in RPMI-1640 and RPMI-1640 complete medium, respectively, both of which were supplemented with 10% fetal bovine serum (FBS, Gibco, 10099-141, Australia) and 1% penicillin–streptomycin in a humidified atmosphere of 5% CO_2 at 37°C .

2.3 Cell Counting Kit 8 (CCK8) assay

Cell viability was assessed using the CCK8 assay (Cell Counting Kit 8, HY-K0301, MedChemExpress). Cells were cultured overnight in 96-well plates at 1×10^4 cells/well in a volume of 100 μ l and then treated with various concentrations of shikonin with 0, 0.3, 0.9, 2.7 and 8.1 μ M, respectively. Subsequently, 10 μ l CCK8 was added to each well and incubated for 20 min at 37°C . The absorbance of each well at 450 nm was recorded by enzyme-labeled instrument, and cell viability was calculated using wells without cells as

blank. IC50 values were obtained from nonlinear regression curve fit analysis by GraphPad Prism 5.0 (Version 7.0, GraphPad Software, Inc).

2.4 Colony-formation assay

Cells were diluted in culture medium and seeded in 6-well plates at a density of 3×10^3 cells per well. After incubation for 24 h, cells were treated with either 3 μ M shikonin or an equal volume of culture medium as a control for 15 d. Grown colonies were stained with crystal violet and photographed. The colony area was calculated using the ImageJ software.

2.5 Wound healing assay

Cells were cultured at 1×10^4 /well in the 6-well plates until they reached the confluence of 90–95%. Then a vertical wound was made with sterile 1 ml pipet tip. The floating cells were washed with PBS, and medium with or without shikonin was added to the remaining cells. The images of wound in each well were taken by microscope at 0 h, 24 h and 48 h. Areas of the wound were measured by Image J and the data are expressed as the ratio of the healed wound area at 24 h and 48 h compared to initial wound area (0 h).

2.6 Reverse transcription polymerase chain reaction (RT-PCR) detection

A549 and H1299 cells were treated with 3 and 5 μ M shikonin for 24 h, respectively. Total RNA was isolated using TRIzol (B511311-0100; Sangon Biotech) and reverse transcribed into cDNA with oligo (dT18) primers using the PrimeScript RT Reagent Kit with gDNA Eraser (Takara; Japan) under the manufacturer's instructions. The relative expression of target genes was determined after normalization to the expression of the internal control *GAPDH* using the $2^{-\Delta\Delta C_t}$ method. The sequences of primers used in RT-PCR are as follows: *DUSP5* forward primer, 5'-TCACCTCGCTACTCGCTTGC-3' and reverse primer, 5'-GATGAGGGCTCTCTCACTCT-3'; *PPP3CC* forward primer, 5'-GTACATGGAGGAATGTAC-3' and reverse primer, 5'-CTCATTGCCATAATCCTCTG-3'; *GADD45B* forward primer, 5'-GGCCGCTCAGCGCCAGGATC-3' and reverse primer, 5'-GAGCGTGAAGTGGATTTGCA-3'; *DUSP9* forward primer, 5'-CAGCCGTTCTGTCACCGTC-3' and reverse primer, 5'-CAAGCTGCGCTCAAAGTCC-3'; *GAPDH* forward primer, 5'-GCCTTCTCCATGGTGGTGA-3' and reverse primer, 5'-ACCGTCAAGGCTGAGAAC-3'.

2.7 Western blot analysis

Cells were cultured in the 6-well plates with various concentrations of shikonin at 3, 6 μ M and 5, 10 μ M, respectively until they reached confluence of 90–95%. Total proteins were extracted by using radioimmunoprecipitation assay (RIPA) buffer, and the concentration was determined using BCA Protein Assay Kit (Beyotime). Equal amounts of protein were separated by 4–12% gradient sodium dodecyl sulfate (SDS)-polyacrylamide gel and transferred to polyvinylidene difluoride (PVDF) membranes (Millipore). The membrane was blocked with 5% milk. The blot was then incubated with primary antibodies overnight at 4 °C and then incubated with anti-rabbit secondary antibodies (1/20,000 dilution)

for 1h on the next day. The immunoreactions were measured by an enhanced chemiluminescence (ECL) system with the manufacturer's instructions.

2.8 RNA preparation and mRNA high-throughput sequencing analysis

To identify the difference of gene expression in response to shikonin exposure, A549 lung cancer cells were cultured over night at 3×10^5 cells/well in 6 cm dish and incubated at 37°C. The cell samples were prepared in triplicate. They were then incubated with 3 μ M shikonin or blank cell medium. After 24 h of incubation, the total RNA was extracted by TRIzol (B511311-0100; Sangon Biotech) to perform high-throughput sequencing. Then, the concentration and quality of purified RNA were detected by using Nanodrop 2000 (Thermo Scientific) and the Bioanalyzer 2100 (Agilent Technologies) for further library preparation. The different expression genes (DEGs) were performed with DESeq2 (version 3.12) in R (version 4.0). The detail profiling methods are referred in our previous study(20). Fold change > 1.5, $P < 0.05$. The accession number for the sequencing data generated in this paper is GSE222640 and can be gained from GEO (Gene Expression Omnibus) datasets. The bioinformatic profiling data for DEG and functional enrichment are all presented in Supplementary Information.

2.9 Statistical analysis

All data are presented as the mean $\pm$ SEM, and each experiment was independently repeated 3 times unless otherwise noted. Statistical analyses were performed using GraphPad Prism 5.0 (Version 7.0, GraphPad Software, Inc) by two-tailed Student t-test. **** represents $p < 0.0001$, *** represents $p < 0.001$, ** represents $p < 0.01$ and * represents $p < 0.05$. $p < 0.05$ were defined as significant.

3. Results

3.1 Shikonin blocked the proliferation and migration of lung adenocarcinoma cells

The chemical structure of shikonin, a potential anticancer agent, is depicted in Fig. 1A. To assess the effect of shikonin on the viability of lung adenocarcinoma cells, we treated A549 and H1299 cells with serial dilutions of shikonin (0, 0.2, 0.4, 0.8, 1.6, 3.2, 6.4 μ M and 0, 0.5, 1.0, 2.0, 4.0, 8.0, 16.0 μ M, respectively) for 24 and 48 h. We determined cell viability using the CCK8 assay, and found that shikonin inhibited cell growth in a dose-dependent manner (Fig. 1B and C); in particular, we detected that the growth of cells declined in a stepwise manner with increasing doses of shikonin (Fig. 1E and F). The respective IC₅₀ values in A549 and H1299 cells were 1.41 μ M and 8.58 μ M at 24 h, and 0.84 μ M and 7.67 μ M at 48 h (Fig. 1D). Interestingly, we noticed that the IC₅₀ values of shikonin on A549 and H1299 cells between 24 and 48 h were not significantly different, indicating that the inhibitory effect of shikonin on A549 and H1299 cells was not time-dependent. However, A549 cells appeared more sensitive to shikonin compared with H1299 cells, as indicated by their IC₅₀ values (Fig. 1D).

Furthermore, we determined how shikonin plays a role on cell proliferation by using colony formation and wound-healing assays. We treated A549 and H1299 cells with 3 and 6 μM shikonin, respectively. As shown in Fig. 2A and B, both A549 and H1299 cells showed high sensitivity to shikonin, as the number and size of colonies were significantly decreased compared with those in the control group (upper panel of Fig. 2A and B). We found that the colony numbers of A549 cells in the control and shikonin treatment groups were 343 and 6, respectively, while the colony numbers of H1299 cells were 72 and 1, respectively (lower panel of Fig. 2A and B). Moreover, we performed a wound healing assay to investigate whether shikonin inhibits cell migration by treating A549 and H1299 cells with 3 and 6 μM shikonin, respectively. We accordingly found that the migration ability of cells was obviously inhibited after shikonin treatment at 24 and 48 h, as indicated in the left panels of Fig. 2C and D. Compared with the control group, we observed a significant reduction in the number of migrated cells in the treated groups at 24 and 48 h (right panels of Fig. 2C and D).

3.2 Shikonin modulated the expression of tumor-suppressor genes through MAPK signaling by high-throughput sequencing

To investigate the fundamental molecular mechanism by which shikonin suppresses the proliferation and migration abilities of lung adenocarcinoma cells, we performed mRNA high-throughput sequencing profiling of A549 cells. As shown in Fig. 3A, a total of 1795 genes were upregulated by shikonin, whereas 1645 genes were downregulated. The top 100 upregulated and downregulated genes are depicted in Fig. 3B. The main molecular function of the top upregulated genes in KEGG profiling was the MAPK signaling pathway shown in Fig. 3C, which was consistent with the findings of previous studies on the shikonin-induced activation of the MAPK pathway(21, 22).

Tumor-suppressor genes are frequently downregulated in various types of cancers. Loss of function of TSGs contributes to carcinogenesis; conversely, rescue of downregulated TSGs might block the progression of cancer cells(14). To investigate whether shikonin activates TSGs in lung adenocarcinoma, we explored the expression of TSGs in lung adenocarcinoma samples from the TSGene database (<https://bioinfo.uth.edu/TSGene/>) and identified that a total of 525 TSGs were downregulated(23, 24). Interestingly, we identified a total of 82 TSGs, including *DUSP5*, *PPP3CC*, and *GADD45B*, among the 1795 upregulated genes after shikonin treatment, indicating that these 82 TSGs that were downregulated in lung adenocarcinoma were in turn activated by shikonin (Fig. 4A). KEGG pathway analysis in Fig. 4B demonstrated that the 82 activated TSGs are mainly involved in cancer and MAPK pathways, which is consistent with the top 100 upregulated genes in Fig. 3C. Moreover, Gene Ontology (GO) profiling showed that these 82 upregulated TSGs are typically related to negatively regulating cell proliferation or positively leading to cell death (Fig. 4C), suggesting that their activation suppresses cell growth and proliferation.

3.3 Expression of tumor suppressors PPP3CC and GADD45B in MAPK signaling was significantly increased by shikonin treatment

To further verify the activation of TSGs by shikonin, we randomly selected 4 TSGs, namely *DUSP5*, *DUSP9*, *GADD45B* and *PPP3CC*, from the 82 upregulated TSGs and detected their expression using RT-PCR. We found that shikonin treatment led to the activation of *GADD45B*, and *PPP3CC* in both A549 and H1299 cells, whereas the expression of *DUSP9* was not significantly changed and *DUSP5* was only upregulated in A549 cells (Fig. 5A and B). Interestingly, among these selected TSGs, *GADD45B* and *PPP3CC* have been reported as tumor suppressors that inhibit cancer cell proliferation and invasion in various cancers; however, their roles in lung adenocarcinoma remain unknown. Therefore, we preferentially selected *GADD45B* and *PPP3CC* as candidate targets for shikonin in lung adenocarcinoma cells. We also performed western blotting to verify the levels of protein expression of *GADD45B* and *PPP3CC* after shikonin treatment. As expected, we detected that the expression of *GADD45B* and *PPP3CC* was obviously increased when A549 and H1299 cells were treated with 3, 6 μ M and 5, 10 μ M shikonin, respectively. Moreover, we observed the activation of marker proteins, such as ERK1/2, JNK, and P38 MAPK, after shikonin treatment, indicating that the MAPK signaling pathway was indeed positively modulated by shikonin.

To determine whether the expression of *GADD45B* and *PPP3CC* was downregulated across various TCGA cancers, we investigated the pancancer gene-expression profiles of *GADD45B* and *PPP3CC* using the UALCAN database (<http://ualcan.path.uab.edu/>)(25). As shown in Fig. 6A and B, the expression of both *GADD45B* and *PPP3CC* was remarkably downregulated in lung adenocarcinoma, as well as in numerous other cancer types, in line with the reported downregulation of *GADD45B* and *PPP3CC* in various cancers(26, 27). Moreover, we wonder whether their expression distributions are correlated with gender, race and tumor stages or not, so we interrogated the RNA-seq expression data of lung adenocarcinoma and the corresponding clinical information from TCGA datasets (<https://portal.gdc.com>). As seen in Fig. 6C and D, the expression of both *GADD45B* and *PPP3CC* was not related to gender, that is, no matter what gender is, their expressions were distinctly lower in lung adenocarcinoma than those of normal ones. The results are the same for races and tumor stages as presented in Fig. 6E-F and 6G-H, respectively.

Taken together, all these results demonstrated that both *GADD45B* and *PPP3CC* were downregulated in lung adenocarcinoma as TSGs and could be activated by shikonin through the MAPK signaling pathway.

3.4 *PPP3CC* and *GADD45B* were correlated with patient prognosis

As *GADD45B* and *PPP3CC* are activated by shikonin to suppress cell proliferation and metastasis in lung adenocarcinoma, we speculated whether their expression was correlated with patient prognosis. Hence, we analyzed Kaplan–Meier (KM) plots of overall survival to confirm the effects of the expression of *GADD45B* and *PPP3CC* on patient survival outcomes. As shown in Fig. 7A and B, patients with LUAD with higher levels of expression of *GADD45B* and *PPP3CC* exhibited better survival outcomes, suggesting that *GADD45B* and *PPP3CC* indeed exhibit a tumor-suppressive effect. Generally, we determined that the high expression of *GADD45B* and *PPP3CC* was correlated with better patient prognosis in KM analysis.

Furthermore, we depicted the schematic diagram of the current research as shown in Fig. 7C. Shikonin activates the expression of *GADD45B* and *PPP3CC* through the JNK/P38/MAPK pathway to suppress the growth and migration abilities of NSCLC cells, providing an alternative mechanism for the inhibitory function of shikonin on NSCLC.

4. Discussion

To investigate their potential for antitumor drug development, we need to understand the underlying molecular mechanism by which natural products modulate the proliferation and invasion of cancer cells. Shikonin is an active naphthoquinone derived from the Chinese traditional medicine Zi Cao (*purple gromwell*) with antitumor, anti-inflammation, and antibacterial functions(28). A Recent study has reported that shikonin suppresses cancer cell proliferation via the MAPK signaling pathway, which mediates the induction of apoptosis(21). However, the detailed molecular mechanisms by which shikonin blocks the proliferation of lung cancer cells through the MAPK pathway are yet to be elucidated. In the current study, we focused on the molecular mechanisms underlying the suppressive effects of shikonin in human lung adenocarcinoma cell lines. Specifically, we found the upregulation of 1795 genes in A549 cells by shikonin, among which 82 TSGs were mainly correlated with the MAPK pathway, indicating a remarkable role of TSGs in NSCLC via the MAPK signaling pathway.

Importantly, TSGs encode critical intracellular regulators that control cell proliferation, cell growth, and DNA damage in cancer. The first TSG *Rb1* (retinoblastoma gene 1), which encodes the retinoblastoma protein that causes RB cancer development, was discovered by Knudson(29). Although both shikonin and inactivation of TSGs are known to be closely related to cancer progression, whether TSGs can be modulated by shikonin remains unclear. High-throughput sequencing revealed that shikonin can activate TSGs in lung cancer cells through the MAPK pathway. Taking TSG *PPP3CC* and *GADD45B*, which are involved in the MAPK signaling pathway, as candidates, we found that the expression of both was upregulated by shikonin treatment, as indicated by qPCR and western blotting assays (Fig. 5). Moreover, KM analysis revealed that the expression of both *GADD45B* and *PPP3CC* was associated with the prognosis of patients with LUAD, consistent with findings from other studies that also reported their inhibitory functions on tumors(30–33). For example, *GADD45B* has been reported to be a promising tumor suppressor in NSCLC and has close association with NSCLC development(32). Meanwhile, downregulation of *PPP3CC* has been linked with an autophagy-related prognostic signature for hepatocellular carcinoma, and suppression of *PPP3CC* by ZEB1 activates NF- κ B signaling, leading to glioma cell proliferation and invasion(33), indicating that *PPP3CC* acts as a tumor suppressor in these cancers. Surprisingly, the present study is the first to demonstrate the suppressive role of *PPP3CC* in NSCLC cells. Hence, *PPP3CC* may serve as a novel therapeutic target for NSCLC.

In general, we have demonstrated that shikonin is capable of blocking the proliferation and invasion of LUAD cells by activating TSGs, such as *PPP3CC* and *GADD45B*, through the MAPK signaling pathway (Fig. 7C). In particular, *PPP3CC* was a new target discovered in NSCLC. To the best of our knowledge, the present study is the first to investigate the relationship between shikonin and TSGs, providing novel

insights into the molecular mechanism by which shikonin inhibits the proliferation of LUAD cells and paving the way for the development of patent medicines based on shikonin to be used in the clinical setting.

Declarations

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Author Contributions

Conceptualization, Y.L. and Y.Y.; methodology, Y.Y. and Y.L.; software, Y.Z; validation, Y.Z. and Y.L.; investigation, Y.Z., Z. F., D.W., and W.L.; data curation, Y.L.; writing—original draft preparation, Y.Z. and D.W.; writing, review and editing, Y.L.; visualization, Y.Z., Z. F., D.W., and Y.L.; supervision, Y.L.; project administration, Y.L.; funding acquisition, Y.L. All authors have read and agreed to the published version of the manuscript.

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Conflicts of Interest

The authors declare no conflicts of interest.

Consent for publication statement

Not applicable.

Ethics approval and consent to participant

Not applicable.

Supplementary Information

All supplementary materials involved in this study are available at *BMC Complementary Medicine and Therapies*.

Data Availability Statement

The datasets supporting the conclusions of this article are included within the article. The original high throughput sequencing data generated in this article can be downloaded at:
<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE222640> with accession number GSE222640 and token number szqrmawqnnsvhcf.

References

1. Campanella A, De Summa S, Tommasi S. Exhaled breath condensate biomarkers for lung cancer. *J Breath Res*. 2019;13(4):044002.
2. Long Y, Wang C, Wang T, Li W, Dai W, Xie S, et al. High performance exhaled breath biomarkers for diagnosis of lung cancer and potential biomarkers for classification of lung cancer. *J Breath Res*. 2021;15(1):016017.
3. Rodak O, Peris-Diaz MD, Olbromski M, Podhorska-Okolow M, Dziegiel P. Current Landscape of Non-Small Cell Lung Cancer: Epidemiology, Histological Classification, Targeted Therapies, and Immunotherapy. *Cancers (Basel)*. 2021;13(18).
4. Mielgo-Rubio X, Uribe-larrea EA, Cortes LQ, Moyano MS. Immunotherapy in non-small cell lung cancer: Update and new insights. *J Clin Transl Res*. 2021;7(1):1-21.
5. Sundahl N, Lievens Y. Radiotherapy for oligometastatic non-small cell lung cancer: a narrative review. *Transl Lung Cancer Res*. 2021;10(7):3420-31.
6. Wei ZC, Chen J, Zuo F, Guo JL, Sun XD, Liu DM, et al. Traditional Chinese Medicine has great potential as candidate drugs for lung cancer: A review. *Journal of Ethnopharmacology*. 2023;300.
7. Zhou Y, Hong T, Tong L, Liu W, Yang XT, Luo JH, et al. Astragalus polysaccharide combined with 10-hydroxycamptothecin inhibits metastasis in non-small cell lung carcinoma cell lines via the MAP4K3/mTOR signaling pathway. *Int J Mol Med*. 2018;42(6):3093-104.
8. Ni MW, Wang HJ, Wang MM, Zhou W, Zhang JY, Wu JR, et al. Investigation on the Efficiency of Chinese Herbal Injections for Treating Non-small Cell Lung Cancer With Vinorelbine and Cisplatin Based on Multidimensional Bayesian Network Meta-Analysis. *Front Pharmacol*. 2021;11.
9. Guo CJ, He JL, Song XMT, Tan L, Wang M, Jiang PD, et al. Pharmacological properties and derivatives of shikonin-A review in recent years. *Pharmacol Res*. 2019;149.
10. Wang HN, Zuo JL. Shikonin Inhibits Non-Small-Cell Lung Cancer H1299 Cell Growth through Survivin Signaling Pathway. *Anal Cell Pathol*. 2021;2021.
11. Boulos JC, Rahama M, Hegazy MF, Efferth T. Shikonin derivatives for cancer prevention and therapy. *Cancer Lett*. 2019;459:248-67.
12. Guo C, He J, Song X, Tan L, Wang M, Jiang P, et al. Pharmacological properties and derivatives of shikonin-A review in recent years. *Pharmacol Res*. 2019;149:104463.
13. Zang F, Rao Y, Zhu X, Wu Z, Jiang H. Shikonin suppresses NEAT1 and Akt signaling in treating paclitaxel-resistant non-small cell of lung cancer. *Mol Med*. 2020;26(1):28.

14. Lipsick J. A History of Cancer Research: Tumor Suppressor Genes. *Cold Spring Harb Perspect Biol.* 2020;12(2).
15. Wang LH, Wu CF, Rajasekaran N, Shin YK. Loss of Tumor Suppressor Gene Function in Human Cancer: An Overview. *Cell Physiol Biochem.* 2018;51(6):2647-93.
16. Chen L, Liu S, Tao Y. Regulating tumor suppressor genes: post-translational modifications. *Signal Transduct Target Ther.* 2020;5(1):90.
17. Foster H, Ruiz EJ, Moore C, Stamp GWH, Nye EL, Li NN, et al. ATMIN Is a Tumor Suppressor Gene in Lung Adenocarcinoma. *Cancer Res.* 2019;79(20):5159-66.
18. Han CC, Li H, Ma ZF, Dong GZ, Wang QY, Wang SW, et al. MIR99AHG is a noncoding tumor suppressor gene in lung adenocarcinoma. *Cell Death Dis.* 2021;12(5).
19. Lin ZK, Liu ZP, Tan XS, Li CH. SH3GL3 functions as a potent tumor suppressor in lung cancer in a SH3 domain dependent manner. *Biochem Bioph Res Co.* 2021;534:787-94.
20. Liang Y, Lu Q, Li W, Zhang D, Zhang F, Zou Q, et al. Reactivation of tumour suppressor in breast cancer by enhancer switching through NamiRNA network. *Nucleic Acids Res.* 2021;49(15):8556-72.
21. Lee JH, Han SH, Kim YM, Kim SH, Yoo ES, Woo JS, et al. Shikonin inhibits proliferation of melanoma cells by MAPK pathway-mediated induction of apoptosis. *Biosci Rep.* 2021;41(1).
22. Lohberger B, Glanzer D, Kaltenegger H, Eck N, Leithner A, Bauer R, et al. Shikonin derivatives cause apoptosis and cell cycle arrest in human chondrosarcoma cells via death receptors and MAPK regulation. *BMC Cancer.* 2022;22(1):758.
23. Zhao M, Sun J, Zhao Z. TSGene: a web resource for tumor suppressor genes. *Nucleic Acids Res.* 2013;41(Database issue):D970-6.
24. Zhao M, Kim P, Mitra R, Zhao J, Zhao Z. TSGene 2.0: an updated literature-based knowledgebase for tumor suppressor genes. *Nucleic Acids Res.* 2016;44(D1):D1023-31.
25. Chandrashekar DS, Karthikeyan SK, Korla PK, Patel H, Shovon AR, Athar M, et al. UALCAN: An update to the integrated cancer data analysis platform. *Neoplasia.* 2022;25:18-27.
26. Wang Q, Wu WH, Gao Z, Li KW, Peng SR, Fan HY, et al. GADD45B Is a Potential Diagnostic and Therapeutic Target Gene in Chemotherapy-Resistant Prostate Cancer. *Front Cell Dev Biol.* 2021;9.
27. Anastasiadou E, Messina E, Sanavia T, Labruna V, Ceccarelli S, Megiorni F, et al. Calcineurin Gamma Catalytic Subunit PPP3CC Inhibition by miR-200c-3p Affects Apoptosis in Epithelial Ovarian Cancer. *Genes (Basel).* 2021;12(9).
28. Sun Q, Gong T, Liu M, Ren S, Yang H, Zeng S, et al. Shikonin, a naphthalene ingredient: Therapeutic actions, pharmacokinetics, toxicology, clinical trials and pharmaceutical researches. *Phytomedicine.* 2022;94:153805.
29. Knudson AG, Jr. Mutation and cancer: statistical study of retinoblastoma. *Proc Natl Acad Sci U S A.* 1971;68(4):820-3.
30. Gong LQ, Cai LQ, Li GD, Cai J, Yi XQ. GADD45B Facilitates Metastasis of Ovarian Cancer Through Epithelial-Mesenchymal Transition. *Oncotargets Ther.* 2021;14:255-69.

31. Barros MC, de Mello JBH, Marchi FA, Pinto CAL, da Silva IC, Damasceno PKF, et al. GADD45B Transcript Is a Prognostic Marker in Papillary Thyroid Carcinoma Patients Treated With Total Thyroidectomy and Radioiodine Therapy. *Front Endocrinol.* 2020;11.
32. Do H, Kim D, Kang J, Son B, Seo D, Youn H, et al. TFAP2C increases cell proliferation by downregulating GADD45B and PMAIP1 in non-small cell lung cancer cells. *Biol Res.* 2019;52(1):35.
33. Wang H, Zhao S, Chen B, Fu C, Dang Y, Fang P, et al. Repression of the expression of PPP3CC by ZEB1 confers activation of NF-kappaB and contributes to invasion and growth in glioma cells. *Jpn J Clin Oncol.* 2018;48(2):175-83.

Figures

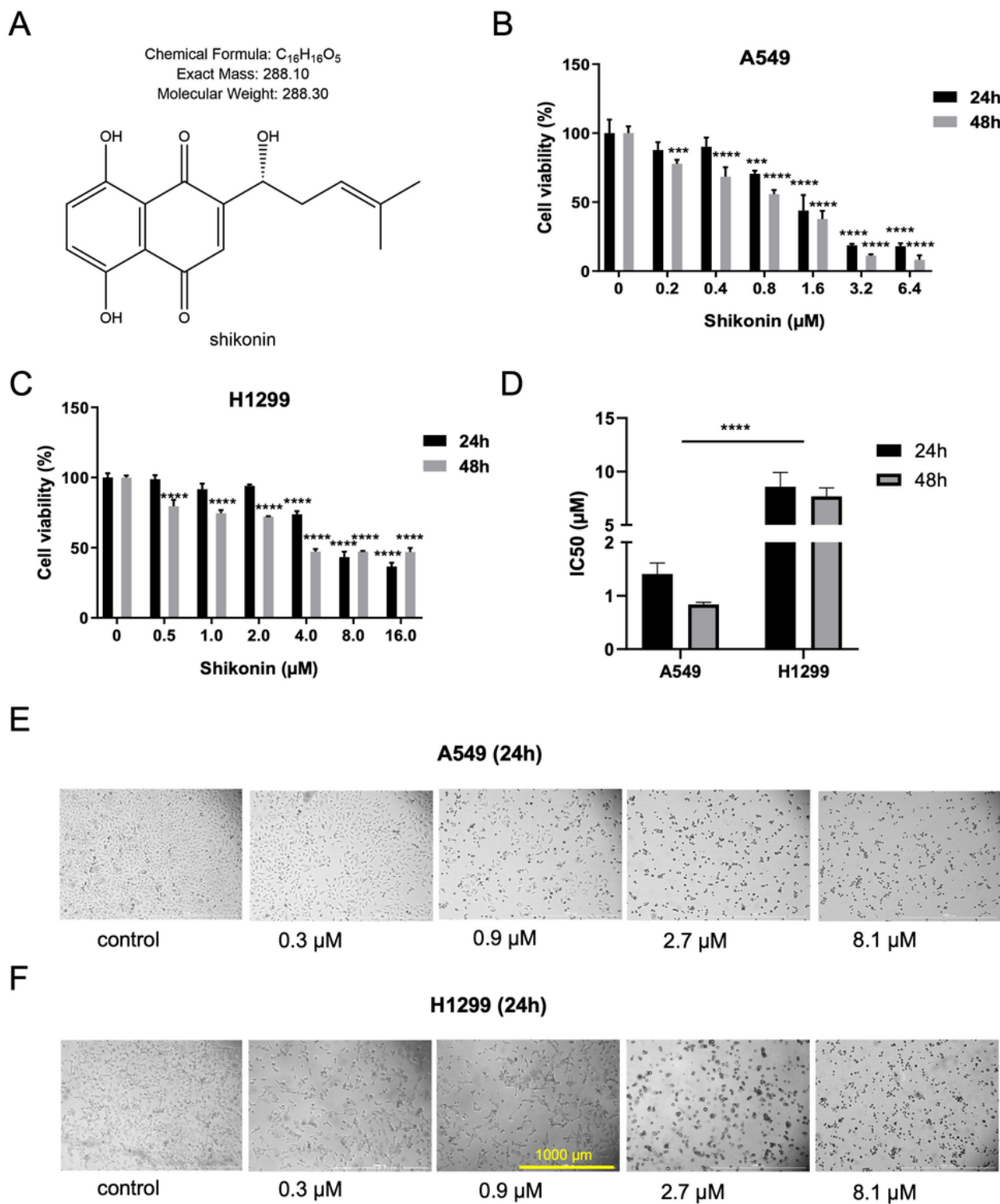


Figure 1

Shikonin inhibits the cell viability of A549 and H1299 cells in dose-dependent manner. **A.** The chemical structure of shikonin. **B-C.** The cell viability of A549 and H1299 cells was determined by CCK8 assay. **D.** The respective IC₅₀ in A549 and H1299 cells was calculated. **E-F.** Shikonin was able to inhibit the cell growth of A549 and H1299 cells in a dose-dependent manner. **** represents $p < 0.0001$, *** represents $p < 0.001$.

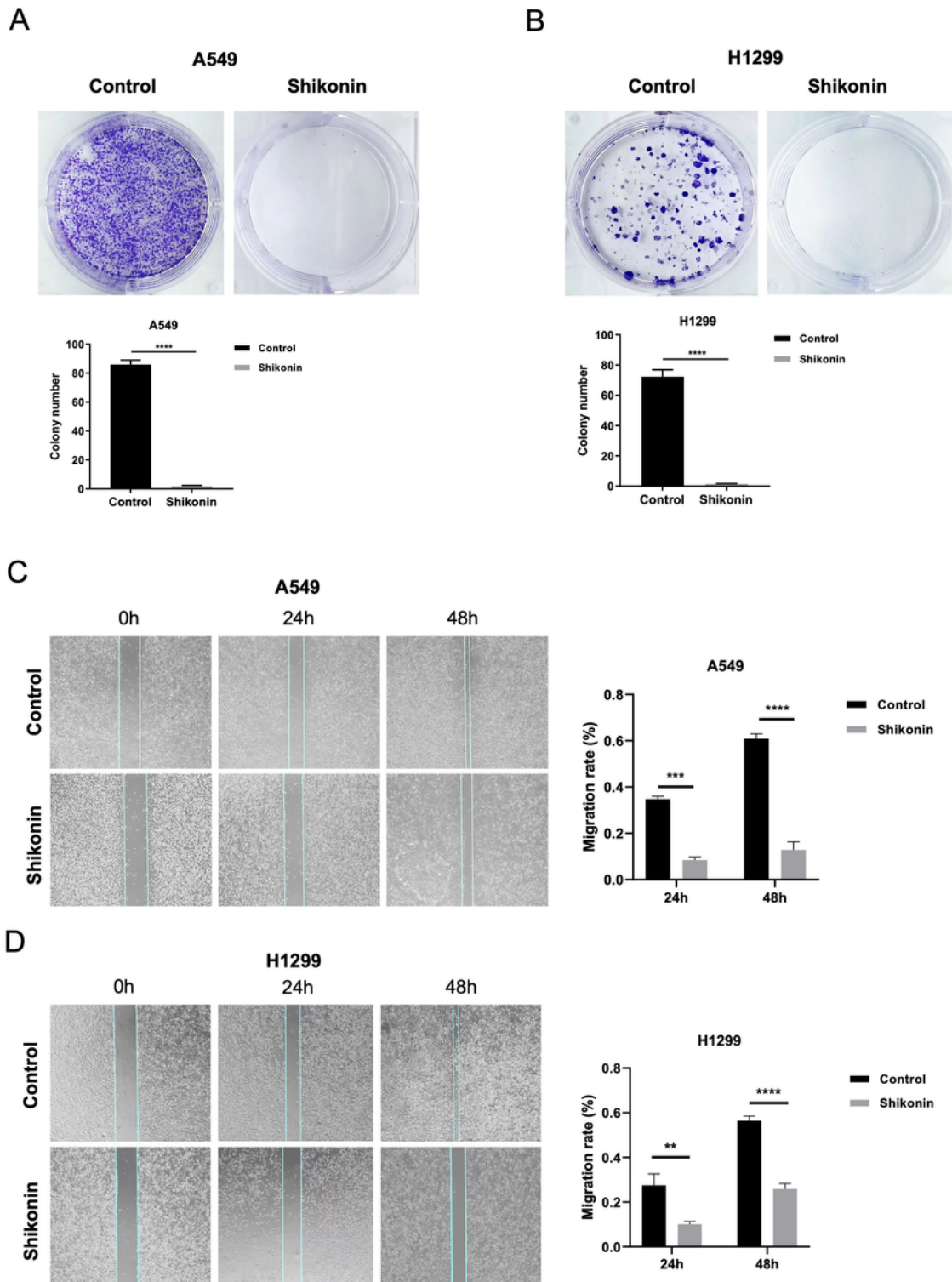


Figure 2

Shikonin inhibits the proliferation and invasion abilities of A549 and H1299 cells. A-B. Shikonin significantly decreased the colony numbers of A549 and H1299 cells in colony formation assays. **C-D.** The invasion viability of A549 and H1299 cells was significantly blocked by shikonin at 24 h and 48 h. Images were gained under magnification x200. **** represents $p < 0.0001$, *** represents $p < 0.001$, ** represents $p < 0.01$.

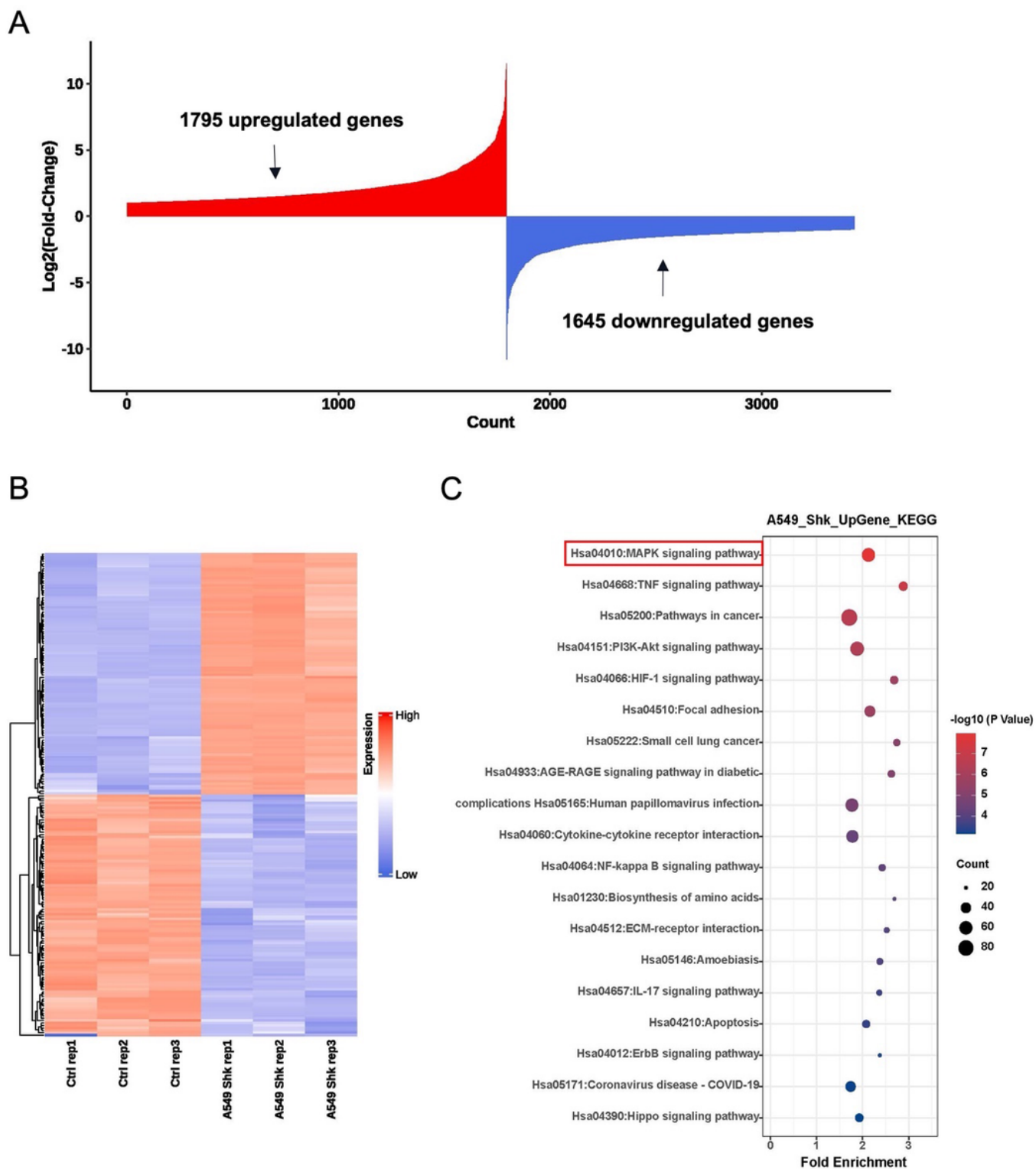


Figure 3

Shikonin is related to TSG expression alterations through MAPK signaling. **A.** A total of 1795 upregulated genes and by 1645 downregulated genes by Shikonin through mRNA sequencing analysis. **B.** Heatmap of the top 100 changed genes by shikonin. **C.** The upregulated genes are mainly involved in MAPK signaling pathway by KEGG profiling.

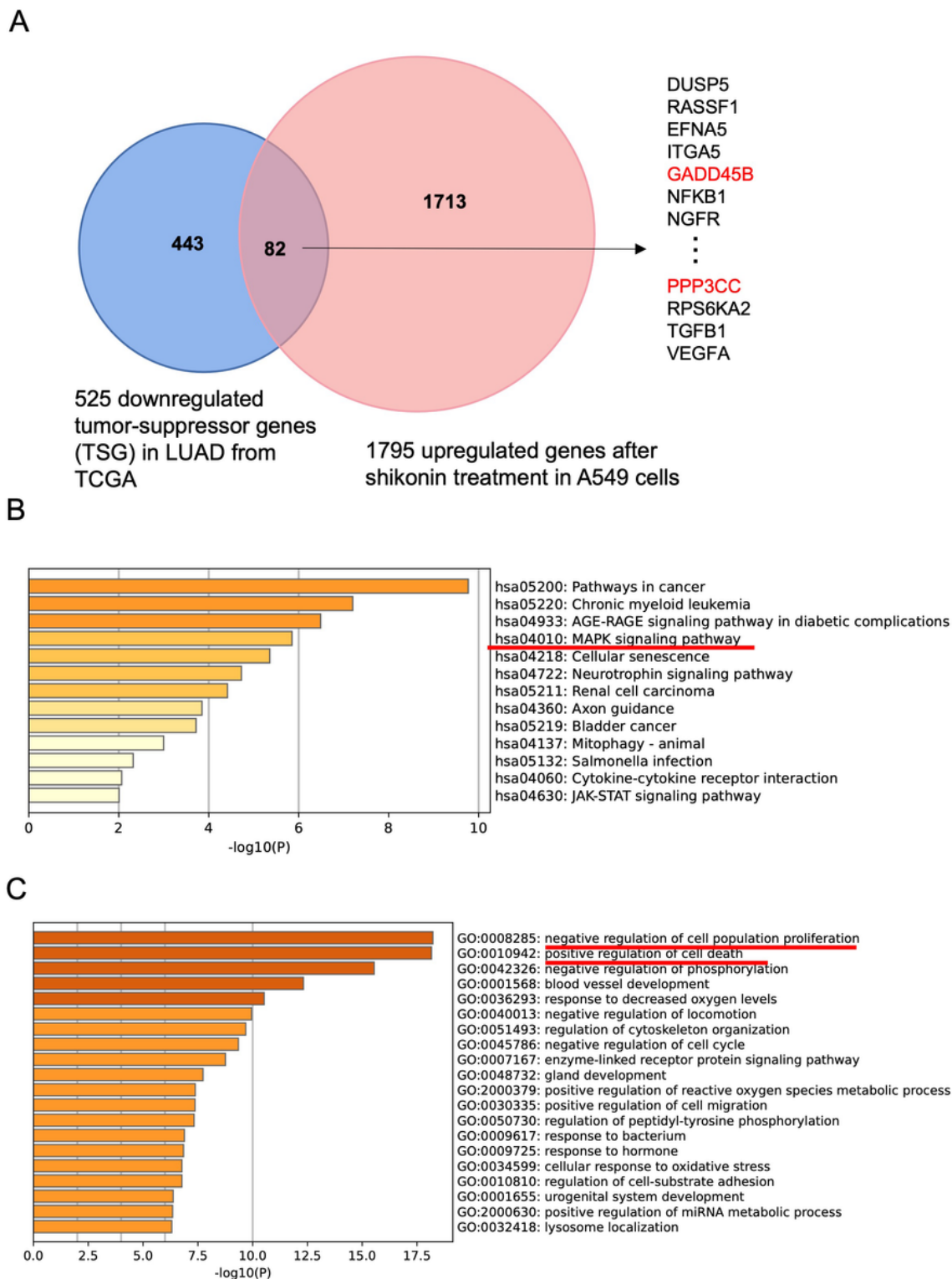


Figure 4

Shikonin may reactivate TSGs in LUAD through MAPK signaling. **A.** A total of 82 activated TSGs among 1795 upregulated genes by shikonin. **B.** KEGG profiling for the 82 upregulated TSGs by shikonin. **C.** GO analysis for the 82 upregulated TSGs by shikonin.

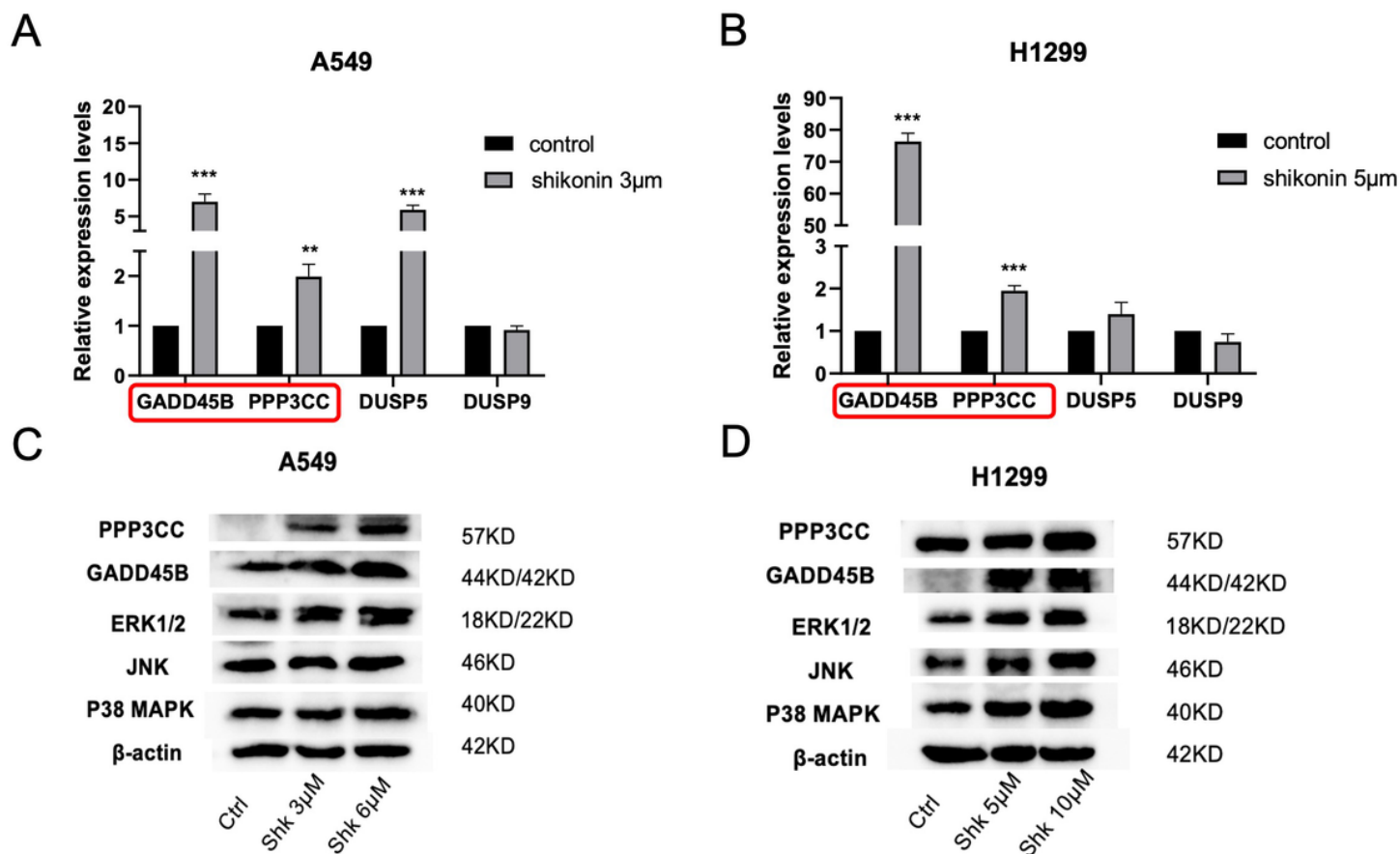


Figure 5

Shikonin increased tumor suppressor genes *PPP3CC* and *GADD45B* in MAPK signaling. A-B. *GADD45B* and *PPP3CC* are activated after shikonin treatment in both A549 and H1299 cells by qPCR detection. **C-D.** The MAPK proteins, *GADD45B* and *PPP3CC* are activated after shikonin treatment in both A549 and H1299 cells by western blotting. The cropped plots are used in the figure to improve the clarity of presentation. *** represents $p < 0.001$, ** represents $p < 0.01$.

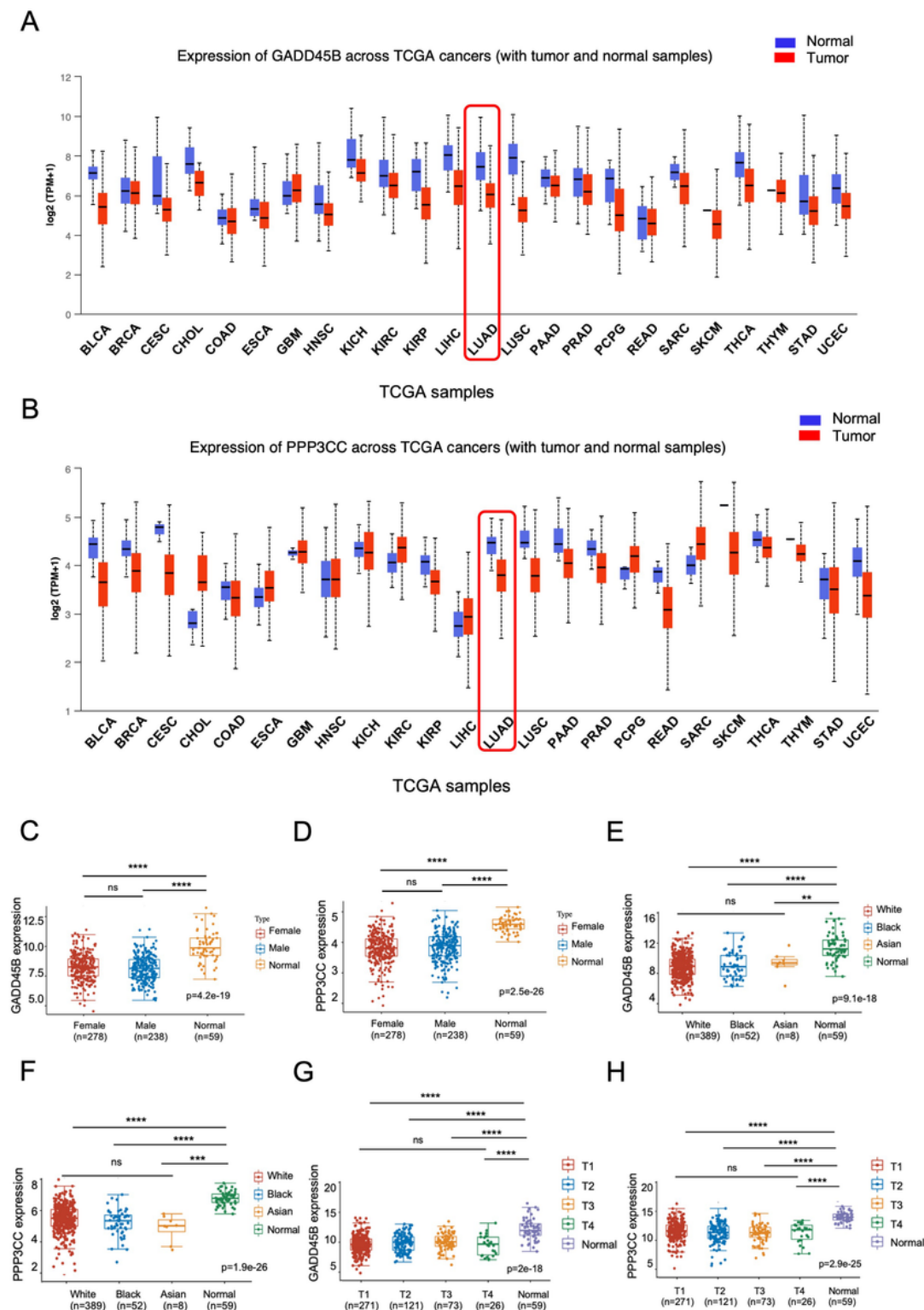


Figure 6

Expression of tumor suppressor genes *PPP3CC* and *GADD45B* in pan-cancer. A-B. Pan cancer gene expression profile of *GADD45B* and *PPP3CC*. Red boxplot depicts expression level in primary tumors, while blue boxplot indicate expression in normal samples. C-H. The expression distribution of *GADD45B* and *PPP3CC* in tumor and normal tissues from TCGA database. The downregulation of *GADD45B* and *PPP3CC* are not related to gender, race and tumor stages. The statistical differences of two and three

groups were tested through the Wilcox test and Kruskal-Wallis test, respectively. **** represents $p < 0.0001$, *** represents $p < 0.001$, ** represents $p < 0.01$.

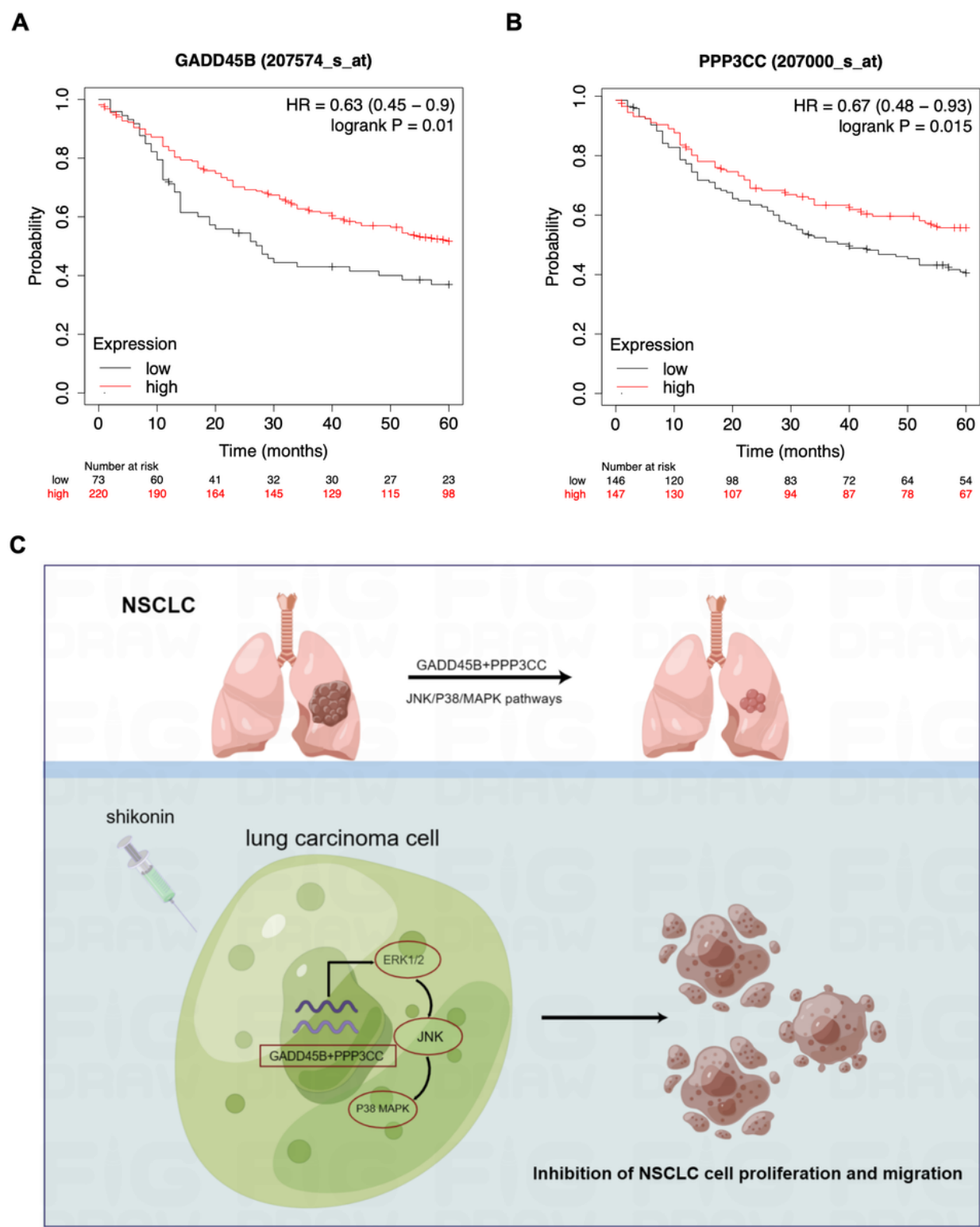


Figure 7

TSGs *GADD45B* and *PPP3CC* are correlated with patient prognosis in clinic. A-B. *GADD45B* and *PPP3CC* expression was positively correlated with patient survival outcomes by Kaplan–Meier plots. C. The

schematic diagram of the present study. Shikonin treatment activates the expression of *GADD45B* and *PPP3CC* through the JNK/P38/MAPK pathway, thus inhibiting the growth and migration of NSCLC cells.

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [supplementarymaterials.zip](#)