

RESEARCH ARTICLE

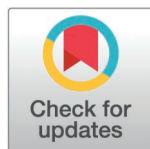
Ethnomedicinal knowledge of plants used in alternative medicine as uterotonics in Lubumbashi, Haut-Katanga, Democratic Republic of Congo

Bashige Chiribagula Valentin^{1*}, Okusa Ndjolo Philippe^{1‡}, Cedrick Shakalenga Mutombo^{2‡}, Many Mboni Henry^{2‡}, Numbi wa Ilunga Evodie^{2‡}, Bakari Amuri Salvius^{2‡}, Jean Claude Nkwanga^{2‡}, Muyumba Singoy Sarah^{1‡}, Mwamba Maseho Faustin^{2‡}, Biayi Benaja Martin^{2‡}, Sangwa Kamulete Gregoire^{2‡}, Mbayo Kalubandika Glaubert^{2‡}, Muhona Melman^{1‡}, Lumbu Simbi Jean Baptiste^{3‡}

1 Department of Pharmacology, Laboratory of Therapeutic Chemistry and Analysis of Natural Substances, Faculty of Pharmaceutical Sciences (UNILU), Commune Kampemba, Lubumbashi, Democratic Republic of Congo, **2** Department of Pharmacology, Laboratory of Pharmacognosy, Faculty of Pharmaceutical Sciences, University of Lubumbashi (UNILU), Commune Kampemba, Lubumbashi, Democratic Republic of Congo, **3** Department of Chemistry, Faculty of Sciences, University of Lubumbashi (UNILU), Commune of Lubumbashi, Democratic Republic of Congo

‡ These authors also contributed equally to this work.

* bashige.chiribagula@unilu.ac.cd



OPEN ACCESS

Citation: Valentin BC, Philippe ON, Mutombo CS, Henry MM, Evodie Nwl, Salvius BA, et al. (2026) Ethnomedicinal knowledge of plants used in alternative medicine as uterotonics in Lubumbashi, Haut-Katanga, Democratic Republic of Congo. PLoS One 21(8): e0355373. <https://doi.org/10.1371/journal.pone.0355373>

Editor: Ramesh Kumar Ahirwar, Guru Ghasidas Vishwavidyalaya, INDIA

Received: March 14, 2026

Accepted: July 20, 2026

Published: August 14, 2026

Peer Review History: PLOS recognizes the benefits of transparency in the peer review process; therefore, we enable the publication of all of the content of peer review and author responses alongside final, published articles. The editorial history of this article is available here: <https://doi.org/10.1371/journal.pone.0355373>

Copyright: © 2026 Valentin et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution,

Abstract

Background

Traditional medicine constitutes a principal component of primary healthcare in Democratic Republic of Congo (DRC), particularly in Lubumbashi. However, ethnobotanical knowledge related to its use remains incompletely documented and underutilized. The present study aimed to systematically document medicinal plants used as uterotonic among residents of Lubumbashi.

Methods

Ethnobotanical data were collected through structured interviews with 207 informants (housewives, $n = 103$; herbalists, $n = 26$; traditional healers, $n = 78$; sex ratio M/F = 0.5; mean age 35.6 ± 4.3 years; mean professional experience 13 ± 4.2 years).

Results

Informants cited 98 medicinal plant species, predominantly trees (38 species), distributed across 43 botanical families, with Fabaceae being the most represented family (16 species). Malaria was the most frequently reported non-uterotonic indication (46 species). A total of 122 traditional oxytocic formulations were recorded; most involved leaves (43 recipes) and were principally administered as cataplasm (44 recipes). To our knowledge, 34 species were reported for the first time with oxytocic

and reproduction in any medium, provided the original author and source are credited.

Data availability statement: All relevant data are within the paper and its [Supporting Information](#) files.

Funding: The author(s) received no specific funding for this work.

Competing interests: The authors have declared that no competing interests exist.

uses, of which eight are documented for the first time as medicinal plants in DRC. Among taxa, *Hylodesmum repandum* (relative citation frequency, RCF=0.20) and *Crassocephalum montuosum* (RCF=0.19) exhibited the highest citation frequencies, while *Securidaca longepedunculata* (use value, UV=0.27) and *Hibiscus sabdariffa* (UV=0.25) showed the greatest overall use values.

Conclusion

These findings provide a baseline inventory for pharmacological, phytochemical, and toxicological studies, and support prioritization of bioassay-guided validation and conservation strategies for culturally important species.

1. Introduction

Childbirth remains a major public health challenge in Sub-Saharan Africa and other low- and middle-income countries, including the Democratic Republic of Congo (DRC). Despite advances in modern medicine, maternal and neonatal mortality rates remain high, with a reported rate of 444 maternal deaths per 100,000 births over the past decade [1–3]. The main causes include hemorrhage, hypertensive disorders, infections, unsafe abortions and obstructed labor. These complications which are common due to socioeconomic inequalities, poverty, limited access to healthcare facilities and a shortage of qualified staff, particularly in rural areas [4,5], could be prevented with rapid access to quality care [5].

Uterotonics are substances that stimulate myometrial receptors, thereby increasing uterine tone and contractions [6]. These include oxytocin, Carbetocin, misoprostol, ergot and prostaglandin derivatives [7]. Concurrently, pregnant women also utilize natural substances to facilitate childbirth [8,9] reflecting their reliance on traditional medicine due to the constraints imposed by modern medical options.

In addition, conventional medicine has advanced obstetric care through technologies such as prenatal imaging, electronic fetal monitoring, and artificial intelligence algorithms that support conventional uterotonics [10,11]. These advances are inaccessible to a significant proportion of the African population, particularly in DRC, where human and material resources are severely limited [12].

In DRC, traditional medicine plays a central role in childbirth management using plants with uterotonic properties. A review of the literature from 1980 to 2010 identified 280 African oxytocic plants [13], while another reported 16 plants [14] of which the following demonstrated experimental uterotonic activity: *Heimia salicifolia* Link, Lythraceae [15], the roots of *Azanza garckeana* (F. Hoffm.) Exell & Hillc., a synonym for *Thespesia garckeana* F. Hoffm, Malvaceae [16], as well as the stem bark of *Viburnum opulus* L., Viburnaceae [17].

In the city of Lubumbashi, to the best of our knowledge, there has been no recorded study systematically documenting uterotonic plants. However, an ethnobotanical investigation conducted by Nama Mwengu (2021) within the Ruashi commune of Lubumbashi documented six plant species traditionally employed

for their uterine effects: *Phyllanthus muellerianus* (Kuntze) Exell, *Hibiscus hiernianus* Exell & Mendonça (syn. *Hibiscus gillettii* subsp. *hiernianus*), *Acacia macrocentra* Walp (Syn. *Mimosa arenosa* var. *arenosa*), *Pseudolachnostylis maprouneifolia* Pax, *Strychnos innocua* Delile, and *Annona senegalensis* Pers [18]. Complementarily, a subsequent survey undertaken in Kipushi in 2017, a locality proximal to Lubumbashi, identified eleven plants reputed to facilitate childbirth. Of these, six species: *Strychnos cocculoides* Baker, *Uapaca kirkiana* Müll.Arg, *Phyllanthus muellerianus*, *Ficus capensis* Thunb (Syn. *Ficus sur* Forssk), *Byrsocarpus orientalis* (Baill.) Baill (Syn. *Rourea orientalis* Baill), and *Aframomum alboviolaceum* (Ridl.) K.Schum, were specifically cited for their putative uterotonic properties [19].

This study aimed to identify medicinal plants and recipes with uterotonic effects, used in Lubumbashi's traditional medicine to facilitate childbirth.

2. Materials and methods

2.1. Study area

The study was conducted in the seven communes of Lubumbashi: Annexe, Kampemba, Katuba, Kamalondo, Kenya, Lubumbashi, and Ruashi, in the Haut-Katanga province in the Democratic Republic of Congo (Fig 1).

Lubumbashi is located between 11°26' and 11°55' South latitude and 27°15' and 27°40' East longitude, at an altitude of approximately 1,230 m above sea level. The region has a tropical climate characterized by a rainy season (November–April) and a dry season (May–October), with a mean annual temperature of 22.4 °C and average annual rainfall of approximately 512 mm. The dominant vegetation type is Miombo woodland, a drought-adapted ecosystem that constitutes an important source of medicinal plant diversity for local communities [20,21].

2.2. Ethnobotanical data collection

Between November 2024 and October 2025, a cross-sectional descriptive ethnobotanical survey was conducted among 207 informants comprising, 103 women who had experienced difficult labor and used medicinal plants to facilitate delivery, 26 herbalists, and 78 traditional healers, through direct, open-ended interviews guided by a Semi-structured questionnaire (S1 Table). This survey was pre-tested with 10 women who had experienced difficult childbirth and 5 traditional healers, which helped improve the informants' understanding and compliance.

To determine the minimum sample size to interview among women who have previously experienced difficulties in childbirth, we used probabilistic sampling, given that their number is unknown but estimated to be substantial. This sample size was calculated using the following Cochran formula:

$$n = \frac{z^2 * p * (1 - p)}{e^2} \quad (1)$$

($z = 1.96$ for 95% confidence, $p = 0.50$, $e = 0.05$), yielding 196 informants. These women were randomly selected from different households in Lubumbashi. Of the 196 women consulted, only 103 (52.5%) reported using plants to accelerate uterine contractions.

The herbalists included in this study were recruited from six major traditional medicine markets in Lubumbashi (Mzee, Kenya, Zambia, Kamalondo, Kalala Banza, and Moise), which serve as dynamic hubs of ethnobotanical knowledge and living plant traditions. We approached all herbalists encountered in these markets who consented to participate. In total, thirty-six practitioners were interviewed. Through direct interviews, they shared the therapeutic recipes they employ, revealing that twenty-six (72%) reported at least one specific uterotonic herbal formulation.

Traditional healers included in this study were initially selected from a pre-established list of 50 practitioners compiled during a previous survey [20]. This process resulted in the identification of 47 traditional healers, of whom only 45 reported

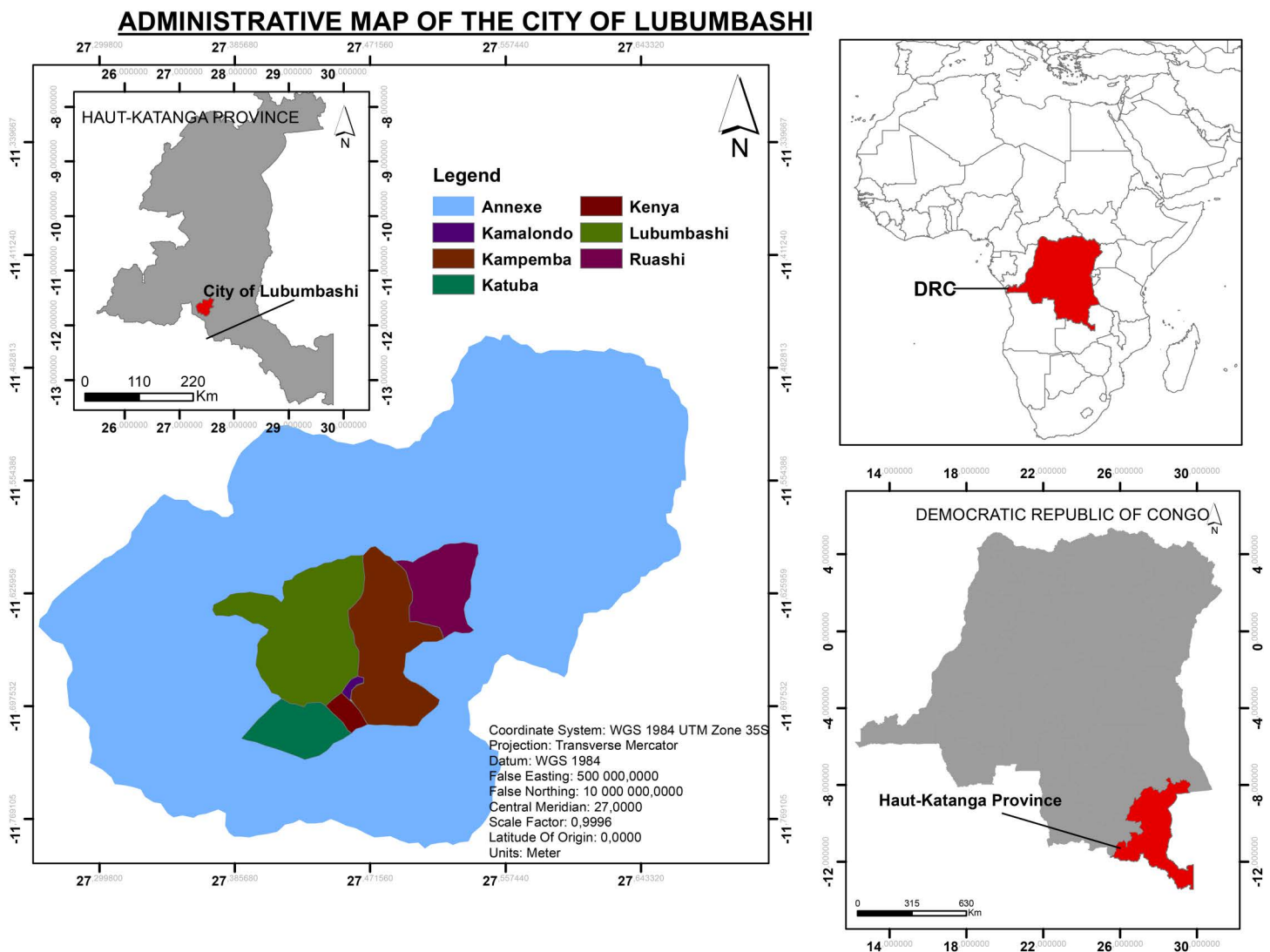


Fig 1. Lubumbashi, Haut-Katanga province in Democratic Republic of Congo.

<https://doi.org/10.1371/journal.pone.0355373.g001>

using uterotonic medicinal plants. To increase the sample size, snowball sampling was subsequently employed, resulting in the inclusion of 33 additional healers, one of whom had already appeared on the initial list.

2.3. Data processing and analysis

Plant specimens were collected in the presence of informants, photographed, pressed, and dried following standard herbarium procedures. Taxonomic identification was performed by qualified botanists at the Kipopo Herbarium (Lubumbashi, DRC). Voucher specimens were deposited under assigned reference numbers. Scientific names were verified and updated using Plants of the World Online (POWO: <https://powo.science.kew.org/>) and World Flora Online databases (WFO: <http://www.worldfloraonline>).

To determine the status of each inventoried taxon in relation to the contemporary scientific record, the accepted botanical name of each species was systematically queried in principle bibliographic databases (Google Scholar, PubMed, Scopus). For every species, we documented the presence or absence of previous ethnobotanical reports of use as a

uterotonic; experimentally verified uterotonic activity; documentation of medicinal use within the Democratic Republic of Congo; documentation of medicinal use specifically from Lubumbashi; and documentation of medicinal use from regions outside the DRC. These categorical assessments were employed to characterize the current state of knowledge for each plant.

Given the current controversy surrounding ethnobotanical indexes [22] We have used only two ethnobotanical indices to assess the significance of the reported species: the Use Value (UV) and the Relative Citation Frequency (RCF). However, apart from these two ethnobotanical indices, we have expressed the results as percentages (%).

The formula determines the use value (UV):

$$UV = \frac{\text{Number of uses mentioned by an informant for a plant}}{\text{Number of informants who cited the plant}} \quad (2)$$

The Relative citation frequency on the plant (organ) (RCF) was calculated by the formula:

$$RCF = \frac{\text{Number of informant who cited the plant}}{\text{Number of informants who took part in the survey}} \quad (3)$$

In this study, the UV index is used to assess the medicinal importance of a plant in the study area; the RCF index measures a plant's popularity among people who use it as uterotonic. This also implies a consensus among informants regarding its use to stimulate uterine contractions during childbirth.

2.4. Ethics considerations

Ethical approval for this study was granted by the Research Medical Ethics Commission of the University of Lubumbashi (UNILU/CEM/035/2024), and we obtained informed verbal consent from all 207 participants after clearly explaining the study's purpose and procedures. Any individual who declined to consent or had no prior experience using medicinal plants to aid childbirth was respectfully excluded. We reassured each informant that their insights would be used solely for academic research, that personal identifiers would be removed, and that no personal information would be shared with third parties.

3. Results

3.1. Profile of inventoried medicinal plants

A total of 98 plant species were identified as being used as uterotonic agents in traditional medicine in Lubumbashi, following ethnobotanical surveys of medicinal plants of traditional practitioners, herbalists, and housewives. Of these species, 61 were reported by traditional practitioners (35 of which were also mentioned by the other categories of informants), 35 by herbalists (24 of which were mentioned by the other categories of informants), and 28 by housewives (19 of which were mentioned by the other categories of informants). The general characteristics of these plants and their status as prior knowledge (bibliographic) in relation to uterotonic use are shown in [Table 1](#).

3.1.1. Taxonomic diversity of medicinal plants recorded. The 98 plant species identified in this survey are distributed across 98 genera and 43 botanical families. The most represented family is Fabaceae, with a relative citation frequency (RCF) of 0.92, encompassing 16 species. This is followed by Asteraceae (RCF: 0.81, 8 species), and Phyllanthaceae (RCF: 0.54; 8 species ([Fig 2](#)).

3.1.2. Variation in geographical, morphological types, and nomenclature. The plant species documented in our ethnobotanical survey are classified into 11 geographical types. The TA type is the most prevalent, with 49 species, and an RCF of 1.0. They exhibit eight distinct morphological types, with the tree form (38 species; RCF: 1.0) as the most

Table 1. General information of plants used as uterotonic agents in Lubumbashi.

Species (Family, HC)	Appellation (Ethnicity) ^{GT, MT}	N ^o Recipe (n=107) ^{UP,P}	N _c	RCF (n=207)	N _u	UV (n=60)	Non uterotonic indication	Ref.
<i>Abrus precatorius</i> L (Fabaceae, KIP000000489)	Buhiti (Lamba) ^{h,p}	R1 ^{w,ε}	9	0.04	9	0.15	Bronchitis, jaundice, hepatic diseases, contraception, tumors, sexual dysfunction, malaria, and snake bites	†: [23,24] ‡: [24,25] ¥: [26]
<i>Adenia gummiifera</i> (Harv.) Harms (Passifloraceae, KIP000000453)	Kimboyi (Lala) ^{k,l}	R2 ^{α,μ}	9	0.04	8	0.13	Infertility, sexually transmitted infections, gastrointestinal diseases, leprosy, respiratory infections, malaria and menstrual problems	†: NR ‡: NR ¥: [26]
<i>Aerva javanica</i> (Burm.f.) Juss. Ex Schult. (Amaranthaceae, KIP000000601)	Kapoka (Bemba) ^{c,q}	R3 ^{x,δ}	8	0.04	9	0.15	Cancer, wound, helminth, atherosclerosis, diabetes, hyperlipidemia, fungal infection, and hepatic diseases	†: NR ‡: [27] ¥: [28] ₯: [28,29]
<i>Aframomum albobolaceum</i> (Ridl.) K.Schum. (Zingiberaceae KIP000001433)	Ntundulu (Kikongo) ^{f,o}	R4 ^{α,δ}	9	0.04	14	0.23	Malaria, helminth, fever, Dysmenorrhea, snake bites, headache, cough, gastritis, amoebic dysentery, hemorrhoids, myoma, and pruritic.	†: NR ‡: NR ₯: [30]
		R5 ^{z,ε}						
<i>Afrothismia mhorana</i> Cheek [Syn <i>Rauvolfia caffra</i> L.] (Burmanniaceae, KIP000000602)	Mutalala (Bemba) ^{b,o}	R6 ^{α,δ}	12	0.06	4	0.07	Fever, headaches, and gastrointestinal diseases	†: NR ‡: NR ₯: [31]
<i>Ageratum conyzoides</i> L. (Asteraceae, KIP000111014)	Uratulu (Kikongo) ^{d,l}	R7 ^{x,δ}	28	0.14	4	0.07	Wounds, skin infections, and respiratory ailments	†: NR ‡: [32] ₯: [33]
<i>Albizia adianthifolia</i> (Schumacher.) W.Wight (Fabaceae, KIP000001031)	Kapeta nzovu (Bemba) ^{f,r}	R8 ^{β,ψ}	29	0.14	11	0.18	Diabetes, eye problems, gastrointestinal diseases, hemorrhoids, headache, neurodegenerative disorders, respiratory ailments, wounds, skin diseases, sexually transmitted infections	†: NR ‡: NR ₯: [33]
<i>Albizia lebbek</i> (L.) Benth (Fabaceae, KIP000111016)	Siriza (Lamba) ^{d,r}	R9 ^{w,ε}	11	0.05	11	0.18	Fever, diarrhea, dysentery, asthma, cold, cough, conjunctivitis, arthritis, dental problems and skin disease	†: NR ‡: NR ¥: [34]
		R10 ^{w,λ}						
<i>Alchornea floribunda</i> Müll.Arg (Euphorbiaceae, KIP000000603)	Lubumbidi (Kiyombe) ^{f,p}	R11 ^{γ,μ}	17	0.08	3	0.05	Inflammatory diseases, fungal ailments, and diarrhea	†: NR ‡: NR ¥: [35]
<i>Allium sativum</i> L (Amaryllidaceae)	Sabolo (Sanga) ^{d,m}	R12 ^{u,δ}	12	0.06	9	0.15	Fungal ailments, cancer, thrombosis, cardiovascular diseases, diabetes, viral diseases and parasitic diseases	†: [36] ‡: [37] ₯: [38]
<i>Aloe chabaudii</i> Schönland (Asphodelaceae, KIP000000604)	Kilambwe ne Byana (Bemba) ^{f,n}	R13 ^{γ,δ}	17	0.08	8	0.13	Skin diseases, Inflammatory diseases, respiratory ailments, urinary diseases, Poisoning, snake bites, cardiovascular diseases	†: NR ‡: NR ₯: [39,40]
<i>Aloe vera</i> (L.) Burm.f (Asphodelaceae, KIP000000605)	Cizimyamuliro (Shi) ^{d,n}	R14 ^{γ,δ}	15	0.07	11	0.18	Skin diseases, Inflammatory diseases, respiratory ailments, urinary diseases, Poisoning, snake bites, cardiovascular diseases, wounds, diabetes, sexually transmitted infections	†: [41] ‡: [42] ₯: [43]
<i>Anchusa ovata</i> Lehm (Boraginaceae, KIP000000606)	Kandasole (Sanga) ^{j,l}	R15 ^{α,μ}	12	0.06	6	0.10	Ulcer, cough, fever, wounds, skin diseases	†: NR ‡: NR ₯: [44]
		R16 ^{α,δ}						
<i>Annona senegalensis</i> Pers (Annonaceae, KIP452120003)	Mulolo (Bemba) ^{i,r}	R17 ^{γ,ε}	7	0.03	7	0.12	Prostate cancers, respiratory diseases, poisoning, rheumatic pains, infertility care and syphilis	†: NR ‡: [32] ₯: [43]

(Continued)

Table 1. (Continued)

Species (Family, HC)	Appellation (Ethnicity) ^{GT, MT}	N ^o Recipe (n = 107) ^{UP, P}	N _c	RCF (n = 207)	N _u	UV (n = 60)	Non uterotonic indication	Ref.
<i>Antidesma venosum</i> E.Mey. ex Tul (Phyllanthaceae, KIP293612638)	Kifubia (Luba-Kat) ^{h,p}	R18 ^{β, ε}	35	0.17	7	0.12	Typhoid, gastrointestinal diseases, diabetes, pneumonia, wounds, and sexually transmitted infections	†: NR ‡: NR T: [33]
<i>Asparagus africanus</i> Lam. (Asparagaceae, KIP000111026)	Atangwali (Hemba) ^{h,p}	R19 ^{γ, ε}	5	0.02	17	0.28	Malaria, helminth, fever, Dysmenorrhea, headache, cough, gastritis, amoebic dysentery, stomach pain, ulcers, hemorrhoids, myoma, syphilis, gonorrhea, and pruritis.	†: [45] ‡: [46,47] T: [48]
<i>Azadirachta indica</i> A. Juss. (Meliaceae, KIP000000607)	Nfwama (Sabwa) ^{d,r}	R20 ^{γ, ε}	4	0.02	13	0.22	Fever, malaria, wounds, headache, ulcers, respiratory diseases, cancer, diabetes, leprosy, dengue fever, chickenpox, and skin diseases	†: [41] ‡: [27] T: [43]
		R21 ^{z, λ}						
<i>Baphiopsis parviflora</i> Benth. ex-Baker (Fabaceae, KIP000000608)	Waase (Tabwa) ^{a,r}	R22 ^{β, ε}	5	0.02	9	0.15	Wounds, coughs, hypertension, erectile dysfunction, skin infections, urinary tract infections, respiratory ailments, and gastro-intestinal diseases.	†: NR ‡: NR T: [49,50]
<i>Bidens pilosa</i> L. (Asteraceae, KIP000000609)	Mbamba ngoma (Luba-Kat) ^{d,l}	R23 ^{γ, θ}	5	0.02	6	0.10	Intoxication, hypertension, dysentery, hepatic diseases, ulcers	†: [41,51] ‡: [52] T: [30]
<i>Bobgunnia madagascariensis</i> (Desv.) J.H.Kirkbr. & Wiersema (Fabaceae, KIP000000610)	Munienze (Luba-Kat) ^{f,r}	R24 ^{β, ε}	6	0.03	9	0.15	Bilharzia, leprosy, cataract, dysentery, headache, cough, diarrhea, and sexually transmitted infections	†: NR ‡: NR T: [30]
<i>Boerhavia diffusa</i> L. (Nyctaginaceae, KIP000000611)	Alini (Bemba) ^{h,l}	R25 ^{γ, δ}	5	0.02	8	0.13	Cough, hypertension, malaria, fever, diabetes, wound, gastro-intestinal diseases	†: NR ‡: NR T: [53]
<i>Bombax ceiba</i> L. (Malvaceae, KIP000000612)	Mulul ilav (Rund) ^{d,r}	R26 ^{γ, ε}	5	0.02	6	0.10	Cholera, coughs, urinary tract infections, dysentery, and gastro-intestinal diseases	†: NR ‡: NR T: [54]
<i>Bothriocline longipes</i> (Oliv. & Hiern) N.E.Br. (Asteraceae, KIP000000613)	Lwibaye (Shi) ^{f,q}	R27 ^{γ, θ}	6	0.03	14	0.23	Fever, malaria, wound, headache, ulcers, respiratory diseases, cancer, diabetes, leprosy, dengue fever, chickenpox, and skin diseases, stomach	†: [55] ‡: [25] T: [56]
		R28 ^{α, δ}						
<i>Cajanus cajan</i> (L.) Huth (Fabaceae, KIP000000614)	Golyolyo (Luba-Kat) ^{d,n}	R29 ^{z, ε}	7	0.03	11	0.18	Diabetes, malaria, ulcers, gastro-intestinal diseases, hemorrhoids, headache, respiratory ailments, wounds, skin diseases, sexually transmitted infections	†: [57] ‡: [58] T: [38]
<i>Capsicum annum</i> L. (Solana-ceae, KIP000000615)	Pilipili (Swahili) ^{d,l}	R30 ^{z, δ}	12	0.06	6	0.10	Cancers, wounds, fever, malaria, stomach	†: NR ‡: [32] T: [59]
<i>Cassia sieberiana</i> DC (Faba-ceae, KIP000000616)	Kafunga nan-shi (Bemba) ^{f,r}	R31 ^{γ, ε}	9	0.04	8	0.13	Fever, indigestion, constipation, Drepanocyto-sis, diabetes, malaria, and skin diseases	†: NR ‡: NR T: [26]
<i>Cissampelos mucronata</i> A.Rich. (Menispermaceae, KIP000000617)	Mukoko (Luba-Kat) ^{h,n}	R32 ^{γ, δ}	7	0.03	6	0.10	Diarrhea, dysentery, sexually transmitted infections, infertility, fever	†: [60] ‡: [58] T: [61,62]
<i>Cissus schmitzii</i> Dewit (Vitaceae, KIP000000618)	Lenda (Luba-Kat) ^{a,p}	R33 ^{α, ε}	9	0.04	7	0.12	Wounds, ulcers, fever, respiratory ailments, sexual dysfunction, diabetes	†: NR ‡: NR T: [63]

(Continued)

Table 1. (Continued)

Species (Family, HC)	Appellation (Ethnicity) ^{GT, MT}	N ^o Recipe (n = 107) ^{UP,P}	N _c	RCF (n = 207)	N _u	UV (n = 60)	Non uterotonic indication	Ref.
<i>Clausena anisata</i> (Willd.) Hook.f. ex Benth (Rutaceae, KIP000000619)	Umusanga (Bemba) ^{f,r}	R34 ^{y,ε}	10	0.05	5	0.08	Hypertension, respiratory ailments, fever, stomach	†: NR ‡: [64] §: [64]
<i>Clematis villosa</i> subsp. <i>Villosa</i> (Ranunculaceae, KIP000000620)	Karhula (Shi) ^{a,n}	R35 ^{y,θ}	11	0.05	7	0.12	Cancer, Pain, Inflammatory diseases, headaches, skin diseases, sexually transmitted infections	†: NR ‡: [65] §: [65]
<i>Cochlospermum planchonii</i> Hook.f. ex Planch (Bixaceae, KIP000000621)	Sowaga (Tabwa) ^{f,q}	R36 ^{y,θ}	15	0.07	6	0.10	Malaria, diarrhea, hepatic diseases, Inflammatory diseases, stomach	†: NR ‡: NR §: [66,67]
<i>Combretum molle</i> R.Br. ex G.Don (Combretaceae, KIP000000622)	Mulema (Bemba) ^{f,p}	R37 ^{y,μ}	14	0.07	8	0.13	Drepanocytosis, diarrhea, helminth, wound, ulcers Poisoning, headaches	†: NR ‡: NR §: [38]
<i>Commelina africana</i> L (Com-melinaceae, KIP000000623)	Idangabani (Bemba) ^{i,n}	R38 ^{y,ε}	13	0.06	6	0.10	Drepanocytosis, urinary tract infections, diarrhea, hemorrhoids, stomach	†: [41] ‡: [68] §: [69]
<i>Crassocephalum montuosum</i> (S.Moore) Milne-Redh. (Asteraceae, KIP437629969)	Cifula (Shi) ^{i,l}	R39 ^{y,δ}	39	0.19	14	0.23	Fever, malaria, wound, headache, ulcers, respiratory diseases, cancer, diabetes, leprosy, rheumatism, dengue fever, chickenpox, and skin diseases	†: NR ‡: NR §: [70]
		R40 ^{y,δ}						
<i>Crassocephalum vitellinum</i> (Benth.) S.Moore (Asteraceae, KIP000000624)	Shungululu (Shi) ^{f,l}	R41 ^{y,δ}	47	0.23	7	0.12	Fever, malaria, sexually transmitted infections, wounds, gastro-intestinal diseases, stomach	†: NR ‡: [25] §: [65]
<i>Cucurbita pepo</i> L (Cucurbitaceae, KIP000000625)	Liboga (Swahili) ^{d,l}	R42 ^{z,δ}	11	0.05	7	0.12	hepatic diseases, diabetes, cancer, ulcer, wound, stomach	†: NR ‡: [71] §: [72]
<i>Cyanthillium cinereum</i> (L.) H.Rob. [syn <i>Vernonia cinerea</i> (L.) Less] (Asteraceae, KIP000000626)	Musubgubulu (Lubakas) ^{a,n}	R43 ^{y,δ}	13	0.06	5	0.08	Urinary tract infections, diarrhea, cough, respiratory diseases, stomach	†: NR ‡: NR §: [38]
<i>Cymbopogon citratus</i> (DC.) Stapf (Poaceae, KIP000000627)	Masela mputu (Lubakas) ^{d,n}	R44 ^{a,ε}	11	0.05	8	0.13	Fever, stomach, headache, malaria, inflammatory diseases, cough, Amoebae	†: [73] ‡: [32] §: [20,33]
<i>Cynodon dactylon</i> (L.) Pers. (Poaceae, KIP000000628)	Lukaka (Sanga) ^{e,l}	R45 ^{x,δ}	12	0.06	6	0.10	Inflammatory diseases, fungal ailments, hypertension and diarrhea, wound	†: NR ‡: [74] §: [75]
<i>Dalbergia boehmii</i> Taub (Faba-ceae, KIP000000629)	Katambo (Zela) ^{f,q}	R46 ^{β,δ}	12	0.06	7	0.12	Diabetes, urinary tract infections, skin diseases, pain, Diarrhea, fever	†: NR ‡: [74] §: [38]
<i>Diplorhynchus condylocarpon</i> (Müll.Arg.) Pichon [Syn <i>Diplorhynchus mossambicensis</i> Benth] (Apocynaceae, KIP361753009)	Mwenge (Sanga) ^{f,p}	R47 ^{y,δ}	14	0.07	6	0.10	Headache, snake bites, hepatic diseases, respiratory diseases, gastrointestinal diseases	†: NR ‡: [76] §: [38,63]
<i>Ekebergia capensis</i> Sparrm (Meliaceae, KIP000000630)	Nyamaru (Bemba) ^{h,r}	R48 ^{z,δ}	13	0.06	9	0.15	Fever, malaria, stomach, skin diseases, wounds, cough, headache	†: [77] ‡: [14] §: [78]
<i>Elaeis guineensis</i> Jacq (Areca-ceae, KIP000000631)	Ekaci (Bembe) ^{f,r}	R49 ^{β,δ}	9	0.04	6	0.10	headaches, pains, rheumatism, cardiovascular diseases, wounds	: NR ‡: [79] §: [38]

(Continued)

Table 1. (Continued)

Species (Family, HC)	Appellation (Ethnicity) ^{GT, MT}	N° Recipe (n = 107) ^{UP, P}	N _c	RCF (n = 207)	N _u	UV (n = 60)	Non uterotonic indication	Ref.
<i>Eleusine indica</i> (L.) Gaertn (Poaceae, KIP000000632)	Palasi (Zela) ^{LI}	R50 ^{y, θ}	4	0.02	6	0.10	asthma, diarrhea, stomach, dysentery, respiratory diseases	†: NR ‡: [52] T: [38]
<i>Erythrina abyssinica</i> Lam (Fabaceae, KIP000000633)	Kisungwa (Bemba) ^{f, r}	R51 ^{y, μ}	5	0.02	12	0.20	Tuberculosis, malaria, HIV/AIDS, diarrhea, cancer, meningitis, inflammatory diseases, urinary tract infections, wounds, diabetes, and skin diseases.	†: NR ‡: [58] T: [38]
<i>Euphorbia hirta</i> L. (Euphorbiaceae, KIP000000634)	Butonvitonvi (Bemba) ^{f, l}	R52 ^{x, ε}	13	0.06	5	0.08	Cancer, Asthma, wounds, gastrointestinal diseases	†: NR ‡: [32, 58] T: [33, 38]
<i>Ficus capensis</i> Thunb [<i>Ficus sur</i> Forssk] (Moraceae, KIP539378804)	Mukiluakilua (Bemba) ^{h, r}	R53 ^{y, δ}	37	0.18	8	0.13	Drepanocytosis, diarrhea, fever, cough, ulcers, skin diseases, stomach	†: NR ‡: [32] T: [38]
<i>Ficus stuhlmannii</i> Warb (Moraceae, KIP000111046)	Mupulampaka (Bemba) ^{f, r}	R54 ^{y, δ}	39	0.19	6	0.10	Drepanocytosis, wounds, hemorrhoids, diabetes, hepatic diseases	†: NR ‡: NR T: [33, 63]
<i>Flueggea virosa</i> (Roxb. ex Willd.) Royle (Phyllanthaceae, KIP000000635)	Kasansubwanga (Bemba) ^{f, p}	R55 ^{y, ε}	12	0.06	5	0.08	Fever, gastrointestinal diseases, skin diseases, stomach	†: NR ‡: NR T: [38]
<i>Grona adscendens</i> (Sw.) H. Ohashi & K. Ohashi (Syn <i>Desmodium adscendens</i> (Sw.) DC] (Fabaceae, KIP000000636)	Busundu (Shi) ^{f, n}	R56 ^{y, δ}	17	0.08	5	0.08	asthma, diarrhea, stomach, dysentery, respiratory diseases	†: NR ‡: NR T: [80, 81]
<i>Gymnanthemum amygdalinum</i> (Delile) Sch. Bip. [Syn <i>Vernonia amygdalina</i> Delile] (Asteraceae, KIP312286606)	Mubirizi (Shi) ^{f, p}	R57 ^{β, ε}	20	0.10	9	0.15	fever, malaria, diarrhea, cough, dysentery, headache, stomach pains, and hepatic diseases	†: [41, 68, 82] ‡: [58] T: [38]
<i>Harungana madagascariensis</i> Lam. Ex Poir (Hypericaceae, KIP000000637)	Kafifi (Bemba) ^{f, r}	R58 ^{y, ε}	8	0.04	6	0.10	Drepanocytosis, malaria, gastrointestinal diseases, diabetes, hepatic diseases	†: NR ‡: NR T: [38]
<i>Hexalobus monopetalus</i> (A. Rich.) Engl. & Diels (Annonaceae, KIP000000638)	Nkoyo (Luba-kat) ^{f, r}	R59 ^{α, μ}	19	0.09	5	0.08	Fever, headache, gastrointestinal diseases, stomach	†: NR ‡: NR T: [63]
<i>Hibiscus sabdariffa</i> L (Malvaceae, KIP000111148)	Mulenda (Swahili) ^{f, l}	R60 ^{y, θ}	8	0.04	15	0.25	Cancers, wounds, fever, malaria, stomach pain, cardiovascular diseases, diabetes, viral diseases, parasitic diseases, gastrointestinal diseases, diabetes, hyperlipidemia, and hypertension.	†: [83] ‡: [27] T: [84]
<i>Hylodesmum repandum</i> (Vahl) H. Ohashi & R. R. Mill [<i>Desmodium repandum</i> (Vahl) Poir.] (Fabaceae, KIP000000638)	Irhuzi (Shi) ^{f, n}	R61 ^{β, ε}	41	0.20	6	0.10	Fever, ulcers, gastrointestinal diseases, malaria, stomach	†: NR ‡: NR T: [85]
<i>Hymenocardia acida</i> Tul (Phyllanthaceae, KIP464414491)	Kapempe (Shi) ^{f, r}	R62 ^{β, μ}	11	0.05	5	0.08	Malaria, skin diseases, hemorrhoids, Parasites diseases, parasitic diseases, and diabetes	†: NR ‡: NR T: [86]
<i>Juglans nigra</i> L (Juglandaceae, KIP000000639)	Nusumbau (Bemba) ^{d, r}	R63 ^{z, μ}	12	0.06	5	0.08	Parasitic diseases, gastrointestinal diseases, diabetes, ulcers	†: NR ‡: NR T: [87]

(Continued)

Table 1. (Continued)

Species (Family, HC)	Appellation (Ethnicity) ^{GT, MT}	N ^o Recipe (n = 107) ^{UP, P}	N _c	RCF (n = 207)	N _u	UV (n = 60)	Non uterotonic indication	Ref.
<i>Khaya nyasica</i> Stapf ex Baker f (Meliaceae, KIP000001372)	Mbamba (Luba-kat) ^{b, r}	R64 ^{y, ε}	36	0.17	8	0.13	Cancers, diabetes, Drepanocytosis, ulcers, hepatic diseases, gastrointestinal diseases	†: NR ‡: NR T: [33,38]
<i>Kigelia aethiopum</i> (Fenzl) Dandy [<i>Kigelia africana</i> subsp. <i>Africana</i>] (Bignoniaceae, KIP000001376)	Kivungwe (Tabwa) ^{f, r}	R65 ^{y, ε}	27	0.13	7	0.12	Wounds, fever, gastrointestinal diseases, skin diseases, respiratory diseases, cancers	†: NR ‡: NR T: [87]
<i>Kigelia africana</i> (Lam.) Benth (Bignoniaceae, KIP000000640)	Kivungwila (Luba-kat) ^{f, r}	R66 ^{β, θ}	51	0.25	10	0.17	Malaria, Wounds, fever, gastrointestinal diseases, skin diseases, respiratory diseases, cancers, ulcers, urinary tract infection	†: NR ‡: [32] T: [87]
<i>Leonotis nepetifolia</i> (L.) R.Br (Lamiaceae, KIP000000641)	Kacumucumu (Shi) ^{g, i}	R67 ^{y, θ}	18	0.09	10	0.17	Fever, cough, malaria, rheumatism, hypertension, gastro-intestinal diseases, skin diseases, respiratory diseases, pain	†: NR ‡: NR ¥: [88]
<i>Leucas martinicensis</i> (Jacq.) R.Br. (Lamiaceae, KIP000000642)	Kanyama-fundo (Shi) ^{f, i}	R68 ^{v, θ}	6	0.03	9	0.15	kidney disorders, rheumatism, cough, diarrhea, fevers, gastro-intestinal diseases, skin diseases, neurodegenerative disorders	†: NR ‡: [58] T: [65]
<i>Luffa cylindrica</i> (L.) M.Roem [<i>Cucumis melo</i> L] (Cucurbitaceae, KIP000111051)	Kikenda mbayo (Luba-kat) ^{f, i}	R69 ^{z, μ}	17	0.08	11	0.18	Respiratory diseases, urinary tract infection, ulcers, Parasites diseases, hypertension, hyperlipidemia, inflammatory diseases, viral diseases, diabetes, wounds	†: [41,89] ‡: [58] ¥: [56]
<i>Malus domestica</i> (Suckow) Borkh (Rosaceae, KIP000000643)	Chungwa nuni (Swahili) ^{d, r}	R70 ^{z, ε}	6	0.03	6	0.10	hypertension, diabetes, hepatic diseases, diarrhea, fevers	†: NR ‡: [52] T: [43]
<i>Milletia griffoniana</i> Baill. (Faba-ceae, KIP000000644)	Umuzimbe (Shi) ^{f, r}	R71 ^{β, ε}	16	0.08	4	0.07	Cancers, viral diseases, Inflammatory diseases	†: [90] ‡: [91] T: [92]
<i>Ochna schweinfurthiana</i> F.Hoffm. (Ochnaceae, KIP000000645)	Ikyoni (Bemba) ^{f, r}	R72 ^{β, ε}	7	0.03	8	0.13	Drepanocytosis, helminth, typhoid fever, malaria, skin diseases, respiratory diseases, wounds	†: NR ‡: [93,94] T: [94]
<i>Ocimum americanum</i> L (Lamiaceae, KIP000000646)	Lueni (Bemba) ^{i, j}	R73 ^{x, μ}	15	0.07	5	0.08	Wound, cough, malaria, cough	†: NR ‡: [32] T: [33]
<i>Paramollugo nudicaulis</i> (Lam.) Thulin (Molluginaceae, KIP000000647)	Eswani (Tabwa) ^{i, j}	R74 ^{x, μ}	9	0.04	5	0.08	Coughs, fever, respiratory diseases, wounds	†: NR ‡: NR T: [95]
<i>Parinari curatellifolia</i> Planch. ex Benth. (Chrysobalanaceae, KIP000001385)	Mupundu (Luba-kat) ^{f, r}	R75 ^{β, δ}	14	0.07	4	0.07	Diabetes, cancer, inflammatory diseases, malaria	†: NR ‡: NR T: [63]
<i>Pentaclethra macrophylla</i> Benth. (Fabaceae, KIP000111062)	Bubala (Kirega) ^{f, r}	R76 ^{y, ε}	11	0.05	9	0.15	Drepanocytosis, wounds, hemorrhoids, diabetes, hepatic diseases, helminth, diarrhea, hyperlipidemia	†: [96] ‡: [32,97] T: [98]
<i>Phyllanthus amarus</i> Schum. & Thonn. (Phyllanthaceae, KIP000111065)	Kavudi (Luba-kat) ^{d, i}	R77 ^{y, ε}	13	0.06	6	0.10	Wounds, stomach diseases, urinary tract infection, hepatic diseases, gastrointestinal diseases	: NR ‡: [32] T: [33]
<i>Phyllanthus microdendron</i> Müll.Arg. (Phyllanthaceae, KIP000000648)	Unono (Luba-kat) ^{f, r}	R78 ^{y, ε}	12	0.06	10	0.17	Wounds, stomach diseases, urinary tract infection, hepatic diseases, gastro-intestinal diseases, diabetes, viral diseases, cancers, malaria	†: NR ‡: NR T: [99]
<i>Phyllanthus muellerianus</i> (Kuntze) Exell (Phyllanthaceae, KIP277202454)	Modere (Sanga) ^{f, p}	R79 ^{α, ε}	7	0.03	11	0.18	Wounds, stomach diseases, urinary tract infection, hepatic diseases, gastro-intestinal diseases, diabetes, viral diseases, cancers, malaria, hypertension	†: NR ‡: [100] T: [30,63]

(Continued)

Table 1. (Continued)

Species (Family, HC)	Appellation (Ethnicity) ^{GT, MT}	N° Recipe (n = 107) ^{UP, P}	N _c	RCF (n = 207)	N _u	UV (n = 60)	Non uterotonic indication	Ref.
<i>Phyllanthus ovalifolius</i> Forssk [Syn <i>Phyllanthus guineensis</i>] (Phyllanthaceae, KIP000000649)	Mupetualupe (Bemba) ^{f, p}	R80 ^{y, δ}	6	0.03	11	0.18	Wounds, stomach diseases, urinary tract infection, hepatic diseases, gastro-intestinal diseases, diabetes, viral diseases, cancers, malaria, hypertension	†: NR ‡: [25] T: [38,63]
		R81 ^{z, δ}						
<i>Physalis peruviana</i> L (Solana-ceae, KIP000000650)	Imbuhu (Shi) ^{d, n}	R82 ^{z, ε}	5	0.02	10	0.17	Cancer, diabetes, ulcers, malaria, asthma, rheumatism, gastro-intestinal diseases, pain, fever.	†: NR ‡: [58] ¥: [26,101]
<i>Piliostigma thonningii</i> (Schum.) Millne-Redhead (Fabaceae, KIP000000651)	Kifumbe (Bemba) ^{f, r}	R83 ^{y, δ}	3	0.01	9	0.15	Coughs, fever, respiratory diseases, wounds, sexually transmitted infections, malaria, typhoid fever, hemorrhoids	†: NR ‡: [32] T: [20,33]
<i>Plumbago zeylanica</i> L (Plumbag-inaceae, KIP000111067)	Daunani (Zela) ^{i, p}	R84 ^{y, δ}	4	0.02	6	0.10	Hemorrhoids, inflammatory diseases, diabetes, hypertension, Drepanocytosis	†: [102,103] ‡: [25,103] ¥: [104]
<i>Psidium guajava</i> L. (Myrtaceae, KIP000301861)	Ipera (Shi) ^{f, r}	R85 ^{β, δ}	7	0.03	12	0.20	Ulcer, cough, fever, wounds, skin diseases, malaria, hepatic diseases, helminth, diarrhea, hyperlipidemia, fungal diseases	†: [105] ‡: [32] T: [38]
<i>Ricinus communis</i> L (Euphorbia-ceae, KIP000000652)	Mubalika (Bemba) ^{b, p}	R86 ^{w, ε}	9	0.04	5	0.08	bilharziasis, gastro-intestinal diseases, hepatic diseases, helminth	†: [106] ‡: [27] T: [87]
<i>Securidaca longepeduncu-lata</i> Fresen (Polygalaceae, KIP000000653)	Lupapi (Bemba) ^{f, r}	R87 ^{β, ε}	8	0.04	16	0.27	sexually transmitted infections, malaria, typhoid fever, hemorrhoids, coughs, epilepsy, gastro-intestinal diseases, headaches, skin diseases, hyperlipidemia, sexually transmitted infections, rheumatism, aphrodisiac, epilepsy, tuberculosis, ulcers, stomach pain	†: NR ‡: [107] T: [38,63]
<i>Senegalia polyacantha</i> (Willd.) Seigler & Ebinger [Syn <i>Acacia polyacantha</i> Willd] (Fabaceae, KIP000000654)	Kibombolo (Tabwa) ^{h, r}	R88 ^{y, θ}	4	0.02	8	0.13	snakebites, malaria, diabetes, wounds, skin diseases, bilharziasis, gastrointestinal diseases	†: NR ‡: [108] T: [43]
<i>Senna occidentalis</i> (L.) Link. (Fabaceae, KIP000000655)	Lukunda bajani (Luba-kas) ^{f, q}	R89 ^{y, ε}	5	0.02	9	0.15	Diarrhea, dysentery, constipation, fever, sex-ually transmitted infections, malaria, diabetes, hepatic diseases	†: [109] ‡: [32] T: [38]
		R90 ^{v, λ}						
<i>Sida acuta</i> Burm.f (Malvaceae, KIP000000656)	Tumfumu (Kikongo) ^{f, l}	R91 ^{y, ε}	21	0.10	7	0.12	Malaria, ulcers, constipation, diabetes, hemor-rhoids, coughs	†: [110] ‡: [111,112] ¥: [88]
		R92 ^{v, λ}						
<i>Solanum incanum</i> L (Solana-ceae, KIP000000657)	Ndulwe (Bemba) ^{i, q}	R93 ^{y, δ}	42	0.20	6	0.10	Viral Parasitic diseases, gastro-intestinal dis-eases, skin diseases, Inflammatory diseases	†: NR ‡: [113] T: [86]
<i>Stereospermum kunthia-num</i> Cham (Bignoniaceae, KIP000000658)	Kasengele (Lamba) ^{f, p}	R94 ^{y, δ}	12	0.06	6	0.10	ulcers, skin diseases, Inflammatory diseases, respiratory diseases, sexually transmitted infections	†: NR ‡: [27] T: [114]
<i>Strychnos spinosa</i> Lam (Logani-aceae, KIP000111072)	Kisongole (Bemba) ^{i, p}	R95 ^{z, δ}	5	0.02	5	0.08	Malaria, diabetes, snakebites, sexually trans-mitted infections	†: NR ‡: [115] T: [20,33,38]

(Continued)

Table 1. (Continued)

Species (Family, HC)	Appellation (Ethnicity) ^{GT, MT}	N ^o Recipe (n = 107) ^{UP,P}	N _c	RCF (n = 207)	N _u	UV (n = 60)	Non uterotonic indication	Ref.
<i>Syzygium cordatum</i> Hochst. ex C.Krauss (Myrtaceae, KIP000111074)	Cinyam-baganyi (Tabwa) ^{h,r}	R96 ^{z,ε}	7	0.03	9	0.15	Malaria, tuberculosis, diabetes, snakebites, sexually transmitted infections, wounds, and respiratory diseases	†: NR ‡: [41,116] T: [33]
<i>Tagetes minuta</i> L (Asteraceae, KIP000000659)	Mbaje mbuni (Zela) ^{d,l}	R97 ^{v,ε}	9	0.04	5	0.08	stomach pain, coughs, gastrointestinal diseases, pain	†: NR ‡: [58] ¥: [65]
<i>Thespesia garckeana</i> F.Hoffm. (Malvaceae, KIP000000660)	Makole (Sanga) ^{f,r}	R98 ^{a,δ}	11	0.05	6	0.10	Fever, headache, stomach pain, diabetes, and inflammatory diseases	†: NR ‡: [19] ¥: [19]
<i>Trema orientale</i> (L.) Blume (Cannabaceae, KIP000000661)	Nalita (Luba-kat) ^{i,p}	R99 ^{v,θ}	34	0.16	6	0.10	Asthma, fever, cough, Inflammatory diseases, malaria	†: [117] ‡: [32] T: [65]
<i>Uapaca kirkiana</i> Müll. Arg (Phyllanthaceae, KIP000000662)	Masuku (Bemba) ^{f,p}	R100 ^{z,μ}	15	0.07	11	0.18	stomach pain, dysentery, sexually transmitted infections, diabetes, wounds, hyperlipidemia, hypertension, Drepanocytosis, helminth, typhoid fever	†: NR ‡: [19] T: [38]
<i>Vitex congolensis</i> var. <i>thomasii</i> (De Wild.) Meerts [syn <i>Vitex thomasii</i> De Wild.] (Lamiaceae, KIP000000663)	Mufutu mukwa (Luba-kat) ^{a,r}	R101 ^{β,ε}	36	0.17	5	0.08	Cancer, Inflammatory diseases, diabetes, gastrointestinal diseases,	†: NR ‡: NR T: [118]
<i>Xylopia aethiopica</i> (Dunal) A.Rich (Annonaceae, KIP000000664)	Njilu luba (Luba-kat) ^{f,r}	R102 ^{β,ε}	7	0.03	11	0.18	rheumatism, diarrhea, dysentery, cough, asthma, hemorrhoids, malaria, diabetes, wounds, sexually transmitted infections	†: [119] ‡: [32] T: [38]
<i>Zanthoxylum chalybeum</i> Engl. (Rutaceae, KIP000001016)	Pupwe (Bemba) ^{f,p}	R103 ^{a,μ}	10	0.05	7	0.12	Drepanocytosis, cancer, malaria, diabetes, tuberculosis, gastrointestinal diseases	†: NR ‡: [25] T: [33,63]
<i>Zanthoxylum zanthoxyloides</i> (Lam.) Zepern. & Timler (Rutaceae, KIP000000665)	Pupwe bimpa (Lamba) ^{f,r}	R104 ^{β,ε}	7	0.03	6	0.10	Drepanocytosis, malaria, fever, tuberculosis, constipation	†: NR ‡: [32] T: [120]
<i>Zingiber officinale</i> Roscoe (Zingiberaceae, KIP000111077)	Tangawisi (Swahili) ^{d,o}	R105 ^{a,θ}	9	0.04	10	0.17	Drepanocytosis, cancer, malaria, diabetes, tuberculosis, gastro-intestinal diseases, wounds, Inflammatory diseases, aphrodisiac	†: [121] ‡: [32,37] T: [33]
<i>Ziziphys abyssinica</i> Hochst. ex A.Rich. (Rhamnaceae, KIP000000666)	Sula (Bemba) ^{f,r}	R106 ^{a,μ}	14	0.07	9	0.15	Drepanocytosis, cancer, diabetes, gastrointestinal diseases, tuberculosis, malaria, wounds, and hypertension	†: NR ‡: [58] T: [38]
<i>Ziziphys mucronata</i> Willd (Rhamnaceae, KIP000000667)	Kankona (Bemba) ^{f,r}	R107 ^{v,θ}	12	0.06	10	0.17	Drepanocytosis, cancer, diabetes, gastrointestinal diseases, tuberculosis, malaria, wounds, hypertension, stomach pain.	†: NR ‡: [122] T: [38,63]

Legend:

† (uterotonic Activity)

‡ (uterotonic uses)

T (medicinal uses in Lubumbashi)

¥ (medicinal uses in DRC outside Lubumbashi)

£ (medicinal uses outside DRC)

UP: used part, P: Preparation

—**Geographical type**— CA: a, EA: b; NA-MA: c; NAT: d; NA-TA-SA-MA: e; TA: f; TA-MA: g; TA-SA: h; TA-SA-MA: i; WA: j; WA-SA: k- NA: North Africa; CA: Central Africa; TA: tropical Africa; SA: southern Africa; MA: Madagascar. **Used part:** Bulb: u; Flowers: v; Seeds: w; Whole plant: x; Stem bark: y; Fruits: z; Root: q; Root bark: β; Leaves: γ **Morphological type** -Annual herb: l; Bulbous geophyte: m; Perennial herb: n; Rhizomatous geophyte: o; Shrub: p; Subshrub: q; Tree: r; Tuberous subshrub: s— **Preparation method**— Cataplasma: δ; Decoction: ε; Infusion: θ; Liniment: λ; Maceration: μ;

(Continued)

Table 1. (Continued)

Ointment: ψ . A **cataplasma** is a medicinal preparation that is applied directly to the skin. The suitability of the plant for this purpose is determined by the condition of the skin. Alternatively, they can be heated in water to soften them, and then gently crushed to extract the active ingredients. **Liniment** is a composition of oil (palm or olive) and plant powder in the proportions of 5:2. The ointment is applied to the vaginal muscle. **Ointment** is a preparation obtained from pork fat with a paste of the plant (parts 3:1). According to informants, on average, childbirth occurs within 30 minutes to two hours after the topical application of the herbal preparation.

<https://doi.org/10.1371/journal.pone.0355373.t001>

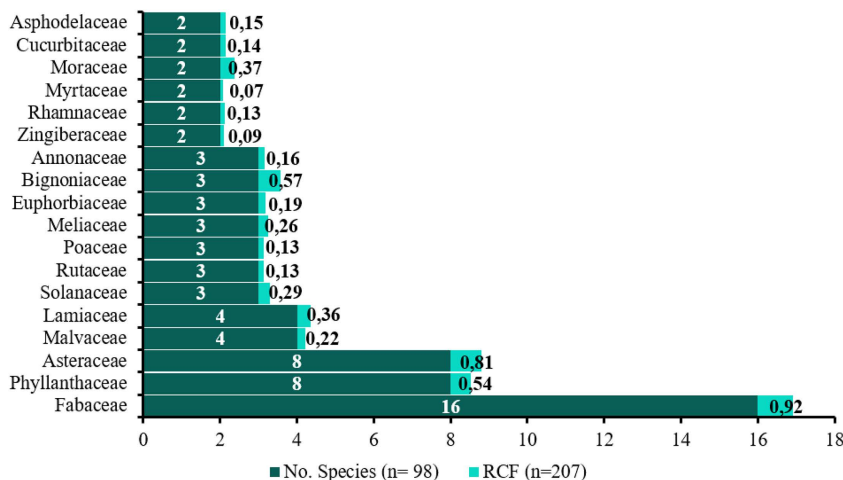


Fig 2. Relative citation frequencies of plant families with $n \geq 2$ plants.

<https://doi.org/10.1371/journal.pone.0355373.g002>

prevalent, followed by annual herbs (21 species; RCF: 1.0) and shrubs (18 species; RCF: 1.0). These species have names in 17 local Congolese languages. The most frequently used language is Bemba (30 species; RCF: 0.99), followed by Luba-Katanga (15 species; RCF: 0.97), and Shi (14 species; RCF: 0.97) (Fig 3).

3.2. Profile of herbal recipes used as uterotonics

The data collected on 98 plant species during ethnobotanical surveys, documented in this study, that are traditionally used in Lubumbashi to facilitate childbirth led to the identification of 122 uterotonics formulations. Among these, 107 recipes use a single plant (Table 1), whereas 15 formulations incorporate two plant species (Table 2).

These preparations utilize nine different plant organs, with leaves being the most frequently used (33 monoherbal recipes and 10 polyherbal recipes), followed by root bark (18 monoherbal recipes and 3 polyherbal recipes) (Fig 4).

There are six different preparation methods for plant organs, with decoction (40 recipes using a single plant, and 3 using two plants) and cataplasma (35 and 9 recipes) being the most used methods (Fig 4).

In polyherbal formulations, no plant species out of 30 is recurrent across multiple distinct preparations. The most frequently utilized formulation, designated as R199, consists of a combination of *Hymenocardia acida* root bark, and *Juglans nigra* fruits, exhibiting a relative citation frequency (RCF) of 0.33. This formulation is followed by R116, which is derived from the mixture of *Ekebergia capensis* roots, and *Erythrina abyssinica* stem bark (Table 2).

3.3. Other human ailments treated by inventoried species

Beyond their role in facilitating childbirth by producing a uterotonic effect, the 98 documented plant species are also utilized in Lubumbashi folk medicine for the treatment of 60 additional ailments. Among these, gastrointestinal diseases,

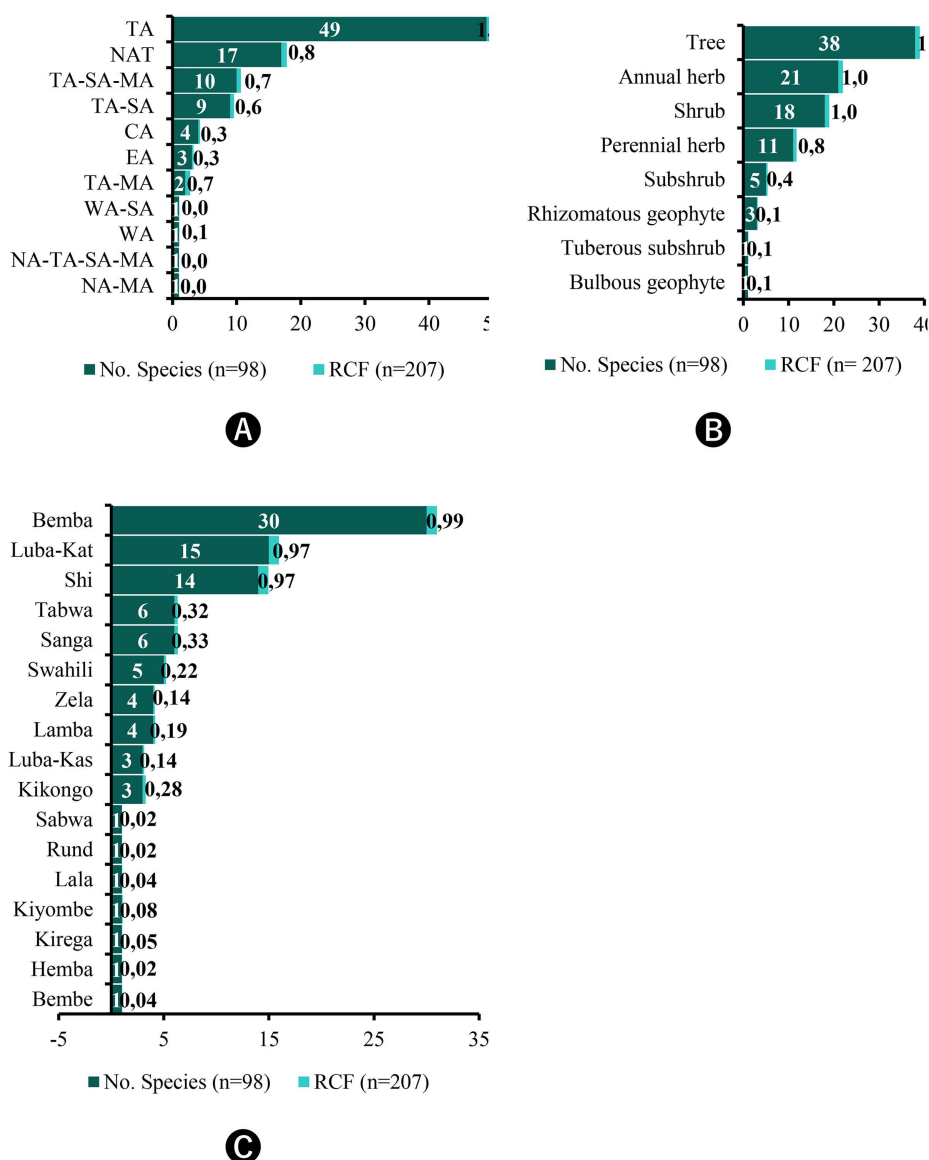


Fig 3. Morphological (a) and geographical (b) type of plants, category of informants (c), and Languages of Local Plant Names (C).

<https://doi.org/10.1371/journal.pone.0355373.g003>

diabetes, wounds, and malaria represent the three primary indications, both in terms of citation frequency (over 100 citations), and botanical diversity, with each condition involving more than 40 plant species from the recorded cohort (Table 3).

3.4. UV and RCF of plant surveyed

The RCF of the plant species documented in this study ranges from 0.01 for *Piliostigma thonningii* to 0.25 for *Kigelia africana*, the species with the highest frequency. This is followed by *Crassocephalum vitellinum* (RCF: 0.23), *Hylodesmum repandum* (RCF: 0.20), and *Solanum incanum* (RCF: 0.20). The use value of the recorded species varies from 0.05 for *Alchornea floribunda* to 0.32 for *Securidaca longepedunculata*, the species demonstrating the highest utilization. Next are *Asparagus africanus* (UV=0.27) and *Hibiscus sabdariffa* (UV=0.25).

Table 2. Traditional polyherbal formulations used as uterotonics.

N°	Specie 1	Specie 2	N° Recipe	UP (Ratio)	Preparation	Size (RCF) (n = 207)
1	<i>Abrus precatorius</i>	<i>Aloe vera</i>	R108	Sd-Lv (2 + 1)	Cataplasm	10(0.05)
2	<i>Annona senegalensis</i>	<i>Ageratum conyzoides</i>	R109	Sb-Wp (1 + 2)	Cataplasm	12(0.06)
3	<i>Aframomum alboviolaceum</i>	<i>Alchornea floribunda</i>	R110	Fr-Lv (2 + 3)	Decoction	15(0.07)
4	<i>Antidesma venosum</i>	<i>Azadirachta indica</i>	R111	Rb-Fr (1 + 1)	Ointment	18(0.09)
5	<i>Boerhavia diffusa</i>	<i>Cajanus cajan</i>	R112	Lv- Fr (3 + 1)	Cataplasm	22(0.11)
6	<i>Cissus schmitzii</i>	<i>Clausena anisata</i>	R113	R-Lv (2 + 1)	Infusion	27(0.13)
7	<i>Combretum molle</i>	<i>Commelina africana</i>	R114	Lv-Sb (2 + 1)	Maceration	30(0.15)
8	<i>Cucurbita pepo</i>	<i>Cynodon dactylon</i>	R115	Fr-Wp (2 + 1)	Decoction	18(0.09)
9	<i>Ekebergia capensis</i>	<i>Erythrina abyssinica</i>	R116	R-Sb (2 + 1)	Decoction	35(0.17)
10	<i>Ficus capensis</i>	<i>Flueggea virosa</i>	R117	Sb-Lv (2 + 1)	Cataplasm	31(0.15)
11	<i>Grona adscendens</i>	<i>Asparagus africanus</i>	R118	Lv-Lv (2 + 3)	Cataplasm	18(0.09)
12	<i>Hymenocardia acida</i>	<i>Juglans nigra</i>	R119	Rb-Fr (3 + 2)	Cataplasm	68(0.33)
13	<i>Khaya nyasica</i>	<i>Kigelia aethiopum</i>	R120	Sb-Sb (3 + 2)	Cataplasm	18(0.09)
14	<i>Malus domestica</i>	<i>Milletia griffoniana</i>	R121	Fr-Rb (3 + 2)	Cataplasm	22(0.11)
15	<i>Ocimum americanum</i>	<i>Piliostigma thonningii</i>	R122	Lv-Lv (3 + 2)	Cataplasm	27(0.13)

Legend – PU-Ratio: Part used and proportion of the mixture; Lv: leaf; R: root; Rb: Root bark; Sb: Stem bark; Fr: fruit; Sd: Seed; N° Recipe. = uterotonic recipe.

<https://doi.org/10.1371/journal.pone.0355373.t002>

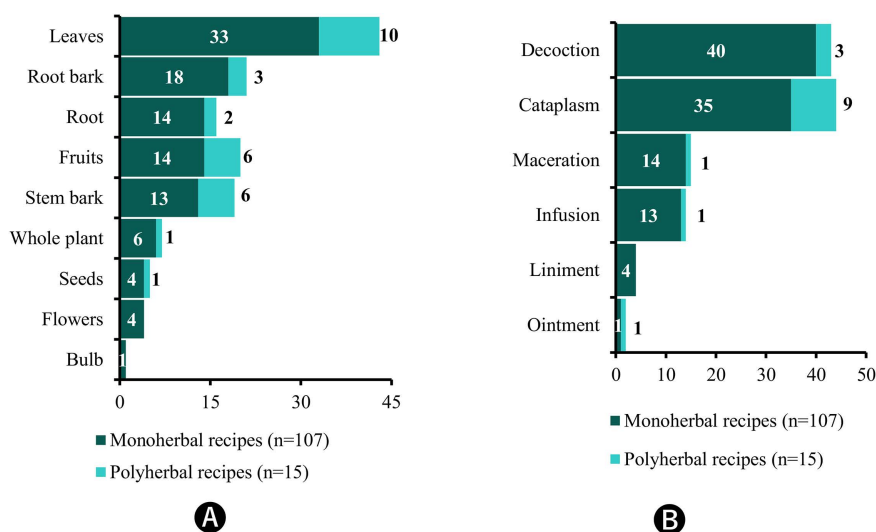


Fig 4. Parts used (A) and mode of preparation of traditional uterotonics formulations (B).

<https://doi.org/10.1371/journal.pone.0355373.g004>

3.5. Socio-demographic profile of informants

The informants surveyed in this study fall into three main categories: housewives (49.8%), herbalists (12.6%) and traditional healers (37.6%). The majority are women (64.7%), with an average age of 41 years, ranging from 17 to 73. They predominantly reside in the Katuba (21.7%) and Annexe (19.8%) municipalities, and most have a secondary education (40.0%). The informants are distributed across six distinct occupational groups, with traditional healers the most numerous

Table 3. Other ailments treated by the surveyed plants.

Ailments (n = 61)	Ns	Ailments (n = 61)	Ns	Ailments (n = 61)	Ns
Atherosclerosis	1	Dengue fever	3	Dysentery	13
Cataract	1	Constipation	4	Urinary tract infection	14
Cholera	1	Gastritis	4	Hypertension	15
Cold	1	Cardiovascular diseases	5	Drepanocytosis	16
Conjunctivitis	1	Fungal ailments	5	Headache	19
Contraception	1	Intoxications	5	Inflammatory diseases	19
Dental problems	1	Leprosy	5	Hepatic diseases	20
Eye problems	1	Typhoid fever	5	Ulcers	20
HIV/AIDS	1	Hemorrhoids	6	Diarrhea	21
Kidney disorders	1	Parasites diseases	6	Sexually transmitted infections	21
Meningitis	1	sexual dysfunction	7	Coughs	25
Thrombosis	1	Asthma	7	Cancers	27
aphrodisiac	2	Hemorrhoids	7	Respiratory diseases	28
Dysmenorrhea	2	Hyperlipidemia	7	Stomach diseases	28
Epilepsy	2	Pain	7	skin diseases	29
Myoma	2	Rheumatism	8	Fever	37
Neurodegenerative disorders	2	Snake bites	8	gastrointestinal diseases	41
Pruritis	2	Tuberculosis	8	Diabetes	44
bilharziasis	3	Viral diseases	8	Wounds	44
Chickenpox	3	Helminths diseases	9	Malaria	46

Legend: Ns: Numbers of species. The intensity of the color ranges from red (low) to green (high), with yellow (medium) in between, corresponding to the number of species identified in the management of the corresponding condition, according to our informants.

<https://doi.org/10.1371/journal.pone.0355373.t003>

(37.5%). Furthermore, a significant proportion (45.4%) have over 11 years' experience applying medicinal plants to facilitate childbirth (Table 4).

4. Discussion

4.1. Diversity of medicinal plants used as uterotonic in Lubumbashi

For millennia, the inhabitants of Lubumbashi have used the flora of the Miombo woodlands to treat various health issues, particularly to facilitate childbirth as uterotonic. In the current investigation, 98 plant varieties were documented according to their application by housewives, herbalists, and traditional healers to support childbirth.

A systematic review of the available literature (Table 1) enabled these taxa to be divided into two main categories, which can be subdivided into five subclasses. The first category comprises 34 species newly documented to have oxytocic uses, while the second category includes 64 species for which oxytocic applications have already been reported in previous studies (Table 1).

The 34 species in the first category were organized into three subclasses:

The first subclasses with nineteen species that are already recognized as medicinal plant in Democratic Republic of Congo, particularly around Lubumbashi, namely: *Aframomum alboviolaceum*, *Albizia adianthifolia*, *Antidesma venosum*, *Bobgunnia madagascariensis*, *Cissus schmitzii*, *Combretum molle*, *Crassocephalum montuosum*, *Cyanthillium cinereum*, *Ficus stuhlmannii*, *Flueggea virosa*, *Harungana madagascariensis*, *Hexalobus monopetalus*, *Hymenocardia acida*, *Juglans nigra*, *Khaya nyasica*, *Kigelia aethiopica*, *Parinari curatellifolia*, *Phyllanthus microdendron*, and *Vitex congolensis* var. *thomasii*;

Table 4. General characteristics of the informants.

Class	Category	Housewives (n = 103)	Herbalists (n = 26)	TH (n = 78)	All Informants (n = 207)
		n _i (%)	n _i (%)	n _i (%)	%
Age	15-25	02(06.3)	00(00.0)	01(07.1)	06.2
	26-35	14(32.9)	01(12.5)	03(21.4)	32.1
	36-45	81(48.6)	05(62.5)	02(14.3)	47.7
	46-55	04(09.5)	01(12.5)	02(14.3)	09.7
	>55	02(02.7)	01(12.5)	06(42.9)	04.3
Experience (Year)	1-5	65(05.4)	00(00.0)	01(07.1)	05.4
	6-10	25(33.7)	01(12.5)	06(42.9)	33.6
	11-15	05(46.2)	04(50.0)	03(21.4)	45.4
	16-20	05(12.0)	02(25.0)	02(14.3)	12.3
	21-25	03(02.7)	01(12.5)	02(14.3)	03.3
Profession	Trade	20(19.4)	00(00.0)	00(00.0)	09.7
	Public sector	21(20.4)	00(00.0)	00(00.0)	10.1
	Liberal	59(57.3)	00(00.0)	00(00.0)	28.5
	Housework	03(02.9)	00(00.0)	00(00.0)	01.5
	TH	00(00.0)	00(00.0)	78(100)	37.7
Gender	Herbalist	00(00.0)	26(100)	00(00.0)	12.5
	Female	77(74.8)	16(61.5)	41(52.6)	64.7
	Male	26(25.2)	10(38.5)	37(47.4)	35.3
Municipality of residence	Annexe	23(22.3)	02(07.7)	16(20.5)	19.8
	Kamalondo	13(12.6)	00(00.0)	02(02.5)	07.2
	Kampemba	15(14.6)	08(30.8)	12(15.4)	16.9
	Katuba	12(11.7)	10(38.5)	23(29.5)	21.7
	Kenya	16(15.5)	02(07.7)	12(02.5)	14.5
	Lubumbashi	18(17.5)	02(07.7)	03(03.8)	11.2
	Ruashi	06(05.8)	02(07.7)	10(12.8)	08.7
Level of studies	None	24(23.3)	02(07.7)	05(06.4)	15.0
	Primary	21(20.4)	08(30.8)	23(29.5)	25.1
	Professional	25(24.3)	03(11.5)	01(01.3)	14.1
	Secondary	26(25.2)	10(38.5)	47(60.2)	40.0
	University	07(06.8)	03(11.5)	02(02.6)	05.8

Legend – TH: Traditional healer. Ni: number of Informants. The percentages for each category are shown in bold, as is the overall percentage. Each time, refer to n to determine it. **Traditional healers and Herbalist:** Unlike herbalists, who are primarily involved in the commercial collection and distribution of plants, traditional practitioners draw on ancestral knowledge to treat illnesses. In addition to botanical resources, they use a wide range of natural therapeutic agents. They may also turn to herbalists to obtain specialized raw materials.

<https://doi.org/10.1371/journal.pone.0355373.t004>

The second subclass with Eight species documented as medicinal elsewhere in DRC, but not previously reported from Lubumbashi: *Adenia gummifera*, *Albizia lebbbeck*, *Alchornea floribunda*, *Baphiopsis parviflora*, *Boerhavia diffusa*, *Cassia sieberiana*, *Hylodesmum repandum*, and *Leonotis nepetifolia*.

The third subclass with seven species whose medicinal use has been established outside the DRC: *Afrothismia mho-roana*, *Aloe chabaudii*, *Anchusa ovata*, *Bombax ceiba*, *Cochlospermum planchonii*, *Grona adscendens*, and *Paramollugo nudicaulis*. Together, these 34 taxa are key to understanding the specific role of Lubumbashi's traditional medicine in

managing difficult deliveries. Their documentation paves the way for targeted pharmacological research to validate the reported oxytocic effects and enrich the local pharmacopoeia with robust scientific evidence.

Although none of the 34 species catalogued in Lubumbashi have, to date, been explicitly reported or validated for oxytocic activity in the accessible literature, several taxa within the same genera exhibit well-documented uterotonic uses. For example, within the genus *Aframomum*, *A. melegueta* K. Schum. (also known as Ataare) is traditionally chewed by the Akoko people of Nigeria to facilitate childbirth [123]. Four species in the genus *Albizia* have been the subject of ethnobotanical investigations: *Albizia zygia* (DC.) J.F. Macbr: Stem bark is used as a purgative in the Ivory Coast to facilitate childbirth [124]; *Albizia chinensis* Lam.: triterpenoid saponins isolated from the roots significantly modulate uterine contractions [125]; *Albizia ferruginea* (Guill. & Perr.) Benth.: a stem bark decoction is used in Nigeria to aid childbirth [126]; *Albizia gummifera* (J. F. Gmel.) C. A. Meyer: The glycoside 'albitocin', isolated from the bark, increases the frequency and amplitude of spontaneous uterine contractions in various animal models [127]. Additional genera have likewise yielded extracts with demonstrated oxytocic activity: aqueous stem-bark extract of *Adenia globosa* Engl. [128]; methanolic leaf extract of *Alchornea laxiflora* (Benth.) Pax: [73]; aqueous leaf extract of *Cassia fistula* L. [129]; aqueous leaf extract of *Cassia italica* Mill [130]; and aqueous stem bark extract of *Vitex doniana* Sweet [131]. These pharmacological evidence of oxytocic activity in plants of the same genus reported in our current investigations suggest that it is possible to identify plants among those in the inventory that also have proven oxytocic potential. This highlights the importance of conducting targeted pharmacological investigations on Lubumbashi inventory species to confirm and characterize their oxytocic potential.

Within the second category, two subclasses were delineated: The fourth subclass (n = 34) comprises taxa that have traditionally been utilized as medicinal plants, yet for which no pharmacological evidence has been documented thus far. The fifth subclass (n = 30) comprises taxa for which uterotonic activity has been documented in the scientific literature. The confirmed uterotonic effects observed in the fifth subclass serve as empirical validation of the ethnobotanical survey data and strongly suggest that a similar proportion of the unstudied taxa may also possess active uterotonic principles.

The fourth subclass comprises 38 species: *Aerva javanica*, *Ageratum conyzoides*, *Annona senegalensis*, *Asparagus africanus*, *Capsicum annum*, *Clausena anisata*, *Clematis scabiosifolia*, *Crassocephalum vitellinum*, *Cucurbita pepo*, *Cynodon dactylon*, *Dalbergia boehmii*, *Diplorhynchus condylocarpon*, *Elaeis guineensis*, *Eleusine indica*, *Erythrina abyssinica*, *Euphorbia hirta*, *Ficus capensis*, *Leucas martinicensis*, *Malus domestica*, *Ochna schweinfurthiana*, *Ocimum americanum*, *Phyllanthus amarus*, *Phyllanthus ovalifolius*, *Physalis peruviana*, *Piliostigma thonningii*, *Securidaca longepedunculata*, *Senegalia polyacantha*, *Solanum incanum*, *Stereospermum kunthianum*, *Strychnos spinosa*, *Syzygium cordatum*, *Tagetes minuta*, *Thespesia garckeana*, *Uapaca kirkiana*, *Zanthoxylum chalybeum*, *Zanthoxylum zanthoxyloides*, *Ziziphus abyssinica*, and *Ziziphus mucronata*.

Many of these species are already recorded in diverse African pharmacopoeias for their uterotonic properties, substantiating an ethnomedicinal consensus on their use to facilitate parturition. This interregional convergence implies the conservation of bioactive uterotonic constituents within these species, and justifies further studies, including *ex vivo* assays on isolated uterine muscle, *in vivo* evaluation in gravid rodent models, and the isolation and structural elucidation of the responsible metabolites. Notably, only five of these species (approximately 13%), those indicated above, have not previously been documented in the traditional lushoie pharmacopoeia. This relatively low proportion highlights the depth and coherence of local botanical knowledge while pinpointing novel candidate species for prioritized pharmacological screening.

4.2. Botanical families, morphology, and endemism of inventoried plants

Of the 98 taxa documented in our ethnobotanical surveys, three species predominate (39%), nearly half are endemic to tropical Africa (49%), and 16% belong to the Fabaceae family. The overrepresentation of Fabaceae trees endemic to tropical Africa corroborates earlier ethnobotanical investigations in the katangense region [20,33,63,86,132,133]. Fabaceae

trees exhibit a suite of physiological and ecological traits conferring resilience in hot intertropical environments. Symbiotic associations with rhizobia enable efficient biological nitrogen fixation, permitting establishment on leached, nutrient-poor tropical soils [134]. A deep, laterally branching taproot system enhances water uptake during prolonged dry seasons. Foliar adaptations, compound leaf architecture, and a thick cuticle, minimize transpirational water loss while preserving high photosynthetic capacity under intense insolation [135,136].

Moreover, pronounced phenological plasticity, characterized by leaf abscission in the dry season followed by rapid foliar flush at the onset of rain, synchronizes growth with favorable moisture regimes, and underpins the ubiquity of Fabaceae in intertropical woody communities. Moreover, extant literature validates the existence of a probable correlation between the characteristic phytochemical composition of Fabaceae – notably their richness in triterpenoid, and steroidal saponins, and estrogenic flavonoids (phytoestrogens) – and their documented uterotonic activity. Indeed, preclinical trials have demonstrated that saponin-rich plant fractions induce, in a dose-dependent manner, an increase in the frequency and amplitude of contractions of isolated uterine smooth muscle, thus replicating the traditional use of these plants to facilitate labor [137]. Furthermore, phylogenetic analyses reveal that the Fabaceae lineages most enriched in estrogenic flavonoids, dedicated to fertility and parturition drugs, coincide with the “warm environments” [138] establishing a chemical basis likely to underpin uterotonic efficacy across this family.

Among the documented plant species, spontaneous, uncultivated varieties account for 91 species (92.8%). These wild plants are predominantly used in traditional medicine, offering substantial advantages in terms of accessibility and affordability. This makes them invaluable for healthcare systems with limited resources, such as those of the Democratic Republic of Congo [139,140]. However, utilizing them presents significant challenges. The chemical composition of these plants varies greatly due to environmental influences, which can affect their therapeutic efficacy and safety. Furthermore, the lack of standardization in most cases increases the risk of toxicity and complicates quality assessment. Furthermore, the excessive harvesting of certain species threatens biodiversity, necessitating the implementation of conservation strategies [141,142]. Therefore, although these plants have significant medicinal potential, their integration into healthcare practices necessitates rigorous scientific research and robust regulatory mechanisms to guarantee their safety and efficacy within a structured framework.

4.3. Uterotonic recipes of medicinal plant survey

In the 122 uterotonic recipes inventoried during our surveys, most of which used a single plant, the leaf (35%) was the most used organ, and decoction (35%) the most prevalent method of preparation in mono-herbal formulation. However, among all the recipes, poultices are the most common method, which is justified by the fact that most recipes are for local application. While decoction is the most common method of preparation in several ethnobotanical studies carried out in the region [63,20, 132–133–,143–149] Various organs seem to be the most popular according to the studies. Both the leaves [33,63,86,132,144,150,151], sometimes roots [86,118,145,146,149] or stem bark [20,152,153].

The use of leaves in traditional medicine offers several significant advantages over other medicinal plant organs, both pharmacologically and ecologically. From a therapeutic point of view, leaves are often rich in the synthesis of most bioactive metabolites [43,154,155]. In addition, their anatomical structure facilitates the extraction of active ingredients, improving the efficacy of traditional preparations [43]. Ecologically, leaf harvesting is generally less destructive than root or stem bark harvesting, contributing to the preservation of plant species and the sustainability of medicinal practices [156]. Moreover, leaves are often available in greater quantities and over a longer period of the year, making them more accessible to local communities. However, their active ingredient content can vary according to season and environmental conditions, requiring rigorous standardization to guarantee their therapeutic efficacy.

During this study, fifteen polyherbal formulations (12%) each combine two plant species, an approach with ambivalent outcomes. On one hand, phytotherapy exploits the synergy of bioactive constituents and mutual masking of adverse effects to achieve a broader therapeutic spectrum with reduced toxicity. On the other hand, the complexity of these

mixtures can prove counterproductive: unforeseen interactions, excessive dilution of key active compounds, and botanical variability related to geographic origin or harvest season all impede the standardization and reproducibility of treatments [33,153]. Added to these challenges are the risks of cross-allergies among multiple species and the uncertain safety of compositions that lack robust clinical validation, which may reveal toxic or mutagenic properties. Nevertheless, the enduring use of most of these formulations for over twenty-five years attests to their acceptable safety profile and adequate efficacy: without a favorable benefit–risk balance, they would have been discontinued long ago.

4.4. Plants highlighted by this survey

In this study, we consider that the plants highlighted are notably the two most frequently cited plants: *Kigelia africana* (RCF: 0.25) & *Crassocephalum vitellinum* (RCF: 0.23); the two most widely used plants: *Securidaca longepedunculata* (UV: 0.27) & *Hibiscus sabdariffa* (UV=0.25) as well as the two plants most frequently cited among those reported for the first time as oxytocic plants thanks to our present study: *Hylodesmum repandum* (RCF: 0.20) & *Crassocephalum montuosum* (RCF: 0.19) (Fig 5).

Indeed, a high degree of citation convergence around a species reflects a high Informant Consensus Index. This homogeneity of knowledge suggests empirical pharmacological validation: the more a plant is cited for its oxytocic properties, the more likely it is to contain bioactive compounds that could account for these properties. Furthermore, the most frequently used plants define the actual Use Value (UV) within the community. In contexts where access to modern healthcare is limited, these species are the pillars of health in this environment. Studying them is a public health emergency, both to validate their safety and to prevent the risks of toxicity associated with non-standardized dosages. However, it should be noted that poorly documented plants are not insignificant; they reflect the endemic nature of knowledge



Fig 5. Plants highlighted in this study in order of appearance: *Kigelia africana*, *Crassocephalum vitellinum*, *Securidaca longepedunculata*, *Hibiscus sabdariffa*, *Hylodesmum repandum*, *Crassocephalum montuosum*.

<https://doi.org/10.1371/journal.pone.0355373.g005>

and the bio-cultural specificity of the environment, which can very easily lead to original research that may result in new knowledge.

If a case had to be made for a single plant per category, such as most cited, most used or new discovery, then it would be the bark of *Kigelia africana*, the leaves of *Securidaca longepedunculata* and the leaves of *Hylodesmum repandum*, which are the plants that stand out most in this study (Fig 5).

The stem bark of *Kigelia africana* (Lam.) Benth. is a key part of African traditional medicine, where it has been used to treat a wide range of illnesses for a long time. As we all know, traditional applications cover a range of dermatological disorders, like eczema and wound healing, as well as gastrointestinal disturbances, including dysentery [30,153]. They also cover critical gynaecological indications, like labour induction and control of postpartum haemorrhage [157,158]. Recent drug studies have shown that these traditional uses are legit and have reported strong antimicrobial, anti-inflammatory, and antioxidant effects, as well as the ability to kill melanoma and renal carcinoma cell lines [159,160]. These bioactivities arise from a range of phytochemicals, such as naphthoquinones (e.g., lapachol and kigelinone), iridoid glycosides (e.g., verminoside and specioside), and polyphenolic compounds (e.g., gallotannins and flavonoids) [161,162]. All this evidence shows that *K. africana* stem bark is important in terms of pharmacology, and that it should be further explored as a source of therapeutic agents.

Although the roots of *Securidaca longepedunculata* are most frequently employed in traditional medicine, the leaves are also used for a wide range of ethnomedical indications, including epilepsy, headache, abdominal pain, infertility, snakebite, toothache, cancer, dermal infections, dislocated jaw, contraceptive purposes, and placental expulsion [153,163]. Pharmacological investigations have provided evidence supporting several bioactivities of leaf extracts, notably antimicrobial activity [164,165], immunomodulatory and anti-inflammatory effects [166], antiparasitic activity [167], antioxidant properties [168], and anticonvulsant activity [169]. These activities have been correlated with the presence of diverse phytochemical classes, principally xanthones, saponins, and flavonoids [164,170], some of which have been reported to possess oxytocic potential [171–175].

The leaves of *Hylodesmum repandum* (syn. *Desmodium repandum*) are a cornerstone of traditional primary healthcare, widely utilized as hemostatic poultices for treating fresh wounds, ulcers, and skin infections, and in decoctions to manage fever, dysentery, and pain. These traditional applications are supported by ethnopharmacological studies demonstrating significant antimicrobial, antioxidant, and anti-inflammatory activities, particularly against pathogens such as *Staphylococcus aureus*, as well as antimalarial activity. This biological efficacy is attributed to a distinct phytochemical profile rich in flavonoids (specifically isoflavonoids), tannins, and alkaloids, groups also reported among those with uterotonic potential [172–174].

5. Limits of the study

The findings of this study should be interpreted in light of several limitations: (i) the absence of quantitative data on harvesting pressure and its potential impact on the population dynamics of highly sought-after taxa; (ii) the lack of clinical validation for interview-derived claims concerning therapeutic indications and mechanisms of action; (iii) the study's emphasis on ethnomedicinal knowledge related to uterotonic uses combined with a limited geographic scope, which may have resulted in incomplete documentation of other local uses of the recorded taxa; (iv) the omission during field surveys of critical usage details, such as administration conditions, contraindications, dosage regimens, and adverse effects, underscoring the need for targeted pharmacological, toxicological, and dosage-finding studies for each reported taxon; and (v) insufficient documentation of herbal formulations and ancillary traditional applications beyond obstetric use, which warrants systematic ethnobotanical, phytochemical, and pharmacological follow-up investigations.

6. Conclusion

The findings of this study support that in Lubumbashi, both traditional healers and women community leaders routinely employ medicinal plants as uterotonic agents in the management of childbirth complications. The taxa identified for

uterotonic use frequently possess additional ethnomedicinal applications, and several of the uses recorded locally correspond with reports from other African pharmacopoeias, indicating regional convergence in traditional obstetric practice.

These ethnobotanical findings provide a rationale for prioritized pharmacological and clinical investigation. Systematic phytochemical profiling, bioassay-guided fractionation, and *in vitro* and *in vivo* evaluation of uterotonic activity are required to validate efficacy and to identify the bioactive constituents responsible for the observed effects. Concurrent toxicological and finding studies, together with documentation of traditional formulations and administration regimens, are essential to assess safety and translational potential.

Finally, given the dual importance of these species for maternal health and local livelihoods, future research should integrate conservation assessments and sustainable use strategies alongside biomedical validation to ensure that any development of plant-derived uterotonics is both scientifically robust and culturally and ecologically responsible.

Supporting information

S1 Table. Survey questionnaire.

(DOCX)

S1 Fig. Graphical Abstract.

(DOCX)

Acknowledgments

The authors would like to thank the Kipopo Herbarium for identifying plant species. They also thank traditional healers, herbalists, and Lubumbashi households for acquiring ethnobotanical data.

Author contributions

Conceptualization: Bashige Valentin Chiribagula.

Data curation: Jean Claude Nkwanga, Muyumba Singoy Sarah, Mwamba Maseho Faustin, Biayi Benaja Martin, Sangwa Kamulete Gregoire, Mbayo Kalubandika Glaubert, Muhona Melman.

Formal analysis: Bashige Valentin Chiribagula.

Investigation: Bashige Valentin Chiribagula, Jean Claude Nkwanga, Muyumba Singoy Sarah, Mwamba Maseho Faustin, Biayi Benaja Martin, Sangwa Kamulete Gregoire, Mbayo Kalubandika Glaubert, Muhona Melman.

Project administration: Bashige Valentin Chiribagula.

Supervision: Okusa Ndjolo Philippe, Lumbu Simbi Jean Baptiste.

Validation: Bashige Valentin Chiribagula, Okusa Ndjolo Philippe, Lumbu Simbi Jean Baptiste.

Writing – original draft: Bashige Valentin Chiribagula.

Writing – review & editing: Cedrick Shakalenga Mutombo, Manya Mboni Henry, Numbi wa Ilunga Evodie, Bakari Amuri Salvius.

References

1. Onambele L, Guillen-Aguinaga S, Guillen-Aguinaga L, Ortega-Leon W, Montejó R, Alas-Brun R, et al. Trends, Projections, and Regional Disparities of Maternal Mortality in Africa (1990-2030): An ARIMA Forecasting Approach. *Epidemiologia (Basel)*. 2023;4(3):322–51. <https://doi.org/10.3390/epi-demiologia4030032> PMID: 37754279
2. Onambele L, Ortega-Leon W, Guillen-Aguinaga S, Forjaz MJ, Yoseph A, Guillen-Aguinaga L, et al. Maternal Mortality in Africa: Regional Trends (2000-2017). *Int J Environ Res Public Health*. 2022;19(20):13146. <https://doi.org/10.3390/ijerph192013146> PMID: 36293727

3. WHO, UNICEF, UNFPA, WBG, UNDESA. Trends in maternal mortality estimates 2000 to 2023. Geneva, Switzerland: World Health Organization. 2025.
4. Osadolor OO, Osadolor AJ, Osadolor OO, Enabulele E, Akaji EA, Odiowaya DE. Access to health services and health inequalities in remote and rural areas. *Janaki Med Coll J Med Sci*. 2022;10(2):70–4. <https://doi.org/10.3126/jmcjms.v10i2.47868>
5. Odjer EK, Mensah EKG. Maternal Mortality and Legal Accountability: Examining Government Responsibility for Preventable Deaths. *Journal of Law and Criminal Justice*. 2025. <https://doi.org/10.15640/jlcj.v13p4>
6. Aydar MZ, Ilina IB, Viktor AM. Uterotonic drugs in obstetric bleeding prevention and treatment. *J Obstet Women's Dis*. 2021;70(1):77–88.
7. Felis S, Quaroni S, De Simone A. Uterotonic Agents. *Am J Med Clin Res Rev*. 2023;2(12):1–14.
8. Haikera HK, Aku-Akai L, Guillaume Aboua Y. Scope of Medicinal Plants for Uterotonic, Tocolytic, and Wellness Effects in Pregnant Women: A Cultural Perspective. *Curative and Preventive Properties of Medicinal Plants*. Apple Academic Press. 2023. p. 341–52. <https://doi.org/10.1201/9781003328544-24>
9. Sumankuuro J, Baatiema L, Crockett J, Young J. Women's use of non-conventional herbal uterotonic in pregnancy and labour: evidence from birth attendants. *BMC Pregnancy Childbirth*. 2022;22(1):600. <https://doi.org/10.1186/s12884-022-04934-2> PMID: 35896986
10. Medjedovic E, Stanojevic M, Jonuzovic-Prosic S, Ribic E, Begic Z, Cerovac A, et al. Artificial intelligence as a new answer to old challenges in maternal-fetal medicine and obstetrics. *Technol Health Care*. 2024;32(3):1273–87. <https://doi.org/10.3233/THC-231482> PMID: 38073356
11. Gallos ID, Yunas I, Devall AJ, Podeseck M, Tobias A, Price MJ, et al. Uterotonic agents for preventing postpartum haemorrhage: a network meta-analysis. *Cochrane Database Syst Rev*. 2025;4(4):CD011689. <https://doi.org/10.1002/14651858.CD011689.pub4> PMID: 40237648
12. Sichalwe MM, Nangi NW, Daniel L, Egboke JO, Tavengana G. Prevalence and determinants of medicinal plants utilization during labour among women of reproductive age in Butiama, Tanzania: A community-based cross-sectional study. *PLoS One*. 2025;20(10):e0334453. <https://doi.org/10.1371/journal.pone.0334453> PMID: 41171764
13. Tripathi V, Stanton C, Anderson FWJ. Traditional preparations used as uterotonics in Sub-Saharan Africa and their pharmacologic effects. *Int J Gynaecol Obstet*. 2013;120(1):16–22. <https://doi.org/10.1016/j.ijgo.2012.06.020> PMID: 23021290
14. Roqaiya M, Begum W, Majeedi SF, Saiyed A. A review on herbs with uterotonic property. *J Phytopharm*. 2015;4(3):190–6.
15. Cobos DS, Luisa M, López N, Alejandrina E, Hernández G, Cruz BV. Uterotonic activity of *Heimia salicifolia* leaves. *J Med Plants Stud*. 2019;7(41653):98–102.
16. Chanda A, Simwinga F, Kaonga P, Gono-Bwalya A, Prashar L. The Uterotonic Screening of the Root Extract of *Azanza garckeana* (Malvaceae) on Isolated Wistar Rat Uterine Smooth Muscles. *Evid Based Complement Alternat Med*. 2020;2020:8873180. <https://doi.org/10.1155/2020/8873180> PMID: 33376504
17. Neuenschwander C, Wu K, Degolier T, Neuenschwander C. An investigation into the use of an herbal labor induction tincture containing black cohosh, cramp bark, partridgeberry, and motherwort on contractile responses produced from isolated strips of mouse uterine tissues. *Pharmacogn J*. 2021;13(6):1478–87.
18. Nama MC, Lunkuntu KH, Malonga KF, Kahumba BJ, Kakoma S. Contribution de la médecine traditionnelle à base des plantes utérotoniques à la santé de la reproduction en RDC. *Int J Soc Sci Sci Stud*. 2021;1(4):1–18.
19. Esther KI, Kleph KZ, Mariette KK, Sylvie YN, Andre WOT, Niclette L, et al. Plants Used by Pregnant Women at Kipushi City in Democratic Republic of Congo: Prevalence and Indications. *Open Access Libr J*. 2017;4(2):1–8.
20. Bashige Chiribagula Valentin, Kasongo Ilunga Pierre, Many Mboni Henry, Félicien Mushagalusa Kasali, Okusa Ndjolo Philippe, Lumbu Simbi Jean Baptiste. Ethnobotanical study of plants used by traditional healers in Lubumbashi (Democratic Republic of Congo) in the management of typhoid fever. *GSC Biol Pharm Sci*. 2022;21(1):265–86. <https://doi.org/10.30574/gscbps.2022.21.1.0403>
21. Valentin BC, Philippe ON, Melman M, Henry MM, Salvius BA, Baptiste LSJ. Ethnomedical knowledge of plants used in alternative medicine to treat hemorrhoidal diseases in Lubumbashi, Haut-Katanga province, Southern Democratic Republic of Congo. *BMC Complement Med Ther*. 2024;24(1):365. <https://doi.org/10.1186/s12906-024-04646-4> PMID: 39394139
22. Leonti M. The relevance of quantitative ethnobotanical indices for ethnopharmacology and ethnobotany. *J Ethnopharmacol*. 2022;288:115008. <https://doi.org/10.1016/j.jep.2022.115008> PMID: 35066067
23. Nwodo OF, Botting JH. Uterotonic activity of extracts of the seeds of *Abrus precatorius*. *Planta Med*. 1983;47(4):230–3. <https://doi.org/10.1055/s-2007-969994> PMID: 6867165
24. Oladimeji AV, Valan MF. The potential therapeutic advantage of *Abrus precatorius* Linn. an alternative to *Glycyrrhiza glabra*: A review. *J Pharm Res Int*. 2021;32(40):79–94.
25. Adia MM, Savina A, Jane N, Joel M, Godwin A, Esezah KK, et al. Medicinal plant species used for contraception and reproductive health care in rural Uganda. *Heliyon*. 2024;11(1):e41518. <https://doi.org/10.1016/j.heliyon.2024.e41518> PMID: 39844971
26. Kasali FM, Kadima JN, Peter EL, Mtewa AG, Ajayi CO, Tusiimire J, et al. Antidiabetic Medicinal Plants Used in Democratic Republic of Congo: A Critical Review of Ethnopharmacology and Bioactivity Data. *Front Pharmacol*. 2021;12:757090. <https://doi.org/10.3389/fphar.2021.757090> PMID: 34776975
27. Maman Noura O, Ilagouma AT, Compaore S, Mahamane Idi Issa A, Bogninou GSR, Ouédraogo R, et al. Ethnopharmacological investigation of plants used for the management of childbirth complications in Niger. *Scientific African*. 2024;24:e02216. <https://doi.org/10.1016/j.sciaf.2024.e02216>

28. Abate L, Tadesse MG, Bachheti A, Bachheti RK. Traditional and Phytochemical Bases of Herbs, Shrubs, Climbers, and Trees from Ethiopia for Their Anticancer Response. *Biomed Res Int*. 2022;2022:1589877. <https://doi.org/10.1155/2022/1589877> PMID: 35155671
29. Tafese Awulachew M. Hand Book of Common Ethiopian Traditional Medicinal Plants: Their Parts and Uses for Human and Animal Treatments. *JDMF*. 2021;7(3):48. <https://doi.org/10.11648/j.jdmp.20210703.11>
30. Valentin BC, Philippe ON, Henry MM, Salvius BA, Suzanne MK, Kasali FM, et al. Ethnomedical Knowledge of Plants Used in Nonconventional Medicine for Wound Healing in Lubumbashi, Haut-Katanga Province, DR Congo. *The Scientific World Journal*. 2024;2024(1). <https://doi.org/10.1155/2024/4049263>
31. Tlhabi DB, Ramaite IDI, Anokwuru CP, Ree TV, Hoppe HC. In Vitro Studies on Antioxidant and Anti-Parasitic Activities of Compounds Isolated from *Rauvolfia caffra* Sond. *Molecules*. 2020;25:3781.
32. Koman SR, Kpan WB, Yao K, Ouattara D. Medicinal uses of plants by traditional birth attendants to facilitate childbirth among djimini women in Dabakala (Center-north of Côte d'Ivoire). *Ethnobot Res Appl*. 2021;21:1–12.
33. Valentin BC, Philippe ON, Melman M, Henry MM, Salvius BA, Baptiste LSJ. Ethnomedical knowledge of plants used in alternative medicine to treat hemorrhoidal diseases in Lubumbashi, Haut-Katanga province, Southern Democratic Republic of Congo. *BMC Complement Med Ther*. 2024;24(1):365. <https://doi.org/10.1186/s12906-024-04646-4> PMID: 39394139
34. Mbuta KKKU, Mwima K, Kavuna M, Kalambayi M, Kazembe K, Booto K. Plantes médicinales de traditions Province de l'Equateur – R.D. Congo. Kinshasa, RDC: Institut de Recherche en Sciences de la Santé. 2012.
35. Agbo MO, Okoye FBC, Ebi GC, Osadebe PO. *Alchornea floribunda* (Müll. Arg.) - A review of its phytochemistry and biological activities. *Tropical Journal of Pharmaceutical Research*. 2020;19(5):1113–20.
36. Gutiérrez AL, Barrantes JB, Navarro MP, Monge MCO, Vargas MJC, Redondo GLM. General aspects about *Allium sativum* – a review. *Ars Pharm*. 2021;62(4):471–81.
37. Kamel N, El Boullani R, Cherrah Y. Use of Medicinal Plants during Pregnancy, Childbirth and Postpartum in Southern Morocco. *Healthcare (Basel)*. 2022;10(11):2327. <https://doi.org/10.3390/healthcare10112327> PMID: 36421651
38. Valentin BC, Philippe ON, Henry MM, Salvius BA, Suzanne MK, Kasali FM, et al. Ethnomedical Knowledge of Plants Used in Nonconventional Medicine for Wound Healing in Lubumbashi, Haut-Katanga Province, DR Congo. *ScientificWorldJournal*. 2024;2024:4049263. <https://doi.org/10.1155/2024/4049263> PMID: 39376217
39. Anywar G, Tugume P, Kakudidi EK. A review of Aloe species used in traditional medicine in East Africa. *South African Journal of Botany*. 2022;147:1027–41. <https://doi.org/10.1016/j.sajb.2021.07.036>
40. Gobvu V, Poshiwaa X, Benhura MA. An ethnoveterinary survey of medicinal plants used to treat poultry diseases in drylands of Zimbabwe. *Ethnobot Res Appl*. 2024;29(31):1–20.
41. Nikolajsen T, Nielsen F, Rasch V, Sørensen PH, Ismail F, Kristiansen U, et al. Uterine contraction induced by Tanzanian plants used to induce abortion. *J Ethnopharmacol*. 2011;137(1):921–5. <https://doi.org/10.1016/j.jep.2011.05.026> PMID: 21645605
42. Hajj MEI, Holst L. Herbal medicine use during pregnancy: A review of the literature with a special focus on sub-Saharan Africa. *Front Pharmacol*. 2020;11:866.
43. Bashige Chiribagula Valentin, Kalavanda Mvurabaar Burck, Okusa Ndjolo Philippe, Lumbu Simbi Jean Baptiste. Ethnomedicinal Knowledge of plants used in Alternative Medicine to treat Acnes in Lubumbashi (Democratic Republic of Congo). *GSC Biol Pharm Sci*. 2025;31(2):137–60. <https://doi.org/10.30574/gscbps.2025.31.2.0186>
44. Jaradat N, Qadi M, Abualhasan MN, Al-Iahham S, Al-Rimawi F, Hattab S, et al. Carbohydrates and lipids metabolic enzymes inhibitory, antioxidant, antimicrobial and cytotoxic potentials of *Anchusa ovata* Lehm. from Palestine. *European Journal of Integrative Medicine*. 2020;34:101066. <https://doi.org/10.1016/j.eujim.2020.101066>
45. El-Ishaq A, Alshawsh MA, Chik ZB. Evaluating the oestrogenic activities of aqueous root extract of *Asparagus africanus* Lam in female Sprague-Dawley rats and its phytochemical screening using Gas Chromatography-Mass Spectrometry (GC/MS). *PeerJ*. 2019;7:e7254. <https://doi.org/10.7717/peerj.7254> PMID: 31355056
46. Mfengwana PMAH, Mashele SS. Medicinal properties of selected *Asparagus* species: A review. *Phytochemicals in human health*. Great Portland Street, London. 2019. p. 1–23.
47. Bussmann RW, Paniagua-Zambrana NY. Ethnobotany in the Andes and the Amazon in a world of Nagoya Protocol and post SARS-CoV-2 pandemic. *Botany*. 2022;100(2):97–108. <https://doi.org/10.1139/cjb-2021-0062>
48. Evans Ngwenah F, Mahamat O, Christopher T. Hypoglycaemic, anti-hyperlipidemic, and anti-inflammatory activities of *Asparagus africanus* (Asparaceae) extract on high-fat diet/streptozotocin-induced diabetes in Wistar rats. *Asian Pacific Journal of Tropical Biomedicine*. 2024;14(12):532–9. https://doi.org/10.4103/apjtb.apjtb_472_24
49. Latham P, Mbuta AK ku. Plantes utiles du Bas-Congo, République Démocratique du Congo. *Inst Rech en Sci la Santé*. 2014;:409.
50. Konda KM, Bavukinana NLP, Mbembe B, Konda KR, Dumu L, Mafuta M, et al. Plantes médicinales de traditions de la RD. Congo - données préliminaires. *Inst Rech en Sci la Santé*. 2015;1:1–438.
51. Adeniyi IA, Olufunke O, Usman IM, Owu DU. Unraveling the reproductive potential of selected flavonoids in *Biden pilosa*: A comprehensive review. *Phytomedicine Plus*. 2025;5(2):100784. <https://doi.org/10.1016/j.phyplu.2025.100784>

52. Jan J, Khare RK, Mir TA. Medicinal plants used during pregnancy and childbirth in Baramulla district of Jammu and Kashmir, India. *Ethnobotanical Research and Applications*. 2021;22:70–80.
53. Chiribagula Valentin B, Ndjolo Philippe O, Mboni Henry M, Mushagalusa Kasali F. Ethnomedicinal Knowledge of Plants Used in Nonconventional Medicine in the Management of Diabetes Mellitus in Kinshasa (Democratic Republic of the Congo). *Evid Based Complement Alternat Med*. 2023;2023:4621883. <https://doi.org/10.1155/2023/4621883> PMID: [37771953](https://pubmed.ncbi.nlm.nih.gov/37771953/)
54. Das G, Shin H-S, Ningthoujam SS, Talukdar AD, Upadhyaya H, Tundis R, et al. Systematics, Phytochemistry, Biological Activities and Health Promoting Effects of the Plants from the Subfamily Bombacoideae (Family Malvaceae). *Plants (Basel)*. 2021;10(4):651. <https://doi.org/10.3390/plants10040651> PMID: [33805546](https://pubmed.ncbi.nlm.nih.gov/33805546/)
55. Katuura E, Kalabika E, Lubega A. Uterotonic Potential of Selected Plants Used by Ugandan Local Communities in the Treatment of Malaria. *EJMP*. 2018;24(2):1–12. <https://doi.org/10.9734/ejmp/2018/40934>
56. Konda K, Bavukinina N, Latham P, Mbembe B, Konda K, Dumu L. Plantes médicinales de la RD. Congo- données préliminaires. *Inst Rech en Sci La Santé*. 2015;1:438.
57. Olatunji-Bello I, Obijeh T, Mojiminiyi F. Tocolytic effect of *Cajanus cajan* (in-vitro studies using the rat uterus). *Nigerian Quarterly Journal of Hospital Medicine*. 2000;10(4):279–81.
58. Kamatenesi-Mugisha M, Oryem-Origa H. Medicinal plants used to induce labour during childbirth in western Uganda. *J Ethnopharmacol*. 2007;109(1):1–9. <https://doi.org/10.1016/j.jep.2006.06.011> PMID: [16901666](https://pubmed.ncbi.nlm.nih.gov/16901666/)
59. Ngalamulume HB, Divengi J-PN, Mbuyi PL, Assumani IZ, Mukeba FB, Mutwale PK, et al. Ethnobotanical, Phytochemical and Pharmacological Studies of *Capsicum annuum* L. (Solanaceae) in the Healing of Traumatic Wounds (Case of Eye Wounds): A Mini Review. *APRJ*. 2022;46–56. <https://doi.org/10.9734/aprj/2022/v10i1182>
60. Lampiao F, Maliwichi-Nyirenda C, Mponda J, Tembo L, Clements C. A preliminary investigation of the effects of labour inducing plant, *Cissampelos mucronata*, on the outcomes of pregnancy using rat models. *Malawi Med J*. 2018;30(3):159–61. <https://doi.org/10.4314/mmj.v30i3.5> PMID: [30627349](https://pubmed.ncbi.nlm.nih.gov/30627349/)
61. Tshibangu JN, Chifundera K, Kaminsky R, Wright AD, König GM. Screening of African medicinal plants for antimicrobial and enzyme inhibitory activity. *J Ethnopharmacol*. 2002;80(1):25–35. [https://doi.org/10.1016/s0378-8741\(01\)00409-3](https://doi.org/10.1016/s0378-8741(01)00409-3) PMID: [11891084](https://pubmed.ncbi.nlm.nih.gov/11891084/)
62. Maroyi A. A synthesis and review of medicinal uses, phytochemistry and pharmacological properties of *Cissampelos mucronata* A. Rich. (Menispermaceae). *J Pharm Nutr Sci*. 2020;10(5):213–22.
63. Numbi Wa Ilunga E, Mbayo Kitambala M, Muya K, Lachenaud O, Mukekwa Maloba J, Lumbu Simbi J-B, et al. Ethnobotanical survey and antimycobacterial activities of plants used against tuberculosis in Lubumbashi, DR Congo. *Trop Med Health*. 2025;53(1):64. <https://doi.org/10.1186/s41182-025-00745-1> PMID: [40319331](https://pubmed.ncbi.nlm.nih.gov/40319331/)
64. Omara T, Kiprop AK, Kosgei VJ, Kagoya S. *Clausena anisata* (Willd.) Hook.f. ex Benth. (Rutaceae): ethnomedicinal uses, phytochemistry, pharmacological activities, toxicity, and clinical application. *Tradit Med Res*. 2022;7(6):51.
65. Bashige CV, Bakari AS, Ndjolo PO, Kahumba BJ, Duez P, Lumbu SJ. Ethnobotanical study of plants used as antimalarial in traditional medicine in Bagira in Eastern RD Congo. *J Pharmacogn Phytochem*. 2020;9(4):1–14.
66. Dall'Acqua S, Kumar G, Sinan KI, Sut S, Ferrarese I, Mahomoodally MF, et al. An insight into *Cochlospermum planchonii* extracts obtained by traditional and green extraction methods: Relation between chemical compositions and biological properties by multivariate analysis. *Industrial Crops and Products*. 2020;147:112226. <https://doi.org/10.1016/j.indcrop.2020.112226>
67. Kabre Z, Yerbanga RS, Meda RN, Haro A, Fofana A, Kam SE. Traditional Uses of Seven Medicinal Plants for Malaria Treatment in Bobo-Dioulasso, Burkina Faso. *Adv Infect Dis*. 2025;15(1):79–95.
68. Attah AF, O'Brien M, Koehbach J, Sonibare MA, Moody JO, Smith TJ, et al. Uterine contractility of plants used to facilitate childbirth in Nigerian ethnomedicine. *J Ethnopharmacol*. 2012;143(1):377–82. <https://doi.org/10.1016/j.jep.2012.06.042> PMID: [22766472](https://pubmed.ncbi.nlm.nih.gov/22766472/)
69. Kibungu Kembelo P, Tuentner E, Vanhove W, Katula HB, Van Damme P, Pieters L. Phytochemical profiling by UPLC-ESI-QTOF-MS of *Commelina africana*, widely used in traditional medicine in DR Congo. *South African Journal of Botany*. 2023;157:325–34. <https://doi.org/10.1016/j.sajb.2023.04.010>
70. Valentin BC, Salvius BA, Jean Baptiste LS. Antibacterial and Antioxidant Activities, Toxicity, and Physicochemical Properties of *Crassocephalum montuosum* (S Moore) Milne-Redh and *Crassocephalum picridifolium* (DC) S Moore. *Adv Pharmacol Pharm Sci*. 2024;2024:9954073. <https://doi.org/10.1155/adpp/9954073> PMID: [39640498](https://pubmed.ncbi.nlm.nih.gov/39640498/)
71. Olarewaju OO, Fajinmi OO, Arthur GD, Coopposamy RM, Naidoo KK. *Bull Natl Res Cent*. 2021;45(1):208.
72. Ndayazi D, Kasali FM, Runigi B, Vianney J, Pierre YY, Kabangu T. Medicinal plants used in complementary medicinal for the treatment of burns in Rutshuru territory (North-Kivu, Democratic Republic of Congo). *J Pharm*. 2018;8(4):72–81.
73. Bafor EE, Omogiade GU. Ex-Vivo Uterine Activity of *Theobroma Cacao* (Malvaceae) Aqueous Seed Extract and *Cymbopogon Citratus* (Poaceae) Aqueous Leaf Extract - A Preliminary Investigation. *Nig J Bas App Sci*. 2020;27(2):108–18. <https://doi.org/10.4314/njbas.v27i2.15>
74. Mercy GT, Amon A, Patrick EO, Casim UT, Clement OA, Esther K. Medicinal plants used in gynecological procedures in Uganda. *J Med Plants Res*. 2020;14(4):185–94. <https://doi.org/10.5897/jmpr2019.6847>
75. Vwakyankazi M, Petit P. Bunganga ya mici: guérisseurs et plantes médicinales à Lubumbashi. *Lubumbashi: Observatoire du Changement Urbain (OCU) – Coopération universitaire au développement (CUD)*. 2004.

76. Maroyi A. The importance of ecological, biological and medicinal properties of *Diplorhynchus condylocarpon* (Müll. Arg.) Pichon (Apocynaceae) in tropical Africa. *Ecol Environ Conserv*. 2021;27:27–35.
77. Sewram V, Raynor MW, Mulholland DA, Raidoo DM. The uterotonic activity of compounds isolated from the supercritical fluid extract of *Ekebergia capensis*. *J Pharm Biomed Anal*. 2000;24(1):133–45. [https://doi.org/10.1016/S0731-7085\(00\)00404-0](https://doi.org/10.1016/S0731-7085(00)00404-0) PMID: 11108547
78. Sadgrove NJ, Mitaine-Offer AC, Khumalo G, Van Wyk BE. An overview of the phytochemistry of medicinal bark (trunk, stem or root) from the most popular southern African species. *Phytochem Rev*. 2025;5:1–21.
79. Soha SAS, Dougnon TJ, Ohouko FHO, Kpodekon MTT, Youssao AKI. Activités biologiques et utilisations de *Elaeis guineensis* (Jacq) et de *Khaya senegalensis* (Desr) en médecine traditionnelle humaine et vétérinaire. *Int J Biol Chem Sci*. 2019;13(1):525–42.
80. Rastogi S, Pandey MM, Rawat AKS. An ethnomedicinal, phytochemical and pharmacological profile of *Desmodium gangeticum* (L.) DC. and *Desmodium adscendens* (Sw.) DC. *J Ethnopharmacol*. 2011;136(2):283–96. <https://doi.org/10.1016/j.jep.2011.04.031> PMID: 21530632
81. Murumba DB, Jeruto P, Charles NN. Ethnobotanical survey of the cultural uses, threats and conservation status of flora in Kiptogot sub-location, Endebess subcounty, western Kenya. *J Med Plants Stud*. 2025;13(1):122–31. <https://doi.org/10.22271/plants.2025.v13.i1b.1797>
82. Kalu IK, Irene II, others. Action of contractile fraction of *Vernonia amygdalina*. Del ethanolic extract on hormonal profile of estrogen and progesterone in female albino Wistar rats. *Glob Res J Sci Nat*. 2015;1(2):1–5.
83. Ogundeko TO, Osagie IA, Bitrus J, Okoye NP, Ogbale EA, Edugbe AE, et al. The uterotonic screening of calyx extracts of *Hibiscus sabdariffa* on estrogenized isolated uterus of Wister strain albino rats. *Int J Front Biol Pharm Res*. 2022;3(1):037–45.
84. Cedrick Mutombo Shakalenga, Henry Many Mboni, Salvator Nsenga Nkulu, Gaël Nzuzi Mavungu, Cynthia Kibwe Mwenya, James Maloba Mwinensege, et al. Mineral elements analysis and total flavonoids content in the fresh leaves from two varieties of *Hibiscus sabdariffa* L. consumed as vegetable in Lubumbashi (DR Congo). *World J Adv Res Rev*. 2021;9(1):147–55. <https://doi.org/10.30574/wjarr.2021.9.1.0512>
85. Many Mboni H, Faes M, Fraselle S, Compaoré M, Salvius BA, Joseph KB, et al. Evaluating phytochemical constituents and in-vitro antiplasmodial and antioxidant activities of *Fadogiella stigmatoloba*, *Hygrophylla auriculata*, *Hylodesmum repandum*, and *Porphyrostemma chevalieri* extracts. *Heliyon*. 2023;9(9):e20103. <https://doi.org/10.1016/j.heliyon.2023.e20103> PMID: 37809863
86. François DO, Eddy NK, James MM, Marsi MK, Justin KK, Lambert MM. Ethnobotanical Studies of Reputed Aphrodisiac Plants Used in Traditional Medicine in Haut-Katanga in DR of Congo. *Int J Innov Sci Res Technol*. 2024;9(5):3317–34.
87. Bashige CV, Bakari AS, Martin BB, Lumbu SJB. Ethnomedical knowledge of plants used in traditional medicine in Mampa village, Haut-Katanga province, Democratic Republic of Congo. *Res Sq*. 2024;:1–50.
88. Pathy Kibungu K. Ethnobotanical study and characterization of medicinal plants used by populations of Kisantu and Mbanza-Ngungu territories, Kongo-Central Province (DR Congo). 2024.
89. Kamatenesi-Mugisha M, Makawiti DW, Oryem-Origa H, Olwa-Odyek, Nganga J. The oxytocic properties of *Luffa cylindrica* (L.) M. Roem. and *Bidens pilosa* L., traditionally used medicinal plants from western Uganda. *Afr J Ecol*. 2007;45(SUPPL. 3):88–93.
90. Wanda GJMK, Starcke S, Zierau O, Njamen D, Richter T, Vollmer G. Estrogenic activity of griffonianone C, an isoflavone from the root bark of *Milletia griffoniana*: regulation of the expression of estrogen responsive genes in uterus and liver of ovariectomized rats. *Planta Med*. 2007;73(6):512–8. <https://doi.org/10.1055/s-2007-967186> PMID: 17486534
91. Banzouzi J, Prost A, Rajemiarimirato M, Ongoka P. Traditional uses of the African *Milletia* species (Fabaceae). *Int J Bot*. 2008;4(4):406–20.
92. Hoosen F, Derek L, Tagatsing N, Maurice F, Emmanuel Y. The anti - cancer potential of active compounds extracted from *Milletia griffoniana* on pancreatic and colorectal cancer cells. *Adv Tradit Med*. 2024;24(3):789–800.
93. Yusuf A, Abdullahi M, A A M. Ethnobotany, phytochemistry and pharmacology of *Ochna schweinfurthiana*: a review. *Univers J Pharm Res*. 2020;4(6):75–9.
94. Bashige CV, Bakari AS, Okusa NP, Longanga OA, Duez P, Lumbu SJ-B. *Ochna schweinfurthiana* F. Hoffm (Ochnaceae). *Phytothérapie*. 2024;22:57–61.
95. Thulin A, Cjbg G, Thulin M, Bidault E, Boulet V, Heidari N. Phylogeny and systematics of *Paramollugo* (Molluginaceae). *Candollea*. 2025;79(2):179–228.
96. Okunrobo L, Nwagwuogbe S, Bafor E. Phytochemical evaluation and in vitro inhibitory effect of the methanol extract and partitioned chloroform fraction of the stem bark of *Pentaclethra macrophylla* Benth (Fabaceae) on non-pregnant rat uterus. *West African J Pharm*. 2012;23(1):19–26.
97. Ajala TO, Abraham A, Keck CM, Odeku OA, Elufioye TO, Olopade JO. Shea butter (*Vitellaria paradoxa*) and *Pentaclethra macrophylla* oil as lipids in the formulation of Nanostructured lipid carriers. *Scientific African*. 2021;13:e00965. <https://doi.org/10.1016/j.sciaf.2021.e00965>
98. Kipute DD, Katayi AL, Luambua NK, Kahindo J-M, Mampeta S, Lelo U, et al. Quantitative Ethnobotany of Multiple-Use Species and Management of the Yangambi Biosphere Reserve in the Democratic Republic of the Congo. *Ecol Evol*. 2025;15(3):e71110. <https://doi.org/10.1002/ece3.71110> PMID: 40065924
99. Mbayo M, Kalonda E, Muya R, Tshisand P, Kanangila A, Bakari S. Test d ' activité antimittotique et étude chimique préliminaire de quelques Euphorbiaceae du Katanga méridional (RDC). *J Adv Bot Zool*. 2016;2012(2):1–13.
100. Doughari JH, Sunday D. Antibacterial Activity of *Phyllanthus muellerianus*. *Pharm Biol*. 2008;20(9):400–5.

101. Tshibangu DST, Kavugho FS, Kabengele CN, Masunda AT, Bongo GN, Kasiama GN, et al. Phytochemical study and evaluation of the antidiabetic and antihyperglycemic activities of the fruit extracts of *Physalis peruviana* L. (Solanaceae). *Phytomedicine Plus*. 2025;5(1):100675. <https://doi.org/10.1016/j.phyplu.2024.100675>
102. Sheeja E, Joshi SB, Jain DC. Antifertility activity of stems of *Plumbago rosea* in female albino rats. *Pharm Biol*. 2008;46(12):920–7.
103. Chhodavadiya N, Vyas K, Patani P. A review on *Plumbago zeylanica* Linn.: As an antifertility agent. *J Cardiovasc Dis Res*. 2023;14(12):2023.
104. Many MH, Keymeulen F, Ngezahayo J, Bakari AS, Kalonda ME, Kahumba BJ, et al. Antimalarial herbal remedies of Bukavu and Uvira areas in DR Congo: An ethnobotanical survey. *J Ethnopharmacol*. 2020;249:112422. <https://doi.org/10.1016/j.jep.2019.112422> PMID: 31765762
105. State E, Ekiti A, State E. Uterotonic effect of methanol extract and fractions of *Psidium guajava* L. (Myrtaceae) stem bark on female white albino rats. *J Pharm Sci*. 2023;2(1):36–44.
106. Kaluwa Kaingu C, Oduma JA, Kanui T. Preliminary investigation of contractile activity of *Ricinus communis* and *Euclea divinorum* extracts on isolated rabbit uterine strips. *J Ethnopharmacol*. 2012;142(2):496–502. <https://doi.org/10.1016/j.jep.2012.05.026> PMID: 22652367
107. Moroole MA, Materechera SA, Mbeng WO, Aremu AO. Medicinal plants used for contraception in South Africa: A review. *J Ethnopharmacol*. 2019;235:19–27. <https://doi.org/10.1016/j.jep.2019.02.002> PMID: 30731180
108. Abba U, Kankara S, Nasir M, Lawal U. Ethnobotanical survey and pharmacognostic features of *Senegalia polyacantha* (Willd.) Seigler & Ebinger. *Niger J Pharm Sci*. 2023;23(2):115–27.
109. Govindaraj Y, Melanaphuru V, Gupta S, Agrahari V, Nema RK. Antifertility activity of the ethanolic extract of *Cassia occidentalis*, *Derris brevipes* variety *Brevipes* and *Justicia simplex*. *World J Chem*. 2009;4(2):118–22.
110. Eze O, Obioma N, Okwesili N, Ogugua V, Parker J. Uterine contractile effect of ethanol extracts of *Sida acuta* Burm F. leaves. *J Investig Biochem*. 2016;5(2):37.
111. Ngolsou F, Obono P, Ndongo MN, Minkande JZ. Analyse phytochimique des extraits aqueux de *Sida acuta* et *Triumfetta cordifolia*, deux plantes utilisées pour faciliter l'accouchement en médecine traditionnelle au Cameroun. *Heal Sci Dis*. 2022;23(2S):14–8.
112. Poli S, Gbekley E, Alognon A, Montant M, Hoinsoy Y, Gbati L, et al. Monographie d'une plante médicinale: *Sida acuta* Burm. *J la Rech Sci l'Université Lomé*. 2023;25(3):9–21.
113. Paniagua-Zambrana N, Bussmann R, Kikvidze Z. *Solanum dulcamara* L. *Solanum lycopersicum* L. *Solanum melongena* L. *Solanum nigrum* L. *Solanum tuberosum* L. Solanaceae. In: Bussmann R, Elachouri M, Kikvidze Z, editors. *Ethnobotany of the Mountain Regions of Eastern Europe: Carpathians*. Cham, Switzerland: Springer Cham. 2024. p. 31–50.
114. Ior LD, Pekuk BN, Otimenyin SO. Behavioural and anticonvulsant properties of methanol extract of the stem bark of *Stereospermum kunthianum* Cham in mice. *J Pharm Bioresour*. 2021;18(1):47–52.
115. Thothela SNS, Kola E, Dalu MTB, Ndhlovu PT. Indigenous Knowledge and Utilisation of *Strychnos spinosa* Lam. in Sub-Saharan Africa: A Systematic Review of Its Medicinal, Nutritional, and Cultural Significance. *Diversity*. 2025;17(4):228. <https://doi.org/10.3390/d17040228>
116. Maroyi A. *Syzygium Cordatum* Hochst. ex Krauss: An Overview of Its Ethnobotany, Phytochemistry and Pharmacological Properties. *Molecules*. 2018;23(5):1084. <https://doi.org/10.3390/molecules23051084> PMID: 29734660
117. Gbolade AA, Adedokun OA, Ibrahim TL. Assessment of contractile effect of *Trema orientalis* (Cannabaceae) root bark extracted with methanol on uterus of non-pregnant rat. *SWJ*. 2025;19(4):955–8. <https://doi.org/10.4314/swj.v19i4.7>
118. Mavungu GN, Mutombo CS, Numbi DM, Nsenga SN, Muyumba WN, Pongombo CS, et al. Smallholders' knowledge about healing goat gastrointestinal parasite infections with wild plants in southern DR Congo. *Front Pharmacol*. 2023;14:1124267. <https://doi.org/10.3389/fphar.2023.1124267> PMID: 36937835
119. Nwafor AV, Ohanme EO, Ofonakara U, Eze CN, Okoroafor FC, Iwe CB. Oxytocic and abortifacient activities of *Xylopia aethiopica* ethanol fruit extract on female wistar rats. *Int J Hum Heal Sci*. 2024;08(03):268–74.
120. Okagu IU, Ndefo JC, Aham EC, Udenigwe CC. *Zanthoxylum* Species: A Comprehensive Review of Traditional Uses, Phytochemistry, Pharmacological and Nutraceutical Applications. *Molecules*. 2021;26(13):4023. <https://doi.org/10.3390/molecules26134023> PMID: 34209371
121. Ghayur MN, Gilani AH. Inhibitory activity of ginger rhizome on airway and uterine smooth muscle preparations. *Eur Food Res Technol*. 2006;224(4):477–81. <https://doi.org/10.1007/s00217-006-0358-1>
122. Adamolekun MM, Akpor OA, Olorunfemi O, Akpor OB. Traditional medicine use during pregnancy and labor in African context: A scoping review. *J Integr Nurs*. 2023;5(1):66–72.
123. Oluwajuyigbe BM, Ige OE. Ethnobotanical plants used for fertility control and delivery enhancement among Akoko and Ose Aborigines in Ondo State, Nigeria. *ijs*. 2025;27(1):129–45. <https://doi.org/10.4314/ijs.v27i1.11>
124. Konan EK, Tie MS. Plantes utilisées dans le traitement des troubles gynéco-obstétriques par les peuples Abbey et Krobou d'Agboville côte d'Ivoire. *Phytothéra*. 2009;09:262–74.
125. Lipton A. An active glycoside from *Albizia* species and its action on isolated uterus and ileum. *J Pharm Pharmacol*. 1963;15:816–24. <https://doi.org/10.1111/j.2042-7158.1963.tb12886.x> PMID: 14134117
126. Antimicrobial and uterine smooth muscle activities of *Albizia ferruginea* extracts. *Boletín Latinoam y del Caribe Plantas Med y Aromáticas*. 2006;5(2):31–5.

127. He Y, Wang Q, Ye Y, Liu Z, Sun H. The ethnopharmacology, phytochemistry, pharmacology and toxicology of genus *Albizia*: A review. *J Ethnopharmacol.* 2020;257:112677. <https://doi.org/10.1016/j.jep.2020.112677> PMID: [32278761](#)
128. Sinei K, Mwangi J. An in vitro study on the oxytocic action of *Adenia globosa* Engl. *East Cent African J Pharm Sci.* 2015;18:96–9.
129. Yadav R, Jabalpur M, Jain GC. Effect of aqueous extract of seeds of *Cassia fistula* on the uterine biochemical milieu of female albino rats. *Pharmacologyonline.* 2017;1:859–67.
130. No A, Bharathi V, Raja M, Students UG. Review article on *Senna italica* Mill. *Int J Pharm Sci Rev Res.* 2018;52(08):44–6.
131. Ladeji O, Udoh FV, Okoye ZSC. Activity of aqueous extract of the bark of *Vitex doniana* on uterine muscle response to drugs. *Phytother Res.* 2005;19(9):804–6. <https://doi.org/10.1002/ptr.1588> PMID: [16220576](#)
132. Kalonji M, Muya RK, Mutombo EK, Cimanga BCC, Numbi EWI, Kahumba JB. Aperçu ethnobotanique de plantes réputées antipaludéennes utilisées dans la ville de Lubumbashi et ses environs, dans le Haut-Katanga en RD Congo. *Ethnopharmacologia.* 2019;61(6):75–84.
133. Bashige Chiribagula Valentin, Alombong Alonie Gracia, Kamwimba Mahungala Arlette, Bakari Amuri Salvius, Okusa Ndjolo philippe. Ethnobotanical study of medicinal plants used in the treatment of sexual dysfunctions in traditional medicine in Kampemba-Lubumbashi, DR Congo. *World J Adv Res Rev.* 2020;7(3):016–32. <https://doi.org/10.30574/wjarr.2020.7.3.0328>
134. Sprent JI, Parsons R. Nitrogen fixation in legume and non-legume trees. *F Crop Res.* 2000;65:183–96.
135. Leal AS, Fidelis A, de Araujo MA, Cozin BB, Martins AR. Belowground systems in tropical savanna: Fabaceae morphoanatomical traits and their relation to fire. *Plant Biol (Stuttg).* 2024;26(7):1118–30. <https://doi.org/10.1111/plb.13730> PMID: [39412517](#)
136. Siddiqui S. Global patterns and drivers of species and genera richness of Fabaceae. *Front Plant Sci.* 2025;16:1581814. <https://doi.org/10.3389/fpls.2025.1581814> PMID: [40761565](#)
137. Fromm O, Degolier T. The contractile capabilities of various herbal constituents on uterine smooth muscle and their shared constituent presence involved with anti-inflammatory/ antioxidant mechanisms. *J Pharmacogn Phytochem.* 2021;10(4):28–37.
138. Thaweepanyaporn K, Thompson JB, Vasudevan N, Hawkins JA. Phylogeny, ethnomedicinal use and the distribution of phytoestrogens in the Fabaceae. *Front Pharmacol.* 2025;16:1583365. <https://doi.org/10.3389/fphar.2025.1583365> PMID: [40492133](#)
139. Alqethami A. Ethnobotanical Assessment of the Diversity of Wild Edible Plants and Potential Contribution to Enhance Sustainable Food Security in Makkah, the Kingdom of Saudi Arabia. *Diversity.* 2025;17(11):785. <https://doi.org/10.3390/d17110785>
140. Mokgehele SN, Du Plooy CP, Araya HT, Amoo SO, Mokgehele SN, Pofu KM. Medicinal plant cultivation for sustainable use and commercialisation of high-value crops. *S Afr J Sci.* 2022;118(7):1–7.
141. Kumari R, Deepali, Bhatnagar S. Biodiversity loss: threats and conservation strategies. *Int J Pharm Sci Rev Res.* 2021;68(37):242–54.
142. Mykhailenko O, Jalil B, McGaw LJ, Echeverría J, Takubessi M, Heinrich M. Climate change and the sustainable use of medicinal plants: a call for “new” research strategies. *Front Pharmacol.* 2025;15:1496792. <https://doi.org/10.3389/fphar.2024.1496792> PMID: [39963365](#)
143. Maloba MJ, Muyumba NW, Kasanya KJ, Mutombo SC, Mbayo KM, Ngoy KE. Ethnobotanical survey and preliminary chemical screening of 14 plants reputed antigestitis in Lubumbashi and surroundings (DR Congo). *Int Eur Ext Enablement Sci Eng Manag.* 2020;8(12):62–9.
144. Kalonda E, Mbayo M, Muhume S, Kasereka M, Mulamba J, Many H, et al. Ethnopharmacological survey of plants used against malaria in Lubumbashi city (D.R. Congo). *J Adv Bot Zool.* 2014;1(2):1–8.
145. Muya K, Tshoto K, Cioci CC, Aseho MM, Kalonji M, Byanga K. Survol ethnobotanique de quelques plantes utilisées contre la schistosomiase urogénitale à Lubumbashi et environs. *Phytothérapie.* 2014. <https://doi.org/10.1007/s10298-014-0877-z>
146. Mbayo KM, Kalonda ME, Tshisand TP, Kisimba KE, Mulamba M, Richard MK. Contribution to ethnobotanical knowledge of some Euphorbiaceae used in traditional medicine in Lubumbashi and its surroundings (DRC). *J Adv Bot Zool.* 2016;4(2):1–16.
147. Bashige CV, Bakari, Amuri S, Mbuyi KS, Kahumba BJ, Duez P, et al. Étude ethnobotanique, phytochimique et évaluation de l'activité antipaludéenne de 13 plantes réputées antipaludéennes dans la commune du Kenya (Lubumbashi, RDC). *Phytothérapie.* 2020;18(2):71–80.
148. Bashige-Chiribagula V, Many-Mboni H, Ntabaza-Ndage V, Numbi Ilunga E, Bakari-Amuri S, Kalonda Mutombo E. Étude ethnobotanique, biologique et chimique de plantes réputées anticariogènes à Lubumbashi – RD Congo. *Phytothérapie.* 2017;15(1):2–9.
149. Amuri B, Maseho M, Simbi L, Duez P, Byanga K. Ethnobotanical survey of herbs used in the management of diabetes mellitus in Southern Katanga Area/DR Congo. *Pan Afr Med J.* 2018;30:218. <https://doi.org/10.11604/pamj.2018.30.218.11718> PMID: [30574237](#)
150. Okombe Embeya V, Lumbu Simbi J-B, Stévigny C, Vandenput S, Pongombo Shongo C, Duez P. Traditional plant-based remedies to control gastrointestinal disorders in livestock in the regions of Kamina and Kaniama (Katanga province, Democratic Republic of Congo). *J Ethnopharmacol.* 2014;153(3):686–93. <https://doi.org/10.1016/j.jep.2014.03.027> PMID: [24657601](#)
151. Bashige-Chiribagula V, Bakari-Amuri S, Mbuyi-Kalonji S, Kahumba-Byanga J, Duez P, Lumbu-Simbi JB. Étude ethnobotanique, phytochimique et évaluation de l'activité antipaludéenne de 13 plantes réputées antipaludéennes dans la commune du Kenya (Lubumbashi, RDC). *Phytothérapie.* 2017. <https://doi.org/10.1007/s10298-017-1152-x>
152. Bashige-Chiribagula V, Many-Mboni H, Ntabaza-Ndage V, Numbi Ilunga E, Bakari-Amuri S, Kalonda Mutombo E, et al. Étude ethnobotanique, biologique et chimique de plantes réputées anticariogènes à Lubumbashi – RD Congo. *Phytothérapie.* 2016;15(1):2–9. <https://doi.org/10.1007/s10298-015-1004-5>
153. Valentin BC, Martin BB, Salvius BA, Baptiste LSJ. Ethnomedicinal Knowledge of Plants Used in Traditional Medicine in Mampa Village, Haut-Katanga Province, Democratic Republic of the Congo. *Sci World J.* 2025;2025(1):2635735.

154. Twaij BM, Hasan MdN. Bioactive Secondary Metabolites from Plant Sources: Types, Synthesis, and Their Therapeutic Uses. *IJPB*. 2022;13(1):4–14. <https://doi.org/10.3390/ijpb13010003>
155. June M, Dar RA, Shahnawaz M, Ahanger MA, Majid I. Exploring the diverse bioactive compounds from medicinal plants: A review. *J Phytopharm*. 2023;12(3):189–95.
156. Devi K, Chaudhary V, Kalia S, Mishra R. Ustainable harvesting of non timber forest produce (NTFPS). *Indian Counc For Res Educ*. 2020.
157. Nabatanzi A, M Nkadameng S, Lall N, Kabasa JD, J McGaw L. Ethnobotany, Phytochemistry and Pharmacological Activity of *Kigelia africana* (Lam.) Benth. (Bignoniaceae). *Plants (Basel)*. 2020;9(6):753. <https://doi.org/10.3390/plants9060753> PMID: 32549404
158. Dossou-Yovo HO, Vodouhè FG, Kindomihou V, Sinsin B. Investigating the Use Profile of *Kigelia africana* (Lam.) Benth. through Market Survey in Benin. *Conservation*. 2022;2(2):275–85. <https://doi.org/10.3390/conservation2020019>
159. Mihaylova R, Bebrivenski N, Zheleva-Dimitrova D, Simeonova R, Singh N, Konstantinov S, et al. Integrative Evaluation of *Kigelia africana* Fruit Extract: Broad-Spectrum Anticancer Activity, Synergism with Cisplatin and Mechanistic Insights in Colorectal Carcinoma. *Molecules*. 2025;31(1):107. <https://doi.org/10.3390/molecules31010107> PMID: 41515404
160. Maiyo ZC, Njeru SN, Toroitich FJ, Indieka SA, Obonyo MA. Ethnobotanical study of medicinal plants used by the people of Mosop, Nandi County in Kenya. *Front Pharmacol*. 2024;14:1328903. <https://doi.org/10.3389/fphar.2023.1328903> PMID: 38313073
161. Murthy SSN, Jainab SIB, Bhuvaneswari G, Subhashini S, Radhakrishnan S, Saravanan S. Phytochemical screening, antioxidant activity and spectral characterization of *Kigelia africana*. *Biochem Cell Arch*. 2025;25(2):1559–70.
162. Priya B, Menkudale A, Gahlot M, Joshi P, Agarwal M. Pharmacognostical study, phytochemical analysis and phenolic content of *Kigelia africana* leaves. *Int J Pharm Pharm Sci*. 2013;5:3–6.
163. Us A, Uh D, Ibrahim H, Bb M. A review on African violet tree (*Securidaca longipedunculata*): A traditional drug with multiple medicinal uses. *Spec J Chem*. 2019;4(3):7–14.
164. Sani A, Ashezi NA, Abubakar US, Garba S. Phytochemical screening and antimicrobial activity of methanolic leaf extract of *Securidaca longipedunculata* Fresen (Polygalaceae). *J Clin Basic Res*. 2023;3(2):921–32.
165. G. HL, J. UK, J. YA, B. RA, U. UA. Comparative Antimicrobial Activity of *Securidaca longipedunculata* (Fresen) Extracts on Selected Human Pathogens. *Saudi J Med Pharm Sci*. 2024;10(01):26–31. <https://doi.org/10.36348/sjmps.2024.v10i01.005>
166. Dermane A, Kporvie AKG, Kindji KP, Metowogo K, Eklü-Gadegbeku K. Immunomodulatory and anti-inflammatory activities of hydro-ethanolic extract of *Securidaca longipedunculata* Fresen leaves. *J Herbmed Pharmacol*. 2024;13(2):280–8. <https://doi.org/10.34172/jhp.2024.49352>
167. Ochora DO, Kakudidi E, Namukobe J, Heydenreich M, Coghi P, Yang LJ, et al. A new benzophenone, and the antiplasmodial activities of the constituents of *Securidaca longipedunculata* fresen (Polygalaceae). *Nat Prod Res*. 2022;36(11):2758–66. <https://doi.org/10.1080/14786419.2021.1925272> PMID: 34000936
168. OBASI TC. FREE RADICAL SCAVENGING ACTIVITY AND TOTAL POLYPHENOL CONTENT OF SECURIDACA LONGIPEDUNCULATA ROOTS AND LEAVES EXTRACTS. *FARMACIA*. 2020;68(1):116–20. <https://doi.org/10.31925/farmacia.2020.1.16>
169. Abdulkadir A, Marakisiyya M, Garba M, Hamza U, Hadiza R, Abdulrazak A. Phytochemical screening and anticonvulsant activities of leaf methanol extract of *Securidaca longipedunculata* (Fresen). *Lapai J Appl Nat Sci*. 2024;9(1):77–83.
170. Shemishere B, Anyebe DA, Bashir YA, John E, Ifie J, Yahaya TO. Phytochemical screening and free radical scavenging activities of methanol leaf and flower extract of *Securidaca longipedunculata*. *FUDMA J Sci*. 2020;4(1):37–42.
171. Gruber CW, O'Brien M. Uterotonic plants and their bioactive constituents. *Planta Med*. 2011;77(3):207–20. <https://doi.org/10.1055/s-0030-1250317> PMID: 20845261
172. Wu C-H, Shieh T-M, Wang K-L, Huang T-C, Hsia S-M. Quercetin, a main flavonoid in onion, inhibits the PGF2 α -induced uterine contraction in vitro and in vivo. *Journal of Functional Foods*. 2015;19:495–504. <https://doi.org/10.1016/j.jff.2015.09.028>
173. Carreiro J da N, Souza ILL de, Pereira JC, Vasconcelos LHC, Travassos R de A, Santos BV de O, et al. Tocolytic action and underlying mechanism of galethin 3,6-dimethyl ether on rat uterus. *BMC Complement Altern Med*. 2017;17(1):514. <https://doi.org/10.1186/s12906-017-2007-6> PMID: 29197370
174. Bafor EE, Kupittayanant S. Medicinal plants and their agents that affect uterine contractility. *Curr Opin Psychol*. 2020;13:20–6.
175. Bello IO, Ayinde BA, Ofeimun JO, Amaechina FC, Kamal N, Gruber CW, et al. Investigation of the uterine modulating activities and mechanisms of *Brachystegia eurycoma* Harms (Leguminosae) methanol stem bark extract and fractions. *Scientific African*. 2024;25:e02298. <https://doi.org/10.1016/j.sciaf.2024.e02298>