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Nano-emulsions based on *Ayapana triplinervis* for the control of *Aedes aegypti*

Alex Bruno Lobato Rodrigues^{1,¶,*}, Rosany Martins Lopes^{2,¶}, Érica Menezes Rabelo^{2,¶}, Rosana Tomazi^{2,¶}, Lizandra Lima Santos^{2,¶}, Lethícia Barreto Brandão^{2,¶}, Cleidjane Gomes Faustino^{2,¶}, Ana Luzia Ferreira Farias^{2,¶}, Cleydson Breno Rodrigues dos Santos^{2,¶}, Patrick de Castro Cantuária^{3,¶}, Allan Kardec Ribeiro Galardo^{4,&}, Sheylla Susan Moreira da Silva de Almeida^{1,2, &}

¹Department of Exact and Technological Sciences, Postgraduate Program in Biodiversity and Biotechnology -Network Bionorte, Federal University of Amapá, Macapá, Amapá, Brazil. Rod. Juscelino Kubitschek, km 02 - Jardim Marco Zero, Macapá - AP, 68903-419.

²Department of Biological and Health Sciences, Federal University of Amapá, Macapá, Amapá, Brazil. Rod. Juscelino Kubitschek, km 02 - Jardim Marco Zero, 68903-419.

³ Amapaense Herbarium, Institute of Scientific and Technological Research of the State of Amapá, Macapá, Amapá, Brazil. Rod. Juscelino Kubitschek, km 10 - 68903-329.

⁴ Laboratory of Medical Entomology, Institute of Scientific and Technological Research of the State of Amapá, Macapá, Amapá, Brazil. Rod. Juscelino Kubitschek, km 10 - 68903-329.

[¶] These authors contributed equally to this work.

22 & These authors contributed equally to this work.

23 *Corresponding author: alexrodrigues.quim@gmail.com, sheyllasusan@yahoo.com.br

25 Abstract

26 *Ayapana triplinervis* is a plant species used in traditional medicine and in
 27 mystical-religious rituals by traditional communities in the Amazon. The aim of this
 28 study is to evaluate the larvicidal activity against *A. aegypti* of nano-emulsions
 29 containing essential oils from *A. triplinervis* morphotypes, and acute oral toxicity in
 30 non-target organism. Essential oils were identified and nano-emulsions were prepared
 31 using the low energy method. The mortality test of *A. aegypti* larvae was performed
 32 according to the protocol recommended by the World Health Organization, and toxicity
 33 in non-target mammals was performed according to the OECD. Phytochemical analyses
 34 indicated the major compounds (E)-Caryophyllene (45.93%) and Thymohydroquinone
 35 Dimethyl Ether (32.93%) in morphotype A, and in morphotype B, Thymohydroquinone
 36 Dimethyl Ether (84.53%) was found. Morphotype A essential oil nano-emulsion
 37 showed a particle size of 101.400 ± 0.971 nm (PdI = 0.124 ± 0.009 and ZP = $-19,300 \pm$
 38 0.787 mV). Morphotype B essential oil nano-emulsion had a particle size of $104.567 \pm$
 39 0.416 nm (PdI = 0.168 ± 0.016 and ZP = $-27,700 \pm 1,307$ mV). Histomorphological
 40 analyses showed the presence of inflammatory cells in the liver of animals treated with
 41 morphotype A essential oil nano-emulsion (MAEON) and morphotype B essential oil
 42 nano-emulsion (MBEON). Congestion and the presence of transudate with leukocyte
 43 infiltration in the lung of animals treated with MAEON were observed. The nano-
 44 emulsions containing essential oils of *A. triplinervis* morphotypes showed an effective
 45 nanobiotechnological product in the chemical control of *A. aegypti* larvae and safe for
 46 non-target mammals.

47 **Keyword:** Nanoinsecticides; Chemical control; Toxicity.

49 Introduction

50 *Aedes aegypti* is an anthropophilic insect vector of contagious infections such as
51 Dengue (DENV), chikungunya (CHIKV), Zika (ZIKV) and Yellow Fever in tropical
52 and subtropical regions of the planet [1]. The spread and survival of the species in
53 human clusters is the result of the destruction of its natural environment, with selected
54 in the wild population the most latent anthropophilic characteristics of the genus [2].

55 In 2015, more than 2 million cases of DENV were recorded in the Americas,
56 including 1.65 million cases reported in Brazil. Of this total, 811 deaths were recorded
57 and the incidence rate was equivalent to 813 cases per 100,000 inhabitants. As for
58 CHIKV, the first autochthonous infection in Brazil occurred in 2014 in the north and
59 northeast regions. In 2015, 38,499 were probable and in 2016 the infection rate
60 increased by about 706.05%, totaling 271,824 cases of CHIKV. In 2016, 215,319
61 probable cases of ZIKV fever were reported in the country, with an incidence rate of
62 105.3 cases per 100,000 inhabitants [3].

63 Brazil invested R\$ 2.3 billion (Reais) from its annual budget in 2016 for the
64 control of *A. aegypti* and its infections; of which 65% were applied in vector combat,
65 19% invested in indirect costs, and 16% applied in direct medical expenses [3].

66 Another point that requires attention is the double and triple co-infections
67 between DENV, ZIKV and CHIKV which can be transmitted simultaneously by *A.*
68 *aegypti* [4]. Therefore, the chemical control applied in the larval phase of *A. aegypti*
69 through the use of plant insecticides is an important tool in stopping mosquito-borne
70 infections and their impacts on public health [5].

Although they are sessile organisms, plants have different forms of defense against phytopathogens and herbivores, including a complex chemical mechanism of secondary metabolites that can be used for chemical control of *A. aegypti*, as the essential oils of the *Ayapana triplinervis* [6].

Originally from South America, *A. triplinervis* (synonym: *Eupatorium triplinervis*) can be found in Brazil, Ecuador, Peru, Puerto Rico and Guyana, besides being adapted in other countries like India and Vietnam [7]. The species is found in Brazil in two morphotypes: Japana-Branca (morphotype A) and Japana-roxa (morphotype B) [8].

A. triplinervis alkaloids and tannins exhibit greater food deterrence in *P. xylostella* and *C. binotalis* agricultural pests, followed by their phenols and flavonoids. *A. triplinervis* extracts disrupted the growth and development of *M. persicae* nymphs and showed significant pest control properties, and plant species may be indicated as potential candidates for further study of their potential as botanical pesticides - an alternative to synthetic insecticides [9].

Despite the significant potential for chemical insect control, secondary metabolites from *A. triplinervis*, especially essential oil (EO), are susceptible to volatility and oxidation by oxygen and light, affecting the biological activity of their chemical constituents, or their limited condition of solubility of the essential oil in the aqueous medium, where *A. aegypti* larvae develop [10].

One way to enable protection against degradation and oxidation, and even improve the solubility in aqueous medium of OE is through the development of a technological nanoemulsification system, which offers advantages such as ease of

handling, stability, protection against oxidation, better distribution, solubility and controlled release [11].

Nano-emulsions are colloidal systems where their particles are submicron size which are carriers of active molecules. Nanocapsulation technology of sparingly soluble bioactives in aqueous medium is useful in the pharmaceutical and food industries to be applied as bioactivity protection and controlled release to improve bioavailability of active compounds [12].

When the particle size distribution of an emulsion is below 80 nm, nano-emulsion has advanced properties compared to conventional sized emulsions as transparent visual appearance, high colloidal stability and a large interfacial area relative to volume [13]. Therefore, the aim of this study is to evaluate the larvicidal activity against *A. aegypti* of nano-emulsions containing essential oils of *A. triplinervis* morphotypes and their acute oral toxicity in non-target organism.

Material and methods

Plant Material

The leaves of *Ayapana triplinervis* morphotypes A and B were collected in the District of Fazendinha (S 0° 03'69.55" and W 51° 11'03.77"), in Macapá, Amapá. Specimen samples were identified by Doctor Patrick Cantuária and deposited in Herbarium Amapaense (HAMAB) of the Institute of Scientific and Technological Research of Amapá (IEPA) under the codes ABLR001-HAMAB e ABLR002-HAMAB.

Essential Oils Extraction

The essential oils were extracted by hydrodistillation using Clevenger apparatus. Dry leaves of morphotype A (2,652 Kg) and morphotype B (0.653 Kg) were extracted for two hours at 100 °C. The essential oils were stored in an amber bottle at -4 °C under light protection [14].

Gas Chromatography/Mass Spectrometry Analysis

The chemical compositions of the essential oils were determined by gas chromatography coupled to the mass spectrometer (GC-MS) [15] on Shimadzu equipment, model CGMS-QP 2010, on a DB-5HT column of the J & W Scientific brand, column 30 m, diameter 0.32 mm, film thickness 0.10 µm and nitrogen as carrier gas. The apparatus was operated under internal column pressure of 56.7 kPa, split ratio 1:20, gas flow in the column of 1.0 mL.min⁻¹ (210 °C), injector temperature 220 °C, and the detector (CG-EM) of 240 °C. The initial temperature of the column was 60 °C with an increase of 3 °C.min⁻¹ until it reached 240 °C, kept constant for 30 minutes. The mass spectrometer was programmed to perform readings in a range of 29 to 400 Da, at intervals of 0.5 seconds, with ionization energy of 70 eV. It was injected 1 µL of each sample with a concentration of 10,000 ppm dissolved in hexane.

A standard mixture of n-alkanes (Sigma-Aldrich C8 - C40) was used to verify the performance of the GC-MS system and calculate the linear retention index (LRI) of each compound in the sample. 1 µL of n-alkanes was injected into the GC-MS system operated under the conditions described above, and their respective retention times (R_T) were used as an external reference standard for calculating the LRI, together with R_T of each compound of interest. The LRI was calculated according to the literature indication and the phytochemical identification was based on the comparison of their pattern of

fragmentation mass spectrum with the library of the equipment and by comparing the calculated linear retention index (LRI) to the literature [16].

Nano-emulsification method

Nano-emulsions were prepared by low-energy method by phase inversion emulsification according to the methodology of Ostertard, Weiss, and McClement [17] and Oliveira et al. [18], which involves the titration of an aqueous phase in an organic phase with constant stirring at room temperature. The organic phase was formed by adding the surfactant to the oil and homogenized for 30 minutes at 750 rpm. Then the aqueous phase was added dropwise and subjected to a magnetic stirrer for 60 minutes. The final essential oil concentration was 25,000 $\mu\text{g.mL}^{-1}$ and the surfactant/oil ratio (SOR) equal 1:1.

Required Hydrophilic-Lipophilic Balance

A mixture of non-ionic surfactants with variable hydrophilic-lipophilic equilibrium value (sorbitan monooleate HLB = 4.3, polyoxyethylene sorbitan monooleate HLB = 15, polyoxyethylene sorbitan monolaurate HLB = 16.7) was used to achieve a range of values of HLB between 8 and 16. The rHLB value of the mixture was calculated with equation 06 [19].

$$rHLB = \frac{[(HLB_1 - m_1) + (HLB_2 - m_2)]}{(m_1 + m_2)}$$

Where rHLB is the required hydrophilic-lipophilic balance, HLB_1 and HLB_2 are the hydrophilic-lipophilic balance values of the mixed surfactants, and m_1 and m_2 are their respective masses.

Particle Size Distribution and Zeta Potential Measurements

Photon Correlation Spectroscopy analysis was performed using Zetasizer Nano Zs (Malvern Instruments, Malvern, UK) equipment with red laser 10 mW ($\lambda = 632.8$ nm). Nano-emulsions were diluted in deionized water (1:25 v/v) and analyzed on the 90° angle. Droplet Size, polydispersity index, zeta potential were evaluated in triplicate and expressed as mean and standard deviation [19].

Evaluation of Larvicidal Activity against *A. aegypti*

A. aegypti larvae from the colony of Institute of Scientific and Technological Research of Amapá – IEPA were maintained in a room (3 m x 4 m) with controlled climatic conditions: temperature of 25 ± 2 °C, relative air humidity of $75 \pm 5\%$, photoperiod of 12 hours.

The methodology was carried out in accordance with the standardized protocol of the World Health Organizations [20] with modification. A preliminary test was conducted at 500, 400, 300, 200 and 100 $\mu\text{g.mL}^{-1}$ in 100 mL of aqueous solution of essential oil with DMSO at 5% or nano-emulsion. Then were added 25 *A. aegypti* third-instar larvae and their mortality was assessed at 24 and 48 hours after exposure.

After the preliminary test, the larvicidal activity of morphotype A essential oil and nano-emulsion were evaluated at 150, 125, 100, 75 e 50 $\mu\text{g.mL}^{-1}$, while morphotype B essential oil and nano-emulsion were evaluated at 100, 80, 60, 40 e 20 $\mu\text{g.mL}^{-1}$. Negative control was 5% DMSO aqueous solution for the essential oil, and nano-emulsion was used surfactant blend at 2.5%. The positive control was BTI in

commercial solution at 0.37 µg.mL⁻¹. The tests were carried out in quintuplicates and the average temperature of the aqueous solution was 25 °C.

After 24 and 48 hours of exposure, dead larvae were counted, taking as dead all those unable to reach the surface. Tests in which the negative control mortality exceeded 20% were discarded and repeated, those with mortality between 5 and 20% were corrected using the Abbott's equation [21].

$$\%Mortality = \frac{((\%M_T - \%M_{NC}) \times 100)}{(100 - \%M_{NC})}$$

%M_T = Test Mortality Percentage;

%M_{NC} = Negative Control Mortality Percentage.

Light and Scanning Electron Microscopy Analysis

Larvae treated in the highest concentration of each nano-emulsion were fixed in 70% ethanol and analyzed under a light microscope model BX41 (Olympus) and photographed in a camera model MDCE 5C. Scanning electron microscopy analyses were evaluated under vacuum using Tabletop Microscope TM3030Plus (Hitachi, Japan) according to Botas and co-workers [22].

Evaluation of Acute Toxicity of *A. triplinervis* Morphotypes Nano-emulsions

This study was approved by the Research Ethics Committee of Universidade Federal do Amapá (CEP – Unifap – 004/2019). All procedures were performed in accordance with international animal care protocols and national animal testing

regulations. The experiments used male and female, 12-week old Swiss albino mice (*Mus musculus*), provided by the Multidisciplinary Center for Biological Research in the Area of Science in Laboratory Animals (CEMIB).

Animals were kept in polyethylene cages at a controlled temperature (25 °C) over a 12-hour period of light or dark cycle and had free access to food and water. The animals had restricted food for three hours before the beginning of the experiment, the food was replaced one hour after the treatment with nano-emulsion.

Acute oral toxicity of the nano-emulsions was performed with 03 experimental groups (morphotype A essential oil nano-emulsion (MAEON), morphotype B essential oil nano-emulsion (MBEON) and negative control), each group with 06 animals (03 males and 03 females).

The test was performed according to the Guideline 423 of the Organization for Economic Co-operation and Development [23] for acute toxic dose class testing. The nanoformulations and negative control were diluted in water at 2,000 mg.kg⁻¹ (limit of 1 mL of the solution per 100 g of body weight of the animals) and administered orally through an appropriate cannula.

The Swiss albino mice (*Mus musculus*) were observed at 30, 60, 120, 240 and 360 minutes after oral treatment and daily for 14 days. Behavioural changes were assessed and recorded: general activity, vocal frenzy, irritability, touch response, response to tail stimulus, contortion, posterior train position, straightening reflex, body tone, grip strength, ataxia, auricular reflex, corneal reflex, tremors, convulsions, anaesthesia, lacrimation, ptosis, urination, defecation, piloerection, hypothermia,

breathing, cyanosis, hyperaemia and death. Water consumption (mL) and food intake (g) were evaluated daily, and body weight every 03 days.

The animals were sacrificed on the fourteenth day in a CO₂ chamber, obeying the ethical principles of animal experimentation. Liver, heart, kidneys and lungs were removed for weighing and macroscopic and histopathological analysis.

Histopathological Analysis

Liver, heart, kidneys and lungs were fixed in 10% formalin buffer solution, routinely processed and embedded in paraffin. Paraffin sections (5 µm) were stained with haematoxylin and eosin. The slides were examined under a light microscope and the magnified images of the tissues' structure were captured for further study [24].

Statistical Analysis

The data were organized into mean and standard deviations. Probit analyses were used to determine the larvae's susceptibility and Anova to determine the statistical difference between treatments using the Statistical Package for the Social Sciences program (version 22, Chicago, Illinois), with a 95% confidence limit.

Results

The morphotype A essential oil showed a translucent colour and yield 0.248% ± 0.037 (m/m). Phytochemical analyses of the MAEO by GC-MS indicated the major compounds (E)-Caryophyllene (45.93%) and Thymohydroquinone Dimethyl Ether (32.93%). The Morphotype B essential oil (MBEO) yield 0.753% ± 0.467 m/m and was

243 purple, Thymohydroquinone Dimethyl Ether (84.53%) was the only compound in
244 greater quantity identified in the MBEO by GC-MS.

245 Thymol Methyl Ether, γ -Gurjunene and (E)-Caryophyllene were exclusively
246 detected in morphotype A, and Valencene and α -Cedrene Epoxide were found only in
247 morphotype B, as shown in Table 1. Additional phytochemical data is available on
248 supplemental material S1-S4.

249 **Table 1.** Chemical composition of essential oils of *A. triplinervis* morphotypes A and B.

Compound	Morphotype A		Morphotype B		LRI Lit.
	Percentege (%)	LRI	Percentege	LRI	
α -Pinene	0.93	948	1.25	948	932
β -Pinene	2.16	943	2.26	943	974
Thymol Methyl Ether	0.68	1231	-	-	1232
α -Gurjunene	0.98	1419	0.41	1419	1409
β -Caryophyllene	45.93	1494	-	-	1419
Thymohydroquinone Dimethyl Ether	32.93	1423	84.53	1423	1424
α -Humulene	1.60	1579	1.11	1579	1452
Aciphyllene	0.53	1490	-	-	1501
β -Selinene	0.88	1469	0.41	1469	1489
α -Muurolene	0.45	1435	-	-	1392
Valencene	-	-	0.53	1474	1496
γ -Gurjenene	0.64	1461	-	-	1475
Caryophyllene Oxide	0.76	1506	0.67	1507	1582

2,5-Di-Tert-Buthyl-1,4-Benzoquinone	4.22	1633	1.82	1633	-
α -Cedrene Epoxide	-	-	0.93	1293	1574

250 LRI = Liner retention index calculated; LRI Lit.= Linear retention index of literature.

251 The content of the α -Pinene and β -pinene monoterpenes added in each
 252 morphotype was equivalent to 3.09% in the MAEO and 3.51% in the MBEO.
 253 Oxygenated derivatives from p-Cymene such as Thymohydroquinone Dimethyl Ether
 254 and 2,5-Di-Tert-Buthyl-1,4-Benzenequinone were found in morphotypes A and B,
 255 while Thymol Methyl Ether was found only in morphotype A. Oxygenated derivatives
 256 of p-cymene totalled 37.00% in morphotype A and 86.35% in morphotype B.

257 Reports in the literature have indicated that the *Eupatorium capillifolium*
 258 essential oil, of which contain 57.1% oxygenated derivatives from p-cymene (36.3%
 259 Thymol Methyl Ether and 20.8% Thymohydroquinone Dimethyl Ether), presented
 260 repellent activity against *A. aegypti* and adulticide activity against *Stephanitis pyrioides*
 261 [25].

262 Carbocyclic and oxygenated sesquiterpenes were found in greater quantity and
 263 diversity in morphotype A than in morphotype B. The sesquiterpene β -Caryophyllene
 264 was predominant only in morphotype A (45.93%). Scientific literature has reported that
 265 antioxidant effect of β -caryophyllene protects rat liver from carbon tetrachloride-
 266 induced fibrosis [26].

267 Studies have shown a variation in the chemical composition of the essential oil
 268 of *A. triplinervis*. However, Thymohydroquinone Dimethyl Ether has been found in
 269 high concentrations. Specimens from Lagoa Grande, Amapá State (Brazil) reported the

presence of the majority compounds Thymohydroquinone Dimethyl Ether with 69.7% and β -Caryophyllene with 19.7% [27]. Another study that analysed the phytochemical composition of *A. triplinervis* essential oil collected on Reunion Island indicated the presence of Thymohydroquinone Dimethyl Ether at 89.9% to 92.8% [7]. However, none of the studies differentiated the chemical composition of the essential oils of each *A. triplinervis* morphotype.

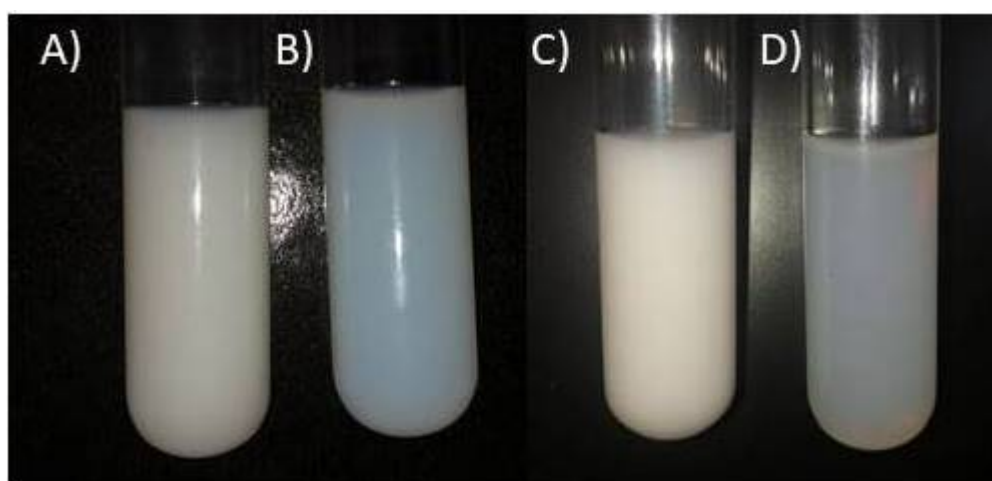
Some species of the Asteraceae family produce Thymohydroquinone Dimethyl Ether as major compounds, among them *Eupatorium marginatum* (64.2%) [28], *Laggera alata* (44.0%) [29], *Pulicania mauritanica* (37.2%) [30], *Laggera pterodonta* (30.5%) [31], *Blumea perrottetiana* (30.0%) [32] and *Sphaeranthus indicus* (18.2%) [33].

Asteraceae are important for their recognized insecticidal activity described in the literature [34, 35]. Alkalamides and coumarins showed insecticidal activity against *Sitophylus oryzae* and *Rhyzopertha dominica* [36], polyacetylenes and terpenes have proven insecticide and repellent activities against larvae and adults of *A. aegypti* [37-39].

Essential oils are good candidates for insecticides, however, their terpenes have low solubility in aqueous media, which represents the first stage of development of culicids such as *A. aegypti*, and are subject to degradation by oxidation and volatility [40]. To improve the solubility of essential oils in water, nano-emulsions were developed by the low energy method using phase inversion composition. This nanoencapsulation technique has advantages: non-heating, solvent free, low cost, and environmentally friendly [41].

The Morphotype A Essential Oil Nanoemulsions (MAEON) produced nanostructures of particle size from 88,830 nm to 333,600 nm, and the Morphotype B Essential Oil Nano-emulsion (MBEON) had particle sizes between 88,830 nm to 333,600 nm on day 0 of analysis (data not shown). Nano-emulsions formed with the hydrophilic-lipophilic balance (HLB) of 10 to 15 were homogeneous and milky, while HLB 15.5 and 16 showed a bluish reflex for both morphotypes, as seen in Fig 1.

Fig 1. *A. triplinervis* morphotype A nano-emulsion in A) HLB 14 e B) HLB 16, and morphotype B nano-emulsion in C) HLB 14 e D) 16 on day 0.



Light scattering on nanoscale colloidal systems causes a characteristic bluish reflection called the Tyndall Effect and represents a parameter indicative of the kinetic stability acquired by nano-emulsions compared to macroemulsions [42]. However, nano-emulsion are susceptible to flocculation, coalescence and sedimentation instabilities [43]. Table 2 shows the physical-chemical parameters of the most promising nano-emulsions of the essential oil of each *A. triplinervis* morphotype.

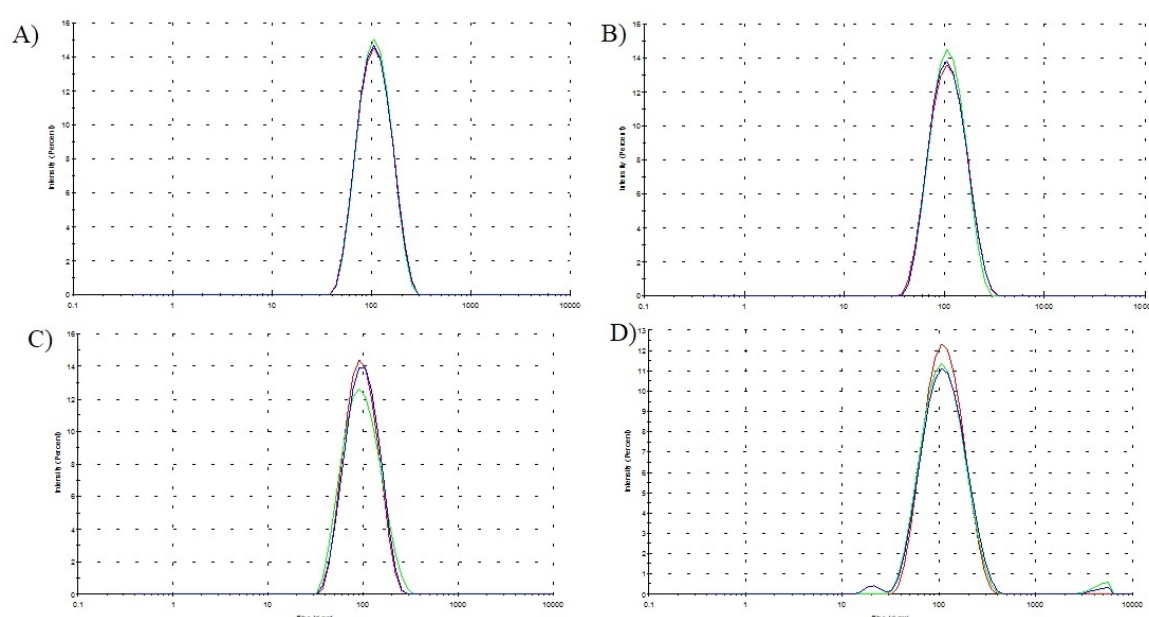
Table 2. Reports of the most promising and optimized nanoemulsion (HLB 16) of each *A. triplinervis* morphotype by particle size (PS), polydispersity index (PdI), zeta potential (ZP), electrophoretic mobility (EM), and electrical conductivity (EC) assessed over time. Data for other HLB values are available in the supplementary material (Fig S5 and S6).

Nano-emulsion	Time (day)	HLB	Particle Size (nn)	Polydispersity Index	Zeta Potential (mV)	Electrophoretic Mobility ($\mu\text{m.cm.V}^{-1}.\text{s}^{-1}$)	Electrical Conductivity (mS.cm^{-1})
MAEON	0	16	88.830 ± 0.948	0.138 ± 0.012	-23.200 ± 0.458	-1.820 ± 0.038	0.025 ± 0.011
	7	16	92.783 ± 0.225	0.120 ± 0.007	-17.667 ± 0.404	-1.385 ± 0.029	0.021 ± 0.004
	14	16	101.167 ± 0.971	0.125 ± 0.009	-20.100 ± 2.787	-1.577 ± 0.220	0.021 ± 0.004
	21	16	99.463 ± 0.242	0.121 ± 0.008	-32.767 ± 1.450	-2.571 ± 0.115	0.014 ± 0.002
MBEON	0	16	99.637 ± 0.529	0.213 ± 0.011	-22.000 ± 1.153	-1.726 ± 0.090	0.020 ± 0.013
	7	16	104.067 ± 0.306	0.186 ± 0.012	-22.400 ± 1.153	-1.756 ± 0.091	0.022 ± 0.001
	14	16	106.567 ± 0.416	0.168 ± 0.016	-26.700 ± 1.308	-2.093 ± 0.100	0.013 ± 0.006
	21	16	99.420 ± 0.519	0.132 ± 0.008	-31.033 ± 2.178	-2.434 ± 0.175	0.020 ± 0.013

MAEON = Morphotype A Essential Oil Nanoemulsion. MBEON = Morphotype B Essential Oil Nanoemulsion; HLB = hydrophilic-lipophilic balance value. Anova did not indicate significant differences between PS, PdI, ZP, EM and EC of MAEON compared to MBEON (p-value <0.001 and F = 772.667).

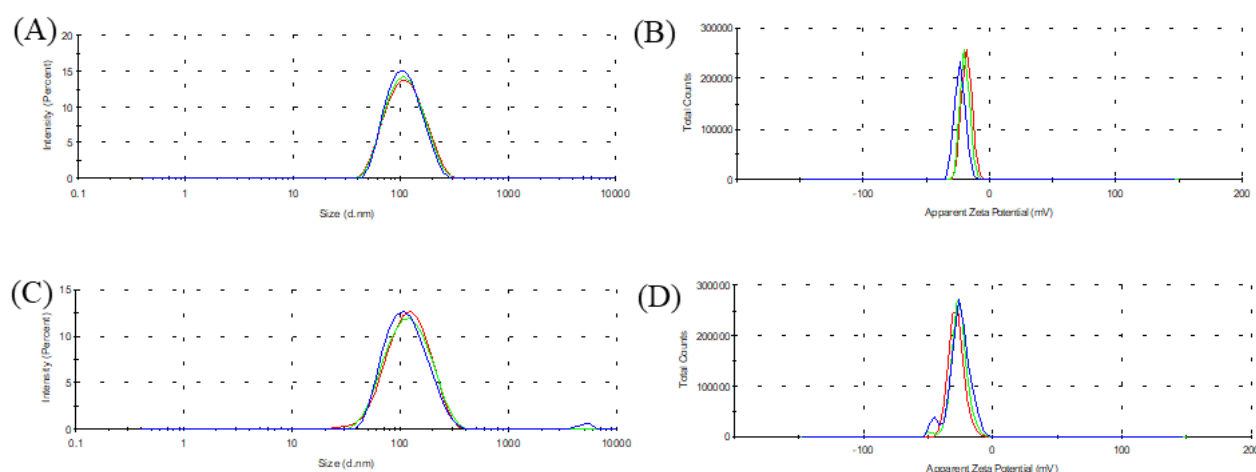
The narrowest emulsions were produced in HLB 16, and their particle size ranged from 88.830 nm to 99.463 nm for the Morphotype A essential oil, and between 99.637 nm to 99.420 nm for the Morphotype B essential oil between days 0 and 21, as shown in the particle size distribution in Fig 2. Emulsions with particle diameters between 20 and 200 nm are called nano-emulsions and possess stability against cremation, flocculation, and sedimentation, but are susceptible to Ostwald ripening over time [43-44].

Fig 2. Particle size distribution of the *A. triplinervis* essential oils nano-emulsion A) morphotype A on day 0; B) morphotype B on day 0; C) morphotype A on day 21; e D) morphotype B on day 21.



The photon correlation spectroscopy analyses indicated that the morphotype A essential oil nano-emulsion (MAEON) had a particle diameter equal to 101.400 ± 0.971 nm ($PdI = 0.124 \pm 0.009$ and $ZP = -19.300 \pm 0.787$ mV). Morphotype B essential oil nano-emulsion (MBEON) showed particle size 104.567 ± 0.416 nm ($PdI = 0.168 \pm 0.016$ and $ZP = -27.700 \pm 1.307$ mV), as indicated in the particle size distribution and zeta potential distribution graph in Fig 3.

Fig 3. Size distribution by intensity and Zeta Potential Distribution of nano-emulsion containing essential oil of *A. triplinervis* morphotype A (A and B) and morphotype B (C and D)



When the particle size distribution is polymodal, i.e with a Polydispersity Index (PI) greater than 0.3, nano-emulsions are susceptible to Ostwald ripening degradation. The mechanism of instability is through the chemical potential difference between smaller droplets and larger droplets caused by mass transport between them, and can cause degradation by coalescence or flocculation [45, 46]. Polydispersity index of morphotype A and B nano-emulsions demonstrated uniform particle size and monomodal distribution, thus is stable against Ostwald ripening [47, 48].

Zeta potential comes from the surface distribution of charges in nano-emulsions and its value of around ± 30 mV indicates a stable system through electrostatic repulsion that prevents flocculation degradation in nano-emulsions [49, 50]. From a zeta potential standpoint, the NOEMA and NOEMB acquired stability over time.

The low solubility of essential oils makes their application in chemical control of *Aedes aegypti* larvae unfeasible, therefore, an O / W nanoemulsion adds advantages as a nanobiotechnological product because it improves solubility of essential oils in aqueous media [51].

The solubility of a nanoemulsion can be assessed in terms of the electrical conductivity resulting from the residues of the surfactant organic groups [52]. MAEON presented electrical conductivity equal to 0.021 ± 0.004 mS.cm⁻¹, and MBEON showed conductivity of 0.022 ± 0.001 mS.cm⁻¹. These results indicate that MAEON and MBEON are soluble in aqueous media [53]. Therefore, it can be concluded that the nanoemulsions containing the essential oils of *A. triplinervis* morphotypes present stability and solubility, and can be used for chemical control of *A. aegypti* larvae.

In insecticidal activity, *A. aegypti* larvae showed atypical behaviour ranging from excitation to lethargy when in contact with essential oil or nano-emulsions, and their mortalities varied as a function of exposure time. Larvae of *A. aegypti* exposed to compounds with neuromuscular action have symptoms such as excitation, convulsions, paralysis, and death [54]. Symptoms observed in the experiment suggest toxic neuromuscular effects from nerve synapses through inhibition of octopamine [55].

Analysis of variance showed differences in the distribution of mean larval mortality caused by nano-emulsions compared to mortality from bulk essential oils (p-value = 0.7340 e F = 0.6247) and indicate that the larvae were more susceptible to nano-emulsions, as shown in Tables 3 and 4.

Table 3. Mortality of 3rd instar larvae of *A. aegypti* of essential oil and nano-emulsion of *A. triplinervis* morphotype A.

	MAEO		MAOEN	
Concentration ($\mu\text{g.mL}^{-1}$)	24 h [*]	48 h [*]	24 h [*]	48 h [□]
150	84.80 $\pm$ 8.67 ^a	91.20 $\pm$ 8.67 ^a	92.80 $\pm$ 3.35 ^a	96.80 $\pm$ 1.79 ^a
125	37.60 $\pm$ 12.84 ^{ab}	47.20 $\pm$ 12.46 ^a	64.00 $\pm$ 29.53 ^a	76.00 $\pm$ 38.78 ^a
100	25.60 $\pm$ 12.84 ^{ac}	39.20 $\pm$ 15.34 ^a	35.20 $\pm$ 20.86 ^{ab}	39.20 $\pm$ 11.10 ^a
75	12.80 $\pm$ 7.69 ^a	20.00 $\pm$ 5.66 ^a	27.20 $\pm$ 8.20 ^{ac}	29.60 $\pm$ 7.27 ^{ab}
50	10.40 $\pm$ 6.07 ^a	16.80 $\pm$ 5.93 ^a	22.40 $\pm$ 8.29 ^a	27.20 $\pm$ 12.13 ^{ac}
0	0.00 $\pm$ 0.00 ^a	0.00 $\pm$ 0.00 ^a	0.00 $\pm$ 0.00 ^a	0.00 $\pm$ 0.00 ^a

Means within a column with different letters are significantly different. Means within a row with different symbols are significantly different (p-value <0.001, F = 13.464).

Table 4. Mortality of the 3rd instar larvae of *A. aegypti* of essential oil and nano-emulsion of *A. triplinervis* morphotype B.

	MBEO		MBEON	
Concentration ($\mu\text{g.mL}^{-1}$)	24 h [*]	48 h [*]	24 h [*]	48 h [□]
100	60.00 $\pm$ 11.31 ^a	64.80 $\pm$ 10.35 ^a	88.80 $\pm$ 5.22 ^a	92.80 $\pm$ 5.22 ^a
80	42.40 $\pm$ 8.29 ^{ab}	49.60 $\pm$ 8.76 ^{ab}	80.00 $\pm$ 6.93 ^a	81.60 $\pm$ 8.29 ^a
60	30.40 $\pm$ 6.07 ^{ac}	40.80 $\pm$ 8.67 ^{ac}	65.60 $\pm$ 15.65 ^a	67.20 $\pm$ 20.28 ^a
40	13.60 $\pm$ 5.37 ^a	22.40 $\pm$ 7.80 ^a	54.40 $\pm$ 33.42 ^{ba}	43.20 $\pm$ 24.23 ^{ba}
20	0.80 $\pm$ 1.79 ^a	3.20 $\pm$ 3.35 ^a	5.60 $\pm$ 2.19 ^{ba}	34.40 $\pm$ 28.93 ^{ba}
0	0.00 $\pm$ 0.00 ^a	0.00 $\pm$ 0.00 ^a	0.00 $\pm$ 0.00 ^{ba}	0.00 $\pm$ 0.00 ^{ba}

Means within a column with different letters are significantly different. Means within a row with different simblos are significantly different (p-value = 0.001, F = 7.984).

Nanoencapsulated system provides essential oil with chemical stability, solubility and bioavailability [56]. Bioavailability is facilitated by passive transport of nano-emulsions into cells when they are less than 200 nn in diameter and may represent an increase in nano-emulsion insecticidal activity [57].

Larvicidal compounds may act by absorption into epithelial tissue, respiratory or ingestion or systemic effect on diffusion by different tissues [58]. The mortality effect of *A. aegypti* larvae in contact with nano-emulsions and bulk essential oils as a function of time can be visualized by lethal concentrations that kills 50% (LC₅₀) and 90% (LC₉₀) exposed larvae in Table 5 and tables S5-S12 of supplemental material.

Table 5. Lethal concentration in order to kills 50% and 90% of 3rd instar larvae of *A. aegypti* in contact with the essential oils or the nano-emulsions of *A. triplinervis* morphotypes, and their lower and upper limits.

	24 h	48 h
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	LC ₅₀ (µg.mL ⁻¹)	LC ₉₀ (µg.mL ⁻¹)	LC ₅₀ (µg.mL ⁻¹)	LC ₉₀ (µg.mL ⁻¹)
MAEO	122.082 (99.630 – 181.737)	235.379 (165.731 – 872.543)	105.979 (84.020 – 146.794)	218.231 (154.044 – 778.960)
MAEON	96.228 (73.912 – 127.785)	202.042 (144.733 – 675.615)	87.434 (65.953 – 110.646)	177.207 (131.862 – 455.481)
MBEO	86.196 (65.812 – 177.738)	200.901 (122.421 – 2351.066)	75.347 (55.442 – 135.702)	202.062 (119.379 – 2097.984)
MBEON	44.755 (30.681 – 57.919)	100.473 (73.997 – 214.623)	35.570 (16.191 – 50.818)	117.450 (74.818 – 629.081)

390

391 The LC₅₀ and LC₉₀ were lower at 48 hours of exposure compared to 24 hours.
392 The variation in larvicidal activity over time may be due to the increased concentration
393 of bioactive compounds that are released over time in a nanoformulation [59]. These
394 results are in line with previous studies showing that micro or nano-emulsions are more
395 effective in chemical control of *Culex pipiens pipiens*, *Culex quinquefasciatus* and
396 *Anopheles stephensi* larvae compared to their bulk essential oils [60 – 62].

397 Comparing the LC₅₀ between nano-emulsion and the bulk essential oil of each
398 morphotype, it can be observed that the insecticidal activity of morphotype B was more
399 effective in the mortality of *A. aegypti* larvae than morphotype A. The most pronounced

insecticidal bioactivity of morphotype B may be due to the higher percentage of Thymohydroquinone Dimethyl Ether.

Contrary to this study, the literature reports that *Laggera pterodonta* essential oil, which contains 20.8% Thymohydroquinone Dimethyl Ether, has moderate adulticidal and larvicidal activity but high repellent activity against *A. gambiae* [63].

Laggera alata essential oil contains 24.4% of Thymohydroquinone Dimethyl Ether and has been described with $LC_{50} = 273.38 \text{ mg.mL}^{-1}$ for *A. aegypti* [64]. *Guateria hispida* essential oils were characterized with 21.0% β -caryophyllene, and *Piper arboreum* essential oil with 10.88% showed LC_{50} of $85.74 \text{ }\mu\text{g.mL}^{-1}$ and $55.00 \text{ }\mu\text{g.mL}^{-1}$ against *A. aegypti* larvae [65, 66]. The results obtained in this study compared to literature data indicate that larvicidal activity may be a result of synergistic action of all secondary metabolites in each morphotype [67].

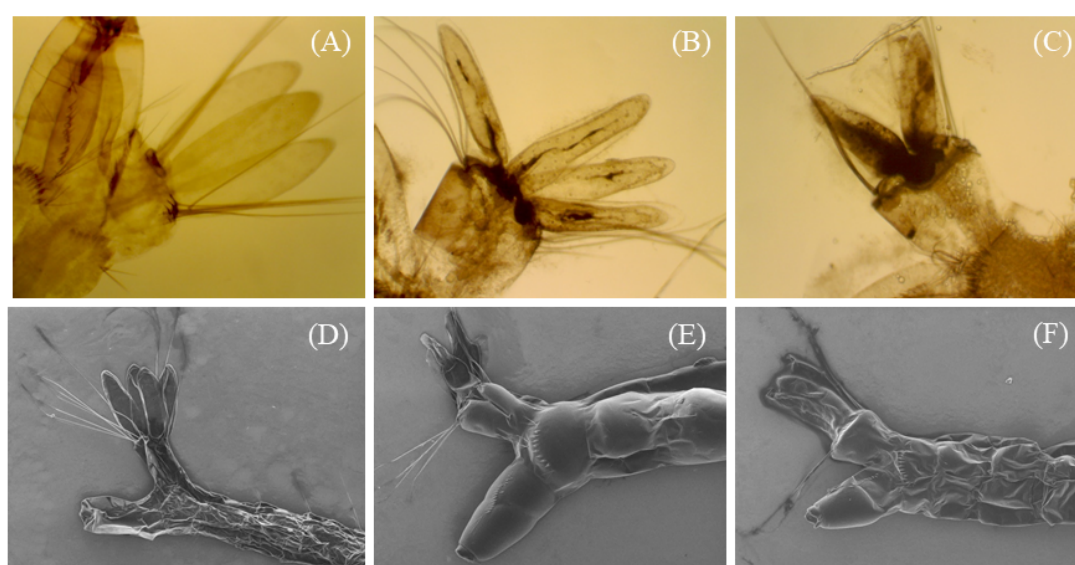
The evaluation of larvicidal activity shows that the essential oil of morphotype A did not show larvicidal activity in *A. aegypti* ($LC_{50} > 100 \text{ }\mu\text{g.mL}^{-1}$), however, the nano-emulsion of morphotype A and the essential oil of morphotype B were active in the control of *A. aegypti* larvae ($LC_{50} < 100 \text{ }\mu\text{g.mL}^{-1}$). Morphotype B essential oil nano-emulsion was the most effective in the chemical control of *A. aegypti* larvae ($LC_{50} < 50 \text{ }\mu\text{g.mL}^{-1}$) [68].

Therefore, it can be concluded that the nano-emulsions containing the essential oils of *A. triplinervis* morphotypes A and B showed nanobiotechnological potential to be applied in the chemical control of *A. aegypti* larvae.

Nano-emulsions show toxicity action that could be observed in larval morphology. The magnitude of toxicity was assessed by a combination of confocal and

scanning electron microscopy. Results indicated deformation and reduction of the volume and surface of anal papillae, and changes deriving from peeling epithelial tissue in the region located in the of the larvae abdomen, as shown in Fig. 4 and supplementary material S11.

Fig 4. Morphological changes in the anal papillae of *A. aegypti* 3rd instar larvae in contact with nano-emulsions of morphotype A essential oil (B e E) and morphotype B (C and F) of *A. triplinervis* compared to control (A e D) by confocal microscopy and scanning electron microscopy.



Essential oils are highly selective for octopaminergic receptors that are distributed in neural and non-neural tissues [69]. Studies have shown that essential oils can modify the activity of neurons by octopamine receptors [70 – 72]. Authors suggest that octopamine acts on insect epithelial tissue to provide plasticity for abdominal distension in breathing and larval movement [73].

Loss of epithelial tissue impairs larval mobility and respiration. Anal papillae morphology and surface dysfunctions affect regulation of osmotic functions and, together with cuticle loss, represent a condition that is detrimental to the survival of *A.*

440 *aegypti* larvae. [54]. These data are in agreement with the results observed for *Pterodon*
441 *emarginatus* nano-emulsion in *Culex quinquefasciatus* larvae [19].

442 In assessment of acute oral toxicity in non-target mammals, mortality was not
443 observed in 2000 mg.Kg⁻¹ of MAEON and MBEON during the 14 days of observation.
444 Hippocratic screening showed that MAEON caused irritability in males and females
445 during the first hour, and corneal reflex in females during the first 3 hours of
446 observation. MBEON caused writhing and irritability in males and females during the
447 first hour of analysis, as shown in Table 6.

448 **Table 6. Effect of oral administration of MAEON and MBEON (2000 mg.kg⁻¹) on**
449 **behavioural parameters of Swiss albino mice (*Mus musculus*) during acute toxicity**
450 **assessment.**

Group	Adults	N° of mice	N° of dead mice	Toxicity Symptoms
Control	Male	3	0	-
	Female	3	0	-
MAEON	Male	3	0	Irritability
	Female	3	0	Irritability and corneal reflex
MBEON	Male	3	0	Contortion and irritability
	Female	3	0	Contortion and irritability

451 In physiological analysis, a decrease in water intake was observed in males and
452 females of the MAEON group, and an increase in water consumption was observed for
453 males and females of the MBEON group. However, these results did not present
454 significant differences when compared with the control group. No changes in food
455 intake were observed in the MAEON and MBEON groups, and the body weight gain

456 did not show significant differences when compared to the control group, as shown in
457 Table 7.

458 **Table 7. Physiological parameters of Swiss albino mice (*Mus musculus*) treated**
459 **with MAEON and MBEON at 2000 mg.kg⁻¹. Values are expressed as mean ±**
460 **standard deviation (n = 3). Statistical significance was calculated using One-way**
461 **Anova followed by Bonferroni's multiple comparison test (p <0.01).**

Parameters	Control		MAEON		MBEON	
	Male	Female	Male	Female	Male	Female
Water (mL)	13.16 ± 1.46	13.35 ± 1.46	10.27 ± 2.19	12.96 ± 1.68	15.34 ± 1.08	15.92 ± 1.26
Food (g)	6.39 ± 0.34	6.30 ± 0.25	6.24 ± 0.51	5.81 ± 0.46	6.16 ± 1.33	5.91 ± 0.34
Body Weight (g)	28.07 ± 1.45	28.85 ± 1.33	29.50 ± 1.44	25.55 ± 1.07	31.47 ± 0.37	24.75 ± 0.39

462 There were no significant differences in relative weight of heart, kidney, and
463 lung when compared to control group organs. However, there was a statistically
464 significant decrease in the absolute liver weight of males treated with MBEON
465 compared to the control group, as shown in Table 8.

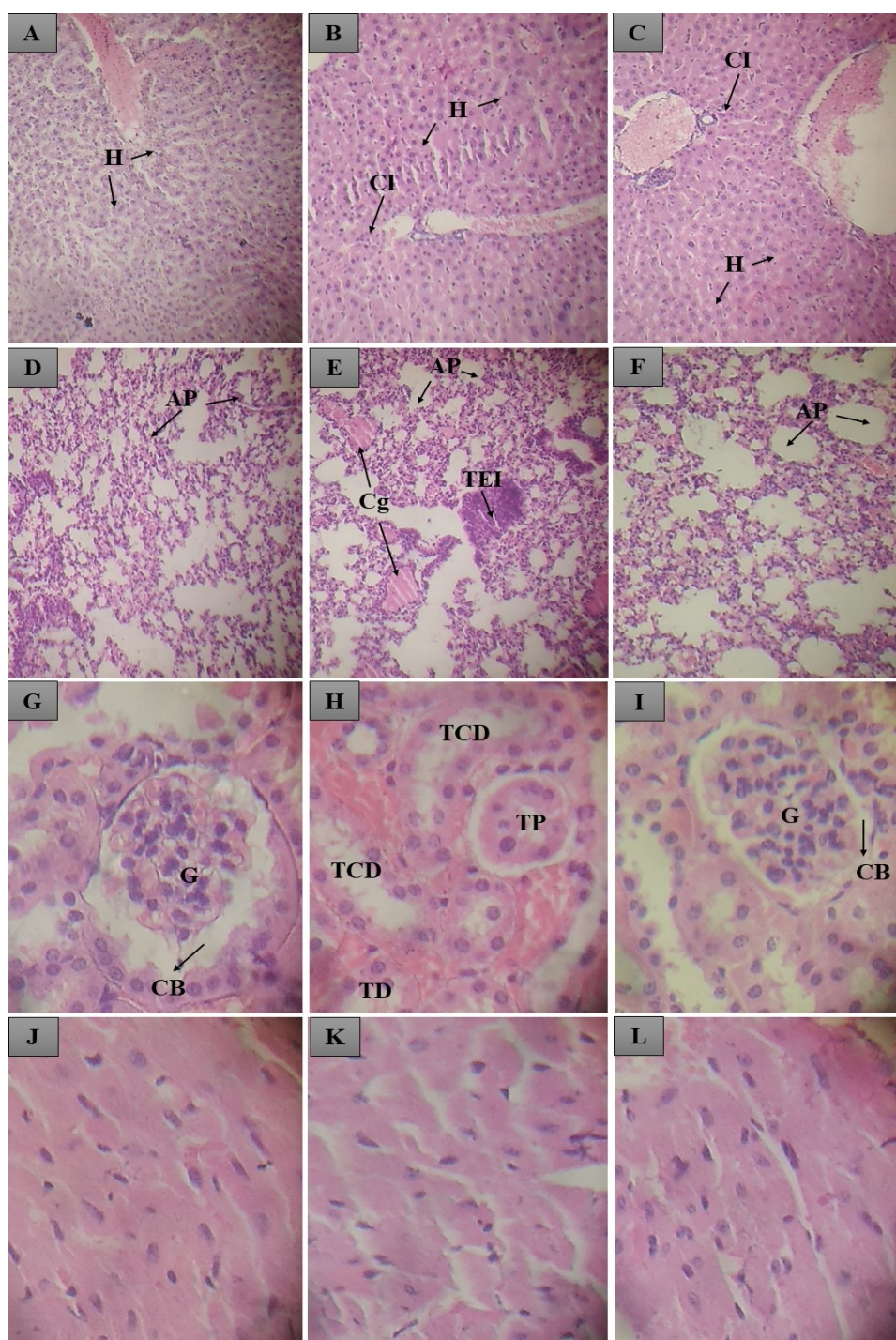
466 **Table 8. Effect of MAEON and MBEON at 2000 mg.kg⁻¹ on the weight of different**
467 **Swiss albino (*Mus musculus*) organs after 14 days of treatment. Values are**
468 **expressed as mean ± standard deviation (n = 3). Statistical significance was**
469 **calculated using One-way Anova followed by Bonferroni's multiple comparison**
470 **test (p <0.01).**

Organs (%)	Control		MAEON		MBEON	
	Males	Females	Males	Females	Males	Females
Liver	5.97 ± 0.42	6.27 ± 0.28	7.55 ± 1.32	7.16 ± 0.50	6.49 ± 0.25*	6.18 ± 0.43
Kidneys	0.81 ± 0.12	0.84 ± 0.12	0.99 ± 0.16	0.76 ± 0.03	0.88 ± 0.05	0.77 ± 0.04
Heart	0.58 ± 0.07	0.54 ± 0.04	0.55 ± 0.09	0.56 ± 0.14	0.49 ± 0.02	0.48 ± 0.01

Lungs	0.73 ± 0.02	0.74 ± 0.02	0.88 ± 0.11	0.85 ± 0.15	0.74 ± 0.15	0.65 ± 0.15
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471 Histological evaluation showed no changes in the liver, heart, lung and kidneys
472 of the control group. However, inflammatory cells were observed in the liver of animals
473 treated with MAEON and MBEON. In the lungs, there was congestion and the presence
474 of transudate with leukocyte infiltration in animals treated with MAEON. The hearts
475 and kidneys of these animals were not affected, as shown in the Fig 5.

476 **Fig 5. Histological section of animals treated orally with control and MAEON and**
477 **MBEON. In A, the normal liver of a control group animal; In B and C, the liver of**
478 **an animal treated with MAEON and MBEON, with normal hepatocytes (H) and**
479 **the presence of inflammatory cells (CI). In D and F, the normal lungs of control**
480 **and MBEON animals, where pulmonary alveoli was observed; In E, the lung of a**
481 **MAEON group animal, where pulmonary alveoli (AP), congestion (Cg) and**
482 **oedematous transudate with leukocyte infiltration (TEI) were observed. In G, H**
483 **and I, the normal kidneys of control, MAEON and MBEON groups with**
484 **glomerulus (G), Bowmam capsule (CB), distal tubule (DT), proximal tubule (TB).**
485 **In J, K and L, normal hearts are observed from the control group, MAEON and**
486 **MBEON.**



Discussion

Few studies are dedicated to reporting the biological activity of *A. triplinervis* in the literature. Studies point to the anxiolytic, antinociceptive and sedative effect on the central nervous system [74], inhibitory activity in the formation of melanin in B16

melanoma cells [75], inhibition of the tyrosinase enzyme [76], antimicrobial activity [77, 78]. Essential oil and Thymohydroquinone Dimethyl Ether isolated from the species demonstrated potential inhibition of ZIKV infection in human cells [79].

The first available chemical data of the *A. triplinervis* in the scientific literature were reported by Semmler in 1908 and indicated 80% of Thymohydroquinone Dimethyl Ether [80]. We report in this study, for the first time in the scientific literature, the phytochemical composition of the essential oils of *A. triplinervis* morphotypes A and B.

Two distinct chemotypes of Thymohydroquinone Dimethyl Ether have been cited in the literature: one in Lucknow (India), which presented Selin-4 (15), 7 (11) - dien-8-one with 36.6%; and the second was found in Tamil Nadu (India), which indicated the presence of 2-Tert-Butyl-1,4-dimethoxybenzene with a percentage of 74.27% [81, 82]. However, no study has differentiated the chemical composition of the essential oil of each morphotype.

The interspecific variability of essential oils produces different chemotypes by a combination of genetic and environmental factors [83] and may contribute to the diversification of ecological interactions between plants and insects [84, 85].

A. triplinervis morphotype A essential oil presented Thymohydroquinone Dimethyl Ether and β -caryophyllene as major compounds. β -caryophyllene is a sesquiterpene that has a wide distribution in species of the family Astereaceae and may have anti-inflammatory, antibiotic, insecticide, antioxidant and anticarcinogenic activity [86]. In morphotype B, the concentration of Thymohydroquinone remained similar to that found in our previous studies.

The chemical composition can vary through climatic and environmental changes [87], and these results can support the understanding of chemical variability in the species and its morphotypes, however, further studies are needed.

The essential oil of morphotype A is the closest to the chemotype described in Brazil by Maia and his co-workers [27], while the essential oil of morphotype B has a greater chemotypic relationship with the specimen described on Reunion Island by Gauvin-Bialacki and Maradon [7]. A different chemotype was reported in India and indicated Selina-4(15),7(11)-dien-8-one as a major compound with a percentage of 36,6%, followed by β -Caryophyllene with 14,7% [81].

Essential oils are considered eco-friendly insecticides, act in low concentrations at the active site, and remain for a short time exposed in nature, however, when inserted into a nanostructured system, they show improvements in insecticidal activity, stability, and safety in their application due to specificity selectivity in insects compared to humans [88].

Our studies have demonstrated eco-friendly, non-heated and solvent-free nano-emulsions that improved the insecticidal activity against *A. aegypti* in relation to bulk essential oils with $LC_{50} < 100 \mu\text{g.mL}^{-1}$ in 24 hours of exposure. This result corroborates other studies published in the literature that showed that the nano-emulsion of *Vitex negundo* is more effective in controlling *A. aegypti* larvae than its bulk essential oil [89], and that the nano-emulsion of *Anethum graveolens* is more toxic to *Anopheles stephensi* than its bulk essential oil [62].

The *A. aegypti* larvae were more susceptible to *A. triplinervis* nano-emulsions than *Baccharis reticularia* (Asteraceae) nano-emulsion with particle size equal to 92.9 nm (PdI = 0.412 e ZP = -20.,4) and $LC_{50} = 221.273 \mu\text{g.mL}^{-1}$ [22].

The larvicidal activity of *A. triplinervis* morphotypes nano-emulsion was more pronounced than *Copaifera duckei* nano-emulsion (PS = 145.2 nm and PdI = 0.378) that showed 70% mortality at $200 \mu\text{g.mL}^{-1}$ [52], and *Pterodon emarginatus* nano-emulsion (PS = 128.0 nm and PdI = 0.250) that demonstrated $LC_{50} = 371.6 \mu\text{g.mL}^{-1}$ in *A. aegypti* larvae [90].

Essential oils are susceptible to deterioration caused by temperature and radiation, which can induce oxidation of their chemical components and cause significant changes in their pharmacological activity [91]. Therefore, it was possible to develop nano-emulsion as a drug delivery system using low energy emulsification method through composition inversion, which uses low-energy input without heating and is solvent-free organic [18] to improve the chemical stability of *A. triplinervis* essential oils [88].

Nano-emulsions can be prepared by the high energy method that uses mechanical devices capable of creating shear stress for formation of finely distributed droplets, and the low-energy method, which depends on spontaneous emulsification within the oil-surfactant-water mixture, altered by changes in phase inversion system or solvent separation [92].

In the low-energy method, spontaneous emulsification is the result of changes in interfacial properties that can be obtained in different ways, such as keeping the composition fixed while the temperature is changing, or keeping the temperature

constant while the composition is changing. Due to the EO's volatility, the increase in temperature may compromise the chemical and pharmacological quality of the nanostructured system, thus, phase inversion emulsification without temperature increase is an appropriate technique. In this process, a catastrophic transition in the O/W to W/O phase inversion is achieved by changing the fraction of slowly added water volume in the system [17].

The literature reports some cases that use the low-energy non-heat method to form an insecticidal nano-emulsions-based delivery system with chemical and kinetic stability in HLB 15. *Baccharis reticular* essential oil, which had a particle size of 138 nm (PdI = 0.453 and ZP = -18.3 mV), and *P. emarginatus* essential oil nano-emulsion with particle size 116.8 nm (PdI = 0.250 and ZP = -19.9 mV) [22, 93].

In addition, *Rosmarinus officinalis* essential oil nano-emulsions were produced in HLB 16.7 with a particle size of 174.1 nm (PdI = 0.136 and ZP was not reported) [51]. The contextualization of these results indicates that the nano-emulsion droplet diameter prepared by the phase inversion method depends on the number of HLB for formulating colloidal stability systems [94].

In this study we also exclusively report colloidal systems with high stability in HLB 16 for the essential oils of *A. triplinervis* morphotypes, which presented monomodal distribution nanostructure and narrow hydrodynamic diameters produced by the low-energy method, non-heat, solvent-free and environmentally friendly with low production costs [19].

The larvicidal activity of *A. triplinervis* essential oils may indicate neuromuscular action by inhibiting the octapaminergic system [95]. Octopamine is a

neuromodulator, neurotransmitter and neurohormone that acts on the insect's nervous system and represents a specific marker of insecticidal activity in essential oils [96].

From the morphological point of view, deformations and reduced volume and surface of anal papillae influence the larval osmoregulation ability and motility [97]. Similar changes in shrinking of the internal cuticle of anal papillae have been described in *A. aegypti* larvae in contact with aqueous extracts of different species of the genus *Piper* and *Argemone mexicana* [98].

The depreciating effects caused by MAEON and MBEON in the anal papillae and in the cuticle affect motility and influence the survival of *A. aegypti* larvae. Similar effects are observed in the morphology of *Culex quinquefasciatus* larvae by the action of *P. emarginatus* nano-emulsion and *A. aegypti* larvae by the action of *B. reticulata* nano-emulsion and may indicate the toxicity mechanism of *A. triplinervis* essential oil nano-emulsions [19, 22].

The development of formulations for the controlled release of terpenes has great potential for reducing side effects and promoting prolonged insecticidal action [99]. Due to the small size that favours rapid absorption and metabolism, the nano-emulsions have the potential to maximize the toxic effect at high concentrations [100], as shown in this study.

Contortion, irritability and corneal reflex indicate action in the central nervous system, and these observations were in accordance with previous studies that demonstrate sedative, anxiolytic and antidepressant effects in different animal models for *A. triplinervis* extract [74].

The oral bioavailability of nano-emulsions increases as a function of the particle size, and its surface charge can influence the potential for acute toxicity through the solubility balance, lipid digestion rate and mucosal penetration rate [101].

The presence of leukocyte infiltrates in the liver, congestion in the alveolar septa and transudate with leukocyte infiltration in the lungs indicate inflammatory processes in these organs caused by MAEON and MBEON [102]. However, the literature has suggested that the application of insecticidal nano-emulsion in non-target mammals is safe [103].

Insecticidal nano-emulsion containing *Eucalyptus staigeriana* essential oil did not show haematological changes in rats [104]. In another study, a nano-emulsion prepared from *Manilkara subsericea* did not show death or toxicological effects on mice [105]. Nano-emulsion containing *Carapa guianensis* did not show cytotoxicity, genotoxicity and hepatotoxicity in mice [106], and Thymohydroquinone Dimethyl Ether, the major metabolite of the samples, showed no toxicity in Zebrafish model [79].

The evaluation of acute toxicity showed that the results found for *A. triplinervis* essential oils nano-emulsions are in accordance with the results described in the literature. The absence of mortality in the nano-emulsion concentration used in this study suggests that the LD₅₀ is superior to the treatment applied (2000 mg.Kg⁻¹) and indicates that nano-emulsions have low toxicity and can be classified in toxicity category 5 [98].

Conclusions

The results indicated that morphotype A presented a different phytochemical composition to that of the major compounds from the morphotype B essential oil. The

essential oil nano-emulsions demonstrated kinetic stability, aqueous solubility, monomodal distribution and particle size 101.400 ± 0.971 for morphotype A, and 104.567 ± 0.416 for morphotype B.

The evaluation of larvicidal activity showed that *A. aegypti* larvae were more susceptible to nano-emulsions than to bulk essential oil. The magnitude of the deleterious effects of nano-emulsions in larvae occurred through the deformation of the anal papillae and the peeling of the cuticle when compared with the control group.

The assessment of acute oral toxicity indicated action on the central nervous system, and histopathological analysis showed damage to the lungs and livers of the animals, however, there was no mortality at the tested dose, indicating that nano-emulsions can be classified in category 5 of toxicity.

In this context, *A. triplinervis* morphotypes essential oils nano-emulsions proved to be an eco-friendly nanobiotechnological product, effective for chemical control of *A. aegypti* larvae and safe for non-target mammals that can be used to control of diseases DENG, CHIKV, ZIKV.

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SUPPORTING INFORMATION

- S1 Fig 1.** Chromatogram of the essential oil of *A. triplinervis* morphotype A.
- S2 table 1.** Chemical composition, retention time (RT), percentage and calculated Linear Retention Index (LRI) and literature of essential oil of *A. triplinervis* morphotype A.
- S3 Fig 2.** Chromatogram of the essential oil of *A. triplinervis* morphotype B.
- S4 table 2.** Chemical composition, retention time (RT), percentage and calculated Linear Retention Index (LRI) and literature of essential oil of *A. triplinervis* morphotype B.
- S5 table 3.** Mortality of *A. aegypti* larvae as a function of the concentration of *A. triplinervis* morphotype A essential oil within 24 hours of exposure.
- S6 table 4.** Mortality of *A. aegypti* larvae as a function of the concentration of *A. triplinervis* morphotype A essential oil within 48 hours of exposure.
- S7 table 5.** Mortality of *A. aegypti* larvae as a function of the concentration of *A. triplinervis* morphotype A essential oil nanoemulsion within 24 hours of exposure.
- S8 table 6.** Mortality of *A. aegypti* larvae as a function of the concentration of *A. triplinervis* morphotype A essential oil nanoemulsion within 48 hours of exposure.
- S9 table 7.** Mortality of *A. aegypti* larvae as a function of the concentration of *A. triplinervis* morphotype B essential oil within 24 hours of exposure.
- S10 table 8.** Mortality of *A. aegypti* larvae as a function of the concentration of *A. triplinervis* morphotype B essential oil within 48 hours of exposure.
- S11 table 9.** Mortality of *A. aegypti* larvae as a function of the concentration of *A. triplinervis* morphotype B essential oil nanoemulsion within 24 hours of exposure.
- S12 table 10.** Mortality of *A. aegypti* larvae as a function of the concentration of *A. triplinervis* morphotype B essential oil nanoemulsion within 48 hours of exposure.
- S13 Fig 3.** Morphological changes in the cuticle of third-insta *A. aegypti* larvae in contact with nanoemulsions containing essential oil of morphotype A (B and E) and morphotype B (C and F) of *A. triplinervis*, and compared to the negative control (A and D) by light microscopy and scanning electron microscopy analyses.