



Complete Genome Sequence of a *Luteibacter* Strain Isolated from a Carnivorous Pitcher Plant (*Sarracenia minor*)

Timothy Yaroshuk,^a  Nikolas M. Stasulli^b

^aDepartment of Forensic Science, University of New Haven, West Haven, Connecticut, USA

^bDepartment of Biology and Environmental Science, University of New Haven, West Haven, Connecticut, USA

ABSTRACT A bacterial isolate of *Luteibacter anthropi*, designated SM7.4, was isolated from decaying insect detritus inside a carnivorous pitcher plant (*Sarracenia minor*). Here, we report a complete genome sequence for *Luteibacter anthropi* strain SM7.4, assembled by combining the Oxford Nanopore MinION flow cell and paired-end Illumina sequencing approaches.

Luteibacter species are Gram-negative, yellow-pigmented bacteria commonly associated with the plant rhizosphere (1–3). The available genomic information for the type strain of *Luteibacter anthropi* is a contig level draft assembly (GenBank accession number [GCA_011759365.1](https://www.ncbi.nlm.nih.gov/nuccore/GCA_011759365.1)) that may not provide the necessary information for studying this bacteria via mutagenesis (4).

Strain SM7.4 was isolated on 17 June 2016 from pitcher plant (*Sarracenia minor*) insect detritus from the North Carolina Botanical Gardens (Chapel Hill, NC, USA; 35.8993°N, 79.0334°W). After incubation at 30°C, the sample was isolated and subcultured on International *Streptomyces* Project 2 (ISP-2) (5) agar plates, made to half-strength (0.5×). Isolate SM7.4 was maintained in 20% glycerol at –80°C. For genomic extractions, a pure isolate was inoculated into 50 mL 0.5× ISP-2 broth and grown overnight at 30°C, with shaking at 160 rpm. DNA was isolated using the Monarch genomic DNA purification kit (New England Biolabs) and quantified via the Invitrogen Qubit double-stranded DNA (dsDNA) high-sensitivity (HS) assay kit, all following the manufacturers' protocols.

For Oxford Nanopore Technologies (ONT) sequencing, library preparation was completed using 200 ng of nonsheared high-molecular-weight DNA with the SQK-RAD004 rapid sequencing kit, according to the manufacturer's instructions, on an R9.4.1 flow cell. MinkNOW v20.06.4 software was used to control the sequencing operations, and Guppy v4.0.9 was used with high accuracy for base calling. For quality control, bases with a score of $\geq Q7$ were sorted into a "fastq_pass" folder and used for subsequent analyses. A total of ~79,570 reads were generated, with 928.64 Mb passing the initial quality filtering (coverage, ~186×; estimated genome size, ~5 Mb). The resulting N_{50} value was 22.63 kb.

The same DNA sample was sequenced with an Illumina NovaSeq instrument through Novogene. For library preparation, DNA was randomly sheared to ~350 bp. Following the manufacturer's protocol, the NEBNext DNA library prep kit was used for end repairing, dA-tailing, and NEBNext adapter ligation. Enrichment via PCR was completed on P5- and P7-indexed oligonucleotides. Paired-end (PE) sequencing generated a read length of 150 bp at each end. Base calling was completed using Casava. Overall, 5,625,594 raw reads were generated, resulting in 1.7 Gb of data (coverage, ~340×). The raw data were filtered by Novogene in-house. Briefly, paired-end reads with >10% uncertain nucleotides, adapter contamination, or >50% of nucleotides with $Q \leq 5$ were discarded. The cleaned reads were used for downstream analysis.

Hybrid assembly was completed using Tricycler v0.5.0 with polishing (6). Adapter trimming was conducted using Porechop v0.2.4 (7) with default parameters. Steps 1 to 7 were performed

Editor David A. Baltus, University of Arizona

Copyright © 2022 Yaroshuk and Stasulli. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Nikolas M. Stasulli, nstasulli@newhaven.edu.

The authors declare no conflict of interest.

Received 22 July 2022

Accepted 28 September 2022

Published 3 November 2022

of the Tricycler instructions on GitHub. For step 1, the assemblers Flye v2.8.3-b1695 (8), Miniasm v0.3-r179/Minipolish v0.1.2 (9), and Raven v1.5.0 (10) were used. One single cluster resulted. One contig of 12 was removed to allow for reconciliation. Medaka v1.4.1 (<https://github.com/nanoporetech/medaka>) was used for polishing with model r941_min_high_360. Pilon v1.24 polishing (11) was conducted 3 times. No custom scripts were used for this assembly. The final assembly was a single circular chromosome of 4,961,663 bp, which was annotated via the DFAST annotation pipeline (12). DFAST was used to determine the highest similarity to *Luteibacter anthrops* strain CCUG25036 and to predict a GC content of 64.2%, 4,255 protein-coding genes (CDs), 6 rRNAs, 57 tRNAs, and 2 CRISPRs. Using ResFinder (13) with default parameters, no antibiotic resistance genes were indicated.

Data availability. This genome project has been deposited at GenBank under BioProject accession number [PRJNA738930](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA738930). The complete genome sequence described here can be found under GenBank accession number [CP077072](https://www.ncbi.nlm.nih.gov/nuccore/CP077072), with the genome described here being version [CP077072.1](https://www.ncbi.nlm.nih.gov/nuccore/CP077072.1). The raw sequences were deposited in the Sequence Read Archive (SRA); the raw ONT-passed fastq files are indexed under accession number [SRR15460238](https://www.ncbi.nlm.nih.gov/sra/SRR15460238), and the paired-end cleaned Illumina reads, as described above, are indexed under accession number [SRR15460237](https://www.ncbi.nlm.nih.gov/sra/SRR15460237).

ACKNOWLEDGMENTS

This work was funded by the University of New Haven, and the funders were not involved in experimental design, execution, or manuscript work. During the initial collection of the sample, N.M.S. was supported by the Training Workforce Development & Diversity Division of the National Institute of General Medical Sciences, National Institutes of Health (K12GM000678). We thank Novogene for Illumina sequencing and initial analysis.

We thank Andrew Wilson at the University of British Columbia for his assistance with the suggestion of programs for sequence assembly. We also thank Jonathan Fehrer for his assistance in the lab. We thank Christine Liloia at the North Carolina Botanical Gardens for coordinating the collection of pitcher plants from their bogs for the initial culturing of this organism.

REFERENCES

- Johansen JE, Binnerup SJ, Kroer N, Mølbak L. 2005. *Luteibacter rhizovicinus* gen. nov., sp. nov., a yellow-pigmented gammaproteobacterium isolated from the rhizosphere of barley (*Hordeum vulgare* L.). *Int J Syst Evol Microbiol* 55: 2285–2291. <https://doi.org/10.1099/ijs.0.63497-0>.
- Akter S, Huq MA. 2018. *Luteibacter pinisoli* sp. nov., a casein degrading bacterium isolated from rhizospheric soil of *Pinus koraiensis*. *Arch Microbiol* 200: 1017–1023. <https://doi.org/10.1007/s00203-018-1515-1>.
- Wang L, Wang G, Li S, Jiang J. 2011. *Luteibacter jiangsuensis* sp. nov.: a methamidophos-degrading bacterium isolated from a methamidophos-manufacturing factory. *Curr Microbiol* 62:289–295. <https://doi.org/10.1007/s00284-010-9707-1>.
- Kämpfer P, Lodders N, Falsen E. 2009. *Luteibacter anthrops* sp. nov., isolated from human blood, and reclassification of *Dyella yeojensis* Kim et al. 2006 as *Luteibacter yeojensis* comb. nov. *Int J Syst Evol Microbiol* 59: 2884–2887. <https://doi.org/10.1099/ijs.0.009100-0>.
- Shirling EB, Gottlieb D. 1966. Methods for characterization of *Streptomyces* species 1. *Int J Syst Evol Microbiol* 16:313–340. <https://doi.org/10.1099/00207713-16-3-313>.
- Wick RR, Judd LM, Cerdeira LT, Hawkey J, Méric G, Vezina B, Wyres KL, Holt KE. 2021. Tricycler: consensus long-read assemblies for bacterial genomes. *Genome Biol* 22:266. <https://doi.org/10.1186/s13059-021-02483-z>.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Completing bacterial genome assemblies with multiplex MinION sequencing. *Microb Genom* 3:e000132. <https://doi.org/10.1099/mgen.0.000132>.
- Kolmogorov M, Yuan J, Lin Y, Pevzner PA. 2019. Assembly of long, error-prone reads using repeat graphs. *Nat Biotechnol* 37:540–546. <https://doi.org/10.1038/s41587-019-0072-8>.
- Wick RR, Holt KE. 2021. Benchmarking of long-read assemblers for prokaryotic whole genome sequencing. *F1000Res* 8:2138. <https://doi.org/10.12688/f1000research.21782.4>.
- Vaser R, Šikić M. 2021. Time- and memory-efficient genome assembly with Raven. *Nat Comput Sci* 1:332–336. <https://doi.org/10.1038/s43588-021-00073-4>.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963–14. <https://doi.org/10.1371/journal.pone.0112963>.
- Tanizawa Y, Fujisawa T, Nakamura Y. 2018. DFAST: a flexible prokaryotic genome annotation pipeline for faster genome publication. *Bioinformatics* 34:1037–1039. <https://doi.org/10.1093/bioinformatics/btx713>.
- Bortolaia V, Kaas RS, Ruppe E, Roberts MC, Schwarz S, Cattoir V, Philippon A, Allesoe RL, Rebelo AR, Florensa AF, Fagelhauer L, Chakraborty T, Neumann B, Werner G, Bender JK, Stingl K, Nguyen M, Coppens J, Xavier BB, Malhotra-Kumar S, Westh H, Pinholt M, Anjum MF, Duggett NA, Kempf I, Nykäsenoja S, Olkkola S, Wiczorek K, Amaro A, Clemente L, Mossong J, Losch S, Ragimbeau C, Lund O, Aarestrup FM. 2020. ResFinder 4.0 for predictions of phenotypes from genotypes. *J Antimicrob Chemother* 75:3491–3500. <https://doi.org/10.1093/jac/ckaa345>.