

Studies

Leaf gas exchange and water relations of the woody desiccation-tolerant *Paraboea rufescens* during dehydration and rehydration

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Abstract

Desiccation-tolerant (DT) plants can withstand dehydration to less than 0.1 g H₂O g⁻¹ dry weight. The mechanism for whole-plant recovery from severe dehydration is still not clear, especially for woody DT plants. In the present study, we evaluated the desiccation tolerance and mechanism of recovery for a potentially new woody resurrection plant *Paraboea rufescens* (Gesneriaceae). We monitored the leaf water status, leaf gas exchange, chlorophyll fluorescence and root pressure of potted *P. rufescens* during dehydration and rehydration, and we investigated the water content and chlorophyll fluorescence of *P. rufescens* leaves in the field during the dry season. After re-watering from a severely dehydrated state, leaf maximum quantum yield of photosystem II of *P. rufescens* quickly recovered to well-watered levels. Leaf water status and leaf hydraulic conductance quickly recovered to well-watered levels after re-watering, while leaf gas exchange traits also trended to recovery, but at a slower rate. The maximum root pressure in rehydrated *P. rufescens* was more than twice in well-watered plants. Our study identified *P. rufescens* as a new DT woody plant. The whole-plant recovery of *P. rufescens* from extreme dehydration is potentially associated with an increase of root pressure after rehydration. These findings provide insights into the mechanisms of recovery of DT plants from dehydration.

Keywords: Desiccation tolerance; Gesneriaceae; leaf hydraulic conductance; *Paraboea rufescens*; rehydration; resurrection plant; root pressure.

Introduction

Desiccation-tolerant (DT) plants can survive in extremely dry environments and withstand dehydration to less than 0.1 g H₂O g⁻¹ dry weight (DW). This equilibrates to 50 % relative air humidity and corresponds to a leaf water potential (Ψ_{leaf}) of about -100 MPa (Gaff 1971; Alpert 2006). Such plants regain normal function after rehydration. Desiccation tolerance is a common phenomenon in lichens, mosses and ferns (Alpert 2006). It is also present in the seeds of most angiosperm species (Liu *et al.* 2008). However, desiccation tolerance is rare in the vegetative tissues of angiosperm species. Only about 135 angiosperm species have been

reported to have desiccation tolerance (Gaff and Oliver 2013). These DT plants can be divided into two types, i.e. homoiochlorophyllous and poikilochlorophyllous (Lüttge *et al.* 2011). Homoiochlorophyllous retain their light-absorbing pigment complexes during dehydration, while poikilochlorophyllous lose their photosynthetic apparatus and thus avoid photodamage (Tuba *et al.* 1998).

One challenge for DT vascular plants during the rehydration process is that they need to refill their xylem after complete embolism during dehydration. The two possible mechanisms that could contribute to xylem refilling in vascular DT plants are capillary rise and positive root pressure. Sherwin *et al.*

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(1998) suggested that capillary rise was the main driving force for xylem refilling in *Myrothamnus flabellifolia*, which is the only known angiosperm DT plant with a woody stem (Moore *et al.* 2007). However, Schneider *et al.* (2000) argued that root pressure, not capillary pressure, was the main driving force for xylem refilling of this species. Root pressure was also found to be an important driving force for xylem refilling of two terrestrial DT ferns (Holmlund *et al.* 2020). However, it is unknown if root pressure is a common mechanism for recovery from complete dehydration in angiosperm DT plants.

Most DT angiosperms are short. That is because the DT plant must remove completely xylem embolism after re-watering from the severely dry state. For example, the height of the tallest woody DT angiosperm plant *M. flabellifolia* is up to 0.8 m (Gaff and Oliver 2013). Desiccation-tolerant plants from Velloziaceae were shorter and showed greater decreases in photosynthetic rate during the dry season than the non-DT species (Alcantara *et al.* 2015). Moreover, clarifying the response of leaf gas exchange and hydraulics in DT plants to dehydration is important for understanding the adaptive strategies of plants to survive in extreme drought conditions.

Gesneriaceae is a large family with 133 genera and more than 3000 species distributed in tropical and subtropical regions (Wang *et al.* 1998). Many species from this family are endemic and endangered, because of their limited distribution area and habitat (Wiehler 1983). Nine genera within the Gesneriaceae have been reported to contain DT species (Gaff and Oliver 2013). Lüttge *et al.* (2011) proposed that there could be DT plants in the genus *Paraboea* (Gesneriaceae); however, there is no direct evidence to support this proposal.

Paraboea rufescens is widely distributed in the karst habitats of southwest China (Wang *et al.* 1998). This species is abundant at the top of karst hills, where water availability can be severely limited in the dry season. Previous studies on this species include reproductive biology (Gao *et al.* 2006) and leaf chlorophyll fluorescence (Huang *et al.* 2012). Although

P. rufescens was referred as a ‘resurrection plant’ by Huang *et al.* (2012), no direct evidence was provided. *Paraboea rufescens* has a woody stem that can be longer than 40 cm, with leaves clustering near the top of the stem (Fig. 1). In the dry season, leaves of *P. rufescens* curl severely and turn their abaxial side outward; however, they unfold and recover completely in the rainy season or after a rain in the dry season. Such characteristics of *P. rufescens* are similar to DT plants. However, there are no reports demonstrating the desiccation tolerance of this species. We hypothesized that (i) *P. rufescens* was a DT plant, able to recover the photosynthetic apparatus from severe dehydration and (ii) after re-watering, root pressure will increase above the value of hydrostatic pressure corresponding to the plant height, which may play an important role in the whole-plant recovery from severe dehydration. We monitored the leaf water status, leaf gas exchange, chlorophyll fluorescence and root pressure of potted *P. rufescens* during dehydration and rehydration, and we investigated the water content and chlorophyll fluorescence of *P. rufescens* leaves in the field during the dry season.

Materials and Methods

Study site and plant material

The study site is located at a karst hill (21°55'N, 101°15'E, 700 m a.s.l.) in Xishuangbanna, Yunnan, China. This region has a monsoon climate with a distinct dry season from November to April. The annual mean precipitation is 1500 mm, and the annual mean temperature is 21 °C. More than 80 % of the rainfall occurs in the rainy season from May to October.

Severely dehydrated leaf and stem samples were collected from each of nine *P. rufescens* individuals in the field during the peak of the dry season in 2016. The water content of these leaves and stems was measured. Two leaves from a similar position on each individual were sampled. One leaf was used to measure leaf water content. The other leaf was rehydrated



Figure 1. Photographs of well-watered and dehydrated *Paraboea rufescens* in the field. (A) A well-watered *P. rufescens* in the rainy season. (B) A dehydrated *P. rufescens* in the dry season. (C) A dehydrated *P. rufescens* with a long stem in the dry season.

by putting the leaf onto a filter paper wet with water in a Petri dish for 24 h, as described by Deng et al. (2003). Maximum quantum yield of photosystem II (PS II) (F_v/F_m) was measured for the rehydrated leaves.

Twenty *P. rufescens* individuals were collected from the top of the karst hill in October 2014 and planted separately into pots (10 cm in diameter × 10 cm depth) for dehydration and rehydration experiments. The potted individuals were put in a shade house with the light level reduced to 50 % by a shade cloth. The shade house was rain-proofed with a plastic roof cover. Plants were well-watered and kept in the shade house for 2 months before water was withheld, and five individuals were well-watered throughout the experiment. During the dehydration process, the mean temperature was 15.5 °C and mean relative humidity was 89 %. The mean light intensity of the shade house was 185 $\mu\text{mol m}^{-2} \text{s}^{-1}$. Dehydration was ended when the leaf water content was lower than 0.10 g $\text{H}_2\text{O g}^{-1}$ DW, which took 27 days. Plants were then re-watered at 20:00 by adding water alone to the soil in the pots, and measurements were conducted at about 10:00 of the next day, which was 14 h after re-watering. Measurements on the rehydration plants were repeated on five continuous days at about the same time. During both dehydration and rehydration processes, leaf water content, water potential, hydraulic conductance, gas exchange rate and quantum yield of photosynthetic system II (Φ_{PSII}) were monitored. Maximum quantum yield of PS II was also measured for well-watered individuals and re-watered individuals on the third, fourth and fifth days after re-watering. Five leaves from each of five individuals were selected for the above measurements. Rehydration was ended when the leaf water content of the re-watered individuals reached the same level as the well-watered control individuals.

Leaf and stem water content

Leaves and 50-mm length of stems with bark removed were weighed to determine fresh weight of samples collected from the karst hill, and then dried at 70 °C for 48 h to determine DW. Leaf and stem water content were expressed as g $\text{H}_2\text{O g}^{-1}$ DW. Leaf water content was measured similarly during the dehydration and rehydration process. The relative water content (RWC, %) was calculated by dividing the absolute water content by the mean value of the saturated leaf water content from the leaf pressure–volume curve, which was 2.69 g $\text{H}_2\text{O g}^{-1}$ DW.

Maximum quantum yield of PS II

Leaf maximum quantum yield of PS II (F_v/F_m) was measured for the field-collected leaves after rehydration, well-watered potted individuals and rehydrated individuals on the third, fourth and fifth day of rehydration with a chlorophyll fluorescence meter (FMS2, Hansatech, Norfolk, UK). Dark-adapting clips affixed to leaves for at least 1 h before measurement. The minimum fluorescence (F_o) was measured firstly, and then the maximum fluorescence (F_m) was measured after a saturating light pulse of 5000 $\mu\text{mol m}^{-2} \text{s}^{-1}$. The F_v/F_m was calculated as $(F_m - F_o)/F_m$ (Maxwell and Johnson 2000).

Leaf pressure–volume curve

Five stems from each of five *P. rufescens* individuals were collected from well-watered potted plants. Stem segments were re-cut at the base and put into water to rehydrate for

2 h. A leaf pressure–volume curve was constructed using a bench-drying method (Turner 1988). Leaf water potential was measured with a pressure chamber (PMS 1000, Corvallis, OR, USA). Leaf weight and positive pressure inside the chamber were measured periodically until the pressure developed in the chamber stopped increasing. The leaves were then dried at 70 °C for 48 h to determine the DW. Prior to evaluation, the curve was visually inspected for indications of excess apoplastic water (plateau effect), which may be an artefact that sometimes accompanies leaf rehydration (Kubiske and Abrams 1991; see Supporting Information—Method S1). Turgor loss point water potential (Ψ_{tlp} , MPa), osmotic potential at full turgor (π_{100} , MPa), leaf water content at turgor loss point, bulk modulus of elasticity along the entire range of positive turgor pressure (ϵ , MPa), and saturated leaf water content (g $\text{H}_2\text{O g}^{-1}$ DW) were then calculated from the pressure–volume curve [see Supporting Information—Method S1, Fig. S4].

Leaf gas exchange and quantum yield of photosynthetic system II

Light-saturated area-based photosynthetic rate (A_a , $\mu\text{mol m}^{-2} \text{s}^{-1}$), maximum stomatal conductance (g_s , $\text{mol m}^{-2} \text{s}^{-1}$) and quantum efficiency of photosynthetic system II (Φ_{PSII}) were measured between 9:00 and 11:00 with a portable leaf gas exchange system embedded with a chlorophyll fluorescence head (LI-6400, Li-COR, Lincoln, NE, USA). The conditions inside the leaf chamber were 400 ppm CO_2 at 25 °C and 50 % relative humidity, with a photosynthetic photon flux density (PPFD) of 1000 $\mu\text{mol m}^{-2} \text{s}^{-1}$. Leaves were equilibrated in the chamber condition for at least 5 min before the leaf gas exchange rate was recorded. Steady-state fluorescence yield (F_t) was measured, followed by the maximum fluorescence in the light (F'_m) after a saturating light pulse of 8000 $\mu\text{mol m}^{-2} \text{s}^{-1}$ light. Φ_{PSII} was calculated as $(F'_m - F_t)/F'_m$ (Maxwell and Johnson 2000). A_a and g_s were measured until the 10th day of dehydration, while Φ_{PSII} was measured until the 12th day of dehydration. After 12 days, the leaves were severely curled, preventing further measurements.

Leaf water potential

Leaf water potential (Ψ_{leaf} , MPa) was measured with a pressure chamber immediately after leaf gas exchange measurements. Leaf water potential was measured until the 13th day of dehydration. Severe shrinkage of the leaf blade and petiole prevented further measurements.

Leaf hydraulic conductance

Leaf hydraulic conductance was measured using an evaporative flux method (Sack et al. 2002). An excised leaf was immediately connected to a pipette via a water-filled tube. The leaf was fixed horizontally onto an open web made with fishing line above a fan and under full sunlight to drive transpiration. The transpiration flow rate (E , mmol s^{-1}) was measured when a steady E was reached. The leaf was then transferred into a moist zip-lock plastic bag. Leaf water potential was measured with a pressure chamber after at least 30 min equilibration within the leaf. The leaf area (LA, cm^2) was measured with a leaf area meter (LI-3000A, Li-COR). Leaf area-based hydraulic conductance (K_{leaf} , $\text{mmol m}^{-2} \text{s}^{-1} \text{MPa}^{-1}$) was calculated as $E/(-\Psi_{\text{leaf}} \times \text{LA})$. K_{leaf} was measured until the 13th day

of withholding water. Severe shrinkage of the leaf blade and petiole prevented further measurements.

Root pressure measurement

Root pressure of four well-watered *P. rufescens* individuals was monitored with digital pressure transducers (PX26-100DV, Omega Engineering, Stamford, CT, USA) following the procedure of [Cao et al. \(2012\)](#). A stem from each individual was cut 2 cm above the ground in the evening. The cut end of the remaining potted plant was immediately connected to a pressure transducer via a water-filled tube. Root pressure was logged every 5 min (CR1000 datalogger, Campbell Scientific, Logan, UT, USA) for 2 days. The value of root pressure of well-watered individuals decreased to negative values and bubbles started to appear in the tube because of daytime transpiration. The tubes were refilled with water and reconnected to the cut end of the stem on the second evening. In addition, root pressure for four individuals from the water-withholding experiment was monitored similarly at the end of the dehydration phase. After re-watering, the pressure transducer was installed immediately to the cut end of the stem on each potted individual. Root pressure data were again logged every 5 min until rehydration ended. Bubbles started to appear in the tubes at the morning of the second day after re-watering (36 h after re-watering). The tubes were refilled with water and reconnected to the cut end of the stem each evening. Root pressure of the re-watered plants was monitored until the sixth day after re-watering.

Statistical analyses

The differences among means across days in leaf water content, Ψ_{leaf} , K_{leaf} , g_s , A_a , Φ_{PSII} and F_v/F_m during the dehydration and re-watering were compared using Tukey's honest significant difference test. The relationships between Ψ_{leaf} and g_s , A_a , K_{leaf} and Φ_{PSII} during dehydration and re-watering were fitted using a generalized additive mode ('gam', [Hastie 2020](#)). All data analyses were done in R ver. 4.0.2 ([R Core Team 2020](#)).

Results

Water status and maximum quantum yield (F_v/F_m) of PS II for *P. rufescens*

The water content of leaf and stem samples of *P. rufescens* collected from the field during the peak of the dry season were 0.079 ± 0.004 g H₂O g⁻¹ DW (mean $\pm$ SE) and 0.062 ± 0.006 g H₂O g⁻¹ DW, respectively. The corresponding RWC for leaf and stem were 3.0 ± 0.1 % and 2.3 ± 0.2 %, respectively. After rehydration by foliar water uptake for 24 h, leaves unfolded completely and F_v/F_m was 0.63 ± 0.02 , which was 90 % of the F_v/F_m in well-watered potted control plants ([Fig. 2](#); see [Supporting Information—Fig. S1](#)). In a pot experiment, after 27 days of dehydration, leaf water content of *P. rufescens* was 0.092 ± 0.006 g H₂O g⁻¹ DW (RWC 3.4 ± 0.2 %), which was similar to that of leaves collected from the field at the peak of the dry season ([Fig. 3A](#)). At this point, plants were re-watered. On the third day of re-watering of dehydrated individuals, the leaf F_v/F_m value was lower than that of the well-watered control plants. However, on the fifth day of re-watering, the F_v/F_m values were no longer significantly different from the well-watered control plants ([Fig. 2](#)).

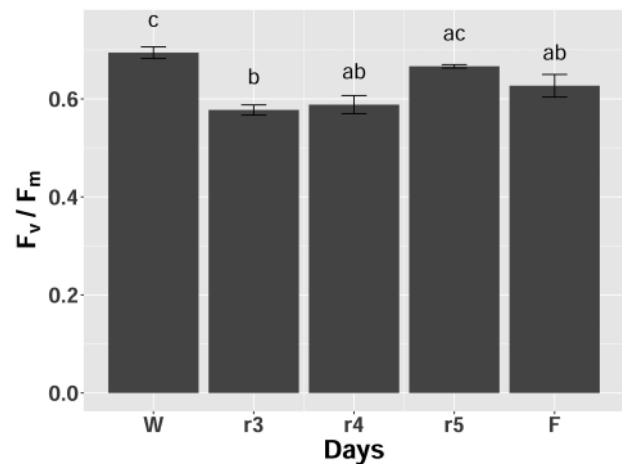


Figure 2. Leaf dark-adapted maximum quantum yield of photosynthetic system II (F_v/F_m) of re-watered and well-watered *Paraboea rufescens*. Means $\pm$ SE ($n = 4-5$) for leaves from well-watered plants (W), for leaves from severely dehydrated potted plants after re-watering for 3 (r3), 4 (r4) and 5 days (r5) and for rehydrated leaves collected from the field at the peak of the dry season (F, $n = 9$) are shown. Different letters above the bars indicate significant differences among means across treatments compared using Tukey's honest significant difference test ($P < 0.05$).

Leaf pressure–volume curve analysis for *P. rufescens*

The water potential at leaf turgor loss was -0.88 ± 0.06 MPa, and the osmotic potential at saturated turgor was -0.62 ± 0.03 MPa. The relative leaf water content at leaf turgor loss point was 78.9 ± 3.3 %, and the bulk modulus of elasticity along the entire range of positive turgor pressure was 3.42 ± 0.75 MPa. The saturated leaf water content was 2.69 ± 0.13 g H₂O g⁻¹ DW.

Effects of dehydration and rehydration on leaf water status, gas exchange and quantum yield of PS II in *P. rufescens*

Leaf water content of potted individuals during dehydration decreased in a stepwise manner. During the first 5 days of dehydration, leaf water content was similar to that of leaves from well-watered control plants, which was 2.47 ± 0.05 g H₂O g⁻¹ DW (RWC: 92 %). Upon further dehydration, leaf water content decreased stepwise to about 1.5 g H₂O g⁻¹ DW from Day 6 to 12, and there was a dramatic drop that eventually reached below 0.1 g H₂O g⁻¹ DW on Day 27 of dehydration ([Fig. 3A](#); see [Supporting Information—Fig. S2](#)). Despite this extreme level of dehydration, leaf water content increased quickly after re-watering. The RWC was 20.5 % after 14 h (first day) of re-watering, and reached 86.4 % after 38 h (second day) of re-watering [see [Supporting Information—Fig. S2](#)]. This high RWC remained stable for the final 3 days of the experiment [see [Supporting Information—Fig. S2](#)].

During dehydration, Ψ_{leaf} dropped gradually ([Fig. 3B](#)). By Day 4 of withholding water, Ψ_{leaf} had decreased to -0.85 ± 0.06 MPa, which was similar to the leaf turgor loss point water potential (Ψ_{tlp} , -0.88 MPa) ($P = 0.615$, t -test). By Day 7 of dehydration, Ψ_{leaf} decreased to -1.69 ± 0.22 MPa, which was significantly lower than the Ψ_{tlp} ($P = 0.020$, t -test). While there was a variation among individuals and slight

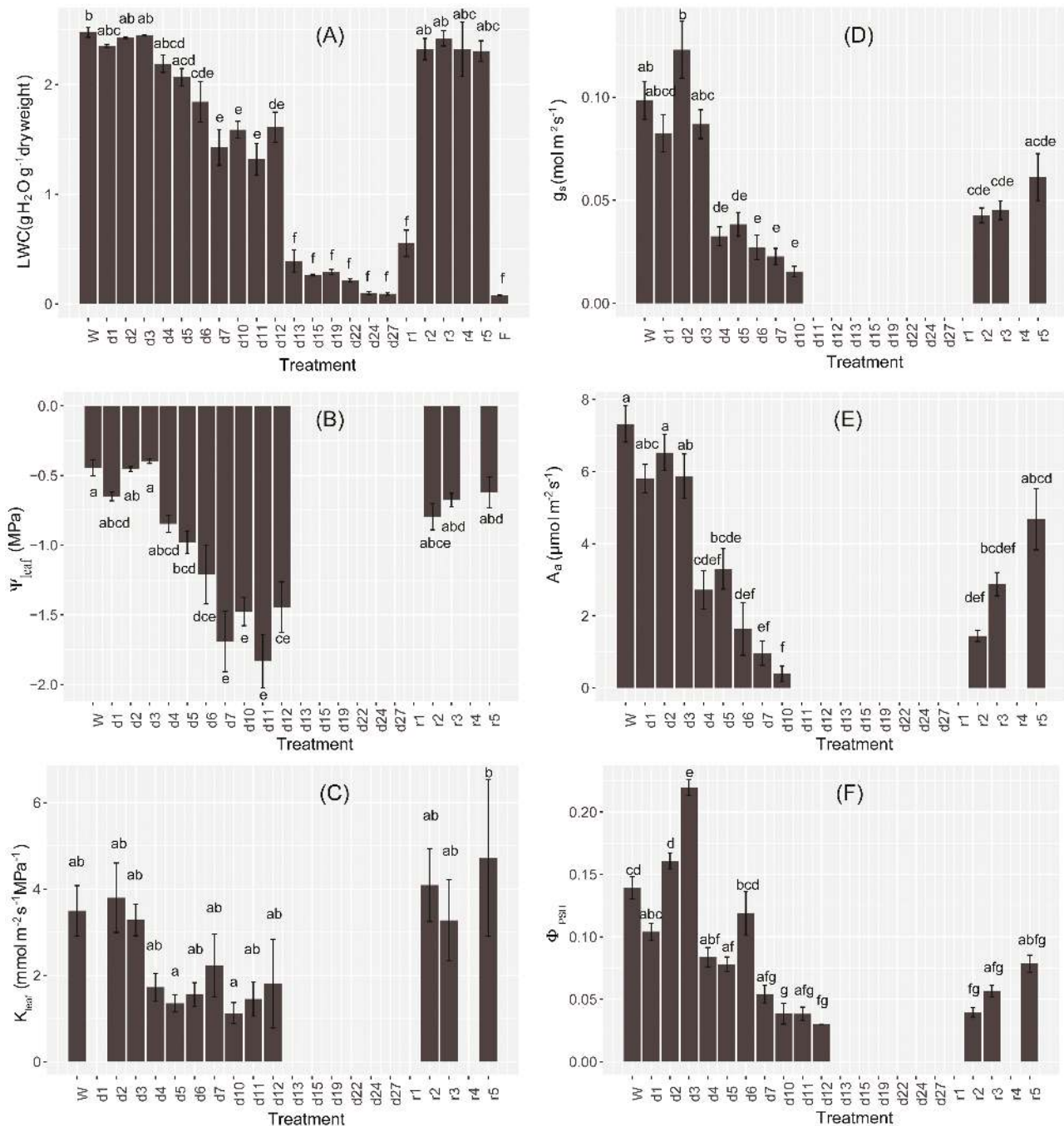


Figure 3. Changes in leaf water status, gas exchange and chlorophyll fluorescence in *Paraboea rufescens* during dehydration and re-watering. (A) leaf water content (LWC), (B) leaf water potential (Ψ_{leaf}), (C) leaf hydraulic conductance (K_{leaf}), (D) maximum stomatal conductance (g_s), (E) leaf area-based photosynthetic rate (A_s) and (F) quantum efficiency of photosynthetic system II (Φ_{PSII}). Values were means $\pm$ SE ($n = 4-5$) taken from well-watered individuals (W), after withholding water for 1–27 days (d1–d27) and after re-watering for 1–5 days (r1–r5). The LWC of leaves collected from the field at the peak of dry season is also given ('F' in panel A, $n = 9$). Different letters above the bars indicate significant differences among means across treatment compared using Tukey's honest significant difference test ($P < 0.05$).

fluctuations among days, Ψ_{leaf} remained stable from Day 7 until Day 12 of dehydration. After the Day 12, Ψ_{leaf} could not be measured due to the strong shrinkage of the leaves. After re-watering, Ψ_{leaf} increased significantly. After re-watering for 38 h (Day 2) there was no significant difference in Ψ_{leaf} between leaves from rehydrated and well-watered control plants (Fig. 3B). After re-watering for 14 h (Day 1), leaves were still curled and folded, which prevented measurement of Ψ_{leaf} , g_s , A_s , K_{leaf} and Φ_{PSII} on that day.

The K_{leaf} was 3.50 ± 0.58 mmol m⁻² s⁻¹ MPa⁻¹ in the well-watered control plants, which decreased to 49 % by Day 4 of withholding water (Fig. 3C). However, K_{leaf} did not decrease further and remained ca. 50 % of the well-watered control plants until the Day 12 of dehydration. After re-watering, K_{leaf} also recovered quickly (Fig. 3C). Although there was some variation among individuals, K_{leaf} recovered to the well-watered control level after 38 h (Day 2) of re-watering and even increased to 135 % of that in leaves of well-watered plants by Day 5 of re-watering.

With the decrease in leaf water content and Ψ_{leaf} , leaf stomatal conductance (g_s), leaf area-based photosynthetic rate (A_a) and quantum yield of photosynthetic system II (Φ_{PSII}) showed a stepwise drop from Day 3 to Day 4 of dehydration (Fig. 3B, 3D–F). Upon dehydration, g_s , A_a and Φ_{PSII} decreased to about 60 % and 33 % of that of well-watered plants on Day 5 and Day 10 of dehydration, respectively. Interestingly, Φ_{PSII} increased to 158 % of the value in the well-watered plants by Day 3 of dehydration. After re-watering, g_s , A_a , Φ_{PSII} increased incrementally, and reached 34 %, 20 % and 28 % of the values in well-watered control plants after 38 h of re-watering (Day 2), respectively. On Day 5 after re-watering, g_s , A_a and Φ_{PSII} were about 60 % of the values in well-watered control plants.

Correlation of Ψ_{leaf} with g_s , A_a , K_{leaf} and Φ_{PSII} during dehydration and rehydration

Stomatal conductance and A_a decreased dramatically with decreasing Ψ_{leaf} (Fig. 4A and B). However, the rate of the decline in g_s decreased when Ψ_{leaf} was below Ψ_{tlp} . The correlation of Ψ_{leaf} with both K_{leaf} and Φ_{PSII} showed a similar trend to that of g_s versus Ψ_{leaf} (Fig. 4C and D), i.e. the rate of decline for both K_{leaf} and Φ_{PSII} decreased when Ψ_{leaf} was below Ψ_{tlp} .

Root pressure in well-watered and re-watered individuals of *P. rufescens*

The root pressure of four well-watered *P. rufescens* individuals exposed to 50 % sunlight remained positive and constant

from evening until about 9:00 of the next day. Root pressure then decreased to negative values in the middle of the day due to transpiration (Fig. 5A). The diurnal pattern of root pressure in the four well-watered individuals showed similar trends over two consecutive days. The maximum root pressure of four well-watered individuals ranged from 4.9 to 11.8 kPa, with a mean value of 7.8 ± 1.7 kPa (Fig. 5A).

Root pressure was measured in a different set of four plants that had been subjected to dehydration and then re-watered (Fig. 5B). In these plants, the diurnal pattern of root pressure fluctuation was similar to that of well-watered individuals on Day 2 of re-watering. Root pressure during the initial phase of rehydration increased continually within 24 h from the start of re-watering. The maximum root pressure during rehydration was generally higher than in well-watered individuals, ranging from 11.5 to 30.4 kPa, with a mean value of 19.8 ± 4.0 kPa (Fig. 5B). The time from re-watering to reach the maximum root pressure was different among the four plants, which were 24 h, 40 h, 42 h and 60 h for plant no. 1 to no. 4, respectively (Fig. 5B). After this initial maximum, the peak root pressure decreased from day to day for the four plants. After re-watering for 72 h, the maximum root pressure approached levels similar to those in the well-watered plants. In addition, the root pressure in all four individuals was positive across most of the daylight hours on the first day of re-watering. However, during daylight hours on subsequent days, the root pressure decreased strongly and could become quite negative (i.e. -20 kPa). Within 24 h of re-watering, parts of the top leaf of all four individuals started to unfold (Fig. 5C). Most leaves at lower position on the four individuals

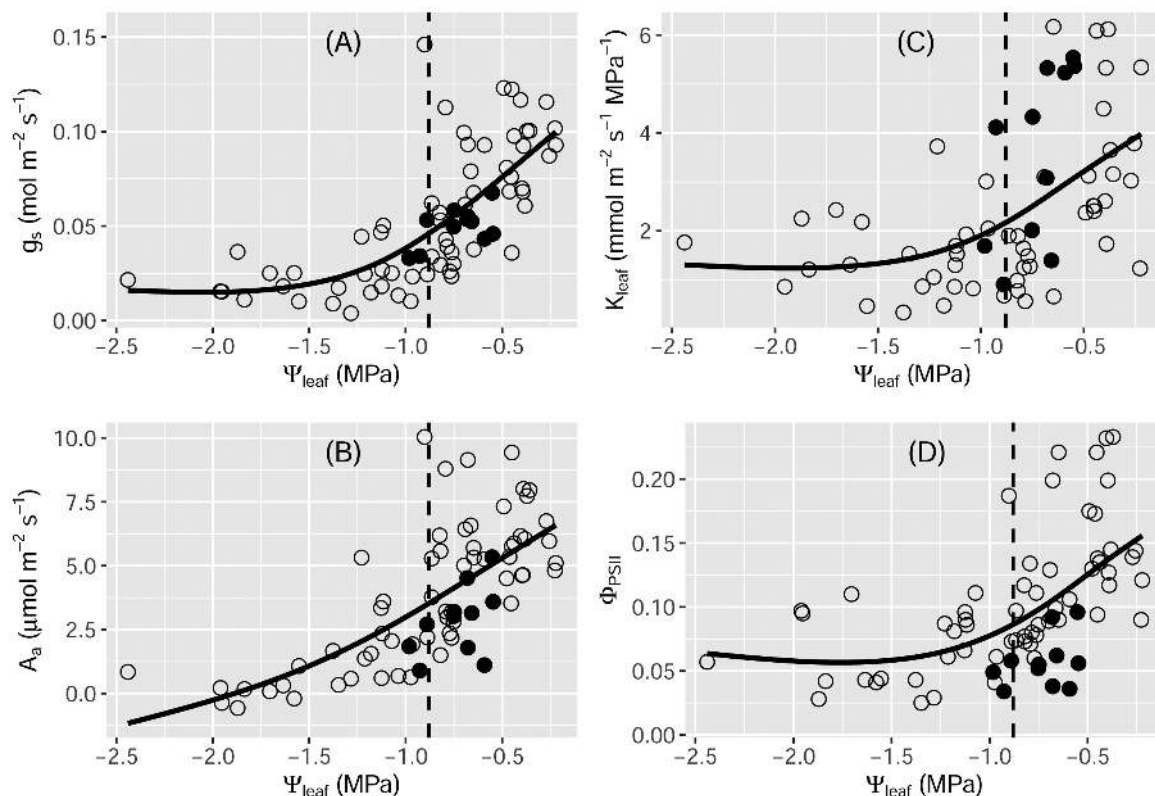


Figure 4. Correlations in *Paraboea rufescens* between leaf water potential (Ψ_{leaf}) and stomatal conductance (g_s) (A), maximum area-based photosynthetic rate (A_a) (B), leaf hydraulic conductance (K_{leaf}) (C) and quantum efficiency of photosynthetic system II (Φ_{PSII}) (D). Data obtained during dehydration (open circles) and during re-watering (closed circles) are shown. Dashed lines indicate the leaf water potential at the turgor loss point (Ψ_{tlp}). Black lines show the trends in the combined data from both plant dehydration and re-watering. Data were fit using the generalized additive model.

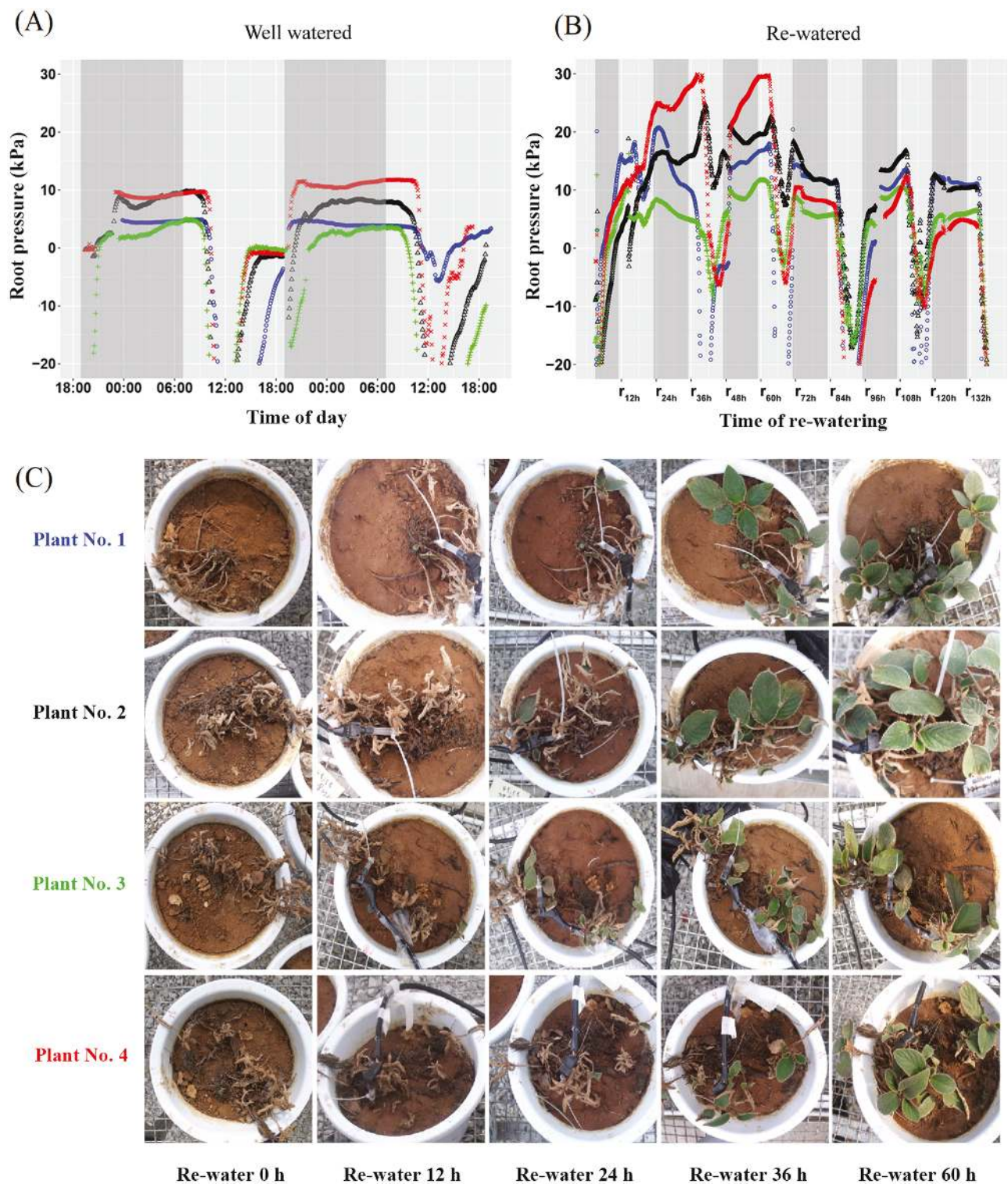


Figure 5. Dynamics of root pressure fluctuations in *Paraboea rufescens* in well-watered individuals (A) and in re-watered individuals recovering from severe dehydration (B), with photos of the four individuals examined during re-watering (C). Different coloured lines and symbols correspond to different individuals. The shaded areas in (A) and (B) indicate the hours of darkness from 19:00 to 7:00.

unfolded and showed recovery after re-watering for 36 h. After re-watering for 60 h, all leaves had recovered completely.

Discussion

Paraboea rufescens is a woody DT plant

Our study provides the first evidence that *P. rufescens* is a DT plant. The leaf water content both in the field and in the

dehydration experiment decreased to less than $0.1 \text{ g H}_2\text{O g}^{-1} \text{ DW}$ without plant mortality (Fig. 3A). After re-watering from the severely dehydrated state, the maximum quantum yield of PS II (F_v/F_m) of detached leaves from the field and leaves of potted individuals both recovered to the levels in well-watered plants (Fig. 2). The recovery of leaf F_v/F_m after rehydration is important evidence for attributing desiccation tolerance to a

plant species (López-Pozo *et al.* 2019). Thus, our study confirmed the speculation of Lüttge *et al.* (2011), where the genus *Paraboea* (Gesneriaceae) was proposed to contain DT plants.

The dehydration and rehydration experiment further confirmed desiccation tolerance of *P. rufescens* (Fig. 3). Leaf water content decreased to less than 0.1 g H₂O g⁻¹ DW at the end of dehydration. However, when water was re-supplied, leaf water status and leaf hydraulic conductance increased quickly, and recovered to the well-watered level within 38 h after re-watering (Fig. 3A–C). Leaf gas exchange and chlorophyll fluorescence traits also trended to recovery, but at a slower rate than that of leaf water status and hydraulic conductance (Fig. 3D–F). Our study suggested that leaf water status and hydraulic conductance of the DT *P. rufescens* could get quickly recovered after rehydration, but the completely recovery of leaf gas exchange might take a longer time.

Although the turgor loss point water potential (Ψ_{tlp}) usually corresponds with the closure of stomata so that plants avoid further dehydration (Bartlett *et al.* 2016), A_a and g_s in *P. rufescens* were still one-third of the levels in well-watered plants on Day 4 of dehydration when Ψ_{tlp} was reached (Fig. 3D and E). Thus, *P. rufescens* could maintain leaf gas exchange even after the turgor loss point. The delay of stomatal closure with dehydration would allow *P. rufescens* to continue carbon assimilation even during water stress. With further dehydration, A_a and g_s decreasing to nearly zero by the Day 10 of withholding water. After re-watering, leaf photosynthetic activity and stomatal conductance recovered gradually. Although A_a and g_s recovered to only 60 % of the well-watered individuals by Day 5 of re-watering, the dark-adapted maximum chlorophyll fluorescence (F_v/F_m) of rehydrated plants on this day was similar to that of well-watered plants (Fig. 2). This observation indicated that the primary photosynthetic process related to the reaction centres of PS II tended to recover faster than the following primary and secondary photosynthetic processes of enzymatic CO₂ assimilation (Fig. 3E and F).

The increase of root pressure after rehydration of *P. rufescens*

We found that the root pressure of rehydrated individuals of *P. rufescens* continually increased during the first 2 days after re-watering (Fig. 5), which could be potentially important for the whole-plant recovery of *P. rufescens* from severe desiccation. Root pressure in rehydrated plants ranged from 11.5 to 30.4 kPa, which was high enough to drive water to a height of 1.17 to 3.10 m, whereas the height the re-watered individuals were less than 0.4 m. High root pressure can successfully refill xylem embolisms and aid the full recovery of two DT ferns (Holmlund *et al.* 2020) and DT *M. flabellifolia* (Schneider *et al.* 2000). Interestingly, the maximum root pressure in rehydrated *P. rufescens* was more than twice that in well-watered plants that were not subjected to desiccation. This large difference in root pressure was similar to that in rice, where re-watered individuals had a root pressure that was 7-fold greater than in well-watered plants before drought (Stiller *et al.* 2003).

Root pressure of re-watered *P. rufescens* plants continued to increase until the leaves were fully unfolded. Once the leaves were unfolded, the root pressure began to decrease during daylight hours due to leaf transpiration (Fig. 5). The increase in root pressure after re-watering might be related to the high ion and solute concentrations that accumulated

during the dehydration process. In addition, the expression of aquaporins during rehydration process might help to speed up the rehydration process. Since the rapid change in water permeability of root over the short term can be mostly accounted by the changes in cell membrane permeability mediated by aquaporins (Javot and Maurel 2002). Besides, Kurowska *et al.* (2019) also found that drought stress and re-watering affected the abundance of aquaporins transcript in barley. Stiller *et al.* (2003) argued that since high root pressure in rice could be maintained for 7 days after re-watering, the increased root pressure could not be caused simply by the passive accumulation of ions. However, the detailed mechanism that generates the high root pressure seen in *P. rufescens* after re-watering of severely dehydrated plants needs further investigation. Understanding the mechanism underlying increased root pressure during re-watering is important for revealing the mechanisms involved in xylem refilling in DT plants.

Capillary force has been confirmed to be an important mechanism for recovery from desiccation in *M. flabellifolia* (Sherwin *et al.* 1998). Our study could not exclude a contribution made by capillary forces to xylem refilling of *P. rufescens* after re-watering. However, root pressure after re-watering was high enough to push water well beyond the top leaves of *P. rufescens*. Guttation was also observed from leaf scars of re-watered *P. rufescens* on the very first day of re-watering [see Supporting Information—Fig. S3B], further indicating that root pressure was in excess of that needed to push water to the top of the plant. Schneider *et al.* (2000) concluded that root pressure rather than capillary pressure was the dominant driving force for the rehydration of *M. flabellifolia*. Holmlund *et al.* (2020) also found that capillary force was insufficient to resurrect distal pinnules in two fern species, perhaps due to reduced xylem transverse area in ferns that limits the resurrection rate. Due to the hydrophobic properties of the inner xylem surface (Zimmermann *et al.* 2004), the contact angle between water and inner xylem surface in dry stem is high, which considerably limits water rise in dry stems of *M. flabellifolia* under capillary pressure (Schneider *et al.* 2000). The capillary force can resurrect proximal pinnules in the DT fern *Pellaea andromedifolia*; however, root pressure is needed for the recovery of distal pinnules (Holmlund *et al.* 2020). However, studies including simultaneous monitoring of root pressure and capillary force are needed to determine the role of capillary force in the rehydration of DT plants after severe dehydration.

Foliar water uptake also played an important role in the recovery of resurrection plants from severe dehydration. The epiphytic DT fern *Pleopeltis polypodioides* could rehydrate the leaf mesophyll and recovered carbon assimilation from foliar water uptake (Prats and Brodersen 2021). In support of the importance of foliar water uptake in plant rehydration, our results showed that placing severely dehydrated leaves from the field on wet filter paper for 24 h allowed the leaves to completely unfold and the dark-adapted F_v/F_m reached 90 % of that observed in well-watered plants. Foliar water uptake could be important for the leave of *P. rufescens* to get quickly recovered and back to normal function from severely dehydration after short-term rainfall events in the field. However, the whole-plant recovery would be more depended on the generation of root pressure, since the recovery of the whole plant would be critical for long-term survival and growth of new leaves for *P. rufescens*.

The ecological significance of *P. rufescens* being a resurrection plant

Being a DT plant is an important adaptive strategy for *P. rufescens*. The annual precipitation in Xishuangbanna is about 1500 mm, of which only 16 % occurs in the dry season from November to April. Due to the low water retention capacity in a karst habitat, plants will experience periodic water deficit, even in the rainy season (Liu et al. 2011). The available soil water in a karst habitat after a heavy rainfall is only sufficient to meet plant transpiration demand for 7–14 days (Li et al. 2008). Since the distribution of *P. rufescens* is restricted to the top of karst hills, it will periodically experience extreme water deficit. According to our measurements, the water content in both leaf and stem in the dry season was below $0.1 \text{ g H}_2\text{O g}^{-1} \text{ DW}$. Leaf water status and hydraulic conductance in *P. rufescens* recovered on the second day after re-watering from severe dehydration (Fig. 3A–C). However, with additional foliar water uptake, the recovery of DT plants can be faster (Holmund et al. 2020). The leaf gas exchange rate for well-watered *P. rufescens* was higher than in *Haberlea rhodopensis* and *Ramonda myconi*, two other resurrection species from the Gesneriaceae, but was lower than that of *Craterostigma plantagineum* and two DT species from Velloziaceae (Nadal et al. 2021). The photosynthetic rate of *P. rufescens* was within the range of tree species growing in the same karst habitat (Fu et al. 2012). Desiccation-tolerant plants have greater chloroplast exposure to intercellular spaces; therefore, they may possess even higher photosynthetic rate than their desiccation-sensitive counterparts when water is not limited (Nadal et al. 2021). Thus, *P. rufescens* has a moderate photosynthetic rate when the water is available, whereas in the dry season, the plant can maintain some level of photosynthesis as Ψ_{leaf} drops below Ψ_{tlp} , and then becomes dormant to avoid damage as it becomes more dehydrated.

Our study showed that *P. rufescens* is a woody DT plant. Leaf water status and hydraulic conductance recovered within 2 days; however, leaf gas exchange could not be recovered to the well-watered status after re-watering for 5 days. The desiccation tolerance trait is likely to be an important adaptive mechanism for *P. rufescens* to survive in the seasonally droughted environment at the tops of karst hills. Our results also showed that root pressure was significantly increased after re-watering of *P. rufescens* from extreme dehydration, which could be important for the whole-plant recovery.

Supporting Information

The following additional information is available in the online version of this article—

Method S1. Data evaluation process of leaf pressure–volume curve.

Figure S1. Photographs of dehydrated *Paraboea rufescens* leaves collected from the field (A) and the same leaves after rehydration on wet filter paper for 24 h (B).

Figure S2. Changes in leaf relative water content (RWC) in *Paraboea rufescens* during dehydration and re-watering. Values are means $\pm$ SE ($n = 4\text{--}5$) taken from well-watered individuals (W), after withholding water for 1–27 days (d1–d27) and after re-watering for 1–5 days (r1–r5). The RWC of leaves collected from the field at the peak of the dry season is also given (F, $n = 9$). Different letters above the bars indicate significant differences among means across treatment compared using Tukey's honest significant difference test ($P < 0.05$).

Figure S3. Guttation (red circles) from leaf scars of *Paraboea rufescens* on a well-watered plant (A) and after re-watering plants from severe dehydration (B).

Figure S4. A typical leaf pressure–volume curve of *Paraboea rufescens*. The x -axis is relative water content (%), and y -axis is the inverse of leaf water potential ($-\text{MPa}^{-1}$). The blue circle indicated the data points after turgor loss point and black circle indicated the data points before turgor loss point. The red line indicated the linear regression line for the data points after turgor loss point. The equation of the linear regression and the R^2 were also shown. Turgor loss point leaf water potential (Ψ_{tlp} , MPa) of this curve was -1.0 MPa , leaf relative water content at turgor loss point was 78 %. The osmotic potential at full turgor (π_{100} , MPa) was -0.71 MPa , and the bulk modulus of elasticity along the entire range of positive turgor pressure (ϵ , MPa) was 2.8 MPa .

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Conflict of Interest

None declared. The authors declare no conflicts of interest.

Contributions by the Authors

P.-L.F. and Y.Z. performed withholding water experiment. P.-L.F. analysed the data and prepared the draft of the manuscript. Y.Z., Y.-J.Z., P.M.F., S.-J.Y. and Z.-X.F. provided important input on the final version of the manuscript.

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Data Availability

The data that support the findings of this study are available from the corresponding authors upon reasonable request.

Literatures Cited

- Alcantara S, de Mello-Silva R, Teodoro GS, Drequeceler K, Ackerly DD, Oliveira RS. 2015. Carbon assimilation and habitat segregation in resurrection plants: a comparison between desiccation- and non-desiccation-tolerant species of Neotropical Velloziaceae (Pandanales). *Functional Ecology* 29:1499–1512.
- Alpert P. 2006. Constraints of tolerance: why are desiccation-tolerant organisms so small or rare? *Journal of Experimental Biology* 209:1575–1584.
- Bartlett MK, Klein T, Jansen S, Choat B, Sack L. 2016. The correlations and sequence of plant stomatal, hydraulic, and wilting responses to

- drought. *Proceedings of the National Academy of Sciences of the United States of America* 113:13098–13103.
- Cao K-F, Yang S-J, Zhang Y-J, Brodribb TJ. 2012. The maximum height of grasses is determined by roots. *Ecology Letters* 15:666–672.
- Deng X, Hu Z-A, Wang H-X, Wen X-G, Kuang T-Y. 2003. A comparison of photosynthetic apparatus of the detached leaves of the resurrection plant *Boea hygrometrica* with its non-tolerant relative *Chirita heterotrichia* in response to dehydration and rehydration. *Plant Science* 165:851–861.
- Fu P-L, Jiang Y-J, Wang A-Y, Brodribb TJ, Zhang J-L, Zhu S-D, Cao K-F. 2012. Stem hydraulic traits and leaf water-stress tolerance are co-ordinated with the leaf phenology of angiosperm trees in an Asian tropical dry karst forest. *Annals of Botany* 110:189–199.
- Gaff DF. 1971. Desiccation-tolerant flowering plants in southern Africa. *Science* 174:10332–11034.
- Gaff DF, Oliver M. 2013. The evolution of desiccation tolerance in angiosperm plants: a rare yet common phenomenon. *Functional Plant Biology* 40:315–328.
- Gao J-Y, Ren P-Y, Yang Z-H, Li Q-J. 2006. The pollination ecology of *Paraboea rufescens* (Gesneriaceae): a buzz-pollinated tropical herb with mirror-image flowers. *Annals of Botany* 97:371–376.
- Hastie T. 2020. *gam: generalized additive models. R package version 1.20*. <https://CRAN.R-project.org/package=gam>, access date: November 10th, 2021.
- Holmlund HI, Davis SD, Ewers FW, Aguirre NM, Sapes G, Sala A, Pittermann J. 2020. Positive root pressure is critical for whole-plant desiccation recovery in two species of terrestrial resurrection ferns. *Journal of Experimental Botany* 71:1139–1150.
- Huang W, Yang S-J, Zhang S-B, Zhang J-L, Cao K-F. 2012. Cyclic electron flow plays an important role in photoprotection for the resurrection plant *Paraboea rufescens* under drought stress. *Planta* 235:819–828.
- Javot H, Maurel C. 2002. The role of aquaporins in root water uptake. *Annals of Botany* 90:301–313.
- Kubiske ME, Abrams MD. 1991. Rehydration effects on pressure-volume relationships in four temperate woody species: variability with site, time of season and drought conditions. *Oecologia* 85:537–542.
- Kurowska MM, Wiecha K, Gajek K, Szarejko I. 2019. Drought stress and re-watering affect the abundance of TIP aquaporin transcripts in barley. *PLoS One* 14:e0226423.
- Li A-D, Lu Y-F, Wei X-L, Yu L-F. 2008. Studies on the regime of soil moisture under different microhabitats in Huajiang karst valley. *Carsologica Sinica* 27:56–62 (in Chinese with English abstract).
- Liu C-C, Liu Y, Guo K, Fan D-Y, Li G-Q, Zheng Y-R, Yu L-F, Yang R. 2011. Effect of drought on pigments, osmotic adjustment and antioxidant enzymes in six woody plant species in karst habitats of southwestern China. *Environmental and Experimental Botany* 71:174–183.
- Liu K, Eastwood RJ, Flynn S, Turner RM, Stuppy WH. 2008. *Seed information database. (Release 7.1, May 2008)*. <http://www.kew.org/data/sid>, access date February 10th, 2022.
- López-Pozo M, Flexas J, Gulías J, Carriquí M, Nadal M, Perera-Castro AV, Clemente-Moreno MJ, Gago J, Núñez-Olivera E, Martínez-Abaigar J, Hernández A, Artetxe U, Bentley J, Farrant JM, Verhoeven A, García-Plazaola JI, Fernández-Marín B. 2019. A field portable method for the semi-quantitative estimation of dehydration tolerance of photosynthetic tissues across distantly related land plants. *Physiologia Plantarum* 167:540–555.
- Lüttge U, Beck E, Bartels D. 2011. *Plant desiccation tolerance*. Berlin, Germany: Springer.
- Maxwell K, Johnson GN. 2000. Chlorophyll fluorescence—a practical guide. *Journal of Experimental Botany* 51:659–668.
- Moore JP, Lindsey GG, Farrant JM, Brandt WF. 2007. An overview of the biology of the desiccation-tolerant resurrection plant *Myrothamnus flabellifolia*. *Annals of Botany* 99:211–217.
- Nadal M, Perera-Castro AV, Gulías J, Farrant JM, Flexas J. 2021. Resurrection plants optimize photosynthesis despite very thick cell walls by means of chloroplast distribution. *Journal of Experimental Botany* 72:2600–2610.
- Prats KA, Brodersen CR. 2021. Desiccation and rehydration dynamics in the epiphytic resurrection fern *Pleopeltis polypodioides*. *Plant Physiology* 187:1501–1518.
- R Core Team. 2020. *R: a language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. <https://www.R-project.org/>, access date: April 10th, 2021.
- Sack L, Melcher PJ, Zwieniecki MA, Holbrook NM. 2002. The hydraulic conductance of the angiosperm leaf lamina: a comparison of three measurement methods. *Journal of Experimental Botany* 53:2177–2184.
- Schneider H, Wistuba N, Wagner HJ, Thurmer F, Zimmermann U. 2000. Water rise kinetics in refilling xylem after desiccation in a resurrection plant. *New Phytologist* 148:221–238.
- Sherwin HW, Pammenter NW, February E, Vander Willigen C, Farrant JM. 1998. Xylem hydraulic characteristics, water relations and wood anatomy of the resurrection plant *Myrothamnus flabellifolius* Welw. *Annals of Botany* 81:567–575.
- Stiller V, Lafitte HR, Sperry JS. 2003. Hydraulic properties of rice and the response of gas exchange to water stress. *Plant Physiology* 132:1698–1706.
- Tuba Z, Protor CF, Csintalan Z. 1998. Ecophysiological responses of homoiochlorophyllous and poikilochlorophyllous desiccation tolerant plants: a comparison and an ecological perspective. *Plant Growth Regulation* 24:211–217.
- Turner NC. 1988. Measurement of plant water status by the pressure chamber technique. *Irrigation Science* 9:289–308.
- Wang W-T, Pan K-Y, Li Z-Y, Weitzman AL, Skog LE. 1998. Gesneriaceae. In: Wu ZY, Raven PH, eds. *Flora of China. Vol. 18. Scrophulariaceae through Gesneriaceae*. Beijing: Science Press, 273–333.
- Wiehler H. 1983. A synopsis of the Neotropical Gesneriaceae. *Selbyana* 6:1–219.
- Zimmermann U, Schneider H, Wegner LH, Haase A. 2004. Water ascent in tall trees: does evolution of land plants rely on a highly metastable state? *New Phytologist* 162:575–615.