

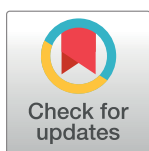
RESEARCH ARTICLE

Self-microemulsifying drug delivery system and nanoemulsion for enhancing aqueous miscibility of *Alpinia galanga* oil

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Abstract

Alpinia galanga oil (AGO) possesses various activities but low aqueous solubility limits its application particularly in aquatic animals. AGO has powerful activity on fish anesthesia. Ethanol used for enhancing water miscible of AGO always shows severe side effects on fish. The present study explores the development of self-microemulsifying drug delivery systems (SMEDDS) and nanoemulsions (NE) to deliver AGO for fish anesthesia with less or no alcohol. Pseudoternary phase diagrams were constructed to identify the best SMEDDS-AGO formulation, whereas NE-AGO were developed by means of high-energy emulsification. The mean droplet size of the best SMEDDS-AGO was 82 ± 0.5 nm whereas that of NE-AGO was 48 ± 1.6 nm. The anesthetic effect of the developed SMEDDS-AGO and NE-AGO in koi (*Cyprinus carpio*) was evaluated and compared with AGO ethanolic solution (EtOH-AGO). It was found that the time of induction the fish to reach the surgical stage of anesthesia was dose dependent. NE-AGO showed significantly higher activity than SMEDDS-AGO and EtOH-AGO, respectively. EtOH-AGO caused unwanted hyperactivity in the fish. This side effect did not occur in the fish anesthetized with SMEDDS-AGO and NE-AGO. In conclusion, SMEDDS and NE are promising delivery systems for AGO.

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Introduction

Alpinia galanga is a plant in family Zingiberaceae. Many parts of *A. galanga* such as rhizome, flower, and young leaf are edible and have been used in Asian folk medicinal and food remedies. *A. galanga* oil (AGO) has wonderful essence that can enhance the flavor of foods and elevate them to a level of brightness. AGO has been extensively used in traditional treatment of stomach ache and diarrhea [1]. Previous studies reported several biological properties of AGO on anti-bacterial, anti-oxidant, and anti-inflammatory activities [2–5]. Interestingly, we have recently found that AGO has excellent action on fish anesthesia [6]. However, the poor water

miscibility of AGO causes limitations in its application, particularly in animals for which water is the essential environment, such as fish.

Absolute ethanol has been used to increase the water miscibility of AGO for fish anesthesia but alcohol causes several side effects in fish, such as hyperactivity, a constellation of congenital and retinal anomalies, and central nervous system (CNS) deficits. Ethanol has also been found to induce stress proteins in zebrafish embryos [7–10]. If we can build up an innovative technology to improve the miscibility of AGO in water and avoid using alcohol, we may solve the two main problems (poor solubility in water of AGO and side effect of alcohol) at the same time. Nanoformulations such as self-microemulsifying drug delivery systems (SMEDDS) and nanoemulsions (NE) have been reported to be able to solubilize and stabilize hydrophobic active ingredients and to enhance the bioavailability of many water insoluble compounds [11–14]. Furthermore, SMEDDS and NE have been shown to enhance the biological activity of natural compounds [15–18].

The objective of the present study was to develop and characterize SMEDDS and NE containing AGO (SMEDDS-AGO and NE-AGO) in order to increase AGO water miscibility and to decrease the use of ethanol in fish anesthesia. The effects of type and amount of surfactant used in the developed SMEDDS-AGO and NE-AGO on the phase behavior of the oil/water systems were investigated. In addition, this study also investigated the anesthetic effect of the developed SMEDDS-AGO and NE-AGO in comparison with AGO ethanolic solution (EtO-H-AGO) on fish using koi (*Cyprinus carpio*) as an animal model.

Materials and methods

Materials

Fresh rhizomes of *A. galanga* were collected in December 2015 from the medicinal garden, Chiang Mai, Thailand. The plant voucher specimen no. 009245 was deposited at the Herbarium, Northern Research Center for Medicinal Plants, Faculty of Pharmacy, Chiang Mai University, Thailand. Polyoxyethylene sorbitan monolaurate (Tween 20) and polyoxyethylene sorbitan monooleate (Tween 80) were from Merck (Hohenbrunn, Germany) while polyoxyethylene octylphenyl ether (Triton X-100) was obtained from Amresco (Cleveland, OH, USA). Absolute ethanol, dichloromethane, and hexane were of analytical grade and were supplied by Merck Millipore (Darmstadt, Germany). Isopropanol was purchased from Labscan Analytical Science (Bangkok, Thailand). Propylene glycol was from Sigma-Aldrich (St. Louis, MO, USA). All other chemicals and solvents were of the highest grade available.

Extraction and chemical analysis of AGO

Fresh rhizomes of *A. galanga* were cut into small pieces and subjected to hydrodistillation for 3 h using a Clevenger apparatus for oil collection. The experiment was done in triplicate. The yield of the oil obtained from each experiment was recorded and the average yield value was calculated. The volatile oil obtained was stored in a refrigerator at -20°C and protected from light until use. AGO was analyzed for its chemical composition using gas chromatography-mass spectrometry (GC-MS). An Agilent 6890 gas chromatograph was coupled to an electron impact (EI, 70 eV) using a Hewlett Packard (HP) mass selective detector (MSD), model HP 5973-MSD and fitted with a fused silica capillary column (HP5-MSI; 30.0 m $\times$ 0.25 mm i.d. $\times$ 0.25 μm film thickness) (Agilent Technologies Inc, USA). The analytical protocol was standardized from previous studies with some modification [19]. Briefly, 1 μL of AGO 1:100 (v/v) diluted with dichloromethane was injected. Identification of the compounds of interest was performed based on comparing retention times to those of reference spectra obtained from two mass spectral databases (Wiley and the National Institute of Standards and Technology;

NIST). The Wiley database is the largest database of its kind and the NIST mass spectral database contains approximately 62,000 mass spectra of an equal number of chemicals [20]. These two mass spectral databases are commonly applied for identifying bioactive compounds in natural plants [21–22]. The percentage of each compound was calculated based on the total area of all peaks obtained from the oil.

Preparation of SMEDDS-AGO

Pseudoternary phase diagrams of AGO were constructed using an aqueous titration method. To obtain the surfactant mixture (Smix), surfactant (Tween 20, Tween 80, or Triton X-100) and cosurfactant (absolute ethanol, propylene glycol, or isopropanol) were mixed at weight ratios of 1:2, 1:1, and 2:1. Then, AGO and Smix were mixed at weight ratios of 0:1, 1:9, 2:8, 3:7, 4:6, 5:5, 6:4, 7:3, 8:2, 9:1 and 1:0. The resulting mixtures were subsequently titrated with water under moderate agitation at ambient temperature. Clear, monophasic liquid samples were classified as microemulsions (ME). All formulations were prepared in triplicate. The pseudoternary phase diagrams were designed using OriginPro8 for Windows (OriginLab Corporation, USA).

Selected formulations of SMEDDS-AGO were then prepared by mixing AGO and several Smix, based on the pseudoternary phase diagrams. Smix was prepared before mixing with AGO using a vortex mixer to the desired weight ratios before transferring to glass vials for further use.

Preparation of NE-AGO

Six formulations of NE-AGO (A, B, C, D, E, and F), composed of 20% AGO, were prepared by means of high-energy emulsification using three different surfactants; Tween 20, Tween 80, and Triton X-100. NE-AGO-A and NE-AGO-B contained 5% and 10% Tween 20, respectively whereas NE-AGO-C and NE-AGO-D contained 5% and 10% Tween 80, respectively and NE-AGO-E and NE-AGO-F contained 5% and 10% Triton X-100, respectively. An emulsion inversion point method (EIP) was used to prepare the pre-emulsions. Briefly, the water phase was added to the oil phase under stirring at 50°C for 5 min before subjecting the mixture to high speed stirring using an Ultra-Turrax T25 (Janke and Kunkel GmbH, Germany) at 16,000 rpm for 5 min. The pre-emulsions obtained (20 g) were then passed through a high pressure homogenizer (HPH) (Avestin Inc., Canada) using various homogenization cycles (1, 3, 5, 7, 10, 15, and 20 cycles) with approximately 1–2 min/cycle and pressures of 5,000 and 10,000 psi at room temperature. The experiments were performed at ambient temperature in triplicate.

Droplet size and zeta potential

The droplet size and size distribution (expressed as polydispersity index, PdI) as well as the zeta potential (ZP) of SMEDDS-AGO and NE-AGO were measured using photon correlation spectroscopy (Zetasizer Nano ZS, Malvern Instruments Ltd., UK). The size measurements were carried out at a fixed angle of 173° at 25°C. All formulations were diluted with distilled water at a ratio of 1:100 (v/v) before measurement. All measurements were carried out in triplicate.

Conductivity study

The electrical conductivity of SMEDDS-AGO and NE-AGO was measured using a conductivity meter EX-20 (Horiba, Japan). The formulations were diluted in distilled water at a ratio of 1:100 (v/v) before measuring. The conductivity meter was calibrated using a standard solution of 1413 $\mu\text{S}/\text{cm}$ (Eutech Instruments) before testing. The experiments were performed at 25°C

by dipping the electrode into the test sample until equilibrium was reached and the readings became stable. All measurements were carried out in triplicate.

Stability study

All SMEDDS-AGO, and NE-AGO formulations were kept at three storage temperatures, 4°C, 20°C, and 40°C, for one month. Changes in optical appearance such as phase separation, color changes, and turbidity were observed visually. The droplet size of the formulations was determined after dispersion the stored samples in distilled water (1:100 v/v) and compared with that of freshly prepared samples investigated in the same manner.

The selected SMEDDS-AGO and NE-AGO kept at different storage conditions (4°C, 20°C, and 40°C) were analyzed for chemical changes using gas chromatography in comparison with AGO. For the SMEDDS-AGO formulations, hexane was added to all samples to get 1:100 dilutions. The diluted samples were mixed using a vortex mixer for 5 min and centrifuged at 13,000 rpm for 1 min. One μL of the supernatant was injected into an Agilent 6890 gas chromatograph equipped with a fused silica capillary column (HP5; 30.0 m $\times$ 0.25 mm i.d. $\times$ 0.25 μm film thickness) from Agilent Technologies Inc, USA, using flame ionization for detection. For the NE-AGO formulations, all samples were firstly diluted with hexane at a ratio of 1:1 (v/v). Subsequently, the hexane layer was separated and further diluted 100 fold with pure hexane. One μL of the diluted sample was injected into the Agilent 6890 gas chromatograph using the same conditions as described above. All experiments were performed in triplicate.

Fish and culture condition

Juvenile *Cyprinus carpio* or koi, with an average weight and length of 9.6 ± 0.1 g and 10.9 ± 0.2 cm respectively, were purchased from a local ornamental fish shop in Thailand, and used in the anesthetic study. The fish were maintained in continuously aerated tanks (500 L) with controlled dechlorinated tap water (temperature 25°C; pH 7.4; total hardness 110 ppm; alkalinity 90 ppm; total ammonia nitrogen and nitrite were negative) for 2–4 weeks before being used in the anesthetic study. 50% of fresh water was changed daily. The fish were fed twice daily with a commercial dry feed (INTEQC Feed, Thailand) and held in natural light conditions. All experimental methods were approved by the Animal Care Committee of the Faculty of Veterinary Medicine, Chiang Mai University (FVM-ACUC no. R3/2555).

Anesthetic effects

Experiments were conducted in glass aquaria (100 $\times$ 100 $\times$ 150 mm), containing 1 L of dechlorinated tap water. EtOH-AGO, 1:10 v/v dilution in absolute ethanol, was used as a positive control. The selected SMEDDS-AGO and NE-AGO at various concentrations (100, 200, 300, and 400 mg/L) of AGO and EtOH-AGO were added to the aquaria. The fish were divided into 12 groups of 20 individual fish for each treatment and concentration ($n = 20$). The fish were selected randomly and used only once. The criteria to determine the anesthetic effects were based on the fish behavioral responses [23]. The surgical stage where the fish reaches stage 3 plane 2 is the most important for evaluating the efficacy of the anesthetics because at this stage the fish stops any swimming activity, shows loss of equilibrium and loss of responsiveness and is thus easy to handle in routine operations, such as netting, weighing, sorting, operations, vaccinations, and transportation. To evaluate the time required for the induction of fish anesthesia into stage 3 plane 2, the fish were individually anesthetized by each formulation with various AGO concentrations. Furthermore, the anesthetic behavioral changes such as hyperactivity (fish exciting and increased swimming activity), jumping (fish jumping out of water) and piping (fish piping for air near water surface) were also recorded by clinical observation. Each fish

was used only once. The maximum observation time was 20 min. When the fish reached the surgical anesthesia stage, they were immediately transferred to anesthetic-free aquaria to measure the recovery time. At the end of the experiment period, the survived fish were maintained in laboratory until they were in healthy condition with no biohazard contamination and then were donated to the aquarist as pet fish.

Statistical analysis

The data are presented as means $\pm$ S.E.M. Kolmogorov-Smirnov's test was used to evaluate normality of the data distribution. Data was then analyzed by an independent t-test or a one-way ANOVA followed by Tukey's post-hoc test. Statistical significance was considered at p -values < 0.05 .

Results and discussion

Chemical analysis of AGO

The yield of AGO from the extraction of the fresh rhizomes of *A. galanga* was 0.10% v/w. The AGO obtained was a clear, pale yellow liquid. Twenty-six compounds representing 91.42% of the total composition obtained from this oil were identified by GC-MS analysis (Table 1). The main constituents of the extract were oxygenated monoterpenes (1,8-cineole, 37.43%) and phenylpropanoids (4-allylphenyl acetate, 25.97%), which is in agreement with previous reports [24, 25]. Slight differences to the reported chemical compositions of AGO [26–28] are caused by geographic, physiological and genetic variations of the plant material [29].

Preparation of SMEDDS-AGO

Pseudoternary phase diagrams were constructed to identify the desired SMEDDS regions and to optimize the ratio of oil (AGO), surfactant, and cosurfactant [30]. Initially, the effect of surfactant without cosurfactant on creating a monophasic area in the phase diagram was investigated. It was found that the areas of monophasic transparent formulations using Tween 20, Tween 80, and Triton X-100 were 32.8 ± 0.3 , 21.2 ± 0.2 , and 27.6 ± 0.2 percent of the total phase diagram, respectively (Fig 1). Thus the results indicate that among the three nonionic surfactants used, Tween 20 created the largest monophasic area in the phase diagrams.

The use of a single surfactant without cosurfactant is usually not sufficient to reduce the interfacial tension between oil and water to form transparent monophasic systems, at least not in a substantial area of the phase diagram [31, 32]. Thus, the addition of a cosurfactant or cosolvent is generally required. To investigate the effect of cosurfactant on supporting the surfactant to create a monophasic area in the phase diagram, three alcohols; ethanol, propylene glycol, and isopropanol, were used in combination with the three different surfactants at a weight ratio of 1:1, 1:2, and 2:1 (termed Smix). It was found that addition of ethanol, which is more hydrophilic than the other two cosurfactants, resulted in the formation of the largest monophasic area in the phase diagrams as shown in Figs 2–4, respectively. After mixing with the cosurfactants, Triton X-100 showed the highest potential to result in a monophasic area in the phase diagrams, followed closely by Tween 80, whereas the use of Tween 20 resulted in the smallest monophasic area. The results also demonstrated that among the three Smix ratios used (2:1, 1:1, and 1:2), the 2:1 mix was the most effective in creating a large monophasic area.

Characterization of SMEDDS-AGO

From the obtained pseudoternary phase diagrams, it was found that SMEDDS-AGO composed of 20% of AGO and 80% of Smix (surfactant:ethanol = 2:1) constituted suitable

Table 1. Chemical composition of AGO.

Retention time (min)	Compound	Amount (%)*
4.08	α -Pinene	4.53
4.39	Camphene	0.19
4.93	Sabinene	0.32
5.03	β -Pinene	3.27
5.33	β -Myrcene	0.79
6.06	α -Terpinene	0.40
6.60	1,8-Cineole	37.43
7.02	1,3,6-Octatriene	0.03
7.35	γ -Terpinene	0.66
8.32	α -Terpinolene	0.33
11.23	Borneol	0.04
11.32	α Terpineol	2.41
11.73	Terpinen-4-ol	3.24
12.65	2-Butenal	0.22
15.23	Chavicol	1.50
18.93	4-Allylphenyl acetate	25.97
19.23	Eugenol	0.14
20.25	Geranyl acetate	0.77
21.43	Methyl eugenol	5.07
22.79	β -Selinene	0.25
23.08	β -Farnesene	0.15
23.90	Germacrene-D	0.24
25.10	β -Bisaboloene	1.10
25.66	β -Sesquiphellandrene	0.78
26.00	Eugenyl acetate	1.46
30.08	δ -Cadinene	0.13
Total		91.42

* Listed amounts are the percentage of each compound calculated based on the total area of all peaks obtained from the oil.

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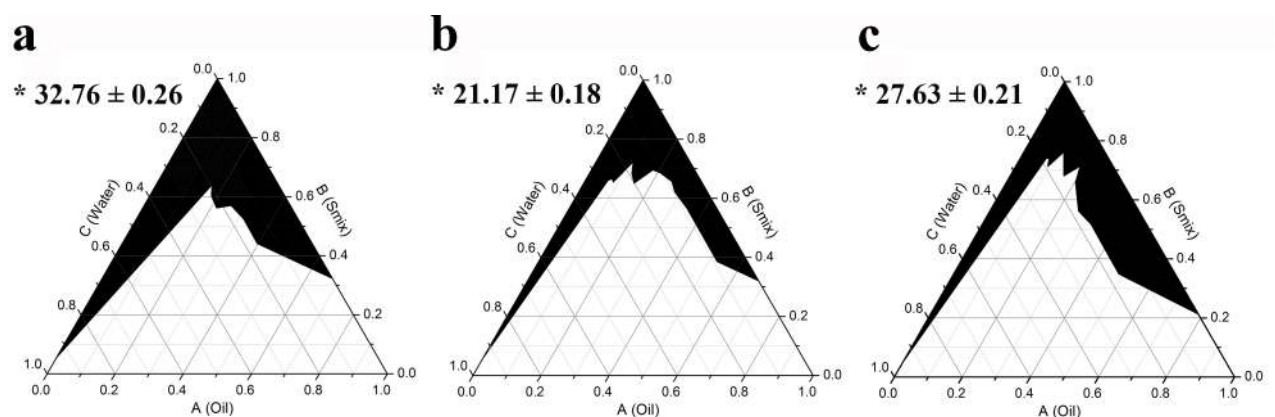


Fig 1. Pseudoternary phase diagrams of AGO containing Tween 20 (a), Tween 80 (b), and Triton X-100 (c). The black area represents the monophasic area. Asterisks (*) indicate the percentage of the monophasic area (means $\pm$ S.E.M.).

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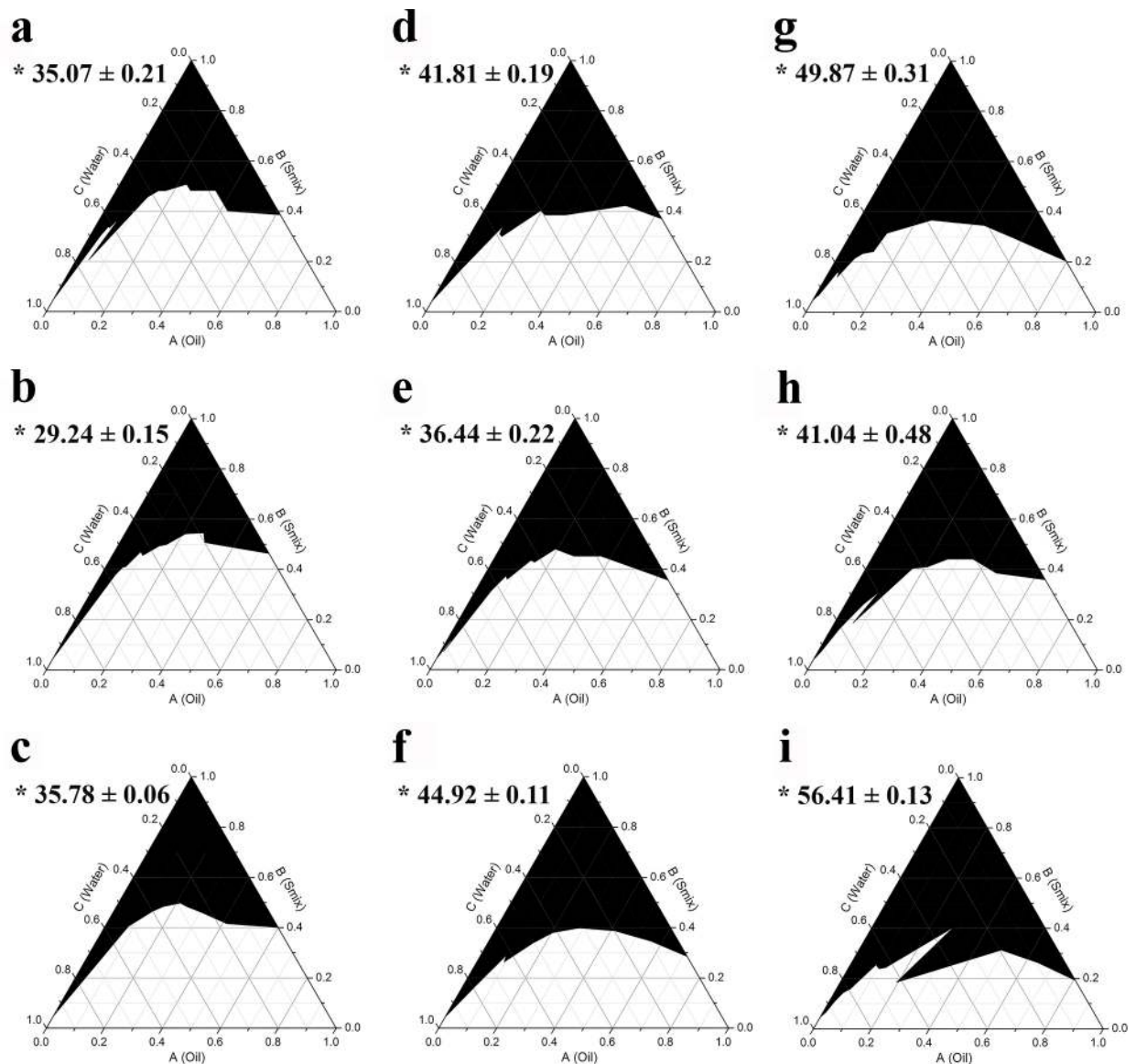


Fig 2. Pseudoternary phase diagrams of AGO using surfactant:ethanol at a weight ratio of 1:1 (a, d, g), 1:2 (b, e, h), and 2:1 (c, f, i) for Tween 20 (a, b, c), Tween 80 (d, e, f), and Triton X-100 (g, h, i). The black area represents the monophasic area. Asterisks (*) indicate the percentage of the monophasic area (means ± S.E.M.).

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formulations for further characterization. Although using Triton X-100 with ethanol at a weight ratio of 2:1 yielded the largest monophasic area in the phase diagrams, Tween 80 was chosen instead of Triton X-100 because it is less irritating and less toxic than Triton X-100 [33]. Thus, SMEDDS-AGO composed of 20% of AGO and 80% of Smix (Tween 80:ethanol = 2:1) was selected for further characterization. The visual appearance of this SMEDDS-AGO formulation was that of a clear, pale yellowish solution. After 100-fold water dilution, it was found that the average droplet size was 82 ± 0.5 nm with a PdI of 0.38 ± 0.02 . The high PdI indicated a fairly broad size distribution in the formulation. Zeta potential, and conductivity of this formulation were -10.40 ± 0.20 mV and 297.7 ± 1.76 μ S/cm, respectively. The zeta potential is one of the factors that influences the physical stability of the dispersed droplets, with high positive or negative values (higher than +30 mV or -30 mV, respectively) usually

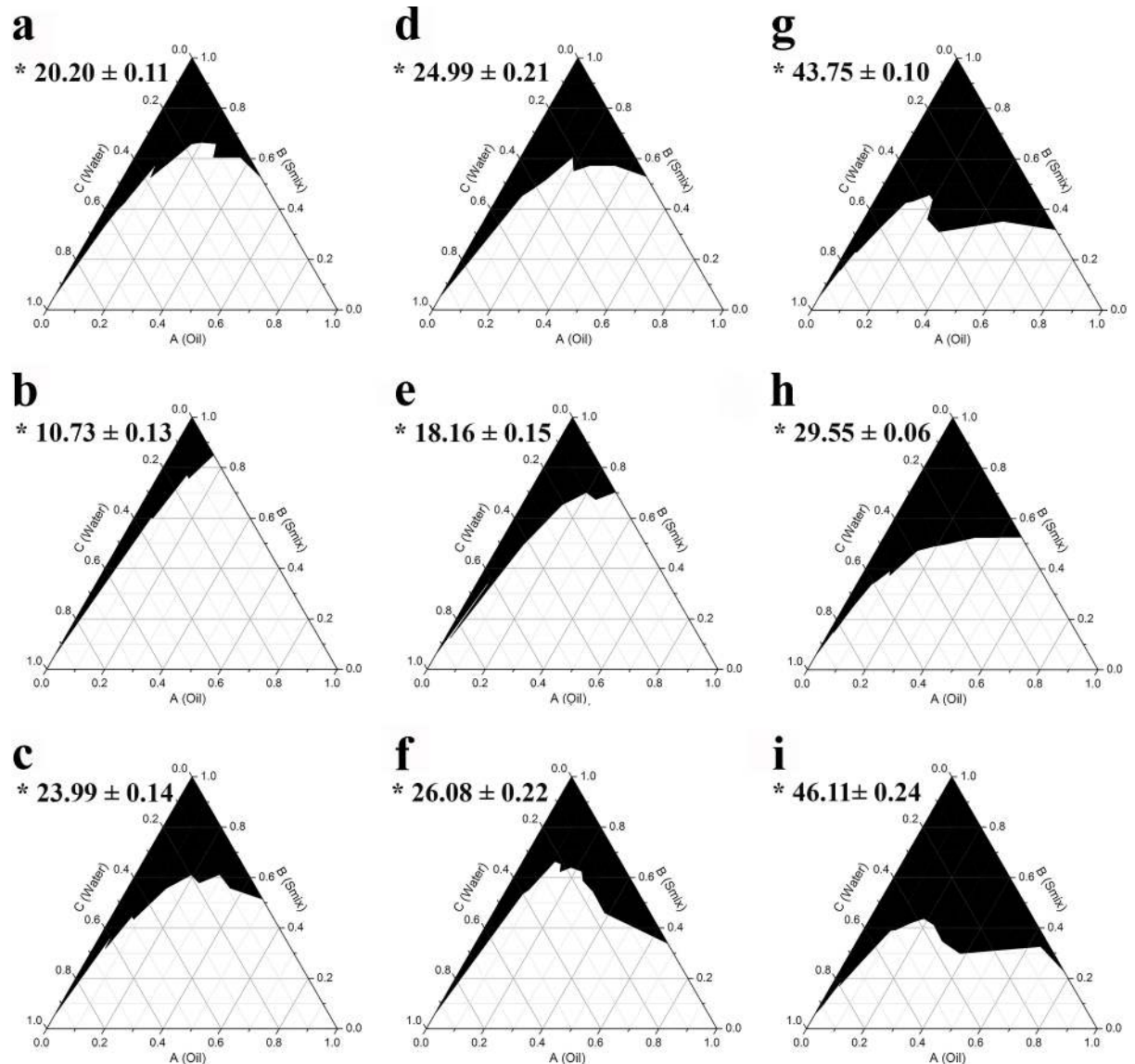


Fig 3. Pseudoternary phase diagrams of AGO using surfactant:propylene glycol at a weight ratio of 1:1 (a, d, g), 1:2 (b, e, h), and 2:1 (c, f, i) for Tween 20 (a, b, c), Tween 80 (d, e, f), and Triton X-100 (g, h, i). The black area represents the monophasic area. Asterisks (*) indicate the percentage of the monophasic area (means ± S.E.M.).

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leading to higher physical stability [34]. In this study, diluted SMEDDS showed zeta potential values of less than -30 mV due to the use of a nonionic surfactant. The physical stability of the dispersed droplets of the diluted SMEDDS was however, still high due to steric stabilization of the nonionic surfactant used [35]. The high conductivity value of this formulation confirmed its expected O/W phase behavior.

Preparation and characterization of NE-AGO

NE formulations are emulsions with a small droplet size in the nanometer range of about 20–200 nm [36]. As can be seen in Table 2, the employed surfactants resulted in different droplet sizes for the various NE-AGO formulations. Generally, an increase in surfactant concentrations led to a decrease in droplet size. After using HPH at a pressure of 10,000 psi for 7 cycles, the largest droplet

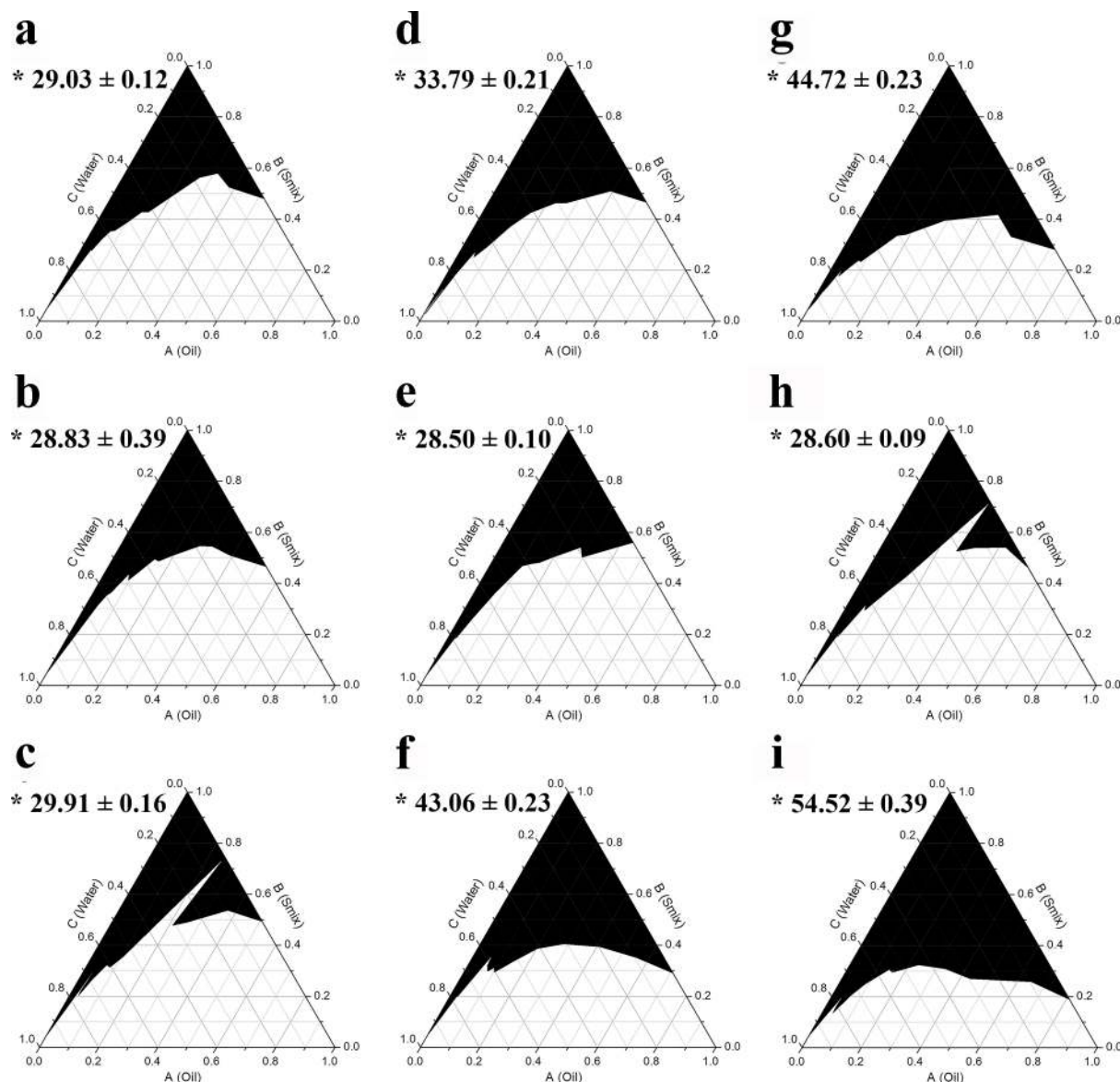


Fig 4. Pseudoternary phase diagrams of AGO using surfactant:isopropanol at a weight ratio of 1:1 (a, d, g), 1:2 (b, e, h), and 2:1 (c, f, i) for Tween 20 (a, b, c), Tween 80 (d, e, f), and Triton X-100 (g, h, i). The black area represents the monophasic area. Asterisks (*) indicate the percentage of the monophasic area (means ± S.E.M.).

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size (130 ± 0.9 nm) was obtained from NE-AGO-E using 5% of Triton X-100 as a surfactant while NE-AGO-F, using 10% Triton X-100, gave the smallest droplet size (33 ± 0.3 nm).

HPH generated intense disruptive forces, resulting in a decreased droplet radius [37]. The droplet size thus decreased by increasing the HPH pressure and the number of HPH cycles (Fig 5) for NE-AGO-D. The largest decrease of the mean droplet size occurred when the pre-emulsions were passed through the HPH (at all pressures) after the first cycle. In addition, the droplet size of the NE-AGO continued to decrease by increasing the number of HPH cycles, until 7–10 cycles, whereafter no further changes in droplet size were measured.

The six formulations of NE-AGO showed a white bluish color or had a translucent, bluish appearances when 10% of Triton X-100 was used as a surfactant. As shown in Table 2, the

Table 2. Characterization of NE-AGO.

NE-AGO	Size (nm)	PdI	Zeta potential (mV)	Conductivity ($\mu\text{S}/\text{cm}$)
NE-AGO-A	63 $\pm$ 1.6	0.38 $\pm$ 0.01	-36.3 $\pm$ 0.52	511.0 $\pm$ 0.33
NE-AGO-B	68 $\pm$ 0.9	0.34 $\pm$ 0.03	-42.70 $\pm$ 0.50	750.7 $\pm$ 0.33
NE-AGO-C	104 $\pm$ 0.5	0.42 $\pm$ 0.00	-35.43 $\pm$ 0.38	571.7 $\pm$ 0.33
NE-AGO-D	48 $\pm$ 1.6	0.43 $\pm$ 0.01	-35.07 $\pm$ 1.82	551.3 $\pm$ 0.33
NE-AGO-E	130 $\pm$ 0.9	0.23 $\pm$ 0.00	-41.83 $\pm$ 0.35	308.0 $\pm$ 1.53
NE-AGO-F	33 $\pm$ 0.3	0.39 $\pm$ 0.01	-31.83 $\pm$ 0.35	499.7 $\pm$ 0.33

Data are presented as means $\pm$ S.E.M.

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smallest droplet size (33 $\pm$ 0.3 nm) was obtained from NE-AGO-F while NE-AGO-E had the largest mean droplet size (130 $\pm$ 0.9 nm). The PdI values of the NE-AGO formulations were approximately 0.4, except for the NE-AGO composed of 5% Triton X-100 where the PdI was about 0.3. Surprisingly, the zeta potential of all NE-AGO formulations were lower than -30 mV. Since non-ionic surfactants were used, the negative zeta potentials were considered to be due to the spontaneous adsorption of common hydroxyl ions in the aqueous system at the o/w interface [38]. The conductivity of all formulations was found to be higher than 100 $\mu\text{S}/\text{cm}$, which, as expected, confirms the O/W nature of the formed NE in the present study.

Stability study

The stability results of the selected SMEDDS-AGO is shown in Table 3. The SMEDDS-AGO showed no phase separation after one month of storage. NE-AGO-A, NE-AGO-B, NE-AGO-E, and NE-AGO-F showed phase separation, except NE-AGO-C and NE-AGO-D which were formulated using Tween 80. These two formulations were further characterized and the results are shown in Table 4. NE-AGO-D which contained 10% Tween 80 showed a mean droplet size smaller than the NE-AGO-C that contained 5% Tween 80 at all storage

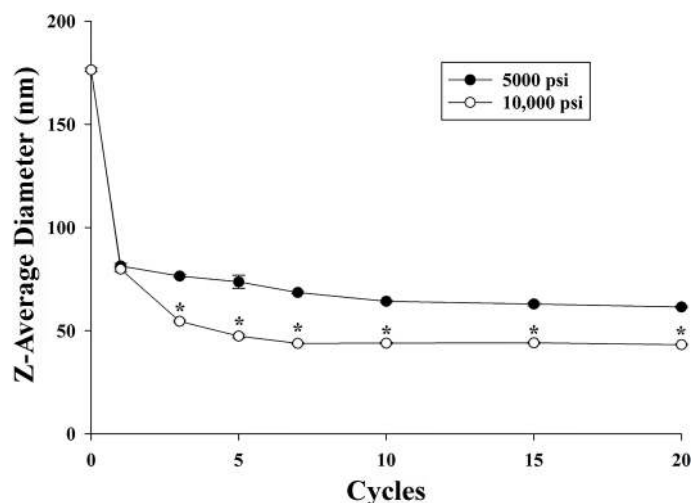


Fig 5. Effect of pressure and number of homogenization cycles on droplet size of the best NE-AGO formulation (NE-AGO-D). Data are presented as means $\pm$ S.E.M. ($p < 0.05$). Asterisks (*) indicate a significant difference between the obtained NE-AGO-D after using a pressure of 5,000 and 10,000 psi ($p < 0.05$). The dataset is available in S1 Dataset.

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Table 3. Characterization of SMEDDS-AGO kept at different temperature for 1 month.

Temperature (°C)	Size (nm)	Pdl	Zeta potential (mV)	Conductivity (μS/cm)
4	142 ± 5.3	0.47 ± 0.08	−12.43 ± 1.01	311.0 ± 0.60
20	92 ± 0.8	0.47 ± 0.02	−8.39 ± 0.51	297.1 ± 2.38
40	106 ± 1.5	0.44 ± 0.03	−9.51 ± 0.44	291.13 ± 1.44

Data are presented as means ± S.E.M.

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temperatures. Stored for one month, the mean droplet size of NE-AGO-D increased from 48 ± 1.6 nm to 78 ± 2.3 nm at 4°C and to 116 ± 1.2 nm at 40°C. However, the mean droplet size of the formulation stored at 20°C changed only slightly from 48 ± 1.6 nm to 66.1 ± 2.4 nm within one month of storage. Due to its small droplet size, NE-AGO-D was selected for further investigation of chemical stability.

For SMEDDS-AGO, during one month of storage at 20°C, the droplet size of the ME formulation obtained from 1:100 v/v aqueous dilutions before size measurement did not change. On the other hand, the ME formulations obtained from the SMEDDS-AGO stored at 4°C and 40°C showed an increase in the mean droplet size compared to freshly prepared samples (82 ± 0.5 nm) to 142 ± 5.3 nm and 106 ± 1.5 nm, respectively. The storage temperature and time had no influence on the zeta potential of both SMEDDS-AGO and NE-AGO.

After storage of AGO, SMEDDS-AGO, and NE-AGO-D at 4°C, 20°C and 40°C for 12 weeks, the content of 1,8-cineole and 4-allylphenyl acetate (the main compounds in AGO) was found to have decreased (Figs 6 and 7) and 58%, 60%, and 46% of the initial levels of 1,8-cineole and 55%, 57%, and 46% of the initial levels of 4-allylphenyl acetate were retained in NE-AGO-D stored at 4°C, 20°C, and 40°C, respectively. The decrease of 1,8-cineole content of NE-AGO-D can be explained by oxidation [39] whereas the decrease in 4-allylphenyl acetate content of NE-AGO-D can be explained by hydrolytic degradation in the presence of water in the NE formulation [39, 40]. Moreover, degradation of 1,8-cineole has been reported when the compound was entrapped in a microencapsulated formulation [41, 42]. In contrast, both compounds; 1,8-cineole and 4-allylphenyl acetate, in AGO and SMEDDS-AGO only slightly decreased by not more than 10% of the initial levels at 4°C and 20°C during 12 weeks of storage as seen in Figs 6 and 7, respectively. Thus the presence of water in the formulation was the main problem for AGO stability and SMEDDS demonstrated to be the most stable of the studied systems.

Table 4. Characterization of NE-AGO kept at different temperature for 1 month.

NE-AGO ^a	Temperature (°C)	Size (nm)	Pdl	Zeta potential (mV)	Conductivity (μS/cm)
NE-AGO-C	4	204 ± 1.7	0.55 ± 0.03	−38.83 ± 0.92	308.3 ± 2.33
	20	144 ± 1.0	0.48 ± 0.02	−21.10 ± 1.14	384.0 ± 1.53
	40	110 ± 1.5	0.46 ± 0.01	−13.30 ± 0.44	484.3 ± 2.19
NE-AGO-D	4	78 ± 2.3	0.50 ± 0.02	−25.50 ± 1.15	321.0 ± 1.15
	20	66 ± 2.4	0.36 ± 0.04	−27.51 ± 0.45	356.0 ± 3.21
	40	116 ± 1.2	0.29 ± 0.01	−22.37 ± 0.82	436.3 ± 2.67

Data are presented as means ± S.E.M.

^a = NE-AGO-A, NE-AGO-B, NE-AGO-E, and NE-AGO-F were not determined due to phase separation.

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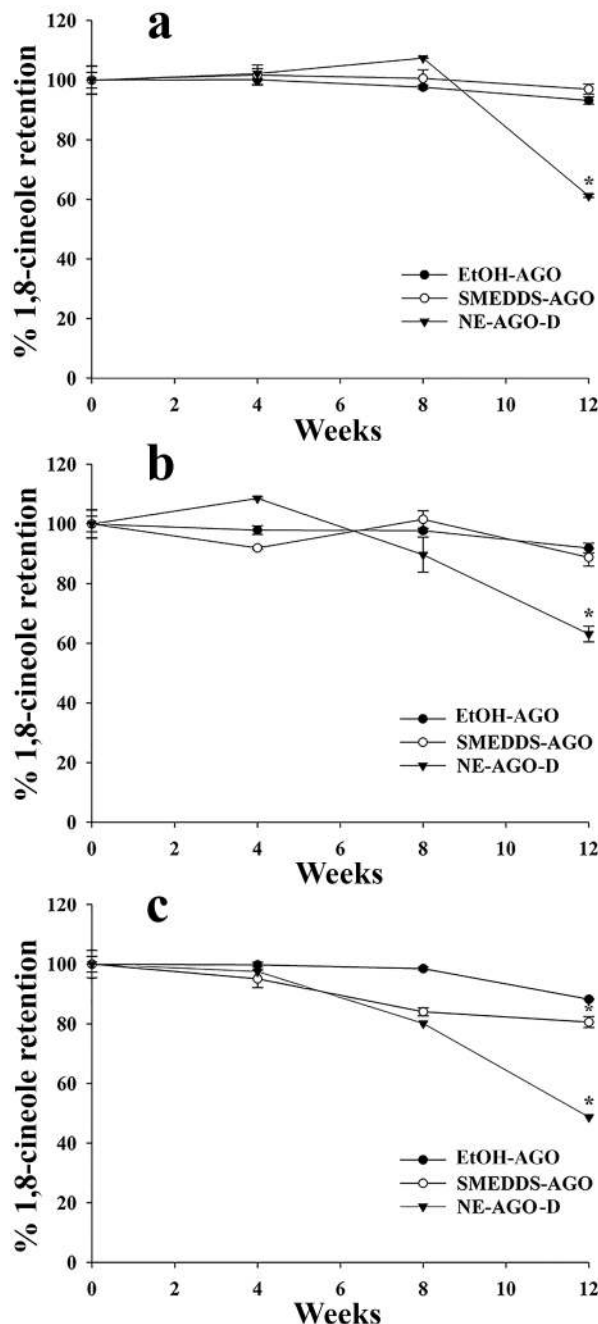


Fig 6. Chemical stability of 1,8-cineole in SMEDDS-AGO, NE-AGO-D, and AGO at 4°C (a), 20°C (b), and 40°C (c). Data are presented as means $\pm$ S.E.M. Asterisks (*) indicate significant difference between AGO and the developed formulations (SMEDDS-AGO and NE-AGO-D) ($p < 0.05$). The dataset is available in [S2 Dataset](#).

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Anesthetic effect

Several reports have shown that SMEDDS and NE are systems that can enhance the biological activity of natural compounds [16, 17, 43–46]. Thus, the anesthetic effect of SMEDDS-AGO and NE-AGO-D on *Cyprinus carpio* in comparison with EtOH-AGO was investigated and the

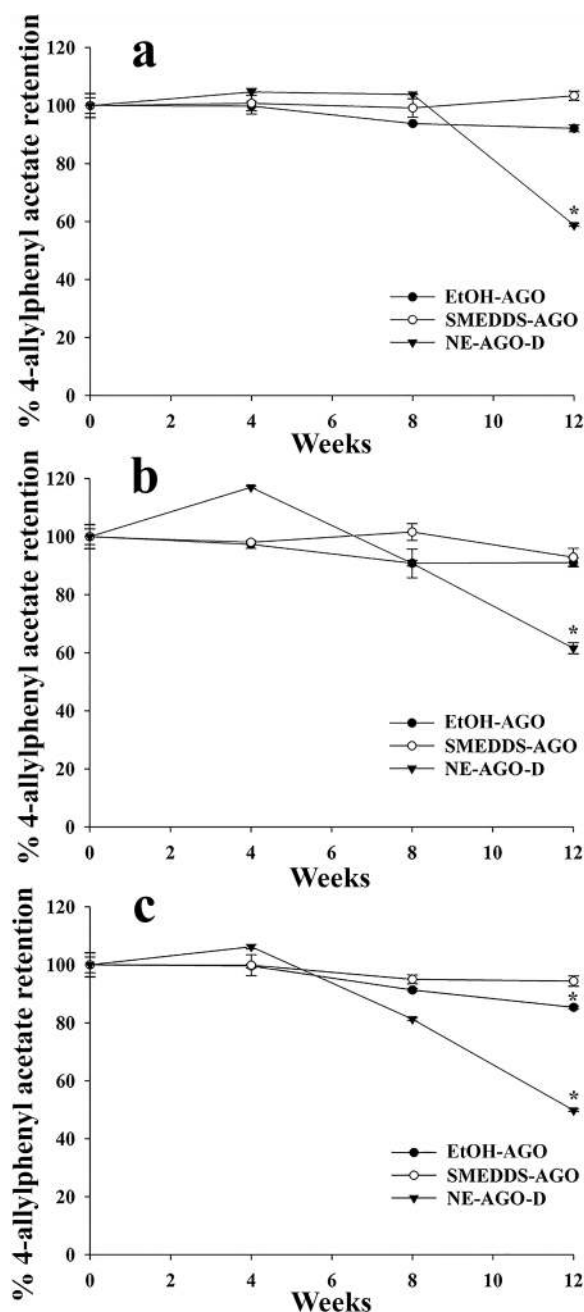


Fig 7. Chemical stability of 4-allylphenyl acetate in SMEDDS-AGO, NE-AGO-D, and AGO at 4°C (a), 20°C (b), and 40°C (c). Data are presented as means $\pm$ S.E.M. Asterisks (*) indicate significant difference between AGO and the developed formulations (SMEDDS-AGO and NE-AGO-D) ($p < 0.05$). The dataset is available in [S3 Dataset](#).

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results are shown in [Fig 8](#). Both of the developed nanoformulations (SMEDDS-AGO and NE-AGO-D) have a higher anesthetic activity than EtOH-AGO as the induction times of SMEDDS-AGO and NE-AGO-D to reach the surgical anesthesia stage were significantly shorter than those of EtOH-AGO ($p < 0.05$) at equal concentrations of AGO (200, 300, and 400 mg/L). However, at a concentration of 100 mg/L, neither EtOH-AGO nor SMEDDS-AGO could induce the fish to stage 3 plane 2 anesthesia which is the surgical stage [23] within an

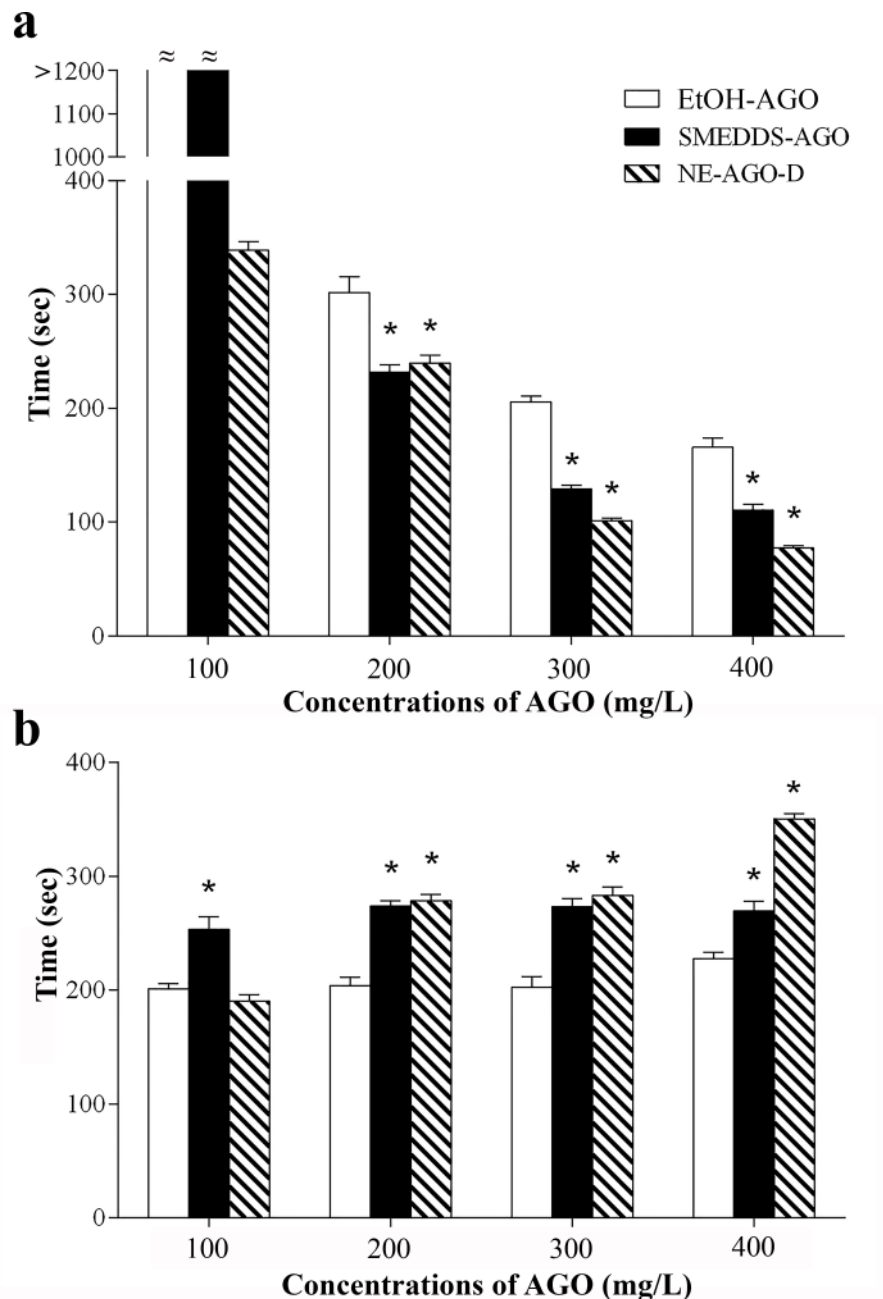


Fig 8. Induction time (A) and recovery time (B) for *Cyprinus carpio* ($n = 20$) exposed to 100, 200, 300, and 400 mg/L of AGO in SMEDDS-AGO and NE-AGO-D in comparison with EtOH-AGO to reach the surgical anesthesia stage. Data are presented as means $\pm$ S.E.M. Asterisks (*) indicate a significant difference between EtOH-AGO and the developed formulations (SMEDDS-AGO and NE-AGO-D) ($p < 0.05$). The dataset is available in [S4 Dataset](#).

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evaluation period of 20 min. Interestingly, at AGO concentration of 100 mg/L using NE-AGO-D, the fish reached the surgical anesthesia stage within approximately 340 sec. At AGO concentrations of 300 and 400 mg/L, NE-AGO-D also showed the highest activity. The fish anesthetized with SMEDDS-AGO or NE-AGO-D at AGO concentrations of 200, 300, and 400 mg/L showed longer recovery times than those anesthetized with EtOH-AGO ($p < 0.05$). The

Table 5. Anesthetic behavioral changes of *Cyprinus carpio* exposed to 100, 200, 300, and 400 mg/L of AGO in SMEDDS-AGO and NE-AGO-D in comparison with EtOH-AGO. The dataset is available in [S5 Dataset](#).

Treatment	Concentration (mg/L)	Anesthetic behavioral changes		
		Hyperactivity	Jumping	Piping
		No. (%)	No. (%)	No. (%)
SMEDDS-AGO	100	0 (0%)	0 (0%)	0 (0%)
	200	0 (0%)	0 (0%)	0 (0%)
	300	0 (0%)	0 (0%)	0 (0%)
	400	0 (0%)	0 (0%)	0 (0%)
NE-AGO-D	100	0 (0%)	0 (0%)	0 (0%)
	200	0 (0%)	0 (0%)	0 (0%)
	300	0 (0%)	0 (0%)	0 (0%)
	400	0 (0%)	0 (0%)	0 (0%)
EtOH-AGO	100	7 (35%)	0 (0%)	0 (0%)
	200	9 (45%)	0 (0%)	0 (0%)
	300	16 (80%)	0 (0%)	0 (0%)
	400	18 (90%)	1 (5%)	3 (15%)

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recovery times of the fish after SMEDDS-AGO and NE-AGO-D exposure were similar for every AGO concentration tested. After monitoring for 2 weeks, no mortality of the fish occurred after exposure to all selected formulations (SMEDDS-AGO, NE-AGO-D, and EtOH-AGO). Interestingly, it was noted that EtOH-AGO caused unwanted hyperactivity in the fish as seen in [Table 5](#). This side effect did not occur in the fish anesthetized with SMEDDS-AGO and NE-AGO formulations. The results of the present study demonstrated that SMEDDS-AGO and NE-AGO-D enhanced water miscibility of AGO and gave higher anesthetic activity with significantly less side effects than EtOH-AGO. The higher anesthetic activity of NE-AGO-D than SMEDDS-AGO was considered to be due to an influence of the droplet size which was significantly smaller for NE-AGO-D than for ME obtained from water dilution of SMEDDS-AGO. No mortality was observed during experimental periods.

Conclusion

We have successfully developed SMEDDS-AGO and NE-AGO formulations for enhancing aqueous miscibility of AGO with the avoidance or limitation of alcohol in the formulations. We demonstrate that surfactant and cosurfactant types as well as their composition ratios play an important role for SMEDDS-AGO and NE-AGO formation and stability. We show that water miscibility of AGO can be significantly enhanced by the developed SMEDDS-AGO and NE-AGO formulations. The most suitable formulation for fish anesthesia of SMEDDS-AGO is composed of 20% AGO and 80% surfactant mixture containing 2:1 ratio of Tween 80:ethanol whereas that of NE-AGO is composed of 20% AGO, 10% Tween 80 and 70% water. These nanoformulations showed extremely higher efficacy and significantly less side effect than EtOH-AGO. We conclude that SMEDDS and NE are the promising delivery systems for AGO to enhance water miscibility, decrease the use of alcohol in the formulations, and enhance biological efficacy of AGO.

Supporting information

S1 Dataset. Droplet size of the best NE-AGO formulation (NE-AGO-D) after passing HPH.
(XLSX)

S2 Dataset. The content of 1,8-cineole in SMEDDS-AGO, NE-AGO-D, and AGO at 4°C, 20°C, and 40°C.
(XLSX)

S3 Dataset. The content of 4-allylphenyl acetate in SMEDDS-AGO, NE-AGO-D, and AGO at 4°C, 20°C, and 40°C.
(XLSX)

S4 Dataset. Induction time and recovery time for *Cyprinus carpio* exposed to 100, 200, 300, and 400 mg/L of AGO in SMEDDS-AGO and NE-AGO-D in comparison with EtOH-AGO to reach the surgical anesthesia stage.
(XLSX)

S5 Dataset. Anesthetic behavioral changes of *Cyprinus carpio* exposed to 100, 200, 300, and 400 mg/L of AGO in SMEDDS-AGO and NE-AGO-D in comparison with EtOH-AGO.
(XLSX)

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References

1. Oonmetta-aree J, Suzuki T, Gasaluck P, Eumkeb G. Antimicrobial properties and action of galangal (*Alpinia galanga* Linn.) on *Staphylococcus aureus*. LWT-Food Sci Technol. 2006; 39: 1214–1220. <https://doi.org/10.1016/j.lwt.2005.06.015>
2. Chudiwal A, Jain D, Somani R. *Alpinia galanga* Willd.—an overview on phyto-pharmacological properties. Indian J Nat Prod Resour. 2010; 1: 143–149.
3. Grzanna R, Lindmark L, Frondoza CG. Ginger—an herbal medicinal product with broad anti-inflammatory actions. J Med Food. 2005; 8: 125–132. <https://doi.org/10.1089/jmf.2005.8.125> PMID: 16117603
4. Prakatthagomol W, Klayraung S, Okonogi S. Bactericidal action of *Alpinia galanga* essential oil on food-borne bacteria. Drug Discov Ther. 2011; 5: 84–89. <https://doi.org/10.5582/ddt.2011.v5.2.84> PMID: 22466145

5. Yang X, Eilerman RG. Pungent principal of *Alpinia galanga* (L.) Swartz and its applications. J Agric Food Chem. 1999; 47: 1657–1662. <https://doi.org/10.1021/jf9808224> PMID: 10564034
6. Pikulkaew S, Khumpirapang N, Chaisri W, Okonogi S. Effects of *Alpinia galanga* oil on anesthesia and stress reduction in *Oreochromis niloticus*. Drug Discov Ther. 2017; 11: 186–192. <https://doi.org/10.5582/ddt.2017.01035> PMID: 28867750
7. Bernstein PS, Digre KB, Creel DJ. Retinal toxicity associated with occupational exposure to the fish anesthetic MS-222. Am J Ophthalmol. 1997; 124: 843–844. [https://doi.org/10.1016/S0002-9394\(14\)71705-2](https://doi.org/10.1016/S0002-9394(14)71705-2) PMID: 9402834
8. Carvan MJ, Loucks E, Weber DN, Williams FE. Ethanol effects on the developing zebrafish: neurobehavior and skeletal morphogenesis. Neurotoxicol Teratol. 2004; 26: 757–768. <https://doi.org/10.1016/j.ntt.2004.06.016> PMID: 15451040
9. Krone P, Sass J, Lele Z. Heat shock protein gene expression during embryonic development of the zebrafish. Cell Mol Life Sci. 1997; 53: 122–129. <https://doi.org/10.1007/PL00000574> PMID: 9117992
10. Mathur P, Berberoglu MA, Guo S. Preference for ethanol in zebrafish following a single exposure. Behav Brain Res. 2011; 217: 128–133. <https://doi.org/10.1016/j.bbr.2010.10.015> PMID: 20974186
11. Chaudhary A, Nagaich U, Gulati N, Sharma V, Khosa R, Partapur MU. Enhancement of solubilization and bioavailability of poorly soluble drugs by physical and chemical modifications: a recent review. J Adv Pharm Educ Res. 2012; 2: 32–67.
12. Holm R, Porter CJH, Edwards GA, Müllertz A, Kristensen HG, Charman WN. Examination of oral absorption and lymphatic transport of halofantrine in a triple-cannulated canine model after administration in self-microemulsifying drug delivery systems (SMEDDS) containing structured triglycerides. Eur J Pharm Sci. 2003; 20: 91–97. [https://doi.org/10.1016/S0928-0987\(03\)00174-X](https://doi.org/10.1016/S0928-0987(03)00174-X) PMID: 13678797
13. Kawabata Y, Wada K, Nakatani M, Yamada S, Onoue S. Formulation design for poorly water-soluble drugs based on biopharmaceutics classification system: basic approaches and practical applications. Int J Pharm. 2011; 420: 1–10. <https://doi.org/10.1016/j.ijpharm.2011.08.032> PMID: 21884771
14. Zhao Y, Wang C, Chow AH, Ren K, Gong T, Zhang Z, et al. Self-nanoemulsifying drug delivery system (SNEDDS) for oral delivery of Zedoary essential oil: formulation and bioavailability studies. Int J Pharm. 2010; 383: 170–177. <https://doi.org/10.1016/j.ijpharm.2009.08.035> PMID: 19732813
15. Bernardi DS, Pereira TA, Maciel NR, Bortoloto J, Viera GS, Oliveira GC, et al. Formation and stability of oil-in-water nanoemulsions containing rice bran oil: in vitro and in vivo assessments. J Nanobiotechnol. 2011; 9: 1–9. <https://doi.org/10.1186/1477-3155-9-44> PMID: 21952107
16. Donsì F, Annunziata M, Sessa M, Ferrari G. Nanoencapsulation of essential oils to enhance their antimicrobial activity in foods. LWT-Food Sci Technol. 2011; 44: 1908–1914. <https://doi.org/10.1016/j.lwt.2011.03.003>
17. Eid AM, El-Enshasy HA, Aziz R, Elmarzugi NA. The preparation and evaluation of self-nanoemulsifying systems containing *Swietenia* oil and an examination of its anti-inflammatory effects. Int J Nanomedicine. 2014; 9: 4685–4695. <https://doi.org/10.2147/IJN.S66180> PMID: 25336948
18. Wang X, Jiang Y, Wang YW, Huang MT, Ho CT, Huang Q. Enhancing anti-inflammation activity of curcumin through O/W nanoemulsions. Food Chem. 2008; 108: 419–424. <https://doi.org/10.1016/j.foodchem.2007.10.086> PMID: 26059118
19. Okonogi S, Chaiyana W. Enhancement of anti-cholinesterase activity of *Zingiber cassumunar* essential oil using a microemulsion technique. Drug Discov Ther. 2012; 6: 249–255. <https://doi.org/10.5582/ddt.2012.v6.5.249> PMID: 23229145
20. Oprean R, Oprean L, Tamas M, Sandulescu R, Roman L. Essential oils analysis. II. Mass spectra identification of terpene and phenylpropane derivatives. J Pharm Biomed Anal. 2001; 24: 1163–1168. [https://doi.org/10.1016/S0731-7085\(00\)00578-1](https://doi.org/10.1016/S0731-7085(00)00578-1) PMID: 11248515
21. Raina V, Srivastava S, Syamasunder K. Essential oil composition of *Acorus calamus* L. from the lower region of the Himalayas. Flavour Fragr J. 2003; 18: 18–20. <https://doi.org/10.1002/ffj.1136>
22. Gardeli C, Vassiliki P, Athanasios M, Kibouris T, Komaitis M. Essential oil composition of *Pistacia lentiscus* L. and *Myrtus communis* L.: evaluation of antioxidant capacity of methanolic extracts. Food Chem. 2008; 107: 1120–1130. <https://doi.org/10.1016/j.foodchem.2007.09.036>
23. Zahl IH, Samuelsen O, Kiessling A. Anaesthesia of farmed fish: implications for welfare. Fish Physiol Biochem. 2012; 38: 201–218. <https://doi.org/10.1007/s10695-011-9565-1> PMID: 22160749
24. Jantan IB, Ahmad FB, Ahmad AS. Constituents of the rhizome and seed oils of Greater Galangal *Alpinia galanga* (L.) Willd. from Malaysia. J Essent Oil Res. 2004; 16: 174–176. <https://doi.org/10.1080/10412905.2004.9698687>
25. Raina V, Srivastava S, Syamasunder K. The essential oil of 'greater galangal' [*Alpinia galanga* (L.) Willd.] from the lower Himalayan region of India. Flavour Fragr J. 2002; 17: 358–360. <https://doi.org/10.1002/ffj.1105>

26. Charles DJ, Simon JE, Singh NK. The essential oil of *Alpinia galanga* Willd. J Essent Oil Res. 1992; 4: 81–82. <https://doi.org/10.1080/10412905.1992.9698016>
27. Mahae N, Chaiseri S. Antioxidant activities and antioxidative components in extracts of *Alpinia galanga* (L.) SW. Kasetsart J Nat Sci. 2009; 43: 358–369.
28. Srivastava B, Singh P, Shukla R, Dubey NK. A novel combination of the essential oils of *Cinnamomum camphora* and *Alpinia galanga* in checking aflatoxin B1 production by a toxigenic strain of *Aspergillus flavus*. World J Microbiol Biotechnol. 2008; 24: 693–697. <https://doi.org/10.1007/s11274-007-9526-0>
29. Chalchat JC, Özcan MM. Comparative essential oil composition of flowers, leaves and stems of basil (*Ocimum basilicum* L.) used as herb. Food Chem. 2008; 110: 501–503. <https://doi.org/10.1016/j.foodchem.2008.02.018> PMID: 26049245
30. Wang Y, Sun J, Zhang T, Liu H, He F, He Z. Enhanced oral bioavailability of tacrolimus in rats by self-microemulsifying drug delivery systems. Drug Dev Ind Pharm. 2011; 37: 1225–1230. <https://doi.org/10.3109/03639045.2011.565774> PMID: 21615281
31. Tenjarla S. Microemulsions: an overview and pharmaceutical applications. Crit Rev Ther Drug Carrier Syst. 1999; 16: 461–521. <https://doi.org/10.1615/CritRevTherDrugCarrierSyst.v16.i5.20> PMID: 10635455
32. Ho HO, Hsiao CC, Sheu MT. Preparation of microemulsions using polyglycerol fatty acid esters as surfactant for the delivery of protein drugs. J Pharm Sci. 1996; 85: 138–143. <https://doi.org/10.1021/js950352h> PMID: 8683437
33. Zhang H, Yao M, Morrison RA, Chong S. Commonly used surfactant, Tween 80, improves absorption of P-glycoprotein substrate, digoxin, in rats. Arch Pharm Res. 2003; 26: 768–772. <https://doi.org/10.1007/BF02976689> PMID: 14560928
34. Saupe A, Gordon KC, Rades T. Structural investigations on nanoemulsions, solid lipid nanoparticles and nanostructured lipid carriers by cryo-field emission scanning electron microscopy and Raman spectroscopy. Int J Pharm. 2006; 314: 56–62. <https://doi.org/10.1016/j.ijpharm.2006.01.022> PMID: 16574354
35. Capek I. Degradation of kinetically-stable o/w emulsions. Adv Colloid Interface Sci. 2004; 107: 125–155. [https://doi.org/10.1016/S0001-8686\(03\)00115-5](https://doi.org/10.1016/S0001-8686(03)00115-5) PMID: 15026289
36. Solans C, Solé I. Nano-emulsions: formation by low-energy methods. Curr Opin Colloid Interface Sci. 2012; 17: 246–254. <https://doi.org/10.1016/j.cocis.2012.07.003>
37. McClements DJ. Edible nanoemulsions: fabrication, properties, and functional performance. Soft Matter. 2011; 7: 2297–2316. <https://doi.org/10.1039/C0SM00549E>
38. Marinova K, Alargova R, Denkov N, Velez O, Petsev D, Ivanov I, et al. Charging of oil-water interfaces due to spontaneous adsorption of hydroxyl ions. Langmuir. 1996; 12: 2045–2051. <https://doi.org/10.1021/la950928i>
39. Turek C, Stintzing FC. Stability of essential oils: a review. Compr Rev Food Sci Food Saf. 2013; 12: 40–53. <https://doi.org/10.1111/1541-4337.12006>
40. Cordes E, Bull H. Mechanism and catalysis for hydrolysis of acetals, ketals, and ortho esters. Chem Rev. 1974; 74: 581–603. <https://doi.org/10.1021/cr60291a004>
41. Krishnan S, Bhosale R, Singhal RS. Microencapsulation of cardamom oleoresin: evaluation of blends of gum arabic, maltodextrin and a modified starch as wall materials. Carbohydr Polym. 2005; 61: 95–102. <https://doi.org/10.1016/j.carbpol.2005.02.020>
42. Mehyar GF, Al-Isamili KM, Al-Ghizzawi HM, Holley RA. Stability of cardamom (*Elettaria Cardamomum*) essential oil in microcapsules made of whey protein isolate, guar gum, and carrageenan. J Food Sci. 2014; 79: 1939–1949. <https://doi.org/10.1111/1750-3841.12652> PMID: 25224989
43. Aboalnaja KO, Yaghoor S, Kumosani TA, McClements DJ. Utilization of nanoemulsions to enhance bioactivity of pharmaceuticals, supplements, and nutraceuticals: nanoemulsion delivery systems and nanoemulsion excipient systems. Expert Opin Drug Deliv. 2016; 21: 1–10. <https://doi.org/10.1517/17425247.2016.1162154> PMID: 26984045
44. Chaiyana W, Saeio K, Hennink WE, Okonogi S. Characterization of potent anticholinesterase plant oil based microemulsion. Int J Pharm. 2010; 401: 32–40. <https://doi.org/10.1016/j.ijpharm.2010.09.005> PMID: 20837121
45. Saeio K, Yotsawimonwat S, Anuchapreeda S, Okonogi S. Development of microemulsion of a potent anti-tyrosinase essential oil of an edible plant. Drug Discov Ther. 2011; 5: 246–252. <https://doi.org/10.5582/ddt.2011.v5.5.246> PMID: 22466371
46. Sakulku U, Nuchuchua O, Uawongyart N, Puttipipatkachorn S, Sootittantawat A, Ruktanonchai U. Characterization and mosquito repellent activity of citronella oil nanoemulsion. Int J Pharm. 2009; 372: 105–111. <https://doi.org/10.1016/j.ijpharm.2008.12.029> PMID: 19162149