



The effect of elevated CO₂ on hyperspectral leaf reflectance in mature trees

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Abstract

Key message Experimentally elevated CO₂ does not significantly alter the overall leaf reflectance of mature *Quercus robur* L., but increases Plant Senescence Reflectance Index (PSRI) suggesting a change in the ratio of chlorophyll to carotene content.

Rising atmospheric CO₂ concentrations, driven by anthropogenic emissions, are projected to reach 550 ppm by 2050. Elevated CO₂ (eCO₂) is expected to have a fertilisation effect on forests, influencing productivity, water relations, and phenology. However, the impact of eCO₂ on leaf reflectance in mature forests remains poorly understood, despite its critical role in radiative transfer processes and remote sensing of forest health. Utilising the Birmingham Institute of Forest Research (BIFoR) Free-Air CO₂ Enrichment (FACE) experiment, we investigated the hyperspectral leaf reflectance of 180-year-old *Quercus robur* L. trees exposed to eCO₂ for 7 years. Our results demonstrate that overall leaf reflectance under eCO₂ is similar to that of leaves exposed to ambient CO₂, but the Plant Senescence Reflectance Index (PSRI) is significantly higher under eCO₂. This index relates to the ratio of foliar chlorophyll and carotene pigments. These findings suggest that *Q. robur* reflectance will not significantly shift under future CO₂ conditions, but the relative content of pigments will change, altering the reflectance of specific wavelengths and providing insights into the leaf level physiological and phenological responses of mature trees to eCO₂.

Keywords Climate change · Forests · Leaf reflectance · Spectroscopy · Vegetation indices · Forest health

Introduction

Since 1850, net anthropogenic CO₂ emissions have reached 2400 (±240) Gt, with concentrations in 2019 surpassing levels observed over the past two million years (Calvin et al. 2023). Under the ‘business as usual’ RCP 8.5 scenario, atmospheric CO₂ concentrations are predicted to reach 550 ppm by 2050 (IPCC 2023). Forests are a major component of the terrestrial carbon sink, absorbing 2.4 (±0.4) petagrams of carbon per year (Pg C year⁻¹) globally (Pan et al. 2024). CO₂ is the carbon input for photosynthesis in plants and elevated CO₂ (eCO₂) has a stimulating effect on photosynthesis, often referred to as the ‘CO₂ fertilisation’ effect (Ciais et al. 2013). CO₂ fertilisation of forests, leading to enhanced productivity, has been observed in recent decades (Zhu et al. 2016; Ruehr et al. 2023). However, the long-term effect of eCO₂ on mature forests and their role as a carbon sink remains uncertain.

The fertilisation response of tree seedlings and saplings to eCO₂ has been extensively experimentally studied

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using Free-Air CO₂ Enrichment (FACE) experiments (as reviewed by Ellsworth et al. 2004; Ainsworth and Long 2005; Maschler et al. 2022). The first generation of FACE experiments provided valuable long-term observations of the effect of eCO₂ on young trees, such as sustained elevated photosynthetic rate (Ainsworth and Long 2005) and increased fine-root production and throughput of root biomass into the soil (Norby and Zak 2011). However, these experiments were conducted in young tree plantations, limiting their application to understanding how mature forests will respond to eCO₂ (Norby et al. 2005; U.S. DOE 2020). Notably, results from the Aspen FACE project revealed that responses observed in seedlings and young trees were not always sustained as the stand matured, emphasising the effect of tree age on eCO₂ responses (Burton et al. 2014). Wood analysis of *Quercus ilex* exposed to a lifetime of eCO₂ (at a natural CO₂ spring) revealed that most growth enhancement occurs when the tree is young (Hättenschwiler et al. 1997), leading to ongoing questions about the growth responses of mature forests to eCO₂.

The second generation of FACE experiments seeks to explore the capacity of mature trees to response to eCO₂ across different biomes (Norby et al. 2016). This study is based at BIFoR FACE, situated in a 180-year-old *Quercus robur* (*Q. robur*) dominated woodland in central England, which has shown increased photosynthetic activity and woody biomass production in the first 7 years of eCO₂ exposure (Gardner et al. 2022a; Norby et al. 2024). These results contrast with the Eucalyptus FACE experiment (EucFACE) based in Australia, where increased carbon uptake did not translate into greater carbon sequestration, likely due to phosphorous limitation (Jiang et al. 2020). As well as the eCO₂ crane experiment in Basel, where after 8 years of eCO₂ treatment, radial stem diameter was not affected in *Quercus petraea* or the other deciduous hardwoods tested (Bader et al. 2013).

In addition to changes in photosynthetic rate and tree growth, eCO₂ has also been shown to affect water relations in trees. In a synthesis of 13 long-term experiments (> 1 year), Medlyn et al. found European forest trees exposed eCO₂ had 21% lower stomatal conductance (Medlyn et al. 2001), although a recent meta-analysis has shown that the eCO₂ induced increase in water use efficiency (WUE) was primarily due to increased photosynthesis rather than decreased stomatal conductance (Gardner et al. 2023). eCO₂ has been shown to increase WUE in mature deciduous forests (Leuzinger and Körner 2007) and *Quercus petraea* saplings (Ofori-Amanfo et al. 2020). After 8 years of eCO₂ treatment, water use was significantly reduced in the Basel deciduous forest experiment (Bader et al. 2013), but at BIFoR FACE although tree's water use was lower on average under eCO₂ this change was not significant (Quick et al. 2025).

At the leaf level of trees, eCO₂ has been shown to decrease chlorophyll content. Onisch et al. reported decreased chlorophyll in young *Picea* and *Fagus* saplings exposed to eCO₂ (Onisch et al. 2016); and Tausz et al. found the needles of young *Picea abies* trees had lower chlorophyll when exposed to eCO₂ (Tausz et al. 1996). In mature, forest-grown trees at the EucFACE experiment mature *Eucalyptus tereticornis* leaves had reduced pigment concentrations (chlorophyll and carotenes) per area (Wujeska-Klaue et al. 2019). In mature *Liquidambar styraciflua* (sweetgum) leaves from the Oak Ridge FACE experiment, after 12 years of eCO₂ exposure leaves had lower chlorophyll (Warren et al. 2015). In the BIFoR FACE experiment, eCO₂ did not affect mass-based chlorophyll concentrations in mature *Q. robur* leaves, but area-based chlorophyll content significantly increased, which was attributed to increased leaf mass per unit area (Gardner et al. 2022b). Similarly, mature *Quercus ilex* and *Quercus pubescens* trees exposed to eCO₂ by a natural spring did not show a significant change in the chlorophyll content of leaves (Schwanz and Polle 1998a).

ECO₂ has been reported to also decrease the nitrogen content of leaves per unit area (Ellsworth et al. 2004; Ainsworth and Long 2005). In mature *Liquidambar styraciflua* trees, foliar nitrogen decreased 17% under long-term eCO₂ (Warren et al. 2015). Reduced foliar nitrogen was also found in *Populus tremuloides*, *Betula papyrifera*, and *Acer saccharum* saplings (Agrell et al. 2000). It has been indicated that reduced foliar allocation of nitrogen to chlorophyll under eCO₂ follows total nitrogen content (Warren et al. 2015; Wujeska-Klaue et al. 2019). ECO₂ has resulted in reduced foliar phenol content in *Quercus* species (Tognetti and Johnson 1999; Watanabe et al. 2021), although some studies have found this effect is only temporary (Dury et al. 1998). In the BIFoR FACE experiment, no significant effect of eCO₂ on the foliar nitrogen content of *Q. robur* foliar has been found (Gardner et al. 2022b).

Studies have also examined the effect of eCO₂ on leaf nutrient content, from the perspective of food sources for insects. In a range of deciduous species, saplings grown under eCO₂ have reduced foliar nutritional quality (Dury et al. 1998). Carbohydrate concentrations, particularly starch, increase in young trees exposed to eCO₂ (Kinney et al. 1997; Agrell et al. 2000; Coley et al. 2002). Combined with reduced nitrogen content, this results in increased foliar carbon to nitrogen (C:N) ratios. For example, at the BIFoR FACE experiment, the foliar carbon content of fresh leaves was found to be significantly higher under eCO₂ (Roberts et al. 2022).

ECO₂ also decreases the antioxidant content of tree leaves. For instance, in mature Mediterranean *Quercus* trees, eCO₂ reduced antioxidant enzyme activity and increased the ascorbate pool (Schwanz and Polle 1998b; Marabottini et al. 2001). In *Q. robur* seedlings, superoxide dismutase content

decreased under eCO₂ (Schwanz et al. 1996) but this has not been investigated in mature *Q. robur* trees. In summary, eCO₂ has been shown to lead to a variety of effects on the phytochemistry of deciduous trees, and on oak trees in particular although there is more limited evidence for the effects on mature trees in forest settings.

Remote sensing of forest health, productivity, and biodiversity relies on the reflectance spectra of leaves (Lausch et al. 2016; Kacic and Kuenzer 2022). Across visible to infrared wavelengths, leaf reflectance is influenced by factors such as pigments, leaf structural composition, and water content (Grant 1987; Baldini et al. 1997; Sims and Gamon 2002). Despite its importance, few studies have examined the effect of eCO₂ on the leaf reflectance of mature trees, which could provide important insight into leaf-level changes underpinning forest fertilisation or acclimation. Hyperspectral reflectance data from forests are widely used to remotely sense leaf traits (Chen et al. 2022), classify species (Puttonen et al. 2010), and monitor forest health (Sonobe and Wang 2017; Torres et al. 2021) under natural conditions. Understanding how leaf reflectance will change under future CO₂ concentrations is, therefore, crucial for accurately monitoring forest health and productivity in a changing climate.

Leaf-level hyperspectral reflectance studies on trees exposed to eCO₂ are limited and have found variable results. Thomas (2005) found an increase in the reflectance of photosynthetically active radiation (PAR; 400–700 nm) and a decrease in chlorophyll in a glasshouse eCO₂ study on tropical *Leguminosae* saplings (Thomas 2005). In contrast, Carter et al. (2000) found no significant effect of eCO₂ on the reflectance of young *Acer saccharum* within a 400–850 nm range in open-trop chambers (Carter et al. 2000). In mature *Quercus pubescens* trees exposed to lifetime eCO₂ via natural springs, normalised difference vegetation index (NDVI) was not affected and nor was Photochemical Reflectance Index (PRI) despite photosynthetic enhancement (Stylinski et al. 2000). Similarly, the Oak Ridge FACE experiment reported no significant changes in reflectance at the leaf or canopy level for a *Liquidambar styraciflua* monoculture under eCO₂ (Wicklein et al. 2012). In the EucFACE experiment, harvested leaf reflectance in the PAR spectrum (400–700 nm) was not significantly affected by eCO₂ treatment (Wujeska-Klaue et al. 2019). However, Pintó-Marijuan et al. successfully used near-infrared (NIR; 1100–2500 nm) reflectance spectroscopy to examine pigment and antioxidant responses of *Quercus ilex* resprouts under eCO₂ (Pintó-Marijuan et al. 2013). Our study aims to expand on these findings by examining a broad continuous range of hyperspectral reflectance (350–2500 nm) of leaves measured in situ in a mature *Q. robur* forest under eCO₂.

The aim of our study is to investigate whether long-term eCO₂ changes the leaf reflectance of mature *Q. robur* trees

in terms of total reflectance, spectral profile, and vegetation indices. The majority of the relevant literature indicates eCO₂ does not significantly affect the leaf reflectance of trees exposed over long time periods, particularly in mature trees. We, therefore, hypothesise that eCO₂ will not significantly affect the leaf reflectance profile or total reflectance but may affect specific regions linked to physiological or structural changes. By examining the entire wavelength range reflectance profile as well as traditional vegetation indices, we expect to find more nuanced changes in leaf reflectance.

Methods

Study site

The Birmingham Institute of Forest Research (BIFoR) has operated a Free-Air Carbon Enrichment (FACE) experiment since 2015 at Mill Haft, Staffordshire, UK (52°48'3.6" N, 2°18'0" W). Mill Haft is 19.1 ha of deciduous woodland dominated by *Q. robur* (pedunculate oak) in the canopy and *Corylus avellana* (common hazel) in the understory, situated in a temperate maritime climate (Hart et al. 2020; MacKenzie et al. 2021). The underlying geology is Helsby Sandstone, and the dominant soil type is Dystric Cambisol, with a sandy clay texture (Norby et al. 2024). The long-term average annual temperature of the site is 9 °C, and the average annual rainfall is 690 mm (Norby et al. 2016). Since the CO₂ fertilisation experiment began in 2017, mean hourly rainfall has been recorded by four rain gauges on the BIFoR FACE meteorological towers (TR-525M, Texas Electronics, Dallas, Texas), alongside daily mean air temperature (HMP155RH, Vaisala, Helsinki, Finland). During the measurement period of this study, the mean daily rainfall was 1.93 mm, and the mean daily temperature was 15.72 °C.

The BIFoR FACE facility is comprised three experimental treatments across nine experimental arrays, each 30 m in diameter. The treatments are three “elevated CO₂” arrays (eCO₂) maintained at 150 ± 38 ppm above ambient CO₂, three control arrays exposed to ambient CO₂ (aCO₂), and three undisturbed woodland areas without fumigation infrastructure. In the present study, only four of the infrastructure arrays were studied due to logistical constraints related to tree canopy access. The fumigation infrastructure arrays have operated from budburst to leaf fall (early April to November) during daylight hours since April 2017, for full details of the experimental design, see Hart et al. (Hart et al. 2020).

Leaf reflectance sampling

We recorded the reflectance spectra of adaxial leaf surface of *Q. robur* in the sun exposed canopy of two aCO₂ and two eCO₂ arrays from July to October 2023. Leaf reflectance measurements were taken on 25th July 2023, 10th August 2023, 24th August 2023, and 6th October 2023. On each measurement day, approximately 30 leaf spectra were taken per tree from two trees per array (60 spectra per array). In total, 932 leaf spectra were recorded, with 469 from aCO₂ arrays and 463 from eCO₂ arrays.

Fully expanded leaves were selected that were in the light exposed crown, avoiding leaves with significant herbivory. From the leaves accessible which met these criteria, every third leaf was sampled until 30 spectra had been taken per tree.

Reflectance spectra were recorded using a high-performance single-beam field spectroradiometer over the range 350–2500 nm using a LC-RP Pro leaf clip with active light source (HR-1024i spectrometer, Spectra Vista Corp, USA). The HR-1024i spectrometer combines three dispersion grating spectrometers which overlap in their wavelength ranges to produce continuous spectra from 350 to 2500 nm. The Very Near Infrared (VNIR) spectrometer measures in the range of 350–1000 nm with a 1.5 nm sampling interval. The first SWIR spectrometer measures in the range of 1000–1890 nm with a 3.8 nm sampling interval. The second SWIR spectrometer measures in the range of 1890–2500 nm with a 2.5 nm sampling interval. The spectrometer was calibrated before use by the NERC Field Spectroscopy Facility. The leaf clip attachment is coupled to the spectroradiometer via a 25° armoured fibre optic. The leaf clip forms a seal around the leaf to exclude external light and is illuminated by an internal lamp. The leaf reflectance is integrated within the field of view, approximately 2 cm diameter. An integrating sphere was not used, this allowed time efficient sampling in the canopy (six times faster) but may lead to overestimation of total reflectance compared to measurements taken with single or double integrating spheres (Hovi et al. 2018). An integration time of 2 s was used for all measurements.

During leaf reflectance measurements, the spectrometer was referenced using the leaf clip's incorporated reflective standard in the leaf clip every 5 min. The HR-1024i applies automatic dark signal baseline correction to every measurement by taking a dark spectrum before each reflectance measurement.

The leaf clip was attached to the adaxial surface in the centre of each leaf, to the left of the midvein, making sure to avoid smaller veins.

Hyperspectral processing

High-resolution spectroscopy represents a computational and analytical challenge due to large datasets and autocorrelation between wavelength variables; consequently, machine learning techniques are increasingly employed to identify key features in spectral datasets (Meza Ramirez et al. 2021). We used supervised and unsupervised machine learning analyses across the whole spectra to identify principal components of variation and regions of the spectra that change under eCO₂, alongside conventional vegetation index analysis.

All leaf reflectance spectra were imported and processed using the SpecDAL package (Lee 2017) in Python 3 (Van Rossum and Drake 2009). The overlapping regions of the spectra produced by the three spectrometers comprising the HR-1024i were stitched to give a continuous spectrum from 350 to 2500 nm. The original reflectance spectra were interpolated to obtain data at 1.0 nm wavelength intervals. Absolute reflectance was calculated by multiplying relative reflectance spectra by the laboratory calibrated reflectance of the leaf clip's reference standard panel (as calibrated by the NERC Field Spectroscopy Facility, panel manufactured by Spectra Vista Corp, USA).

Analysis of the hyperspectral dataset was carried out in R version 4.3.2 (R Core Team 2023) using the following packages: hyperspec (Beleites and Sergo 2017), hyperSpec. utils (Mayer 2024), readr (Wickham et al. 2024a), dplyr (Wickham et al. 2023), tidyr (Wickham et al. 2024b), factoextra (Kassambara and Mundt 2020), caret (Kuhn 2008), FactoMineR (Lê et al. 2008), tibble (Müller and Wickham 2023), scutr (Ganz 2023), kableExtra (Zhu 2024), stringr (Wickham 2023), and effectsize (Ben-Shachar et al. 2020). The following packages were used for data visualisation: ggplot2 (Wickham 2016), viridis (Garnier et al. 2024), and ggpubr (Kassambara 2023).

Hyperspectral analysis

We calculated the mean total leaf reflectance for eCO₂ and aCO₂ exposed leaves by integrating reflectance across the wavelength range and assessed the effect of CO₂ treatment via linear mixed effect modelling.

We calculated the first ten principal components of the hyperspectral dataset and evaluated the percentage of variance explained by each component. Principle component analysis is an unsupervised method of dimension reduction. We then analysed the difference in values of the first four components between CO₂ treatments by visualisation, followed by multivariate mixed effect modelling.

To assess the degree of difference between leaf spectra exposed to elevated versus ambient CO₂, we tested the ability of a machine learning model to predict CO₂ treatment from

leaf reflectance spectra. The spectral data were pre-processed with centring and scaling. Partial least squares discriminant analysis (PLSDA) models were trained on 70% of the leaf reflectance measurements (evenly split between leaves measured in ambient and elevated arrays), using tenfold cross-validation. PLSDA models were calculated with up to 60 components, and the accuracy of models with different numbers of components was evaluated to select an optimum number of components. The PLSDA model with the optimum number of components was then applied to unseen test spectra, and we evaluated the accuracy of the model's predictions of which CO₂ treatment leaves had been exposed to. We used the cumulative wavelength importance to understand which wavelength regions were most important for class separation in the PLSDA model.

Vegetation indices

We calculated a range of vegetation indices from the leaf reflectance spectra and compared the average vegetation index values of leaves exposed to elevated versus ambient CO₂ using mixed effect modelling. The names, formulas, and original references of the vegetation indices are listed in Table 1.

Statistical analysis: mixed effects modelling

Our dataset contains a hierarchical structure, individual leaves belong to separate trees, arranged in arrays, which then receive different CO₂ treatments. We also took measurements over

time, introducing a time series element to our dataset. To explicitly include these sources of variation in our analysis of the effect of eCO₂ on different elements of leaf reflectance, we used linear mixed effects modelling. Mixed effects modelling models the effect of fixed effects (such as CO₂ treatment) on response variables, as well as modelling the variation in response variable explained by random effects (such as hierarchical structure or time-series). Throughout our analysis we used variations of the model:

$$\text{Response variable} \sim \text{CO}_2 + (1|\text{Tree}) + (1|\text{Tree} : \text{Date})$$

The tree ID was used as a random effect throughout because it explained significant amounts of variation, whereas the array number did not explain any further variance when included alongside tree ID. Date of measurement for a given tree also explained variance and so was included as a nested random effect. Model estimation was carried out using Restricted Maximum Likelihood (REML), with Satterthwaite's approximation for degrees of freedom. Multivariate mixed effects modelling was used to model the response of the first four principal components to CO₂ treatment.

When comparing CO₂ treatments, density plots are used instead of traditional box plots to display the data distribution.

Table 1 Formula of common vegetation indices calculated from leaf reflectance

Vegetation index	Equation	References
Normalised Difference Vegetation Index	$\text{NDVI} = \frac{\rho_{800} - \rho_{670}}{\rho_{800} + \rho_{670}}$	Rouse et al. (1974)
Modified Chlorophyll Absorption Ratio Index	$\text{MCARI} = [(\rho_{700} - \rho_{670}) - 0.2 * (\rho_{700} - \rho_{550})] * \left(\frac{\rho_{700}}{\rho_{670}}\right)$	Daughtry et al. (2000)
Photochemical Reflectance Index	$\text{PRI} = \frac{\rho_{570} - \rho_{530}}{\rho_{570} + \rho_{530}}$	Gamon et al. (1997) Peñuelas et al. (1995a)
Plant Senescence Reflectance Index	$\text{PSRI} = \frac{\rho_{680} - \rho_{500}}{\rho_{750}}$	Merzlyak et al. (1999)
Normalised Difference Nitrogen Index	$\text{NDNI} = \frac{\log\left(\frac{1}{\rho_{1510}}\right) - \log\left(\frac{1}{\rho_{1680}}\right)}{\log\left(\frac{1}{\rho_{1510}}\right) + \log\left(\frac{1}{\rho_{1680}}\right)}$	Serrano et al. (2002), Fourty et al. (1996)
Normalised Difference Lignin Index	$\text{NDLI} = \frac{\log\left(\frac{1}{\rho_{1754}}\right) - \log\left(\frac{1}{\rho_{1680}}\right)}{\log\left(\frac{1}{\rho_{1754}}\right) + \log\left(\frac{1}{\rho_{1680}}\right)}$	Serrano et al. (2002), Fourty et al. (1996), Melillo et al. (1982)
Normalised Difference Water Index	$\text{NDWI} = \frac{\rho_{860} - \rho_{1240}}{\rho_{860} + \rho_{1240}}$	Gao (1996)
Normalised Phaeophytinisation Quotient Index	$\text{NPQI} = \frac{\rho_{415} - \rho_{435}}{\rho_{415} + \rho_{435}}$	Peñuelas et al. (1995b), Barnes et al. (1992), Ronen and Galun (1984)

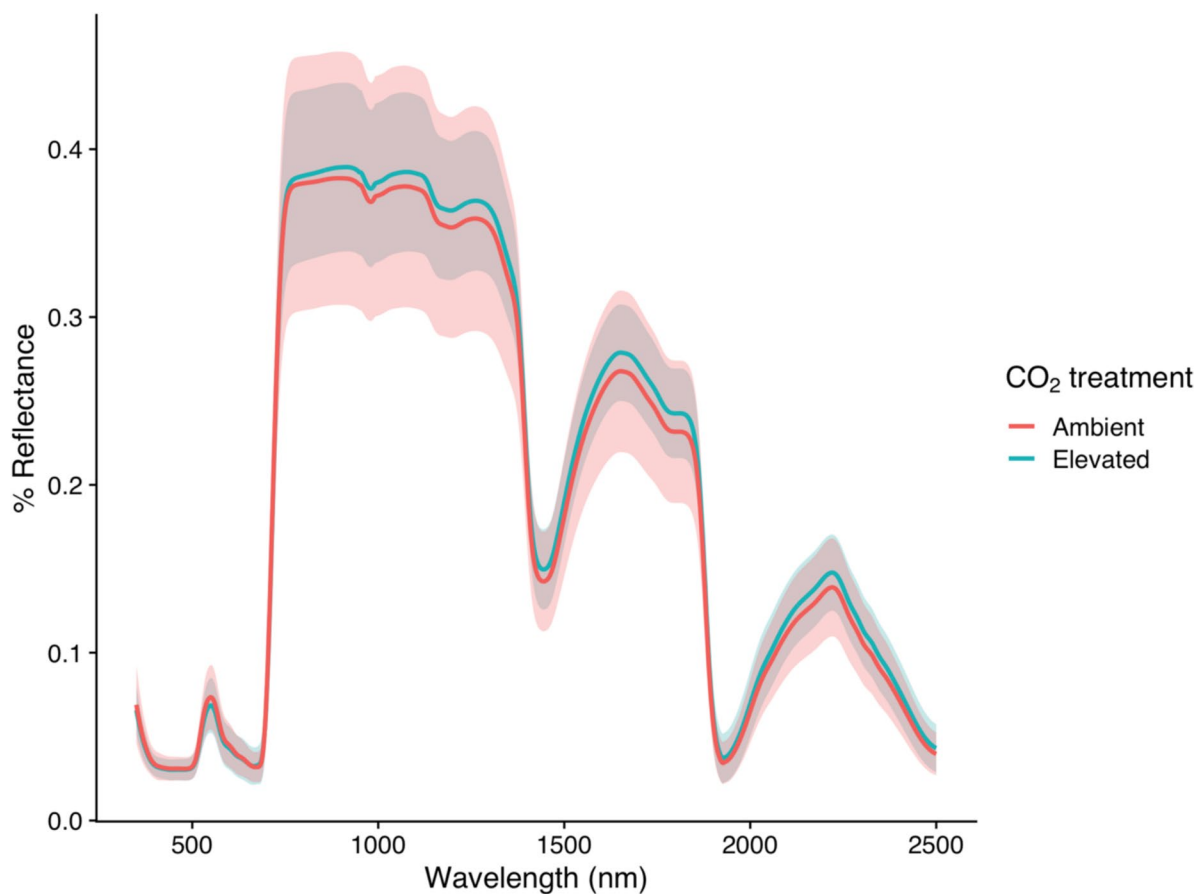


Fig. 1 Average percentage leaf reflectance spectra across the range of 350–2500 nm of oak leaves exposed to ambient (red) versus elevated (blue) CO₂ treatments at the BIFoR FACE experiment. Shaded areas

represent $\pm$ one standard deviation, with red shading for ambient CO₂ and blue shading for elevated CO₂

Results

Integrated leaf reflectance

The average reflectance spectra of leaves exposed to aCO₂ versus eCO₂ are visualised in Fig. 1. Visually, the reflectance of leaves exposed to eCO₂ was higher than those exposed to aCO₂, except for the green reflectance peak. At the green reflectance peak (540 nm), leaves exposed to eCO₂ had slightly lower reflectance than leaves exposed to aCO₂. The position of the Red-Edge does not visually differ between CO₂ treatments. The infrared reflectance peak between 2200 and 2400 nm was elevated and slightly shifted towards lower wavelengths under eCO₂.

The mean integrated reflectance for leaves exposed to eCO₂ was 436.67 (95% CI: 432.40–440.93), whereas under aCO₂ treatment it was 423.18 (95% CI: 416.34–430.02). Using a linear mixed effects model, the average integrated leaf reflectance under aCO₂ was an intensity of 423.00 (Standard Error (SE) = 6.93). Under eCO₂ integrated leaf reflectance was on average 13 units higher or 3.07%

(SE = 9.81, Satterthwaite's degrees of freedom (DF) = 5.97, Satterthwaite's P = 0.23), though this was not statistically significant. Random effects analysis of the date of measurement and tree ID showed that tree-to-tree variability in integrated leaf reflectance accounted for 72.77 variance (Standard Deviation (SD) = 8.53), while date of measurement accounted for 356.67 variance (SD = 18.89). The residual variance in integrated leaf reflectance was 3523.52, indicating large variability which was not explained by CO₂ treatment, the random effects of tree-to-tree, or date of measurement variability. Inclusion of the array number as a random effect in the model did not explain any further variance.

Leaf reflectance principal component analysis

The first four principal components (PCs) cumulatively explained 97.2% of the total variance in hyperspectral leaf reflectance (Fig. 2b, PC1: 64.66%, PC2: 23.33%, PC3: 7.04%, PC4: 2.31%). The first component (PC1) is loaded negatively by wavelengths around the red edge (Fig. 2a), while the second component (PC2) is positively

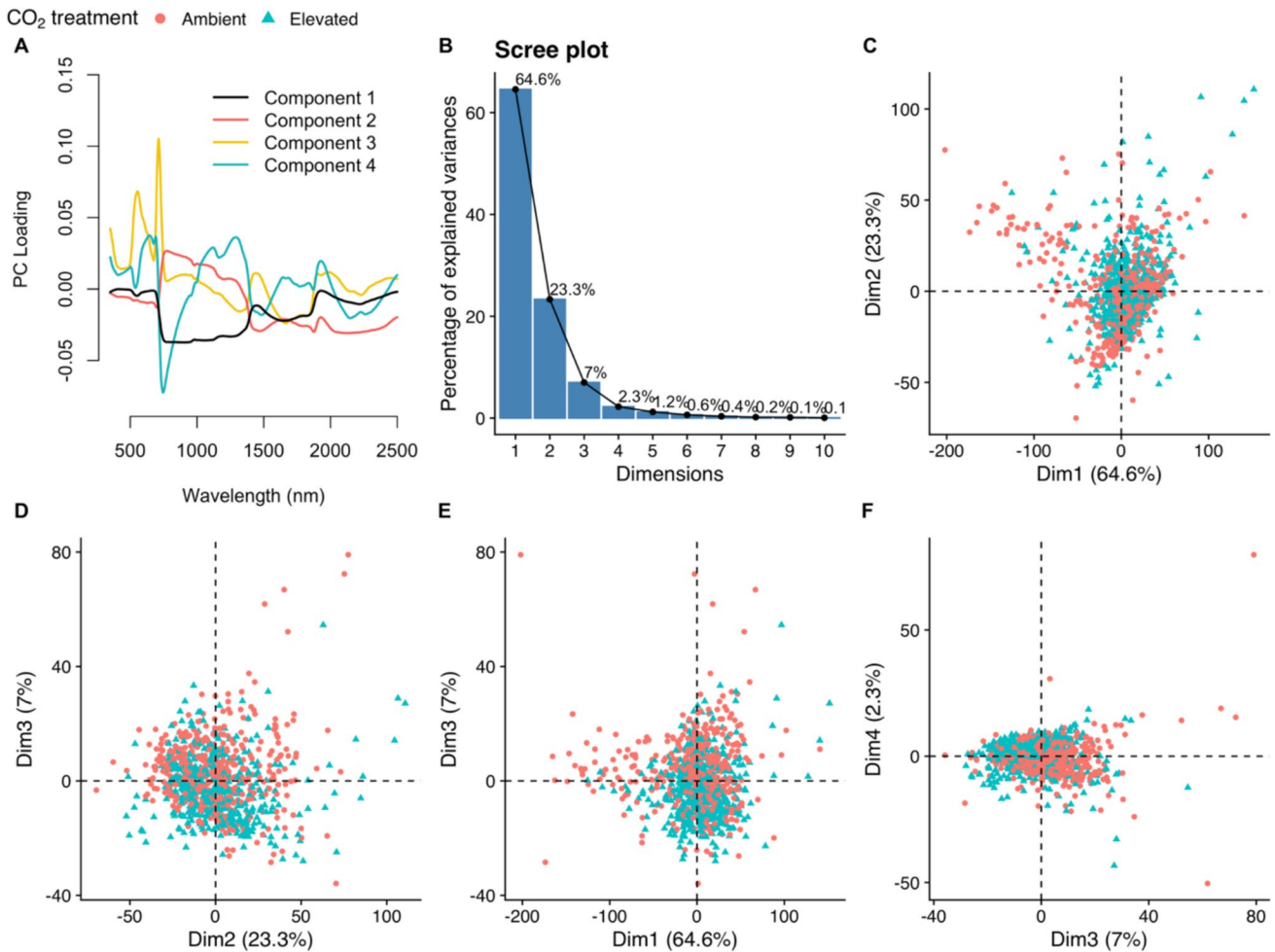


Fig. 2 Principal component analysis (PCA) of the leaf reflectance spectra in the range 350–2500 nm of oak trees exposed to ambient (red) versus elevated (blue) CO₂ treatments at the BIFoR FACE experiment. **a** Wavelength (nm) loading of the first four principal

components. **b** Scree plot of variance explained by increasing the number of principal component dimensions. **c–f** Leaf reflectance of trees exposed to ambient (red) versus elevated (blue) CO₂ treatments, plotted against combinations of the first four principal components

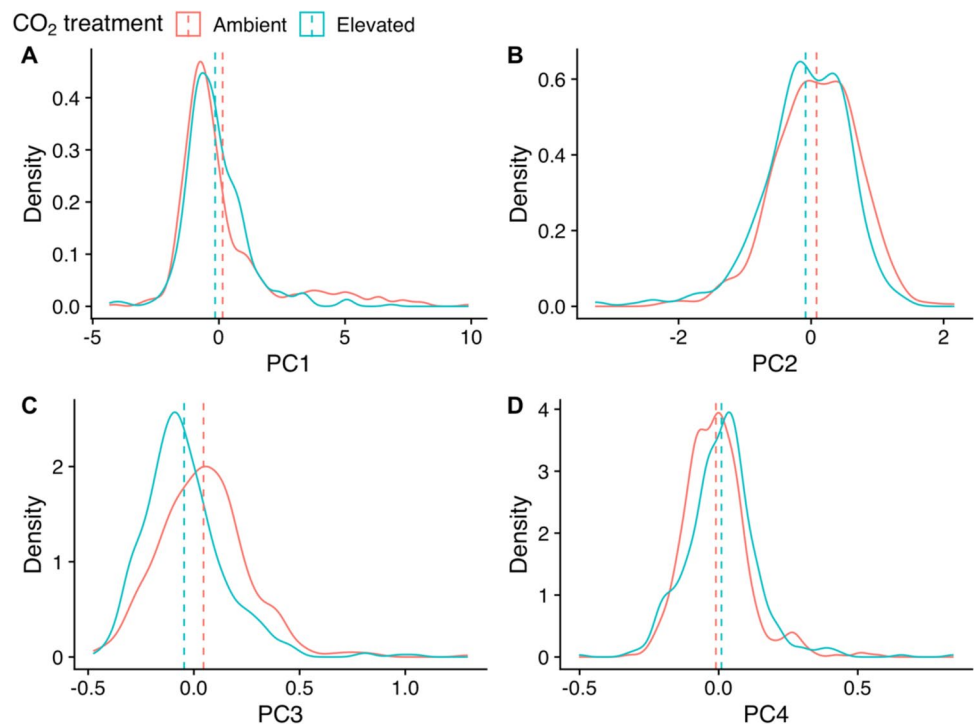
loaded by the red edge and wavelengths between 750 and 1500 nm. The third principal component (PC3) is strongly positively loaded by wavelengths characterising the green peak (550 nm) and red edge (680–750 nm), and the fourth component (PC4) is strongly negatively loaded by wavelengths characterising the red edge (680–750 nm) and positively loaded by longer wavelengths. When individual leaf reflectance spectra are plotted against combinations of the first four principal components (Fig. 2c–f), the spectra show limited clustering by CO₂ dosage. Clustering by CO₂ dosage is clearer against components two and three (Fig. 2d), and three and four (Fig. 2f). As an unsupervised dimensionality reduction method, principal component analysis reveals that CO₂ treatment has a limited diverging effect on values of the first four principal components of leaf reflectance. Figure 3 shows the difference in density distribution of principal component scores between CO₂ treatments, the

clearest difference is in component three which has a narrower spread of lower scores under elevated CO₂ (Fig. 3c).

To assess whether CO₂ treatment significantly influenced leaf reflectance spectra as summarised by the first four principal components, we fitted a multivariate linear mixed-effects model with CO₂ treatment as a fixed effect and random intercepts for tree ID and nested date of measurement.

The model had an intercept of 0.066 (SE = 0.063) for ambient CO₂ and the fixed effect of eCO₂ treatment was estimated at –0.1324 (SE = 0.089). However, this difference was not statistically significant (DF = 5.9, $P = 0.19$). Random effects analyses indicated low between-group variation. The tree-to-tree variance was 0.01 (SD = 0.10) and among the tree-by-date combinations 0.016 (SD = 0.13), while the residual variance (i.e., variation among individual leaves) was substantially larger (0.79; SD = 0.89). Adding the array as an additional level of random effect did not explain any

Fig. 3 Density plots showing the distribution of values of the first four principal components (PC1–4) in leaf reflectance spectra of oak trees exposed to ambient (red lines) versus elevated (blue lines) CO₂ treatment. Vertical dashed lines indicate mean principal component value per treatment (ambient in red, elevated in blue). Panel **a** represents principal component 1, panel **b** represents principal component 2, panel **c** represents principal component 3, and panel **d** represents principal component 4. Data taken from mature *Quercus robur* trees at the BIFoR FACE experiment, Staffordshire



further variation in the principal components of leaf reflectance spectra.

Leaf reflectance partial least squares discriminant analysis (PLSDA)

Tenfold cross-validation of the PLSDA model on the training dataset had the highest accuracy with 44 components (Fig. 4, accuracy = 0.89). The 44-component PLSDA model had an accuracy of 0.88 (95% confidence intervals: 0.84, 0.92) for assigning unknown leaf spectra to the correct CO₂ treatment, sensitivity was 0.89, and specificity was 0.87. Compared to the ‘no information rate’ (assigning all spectra to aCO₂ class), the PLSDA model was significantly more accurate (‘no information rate’ accuracy = 0.50, $P < 2e-16$). Between the two classes (aCO₂ and eCO₂), error rates were not significantly different (McNemar’s Test, $P = 0.73$). Figure 5 visualises the cumulative importance of different wavelengths in separating the reflectance spectra of aCO₂ leaves from eCO₂ leaves. Key regions of importance include the upper edge of the green reflectance peak (565–580 nm), the water absorption region at 1400 nm, and the shortwave infrared peak at 1600–1720 nm. The 44-component PLSDA model was able to accurately separate leaf spectra exposed to aCO₂ from those exposed to eCO₂.

Vegetation indices

Linear mixed-effects models were fitted to examine the impact of CO₂ treatment on each vegetation index, with random intercepts for tree and for tree-by-date to account for the hierarchical structure. The results of these models are presented in Table 2. CO₂ treatment did not have a significant effect on the value of most of the vegetation indices tested except PSRI. Random effect analysis revealed moderate tree-to-tree variation in all vegetation indices, and moderate variation between dates of measurement, although the majority of variation resided at the residual level indicating the predominant source of variation in most vegetation indices in this experiment was at the leaf level and was not affected by CO₂ treatment. Figure 6 visualises the distributions of vegetation index values of leaves exposed to ambient or elevated CO₂. Although MCARI did not differ significantly between CO₂ treatments, the distribution of values under eCO₂ is much narrower than under aCO₂ indicating reduced variance in MCARI values under eCO₂. Similarly, there is a narrower distribution of values around the median NDNI and NPQI values in leaves exposed to eCO₂.

PSRI was significantly higher in leaves exposed to eCO₂ compared to aCO₂ (see Table 2, $+2.6 \times 10^2$ PSRI relative to aCO₂, $t(5.97) = 2.5$, $P = 0.049$). Random effects analysis revealed moderate variability among trees (variance = 0.14, SD = 0.012) and dates (variance = 0.24, SD = 1.5×10^{-4}), although most of the variation resided at the residual level (variance = 1.8×10^{-3} , SD = 0.043). Leaves exposed to

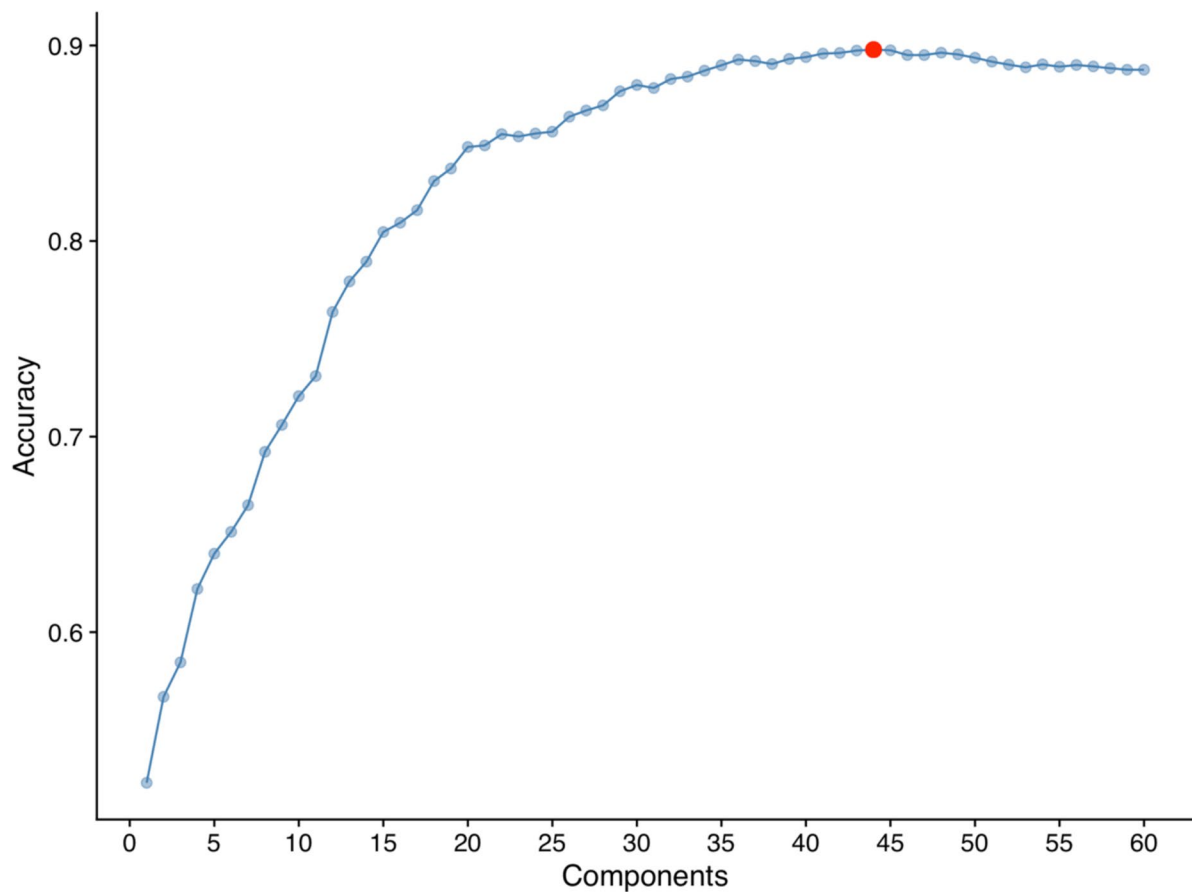


Fig. 4 Cross-validation tuning of a partial least squares discriminant analysis (PLSDA) model to separate leaves exposed to elevated versus ambient CO_2 by their reflectance spectra. Model accuracy is plot-

ted against number of model components, with the optimum number of model components for maximum accuracy highlighted as a red circle (44 components, 0.89 accuracy)

eCO_2 exhibited higher PSRI values throughout the measurement period, whereas the PSRI of leaves exposed to aCO_2 increased steadily from July to October. Figure 6i shows how the average PSRI per CO_2 treatment changed over time.

Discussion

Our analysis shows that most regions of mature *Q. robur* leaf reflectance spectra and derived vegetation indices were not significantly affected by eCO_2 treatment after 7 years of eCO_2 exposure, except in the case of PSRI which was higher in eCO_2 exposed leaves.

In our study, leaves exposed to eCO_2 showed a non-significant trend of increased the integrated leaf reflectance, 3.2% higher than leaves exposed to aCO_2 . Leaf reflectance measurements taken with an integrating sphere give a more accurate measurement of total leaf reflectance and any albedo effects by including all possible angles of reflectance, however, when taking many measurements in the forest canopy the speed and practicality of leaf clip reflectance

measurements outweighed any benefits an integrating sphere may have offered. Thomas et al. who found a significant leaf reflectance under eCO_2 in saplings of tropical tree species using an integrated sphere, although the extent of reflectance increase varied from 9% to 23% by species (Thomas 2005). It has been suggested that such changes could affect forest albedo (Thomas 2005). Forest albedo, which refers to the amount of solar radiation reflected back to the atmosphere by forests, plays a critical role in regulating the Earth's climate and is a source of uncertainty in the radiation budget of climate models (Alibakhshi et al. 2020). Forest albedo is influenced not only by forest structure but also by leaf-level optical properties (Hollinger et al. 2010; Meunier et al. 2022; Henniger et al. 2023). For example, in China, forest greening between 2002 and 2019 was accompanied by an increase in shortwave albedo (Yan et al. 2021). Luyssaert et al. highlighted the lack of consideration of changes to forest albedo in the management of forests aimed at offsetting CO_2 emissions (Luyssaert et al. 2018). Despite this, the effect of eCO_2 on forest albedo remains understudied. Our results suggest that in the long-term eCO_2 may not have a significant effect

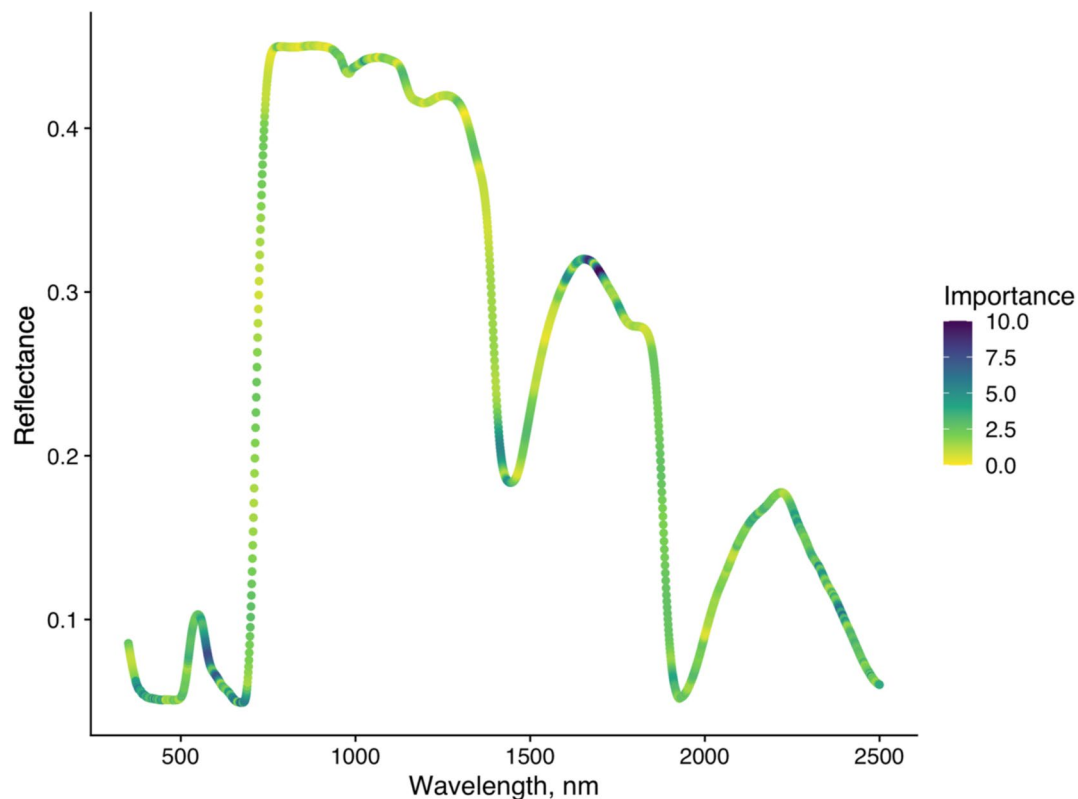


Fig. 5 Cumulative wavelength (nm) importance for class separation of CO₂ treatments (ambient CO₂ versus elevated CO₂) by leaf reflectance spectra in partial least squares discriminant analysis (PLSDA)

model. Colours indicate the importance of each wavelength to the discrimination model

on the total leaf reflectance of mature *Q. robur*, but more detailed analysis of total leaf reflectance and transmission using an integrating sphere would be necessary to predict the affect of eCO₂ on oak forest albedo. Further analysis is needed to understand how the total reflectance of different mature tree species will respond to future CO₂ concentrations, as well as the effect on temperate forest albedo.

As hypothesised, the spectral reflectance profile of leaves exposed to eCO₂ was similar to that of leaves exposed to aCO₂. The spectral signature as summarised by the first four principal components was not significantly affected by CO₂ treatment after accounting for hierarchical structure. The large residual in the mixed effects model of principal components suggests that most of the variation in leaf reflectance spectra arose at the leaf level rather than at the tree or temporal level. Most of the vegetation indices examined also did not respond to CO₂ treatment, although the reduced spread of MCARI, NDNI, and NPQI values under eCO₂ could indicate some form of directional response. The majority of previous studies which have examined the effect of eCO₂ on the leaf reflectance of trees have also not found a significant eCO₂ effect in mature *Quercus pubescens* (Stylinski et al. 2000), in young *Acer saccharum* (Carter et al. 2000), in mature *Liquidambar styraciflua* (Wicklein et al. 2012),

nor in mature *Eucalyptus tereticornis* (Wujeska-Klaue et al. 2019). Significantly increased reflectance in the green peak and red edge was reported in *Pinus strobus* L exposed to eCO₂ (Carter et al. 1995). eCO₂-induced spectral changes have also been reported in crops (Gray et al. 2010; Tormena et al. 2019). On balance, the lack of eCO₂ response in the spectral profile of *Q. robur* in this study is in keeping with previous research in deciduous tree species. This suggests the spectral profile of future deciduous forests will not be broadly altered by future CO₂ concentrations.

Our PLSDA model is useful in highlighting the regions of the spectra which differed most between CO₂ treatments and so answering the question “how does eCO₂ affect leaf reflectance?” in a nuanced way. The key regions of differentiation were the upper edge of the green reflectance peak, the water absorption region at 1400 nm, and the shortwave infrared peak around 1650 nm. First, the green reflectance peak is affected by the overlapping reflectance peaks of different chlorophyll, carotene, and anthocyanin pigments (Sims and Gamon 2002), so changes in the upper edge green peak reflectance could reflect a change in one or multiple pigment concentrations.

The 1400 nm water absorption peak was also important in differentiating between leaves exposed to different CO₂

Table 2 Summary of linear mixed effect model results for vegetation indices of leaves exposed to ambient (aCO₂) or elevated CO₂ (eCO₂)

Index	aCO ₂		eCO ₂ effect		SE	T value	P value	Tree variance		Date variance		Residual variance		Figure
	Mean value	SE	Mean effect	SE				Value	SD	Value	SD	Value	SD	
NDVI	8.4 × 10 ⁻¹	7.9 × 10 ⁻³	+ 4.1 × 10 ⁻³	1.1 × 10 ⁻²	1.1 × 10 ⁻²	3.5 × 10 ⁻¹	7.4 × 10 ⁻¹	1.9 × 10 ⁻⁴	1.4 × 10 ⁻²	1.9 × 10 ⁻⁴	1.4 × 10 ⁻²	2.8 × 10 ⁻³	5.3 × 10 ⁻²	6a
MCARI	1.1 × 10 ⁻¹	1.6 × 10 ⁻²	- 1.0 × 10 ⁻²	2.2 × 10 ⁻²	2.2 × 10 ⁻²	- 4.8 × 10 ⁻¹	6.5 × 10 ⁻¹	6.6 × 10 ⁻⁴	2.6 × 10 ⁻²	1.1 × 10 ⁻³	3.3 × 10 ⁻²	2.3 × 10 ⁻³	4.8 × 10 ⁻²	6b
PRI	- 1.1 × 10 ⁻²	4.5 × 10 ⁻³	+ 5.1 × 10 ⁻⁴	6.4 × 10 ⁻³	6.4 × 10 ⁻³	8.1 × 10 ⁻²	9.4 × 10 ⁻¹	6.8 × 10 ⁻⁵	8.3 × 10 ⁻³	4.2 × 10 ⁻⁵	6.4 × 10 ⁻³	2.5 × 10 ⁻⁴	1.6 × 10 ⁻²	6c
PSRI	- 3.1 × 10 ⁻³	7.3 × 10 ⁻³	+ 2.6 × 10 ⁻²	1.0 × 10 ⁻²	1.0 × 10 ⁻²	2.5 × 10 ⁻⁰	4.9 × 10 ⁻² *	1.4 × 10 ⁻⁴	1.2 × 10 ⁻²	2.4 × 10 ⁻⁴	1.5 × 10 ⁻²	1.8 × 10 ⁻³	4.3 × 10 ⁻²	6d
NDNI	1.1 × 10 ⁻¹	4.7 × 10 ⁻³	+ 2.8 × 10 ⁻³	6.7 × 10 ⁻³	6.7 × 10 ⁻³	4.2 × 10 ⁻¹	6.9 × 10 ⁻¹	7.7 × 10 ⁻⁵	8.8 × 10 ⁻³	3.8 × 10 ⁻⁵	6.1 × 10 ⁻³	2.3 × 10 ⁻⁴	1.5 × 10 ⁻²	6e
NDLI	3.5 × 10 ⁻²	1.5 × 10 ⁻³	- 3.1 × 10 ⁻⁴	2.2 × 10 ⁻³	2.2 × 10 ⁻³	- 1.4 × 10 ⁻¹	8.9 × 10 ⁻¹	8.3 × 10 ⁻⁶	2.9 × 10 ⁻³	3.0 × 10 ⁻⁶	1.7 × 10 ⁻³	2.5 × 10 ⁻⁵	5.0 × 10 ⁻³	6f
NDWI	3.1 × 10 ⁻²	4.7 × 10 ⁻³	- 7.0 × 10 ⁻³	6.6 × 10 ⁻³	6.6 × 10 ⁻³	- 1.1 × 10 ⁻⁰	3.3 × 10 ⁻¹	6.8 × 10 ⁻⁵	8.2 × 10 ⁻³	6.0 × 10 ⁻⁵	7.7 × 10 ⁻³	5.0 × 10 ⁻⁴	2.2 × 10 ⁻²	6g
NPQI	1.3 × 10 ⁻²	1.7 × 10 ⁻³	+ 2.0 × 10 ⁻³	2.4 × 10 ⁻³	2.4 × 10 ⁻³	7.6 × 10 ⁻⁰	8.3 × 10 ⁻¹	4.1 × 10 ⁻⁶	2.0 × 10 ⁻³	2.5 × 10 ⁻⁵	5.0 × 10 ⁻³	1.3 × 10 ⁻⁴	1.2 × 10 ⁻²	6h

Note(s): In all model the degrees of freedom were 5,97, using Satterthwaite's method. P values marked with * indicate a statistically significant effect ($\alpha=0.05$). All values are given to two significant figures in standard form

treatments. In a different species of oak (*Quercus agrifolia*), the 1650 nm peak has also been shown to correspond to leaf water status (Pu et al. 2003), but it has not been explicitly characterised in *Q. robur*. In a global meta-analysis, eCO₂ has been shown to increase leaf water content under drought conditions (Wang et al. 2022). Generally, eCO₂ has been found to increase vegetation's water use efficiency (WUE) as a result of stomatal closure and/or increased photosynthetic activity (Gilbert et al. 2011; Gardner et al. 2023). Additionally, in the BIFoR FACE experiment, there were no significant changes to water use under eCO₂.

Wavelengths in the 1650 nm reflectance peak were the most important in the PLSDA model's differentiation between CO₂ treatments. Tormena et al. (2019) also identified a response to eCO₂ in coffee leaf reflectance bands around 1657 nm and 1698 nm via PLSDA (Tormena et al. 2019). The 1650 nm peak is an indirect reflectance band, due to chemicals such as cellulose (Curran 1989) and affected by leaf water content (DANSON et al. 1992; Tian et al. 2001). The cell wall hemicellulose content increased in *Betula pendula* leaves exposed to eCO₂ (Oksanen et al. 2005), but the effect of eCO₂ on leaf cellulose content in oak species has not been well examined. More broadly, eCO₂ has been shown to increase leaf carbohydrate content (Kinney et al. 1997; Agrell et al. 2000; Coley et al. 2002), and at the BIFoR FACE experiment foliar carbon content in fresh leaves was significantly higher under eCO₂ (Roberts et al. 2022), which could include changes to cellulose content. While firm conclusions cannot be drawn from the PLSDA analysis, the importance of wavelengths in the upper green reflectance, water absorption band, and 1650 nm peak are useful to highlight as directions for future investigation of the foliar effects of eCO₂ in mature deciduous trees.

The one vegetation index which CO₂ treatment had a significant effect on was PSRI. PSRI is sensitive to the ratio of carotene to chlorophyll pigments in the leaf and is used as a quantitative measure of senescence-induced degradation of chlorophyll (Merzlyak et al. 1999). PSRI was significantly higher under eCO₂ and was elevated early in the growing season compared to trees exposed to aCO₂. This indicates changes in the relative content of carotenes and chlorophyll under eCO₂. Since no changes in chlorophyll content have been found with eCO₂ treatment at BIFoR FACE (Gardner et al. 2022b), the increase in PSRI is more likely to be due to increased carotene content than decreased chlorophyll. Reductions in carotene content with eCO₂ treatment have been found previously (Loladze et al. 2019). In the only study on *Q. robur*, the decrease in carotenoid content under eCO₂ was not significant (Schwanz and Polle 2001), but this study was on seedlings in pots so may not be transferable to adult trees in a mature forest. Norby et al. (2024) reported leaf loss started earlier in some years under eCO₂ at the BIFoR FACE experiment, which could also lead to

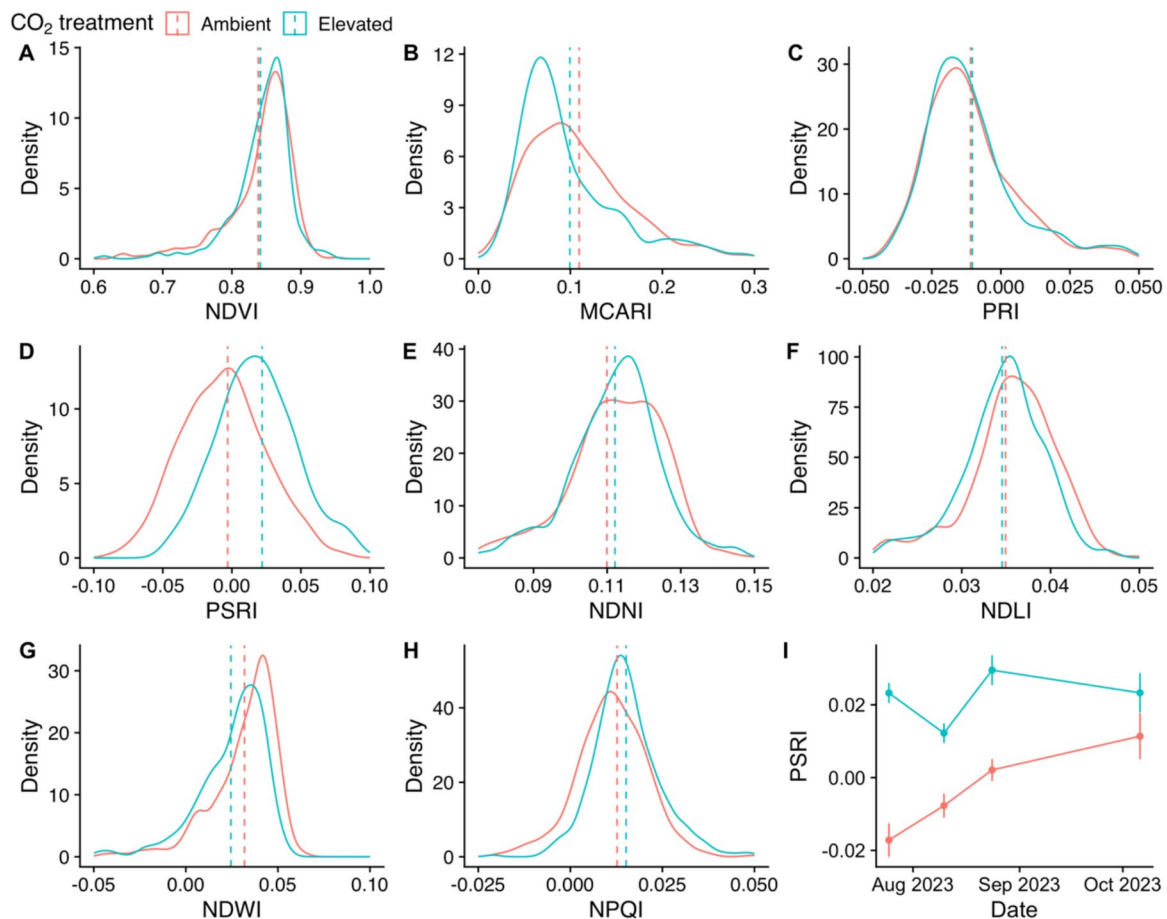


Fig. 6 a–h Density plots showing the distribution of values of range of vegetation indices of oak leaves exposed to ambient (in red) or elevated (in blue) CO₂ treatment. Dashed lines indicate the mean value

of each vegetation index for each CO₂ treatment. **i** Time series of mean Plant Senescence Reflectance Index (PSRI) for each CO₂ treatment

earlier chlorophyll breakdown and elevated PSRI (Norby et al. 2024). Other FACE experiments have also reported extended senescence (Sigurdsson 2001), although in popular senescence was delayed by eCO₂ (Cotrufo et al. 2005; Taylor et al. 2008). In tree seedling experiments white birch and basswood, autumn senescence accelerated under eCO₂ (Li et al. 2019; Tedla et al. 2020). Ontogeny is easier to study in herbaceous plants, but there are also mixed effects of eCO₂ on herbaceous plant senescence (Curtis et al. 1989; de la Mata et al. 2012). We would encourage future work to examine the pigment concentrations of leaves from the BIFoR FACE experiment via HPLC, to quantify the changes in carotenes indicated by our spectral data and how pigments change seasonally in relation to senescence.

In large scale ecosystem manipulation experiments, like the BIFoR FACE experiment, engineering and financial constraints limit the number of replicate arrays and thus limit the “n” of the study. With small numbers of replicate arrays, very large changes are needed between treatments to produce statistically significant results using traditional

statistical methods (Norby et al. 2024). Mixed effect modeling goes some way to address this issue. However, the value of observations from large ecosystem manipulation experiments should not be dismissed on account of the difficulty in producing statistically significant results while avoiding pseudo-replication (Davies and Gray 2015).

Conclusion

Long-term exposure to eCO₂ did not significantly change the intensity of leaf reflectance or the overall spectra of mature *Q. robur* trees. However, the Plant Senescence Reflectance Index (PSRI) increased under eCO₂ conditions, indicating a change in the ratio of chlorophyll to carotene pigments. Our analysis also highlighted reflectance changes in the infrared spectrum, which may correspond to alterations in leaf water content and carbohydrate content. Hyperspectral leaf reflectance is a useful tool for understanding the consequences

of future atmospheric CO₂ concentrations on foliar features such as pigments.

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Author contributions ALJ, EJ, and CP conceived the field experiment. ALJ collected, processed, and analysed the data. AG contributed to the interpretation of the results by ALJ. ALJ wrote the first draft. All authors contributed to writing of the final draft.

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Data availability The spectral dataset and analysis code used in this research will be freely available to download at <https://doi.org/https://doi.org/10.5281/zenodo.15323740> upon publication.

Declarations

Conflict of interest The authors have no relevant financial or non-financial interests to disclose.

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