

An investigation of the fluid-holding cavities in a lignocellulose-based bamboo matrix via a combined X-ray microtomography and proton time-domain NMR approach

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Abstract

Bamboo (*Dendrocalamus giganteus*) is a functionally graded material with well-organized hierarchical structures. Its micrometer-sized vascular bundles and parenchymatic living cells allow an efficient upward flow of water and nutrients, endowing the organism with remarkably fast growth. As demonstrated recently, the hollow microstructure channels can be explored as a natural template for microfluidics applications in chemical synthesis, analytical detection, solar steam generation, and electrochemical devices. Thus, the knowledge of the kinetics of the imbibition and spatial distribution of fluid through the microcavities of the bamboo vegetal tissue became of interest. Here, we employed a combination of X-ray microtomography (μ CT) and proton time-domain NMR (TD-NMR) to identify, measure, and investigate empty volumes embedded in the bamboo's tissue as experienced by different organic and inorganic fluids, namely dimethyl sulfoxide (DMSO) and distilled water (H_2O). Results have shown that the extensive communication between the voids (parenchymal cells and vascular channels) does not prevent the individuation of fluid reservoirs with different behaviors, as evidenced by the H_2O desorption profile. Bamboo parenchyma was the tissue that retained more residual H_2O after desorption. The difference in wettability of bamboo tissues by DMSO and H_2O was detected.

Introduction

The bamboo (*Dendrocalamus giganteus*) plant tissue matrix is chemically and morphologically complex. It is considered an intelligent functionally graded material with oriented, hierarchical, and shaped biological tissues that developed over long-term evolution and selection (Nogata e Takahashi 1995). At the molecular level, it consists mainly of cellulose, a linear polysaccharide, highly organized due to hydrogen bonds between the abundant hydroxyl moieties of β -(1–4)-D-glucopyranose monomers; hemicellulose, a branched polysaccharide of low crystallinity consisting of monosaccharide units of 5 to 6 carbons (mainly glucose, xylose, galactose, and mannose) and lignin, a heterogeneous phenolic macromolecule. Both cellulose and lignin are responsible for the high mechanical properties displayed by the plant and are found in large quantities (55% and 25%, respectively) in the bamboo matrix (Ji et al., 2013; Youssefian and Rahbar, 2015).

At the micrometric level, the bamboo tissues concentrate cavities where liquids can flow and be stored, being greatly responsible for its fast growth. In the longitudinal direction of the bamboo's culm, straight and parallel channels with diameters varying from a few tens to a few hundred micrometers compose the phloem, protoxylem, and metaxylem vascular systems. These vascular bundles are mechanically protected by the sclerenchymatous tissue, mainly constituted of highly oriented, crystalline, and hierarchical ligno-carbohydrate complex (LCC) (Hao et al. 2018). Also present in the culm, polyhedral-shaped micrometric cells, with highly lignified walls interconnected via micro- and nanostructures called pits, constitute the parenchyma tissue (Xu et al. 2011; Ji et al. 2013; Peng et al. 2013; Youssefian and Rahbar 2015; Pandoli et al. 2015).

Functional gradients, heterogeneities, and highly anisotropic morphology of biological materials have recently attracted much attention in investigations concerning novel sustainable and high-performance materials (Liu et al. 2017). Bamboo, less than wood, has been explored for new challenging applications adding mechanical, optical, and chemical properties. Zhu *et al.* described the delignification and subsequent polymer infiltration of basswood samples with a relevant increase in strength and elastic modulus compared with neat basswood (Zhu et al. 2016). Similarly, Li *et al.* (Li et al. 2017d) reported transparent wood composite materials with lower thermal conductivity and higher work-to-fracture values when compared to similar-sized glass samples. The inclusion of quantum dots (QD) in transparent wood composites by Fu and collaborators gave rise to luminescent biocomposites, readily combining multiaxial load-bearing performance with novel optical features (Fu et al. 2018). Balsa wood was made highly flame-resistant by incorporating colloidal montmorillonite clay into its structure to circumvent the flammability of wood materials employed in architecture and engineering (Fu et al. 2017).

Other works also relied on the anisotropic structure of the network of channels present in plants, through which passive forces can transport fluids. Jia and collaborators explored the presence of mesoporous hydrophilic channels inside several plants. They showed that, in some cases, a simple wood delignification procedure is enough to transform them into a powerful microfluidic platform capable of efficiently moving volumes of liquids that may contain even colloidal particles, such as carbon nanotubes (Jia et al. 2018). Richardson and Lang explored the ability to transport and internalize small molecules exhibited by vegetal tissue in synthesizing two metal-organic frameworks (MOFs) within plant tissues. The nano-biohybrids obtained showcased the potential of the new materials in sensing small molecules (Richardson and Liang 2018). Stavriniidou *et al.* developed an electrical circuit based on electrically conducting polymers anchored in the framework formed by the xylem of *R. floribunda* specimens, reaching electrical conductivities of the order of 0.1 S.cm^{-1} (Stavriniidou et al. 2015). Composite materials based on basswood infiltrated with an Sn-Bi alloy showed thermal, mechanical, and electrical conductivity properties dependent on the anisotropic structure presented by this wood (Wan et al. 2017).

In the mentioned studies, the internal channel structures of the vegetal tissue were often harnessed to enable the chemical modifications necessary for developing the new materials. In the literature reports concerning bamboo specimens, most chemical modifications were conducted on the external surface, affording materials with new properties. Superhydrophobic bamboo surfaces with magnetic and electrical conductivity properties were achieved by Jin *et al.* by coating the specimens with *in situ* generated silver and $\gamma\text{-Fe}_2\text{O}_3$ nanoparticles and further treatment with fluoralkoxysilanes. The new materials showed very obtuse water contact angles, resistance to change in pH and ionic force of the external medium, and, in the case of $\gamma\text{-Fe}_2\text{O}_3$ coated specimens, superparamagnetic behavior (Jin et al. 2015a, b). Li et al. made the bamboo surface mold-resistant by incorporating ZnO nanoparticles fabricated *in situ* via a wet chemical method. It is worth noting that in these past instances, the internal architecture of the bamboo plant never played an essential role in the product's performance or feasibility (Li et al. 2017a). Nanostructured Copper oxide (CuO and Cu_2O) imbibed onto bamboo surfaces were

explored for heterogenous catalytic click-chemistry reactions in several organic solvents (De Sá et al. 2022).

Few reports have been published exploring the internal structure of bamboo as bio-templates for chemical platforms, steam generation, and electrochemical devices. Kuan and co-workers developed the first lignocellulose-based analytical devices (LADs) as an analytical platform for chemical detection through a colorimetric dosage of several analytes in water, milk, and urine (Kuan et al. 2015).

Several papers outlining ways to harness the internal structure of bamboo tissues have been recently published, describing the filling and chemical functionalization of the parenchyma and/or vascular bundles with transition metal salts or nanoparticles, such as copper and silver. Copper (I)-catalyzed 1,3-dipolar cycloaddition reactions between azides and terminal alkynes using an appropriately functionalized bamboo-based microfluidic reactor were achieved (De Sà et al. 2019). Regioselective distribution of silver nanoparticles capped with negatively and positively charged surfactants to the parenchyma and vascular channels to prevent fungal proliferation in bamboo specimens was described (Pandoli et al. 2016; Ginoble Pandoli et al. 2019). The development of unprecedented bamboo-based devices with ultra-highly conductive hollow channels guided by a bamboo bio-template for electric and electrochemical applications was also reported. Colloidal silver ink impregnation into the microchannels allowed to obtain the highest conductivity cellulose-based ordered material for a new generation of bambootronics devices, such as 3D electric circuits, microfluidic heaters, and fully integrated micro electrochemical cells (Pandoli et al. 2020). Jinpeng Li and co-workers have recently incorporated catalytic Ag-NPs into the bamboo vascular bundles to obtain a stable and high-efficiency catalytic microreactor for nitroaromatics reduction (Li et al. 2022).

A plasmonic bamboo device for solar steam generation has been developed to explore water's capillary transportation through micro-sized channels (Sheng et al. 2020). In this case, the bamboo matrix decorated with plasmonic Ag and Pd-NPs can convert the light energy into heat energy. Seawater flowing upwards through the vascular bundles is vaporized to form steam and, consequently, desalinated water.

Recently, the anisotropic behavior of water vapor sorption into the hierarchical structure of bamboo was demonstrated, revealing different sorption directions, longitudinal (L) and transversal (T), for a saturated salt solution and water vapor (Chen et al. 2021). Thus, the need for a deeper understanding of how vascular channels and cell tissues interact with different fluids mainly used in organic chemistry has become evident.

X-ray microtomography (μ CT) is a non-destructive technique widely used for the characterization of the microstructure of plants and has been used for bamboo as well (Palombini et al. 2020). Time-domain nuclear magnetic resonance (TD-NMR) is an efficient, fast, and non-destructive way to probe fluid behavior in confined environments (Steele et al. 2016; Singer et al. 2018). It is widely used in the study of oil reservoir rock matrices (Muhammad e Azeredo 2014; Plastino et al. 2017; Rudsuck et al. 2019), water trapped in hydrogels (Rodrigues et al. 2016, 2017; Ratzsch et al. 2017) and to study the molecular

dynamics of liquids during the hardening of cementitious materials (Valckenborg et al. 2001; Jansen et al. 2018).

However, TD-NMR has few reported uses in the literature for studying liquids confined in vegetal tissue apart from the quantification of oil in seeds (Prestes et al., 2007; Colnago et al., 2011, 2014). Zhang *et al.* used TD-NMR methods to characterize hierarchical pore structures in cotton fibers by measuring the different relaxation behaviors of the liquids imbibed in the cellulose matrix (Zhang et al. 2016). Mohebbi, Claussen, and Blümich used a single-sided TD-NMR apparatus to quantify the amount of water in open foams and fibrous materials like paper (Mohebbi et al. 2019). Li, Wang, and Zhang determined the rate of humidity desorption in wood (*Populus platyphylla* and *Pinus sylvestris*) samples via TD-NMR (Li et al. 2017b). Mikac *et al.* used magnetic resonance imaging (MRI) and TD-NMR methods to investigate the behavior of moisture content in *Fagus sylvatica* branches (Mikac et al. 2021). Recently, Li and Ma. proposed the investigation of the cell wall water of wood in the hygroscopic range using 2D TD-NMR, successfully differentiating the water mobility in cellulosic material (Li and Ma 2022).

The interest in investigating the behavior of fluids in the cavities of harvested and treated bamboo emerges when its use as a renewable platform for carrying out physicochemical processes becomes evident. However, morphological heterogeneities in the cavities that retain and transport fluids inside the living plant can be found in each internode. Therefore, the structure-properties relationship in this context is highly relevant for the current and future applications of this class of renewable porous material.

In this work, we analyzed the behavior of organic (dimethyl sulfoxide) and inorganic (distilled water) fluids in the lignocellulosic matrix of bamboo (*Dendrocalamus giganteus*) through proton TD-NMR experiments, with the support of qualitative and quantitative structural data extracted from μ CT analysis.

Experimental

Bamboo sampling

Bamboo specimens consisted of cylinders of 1 cm in height and 0.6 cm in diameter, extracted from internode number 3 closer to the ground. The sampling process is illustrated in Fig. 1. Considering the radial direction from the outer to the inner wall of the internode, we cut specimens at the external (close to the bark), intermediate and internal parts. The bamboo samples will be referred to as External for the outermost fraction, Middle for the intermediate fraction, and Internal for the innermost fraction. To eliminate organic residues, samples were washed under magnetic stirring in distilled water at 60°C for two hours. The samples were dried in an oven at 100°C until constant mass.

Probe fluids

Initially, we selected a wide range of organic and inorganic fluids to serve as a relaxation probe in the experiments. However, due to issues such as high volatility (methanol, ethanol, chloroform,

tetrahydrofuran), which could prevent some of the lengthier experiments, we finally chose two solvents with high boiling points. Distilled water (H₂O) (resistivity of 18 MΩ, filtered through a 0.22 μm mesh membrane, Milli-Q Gradient System A10, Millipore, USA) and dimethyl sulfoxide (DMSO) (ACS Reagent, Sigma-Aldrich, St. Louis, USA) were selected due to their low volatility, varying polarity values and different capacity for establishing physicochemical interactions with the lignocellulosic matrix of the bamboo plant.

Probe fluids saturation of the bamboo sample fractions

We conducted the saturation of the bamboo samples (External, Middle, and Internal) with each probe fluid using a vacuum chamber (Fig. 2). First, the samples were oven-dried at 100 °C until constant mass. Then, they were submerged in a given fluid volume (usually 5 mL of DMSO or H₂O) inside a Schlenk flask connected to a vacuum line. At this point, the samples remained afloat in the liquid. The system remained under reduced pressure (760 mmHg) for two hours. When the vacuum was broken, the samples sank into the fluid, suggesting extensive filling of their structural cavities. The samples were then surface-dried with absorbent paper, weighted again, and taken to the proton time-domain nuclear magnetic resonance equipment for analysis. The samples were taken to an oven at 100°C for drying to constant mass before changing the type of probe fluid.

X-ray microtomography (μCT)

The 3D images of neat bamboo samples were obtained using a Zeiss Xradia 510 Versa microtomograph to visualize and quantify the 3D structure. The experimental parameters for all samples can be found in Table 1.

Table 1 X-ray microtomography (μCT) experimental parameters.	
Objective Lens	4X
Pixel Size (μm)	4.49
Voltage (kV)	50
Power (W)	4
Exposure time (s)	1.5
Number of projections	1601

The images were processed using the CTan software (version 1.18.8.0, Bruker, 2018) and Avizo© software (version 9.0.1, FEI, 2018). Image pre-processing using a non-local means filter was used to improve the signal-to-noise ratio. Smaller regions of interest were delimited around the sample's image to

remove the background and reduce computational requirements. Watershed segmentation (ThermoFisher Scientific 2018) as used to automatically separate the structures of interest (sclerenchyma, parenchyma, and vascular channels). Segmented images were then used for quantification.

¹H time-domain nuclear magnetic resonance (TD-NMR)

All measurements were performed on a MARAN Ultra (Oxford Instruments®) low-resolution NMR spectrometer operating at 0.54 T (23 MHz, ¹H Larmor frequency) equipped with an 18 mm diameter probe at 30 ± 1°C. The 90° (p90) RF pulse was calibrated to 7.5 µs, and the 180° (p180) pulse was calibrated to 15 µs. Coil dead time and filter stabilization time were set to 6.0 and 3.0 µs, respectively.

We employed the CPMG (Carr-Purcell-Meiboom-Gill) pulse sequence with 2048 echoes, with a time between echoes (2τ) of 600 µs. We carefully chose this interval between echoes to exclude the bulk of the relaxation signal from the lignocellulosic matrix's protons. The variation in the transverse relaxation time (T_{2*}) of the fluids' molecules as a function of the bamboo's microcavities saturation level and morphology works as the primary investigation probe.

For measurements of the fluids' molecular dynamics in non-dynamic conditions, the samples saturated with each fluid were inserted in the TD-NMR tubes, and CPMG curves were acquired and stored for later processing.

For the fluid desorption experiment, a given sample was filled with H₂O with the aid of the vacuum chamber, then superficially dried with a paper towel, and then left inside an open TD-NMR tube at 30°C. The equipment was then set to acquire and store a CPMG curve immediately after the sample insertion and then at 2 hours intervals for five days (120 h).

Data processing was conducted with the aid of commercial software. For the Inverse Laplace Transform (ILT), we employed WinDXP® Toolbox 1.8.1.0 (Resonance Instruments®), with a smoothing parameter set at 2×10⁻⁴ and tolerance at 10⁻⁷. A maximum of 50 iterations was allowed. No point in the relaxation curve was discriminated, and all distribution curves were constructed with 256 points between 10 µs and 10 s. All curve fitting procedures were made in Origin 8.5 (OriginLab®).

The Solid-Echo sequence (SEPS) was used to evaluate the influence of the presence of the fluid inside the polymer matrix of the inner walls of the bamboo sample. The decay signal was acquired with 3072 points spaced by 1 µs, with a receiver gain of 40 dB and a recycle delay of 2 s. The obtained FID signal was adjusted according to the following function:

$$A(t) = F_R \exp \left[- \left(\frac{-t}{T_{2*}^R} \right)^2 \right] + F_{SR} \exp \left[- \left(\frac{-t}{T_{2*}^{SR}} \right) \right] + F_M \exp \left[- \left(\frac{-t}{T_{2*}^M} \right) \right] + k(1)$$

where, $A(t)$ is the total amplitude of the FID signal, F_R is attributed to highly rigid molecular fractions, F_{SR} is associated with molecular fractions with intermediate molecular mobility and F_M is the term

related to highly mobile molecular fractions. The different T_{2*} in Eq. 1 are related to the transverse relaxation time of each fraction, and k is an offset parameter. Physical interpretations of the fractions and their respective relaxation times are given in the discussion section.

From the decomposition of the signal into the respective components, the average residual dipole coupling $\langle M_2 \rangle$ was extracted from the adjustment of the component Fr with the function below:

$$G(t) = \exp\left(-\frac{1}{2M_2t^2}\right) \quad (2)$$

Furthermore, the theoretical background for attributions of the behavior of liquid molecules in confined spaces such as pores, voids, and channels is given below. The ILT can be applied to the NMR decay signal to detect and discriminate the relaxation behavior of water protons in different families of pore sizes. The transverse relaxation rate of a liquid within porous materials is governed by the rates of free solvent relaxation (T_{2F}), surface relaxation (T_{2S}), and diffusion resulting from molecular diffusion (T_{2D}) (Zhang et al. 2016):

$$\frac{1}{T_2} = \frac{1}{T_{2F}} + \frac{1}{T_{2S}} + \frac{1}{T_{2D}} \quad (3)$$

Considering that $T_{2F} \gg T_{2S}$ and a system within a fast diffusion limit, the equation can be approximated to (Al-Mahrooqi et al. 2006; Capitani et al. 2012):

$$\frac{1}{T_2} = \frac{1}{T_{2S}} = \rho \frac{S}{V} \quad (4)$$

where S is the pore surface area, V is the pore volume, and ρ is the relaxivity constant of the confined fluid. For a given ρ , as $\frac{S}{V}$ is proportional to $\frac{1}{R}$ for a sphere, one sees that the transverse relaxation rate is directly proportional to the pore size.

Results And Discussion

The highly anisotropic 3D architecture of the bamboo vegetal tissues was investigated by μ CT, as shown in Fig. 3. The variation and spatial distribution of the different biological tissues were analyzed through segmentation and post-processing μ CT images. Sclerenchyma tissue with its lignin-rich fibers (green, Fig. 3b) presents a variation in morphology and density along the radial direction from the outer to the inner wall. Accordingly, the diameters of longitudinally-oriented vascular channels (blue, Fig. 3d) imbibed in the sclerenchyma tissue varied from 121 ± 27 to 192 ± 32 μ m. Here it is helpful to note that we call the plant's phloem, xylem, and metaxylem channels collectively "vascular bundle" and "vascular channels." The parenchyma (yellow, Fig. 3c) is distributed throughout the volume of the sample. The volume occupied by parenchyma tissue cells and vascular channels in bamboo samples from each fraction is described in Table 2. Together, the vascular channels and the living cells of the parenchyma represent

microcavities of different sizes and morphologies and are the focus of the present work. These structures play fundamental roles in the growth, mechanical strength, transportation, and storage of organic and inorganic materials needed to maintain vegetal homeostasis.

Table 2

Volumes occupied by the microcavities in each fraction of the bamboo sample and diameter of the structures with voids in the plant tissues as obtained via μ CT

Sample fraction	Vascular channels volume (mm ³)	Parenchyma cortex volume (mm ³)	Vascular bundle diameter (μm)	Parenchyma cells diameter (μm)
Internal	3.25	14.14	192 ± 32	38 ± 10
Middle	3.01	11.76	175 ± 39	33 ± 7
External	2.20	8.03	121 ± 27	26 ± 5

The cells of the parenchyma cortex and the fibrous bundles of the sclerenchyma, where the vascular channels are found, increase in size from the external to the internal fraction (Pandoli et al. 2016). This change directly affects the total fluid's mass that a given fraction of comparable dimensions can hold when fully saturated, as described in Table 3.

Table 3

Fluid's total mass in saturated bamboo samples

Sample fraction	H ₂ O mass (g)*	DMSO mass (g)*
Internal	0.1412 ± 1.5 × 10 ⁻⁴	0.1024 ± 1.1 × 10 ⁻⁴
Middle	0.1169 ± 1.1 × 10 ⁻⁴	0.0867 ± 5.7 × 10 ⁻⁵
External	0.0686 ± 5.8 × 10 ⁻⁵	0.0502 ± 5.8 × 10 ⁻⁵

*- Masses are reported as the mean of triplicates with standard deviations.

The discrepancy found in the masses of H₂O and DMSO in the saturated samples can be attributed to the different wettability capacities of the molecular structure of bamboo tissues composed of cellulose, hemicellulose, and lignin. The filling fluids used in this study differ in molecular mass, polarity, and the number of hydrogen bonds they can establish, with H₂O having a greater physicochemical affinity for the bamboo plant structure (Reichardt e Welton 2010).

The observation of cross sections obtained from μ CT images (Fig. 4) of the three fractions indicates varying degrees of contact between vascular channels and parenchyma cells. The cells' wall of the parenchyma cortex is populated by pit holes, cell-wall openings hundreds of nanometers in diameter, responsible for exchanging materials in the living plant. In the case of internal and middle fractions, the

parenchyma cells present extensive contact with the vascular channels. In the external fraction, the extent of the contact between the vascular channels and the parenchyma is smaller than in other fractions. In addition, the external fraction exhibits more massive fibrous bundles, unlike the innermost fractions of bamboo.

TD-NMR is very useful in probing the molecular mobility of a liquid phase embedded in a solid matrix. It can provide very accurate data about the kind of environment molecules in a liquid are occupying, revealing different degrees of confinement. In Fig. 5a, we have the domain curves, obtained by the CPMG pulse sequence, of water molecules in four different environments. The free water curve corresponds to the behavior expected from a fluid when it is free from spatial confinement; that is, most of its molecules can undergo free rotation and diffusion processes. This behavior translates into a narrow and well-defined domain curve, with an average relaxation time value close to 3 s for water. A fluid confined in a solid matrix suffers the effects of interaction between its molecules and the material wall (Al-Mahrooqi et al. 2006). These effects interfere with the magnetic relaxation of the fluid molecules, widening the bases of the domain curves and significantly reducing the relaxation time values, as seen in Fig. 5.

The area under a relaxation domain curve can be related to the number of protons that relax in that time interval. We observed that the water molecules confined in bamboo cavities have at least two large relaxation domains: one at shorter times (tens of milliseconds) and the other at longer times (around 100 milliseconds). Relaxation time values are inversely proportional to the confinement experienced by the fluid molecules (De Sá et al., 2022). Therefore, we can attribute the shortest relaxation times to the water molecules occupying the parenchyma cavities (left side) and the longest times to the water molecules occupying the vascular channels (right side).

There is a clear difference in the populations of fluid molecules in each of the bamboo fractions. In the internal fraction, the largest population is found inside the vascular channels; in the middle fraction, the population relative to water in the parenchyma starts to increase and, finally, in the external fraction, the amount of water contained in the parenchyma is slightly larger than that inside the vascular channels. Therefore, the importance of choosing which region of the plant becomes evident when technological applications of the material are envisioned. The results show that each fraction has a distinctive way of distributing fluid throughout its volume (Table 4).

Table 4
Distribution of H₂O and DMSO molecules in each saturated bamboo fraction

Sample	Parenchyma cortex (%)		Vascular channels (%)	
	H ₂ O	DMSO	H ₂ O	DMSO
Internal fraction	36	27	64	73
Middle fraction	42	31	58	69
External fraction	52	38	48	62

From the distributions of transverse relaxation domains, two relaxation domains are observed, both for the water-saturated bamboo and the DMSO-saturated bamboo.

The domain with the lowest T_2 value (approximately 2×10^{-3} s) is related to the solvent present in more confined spaces of the bamboo matrix, such as the parenchyma and sclerenchyma and the internal channels that connect the different regions of the bamboo. In addition, there is the fraction of the solvent on the surface of the inner walls of the vascular channels, where the effect of surface relaxation is more pronounced, which promotes the reduction of the T_2 value compared to the free solvent (bulk) molecules.

The domain with the highest T_2 value (approximately 0.1 s) is related to the solvent present in the vascular channels, more specifically in their central region, away from the internal walls. In this region's cross-section, the solvent molecules can be found in an unimpeded environment, translating to a relaxation regimen closer to their bulk state.

Similar trends are observed when we compare the internal, intermediate, and external bamboo fractions distribution for the water- and DMSO-logged samples. The domain with the lowest T_2 value shifts to the shorter relaxation times for the outer fraction of the bamboo. However, the domain with the highest T_2 value shifts to the right, increasing its value for the outermost fraction of the bamboo. The shortening of the relaxation times corroborates the data obtained from microtomography. As noted, parenchyma and sclerenchyma have the smallest pore sizes, which become even smaller as we advance from the inside to the outside of the bamboo. This higher confinement of the fluid translates into a reduction in the value of T_2 .

The increase in the T_2 value for the domain around 0.1 s can be explained by the more significant number of bundles of vascular channels present in the outermost fraction, as highlighted in Fig. 3. Although the diameters of the bundles are smaller in the outermost region of the bamboo, the channels have three-dimensional cylindrical structures, and each cylinder has a central region in which the fluids' molecules can relax their magnetization with less encumbrance; that is, they do not experience the effect of surface relaxation of the channel walls. Since the outermost fraction has more vascular channels, the amount of water in the bulk state is larger than in the innermost fraction of the bamboo. This, in turn, translates into an increase in the T_2 value for the second domain shown in Fig. 5.

The difference in distribution profiles between water and DMSO should be highlighted. While the first solvent has narrower and more well-defined domains, the transverse relaxation distribution for DMSO shows a broader profile. From the presented data, we can infer that, while the bamboo tissues' water-saturation capabilities are extensive, as expected, the DMSO molecules can permeate with ease through the plant matrix. This information may be relevant when choosing the best solvent in a microfluidics process.

The Solid-echo pulse sequence (SEPS) was employed to investigate the effects of the saturation of water or DMSO (Fig. 6) molecules on the structure of the bamboo matrix. Solvents of different polarities act on

the lignocellulosic chains. These observations seek the best performance of bamboo in microfluidics operations.

The FID measurements obtained by SEPS reveal three well-defined regions in the water-saturated (Fig. 6, on left side) or DMSO-saturated bamboo structure (Fig. 6 on the right side). The rigid fraction (F_r) is attributed to the sclerenchyma (crystalline cellulose), the semi-rigid fraction (F_{sr}) is associated with the parenchyma, and the high mobility fraction (F_m) refers to the inner walls of the vascular bundles. These vessels contain the solvent molecules relaxing closest to the bulk state, with high T_{2*} values.

Relaxation signals for dry bamboo were obtained to evaluate the effect of the presence of water or DMSO on the three (F_r , F_{sr} , and F_m) relaxation regions of the bamboo. In the absence of solvent molecules, the bamboo polymeric matrix displays rigidity, as observed in the fast decay regimen of the NMR signal (Fig. 6, middle column). The signal was fitted by a Gaussian function with a transverse relaxation time of about 10 μ s and a residual dipole coupling (M_2) close to 9000 $\text{rad}^2.\text{ms}^{-2}$.

As expected, the presence of liquid molecules with chemical affinity with the natural polymer matrix increases the molecular mobility of the lignocellulosic structures constituting the samples, leading to the observed increase in the T_{2*} values. Similar behavior was observed in the magnetic relaxation of wood samples saturated with different fluids (Zhang et al. 2016).

For the samples saturated with water, a rising trend in the F_r contribution to the total magnetization in each sample is observed when we move from inner to outer fractions of the sample (35 to 55% of F_r). As seen in the μ CT results (Figs. 3 and 4), the external fraction contains larger quantities of sclerenchyma, which displays very little permeability. The percentage of F_{sr} remains higher than F_m since the parenchyma volume is larger than the vascular bundles in the samples, as seen in Table 02.

Comparing the values of M_2 for the dry ($M_{2\text{average}} = 9241 \text{ rad}^2.\text{ms}^{-2}$) and water-saturated bamboo samples ($M_{2\text{average}} = 8159 \text{ rad}^2.\text{ms}^{-2}$) for each of the sampling fractions, the internal fraction shows a reduction of 16% in this parameter, the highest variation in all three sampling fractions. This reduction reflects a small volume of sclerenchyma tissues (Table 4), implying a higher water-soaking ability. Furthermore, the analysis of the M_2 values informs that solvation of the sclerenchyma with water and DMSO molecules has happened. We tentatively attribute this to the micro and nanometric pits interconnecting vascular bundles and sclerenchyma tissues, allowing for solvent flow between these regions.

The bamboo samples saturated with DMSO also display three well-defined regions, as described above. The F_r values (from 47 to 65%) are higher when compared to the results obtained with distilled water (from 35 to 55%). This feature, associated with the higher values of M_2 , suggests that DMSO has lower permeability in the sclerenchyma than water. We attribute the rise in the F_r percentage from 47% (internal) to 65% (external) to the higher volume of sclerenchyma tissue in the external fraction.

Another important aspect is the T_{2*} values for the region of high mobility (F_m) of the bamboo samples saturated with DMSO ($T_{2*average}=1563$ s), which are higher than those obtained with the water ($T_{2*average}=1089$ s). This suggests that DMSO molecules tend to occupy, preferably, the vascular bundle.

Therefore, it is observed that ultrapure water, as expected, has a greater capacity to diffuse through the bamboo's internal structures, mainly due to polarity and molecular size and the ability to perform hydrogen bonds. Both solvents remain primarily in the parenchyma and vascular channels, but water also has a small capacity to permeate the sclerenchyma.

The evolution of the magnetic relaxation domain curves in TD-NMR as a function of time can be represented through contour surface maps (Rodrigues et al. 2018). The intensities and relaxation time values of the domains at each moment represent the dynamics of the water evaporating inside the bamboo cavities as a function of time. The water evaporation from bamboo samples was monitored in real-time by TD-NMR to describe the behavior and distribution of the water molecules inside the bamboo microcavities. Figure 7 shows the results obtained. Initially, 120 h of process monitoring was recorded, but from 72 h onwards, the remaining amount of water becomes very small, and the signal-to-noise ratio greatly decreases. This compromises the quality of the Inverse Laplace Transform that generates the domain curves, which significantly reduces the reliability of the results.

A qualitative analysis of each contour surface map shows that the behavior of confined water during evaporation is different for each bamboo fraction. There are two large populations of water molecules represented by the two trails on the map: one at longer relaxation times relative to water in vascular channels and one at shorter relaxation times relative to the environment of the parenchyma cortex. The coalescence of the confined water molecules' transverse relaxation domains progressively increase from the outer to the inner fraction, again indicating the existence of varying degrees of connection between the two cavities in the tissue. We propose that this behavior becomes apparent in TD-NMR experiments from a certain threshold of water saturation, such as those used in this work.

Previous experiments with wood samples containing varying amounts of water did not report the present findings, as they operated inside a framework of water vapor saturation or natural humidity present in their samples (Mikac et al. 2021; Cocusse et al. 2022; Li and Ma 2022). The extent of communication between the parenchyma cavities and the vascular channels became evident during the desorption experiment. The population of water molecules in vascular channels disappears after 48 h in the three fractions, albeit at different evaporation rates for each fraction. The water present in the smaller cavities of the parenchyma does not disappear completely, although the amplitude of these domains decreased considerably compared to the beginning of the evaporation process. It has been demonstrated that a residual amount of water is retained in lignocellulosic materials due to hydrogen interactions with the cellulose chains in plant tissue (Zhang et al. 2016; Li et al. 2017c).

Both the coalescence of the relaxation domains of water molecules and the time until the amplitude of these domains is considerably reduced by evaporation substantiate the results of μ CT. Previous studies

have shown that the interconnected pore structure constitutes a highly specialized fluid transportation system (Liu et al. 2021). The efficiency of this system is evident when we examine the behavior of the confined water volume in the samples of the internal and middle fractions. In highly confined environments, the trapped water molecules have their evaporation rate delayed at temperatures close to room temperature, like those used in our experiments, which may explain the remaining amplitude of the observed relaxation domains even though it has long test times. We demonstrated that the efficiency of fluid exchange between the two types of microcavities (vascular bundles and parenchyma cells) becomes a relevant factor affecting the fluid's evaporation rate.

Conclusions

μ CT and TD-NMR showed great potential for analysis of the internal structure of bamboo, as well as the distribution of solvents of different polarities and their effects on the molecular mobility of the fully solvent-saturated bamboo structure. The μ CT results supported the reasoning used in the interpretation of the TD-NMR data. Most of the interpretations of the behavior of the molecular dynamics of the fluids confined in the bamboo structure were only possible with the aid of this technique's qualitative and quantitative information.

The CPMG sequence, through the values of T_2 and the area under the distribution of domains, can map the distribution of solvents in the parenchyma and vascular channels, being sensitive to the morphology changes as a function of the bamboo cut region (internal, intermediate, or external), corroborating the data obtained by microtomography. Ultrapure water and DMSO showed similar distribution profiles. Monitoring water desorption indicates that it takes 40 to 48 hours for almost all of the solvent to leave the bamboo matrix, leaving only a small fraction confined in the parenchyma.

The Solid Echo sequence, through the T_{2*} values and the percentages of rigid, semi-rigid, and mobile fractions, was sensitive to the mobility variations of the lignocellulosic chain segments in the presence of ultrapure water or DMSO. The results indicate that water has a greater permeation capacity and affects the rigid fraction of bamboo more, while DMSO settles more easily in vascular channels.

We believe that the data found in this work may be fundamental in choosing the most suitable solvent for a particular type of bamboo during a microfluidic process, as well as its effects on the structure of the segments of the bamboo chain.

In general, the results indicate dependence between the physicochemical characteristics of the fluids and lignocellulose-based microfluidic reactors with hierarchical architecture, such as bamboo. Studies with other fluids and ionic liquids are being conducted by our group to understand better the molecular behavior in confined environments made up of natural polymers.

Declarations

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Author contributions. Conceptualization: OGP, EJRR, Data Curation: OGP, EJRR, RPCN, HDLA, Funding acquisition: OGP, SP, KG, Investigation: OGP, EJRR, RPCN, HDLA, DSS, Project administration: OGP, SP, KG, Supervision: OGP, SP, KG, RPCN, Validation: OGP, EJRR, RPCN, Visualization: EJRR, RPCN, HDLA, DSS, Writing original draft: OGP, EJRR, RPCN, DSS, Writing-review and editing: OGP, EJRR, RPCN, DSS, SP, KG.

Conflict of interest. There are no conflicts of interest/competing interests.

Data availability. All data generated or analyzed during this study are included in this published article.

Ethical approval No results of studies involving humans or animals are reported.

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Figures

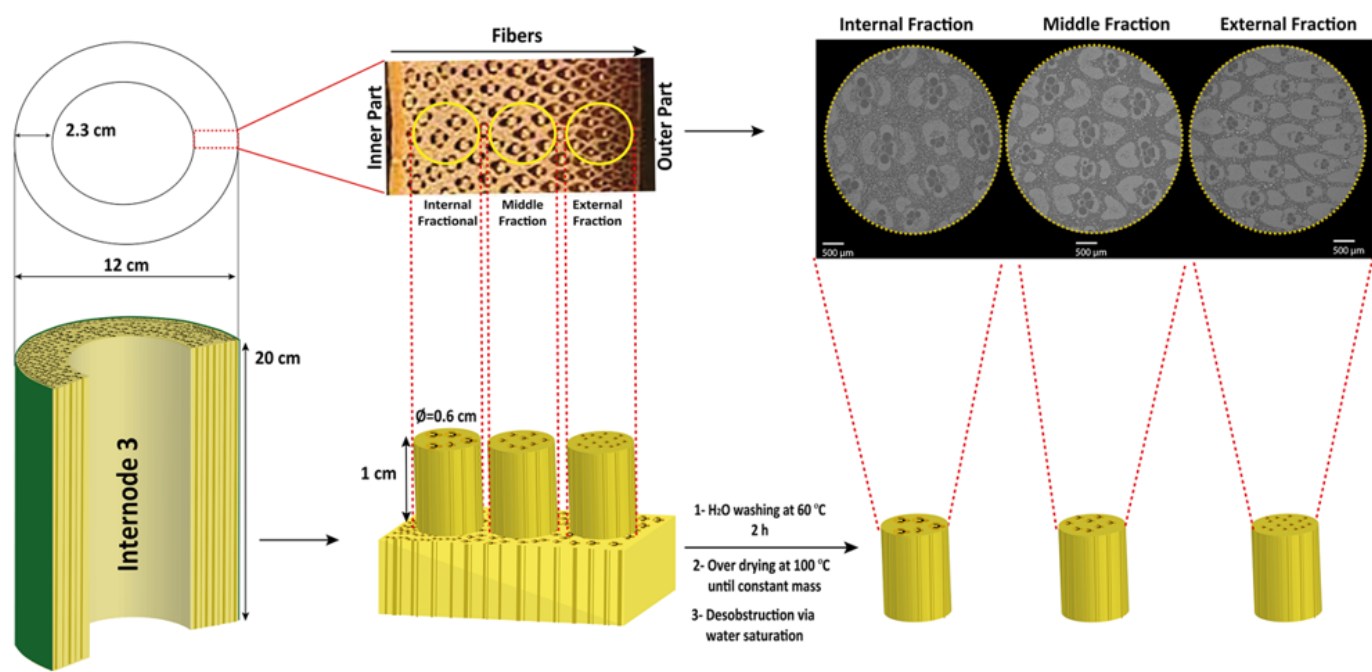


Figure 1

Bamboo sampling, conditioning, washing, and drying steps.

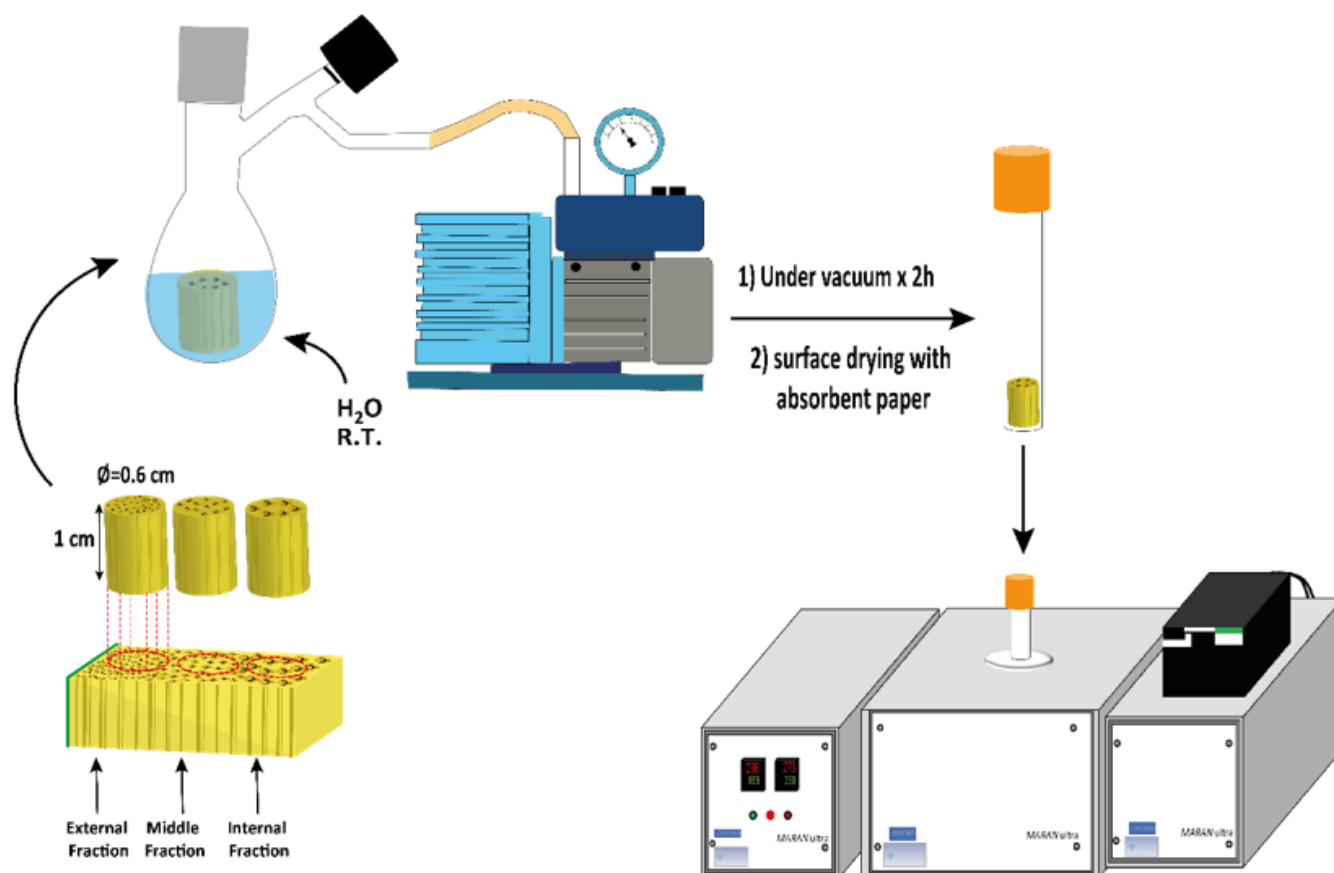


Figure 2

Distilled water saturation of the bamboo fractions for TD-NMR analysis.

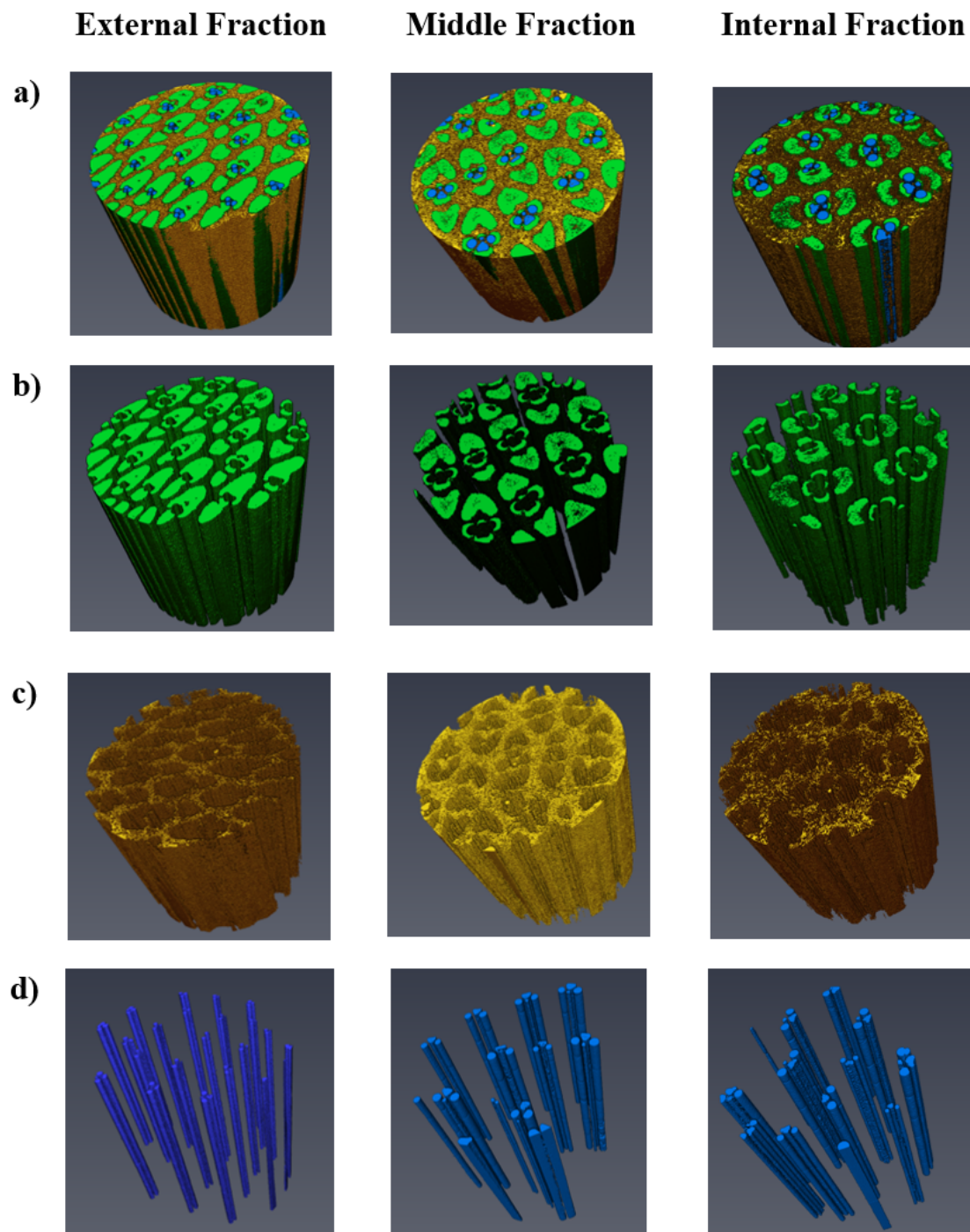
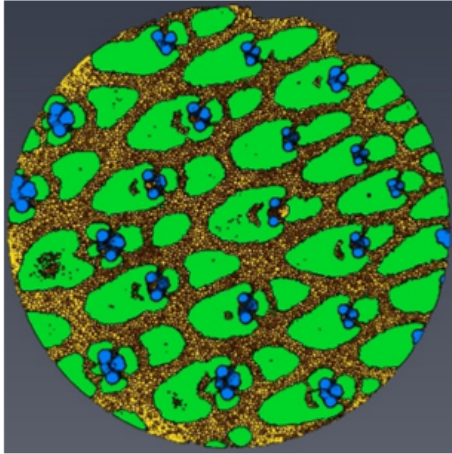
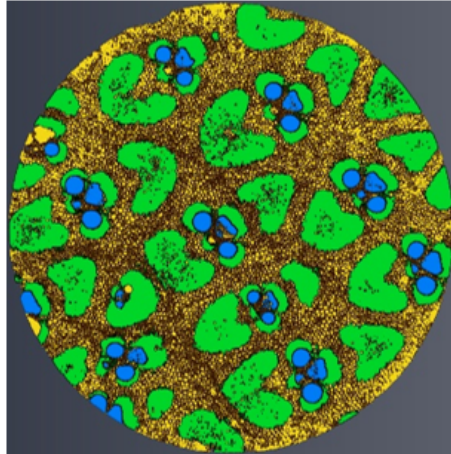
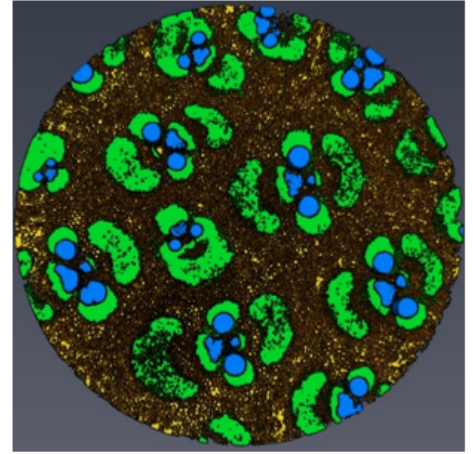


Figure 3

μ CT 3D images of the fractions sampled from the bamboo internode. In each group, the following structures are displayed: a) the whole sample; b) the sclerenchyma; c) the parenchyma and; d) the bundles of vascular channels.

External Fraction**Middle Fraction****Internal Fraction****Figure 4**

μ CT images of the transverse section of each bamboo sample fraction. Several heterogeneities in the volume and spatial distribution of the fluid-holding compartments can be observed. The vascular channels are shown in blue, and the parenchyma is shown in yellow. Furthermore, the fibrous shafts of the sclerenchyma (shown in green) display an increasing number of hollow sections from the External to the Internal fraction.

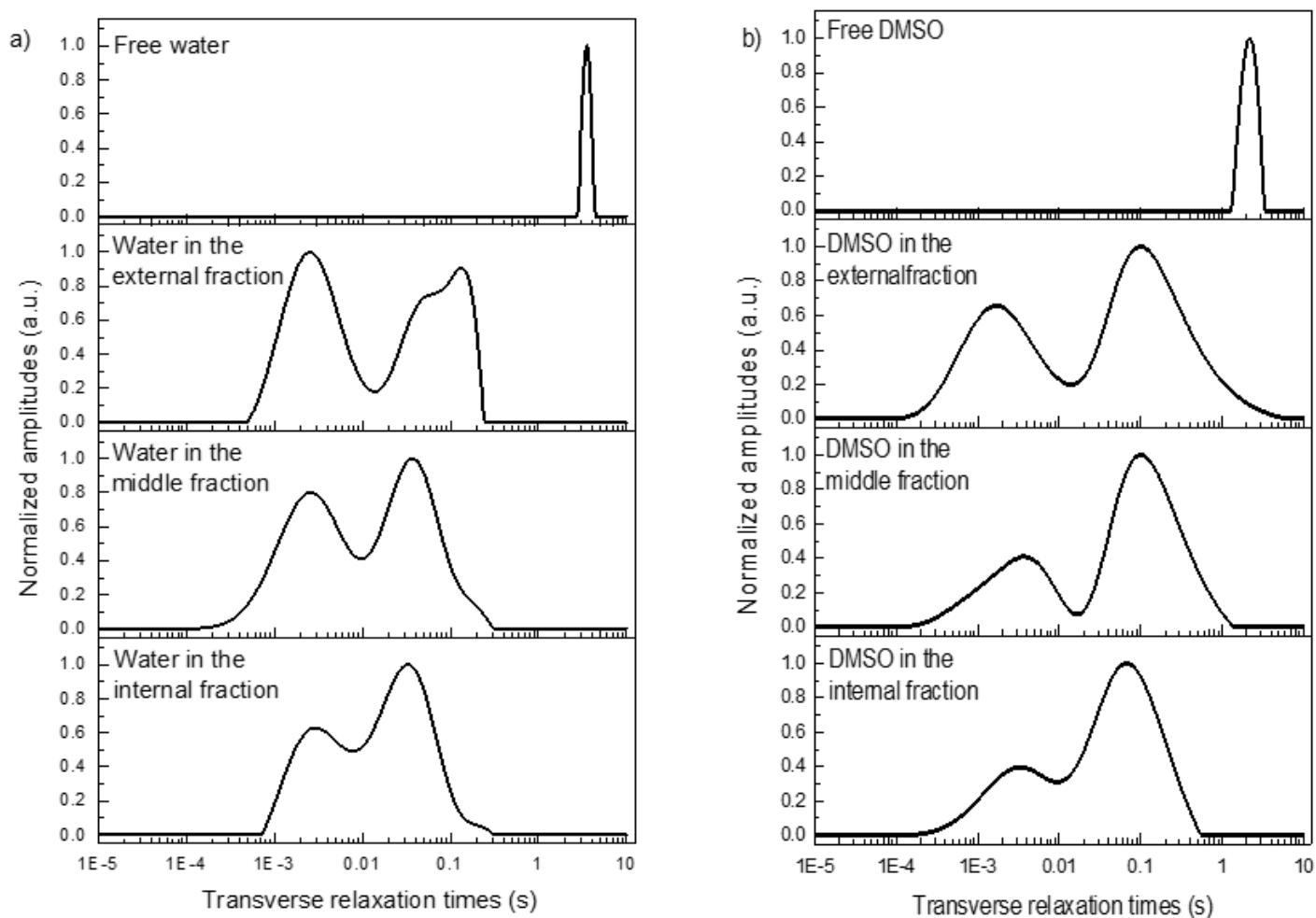


Figure 5

Relaxation distribution curves, obtained via ILT on CPMG data, displaying the effect of physical confinement on water (a) and DMSO (b) molecules.

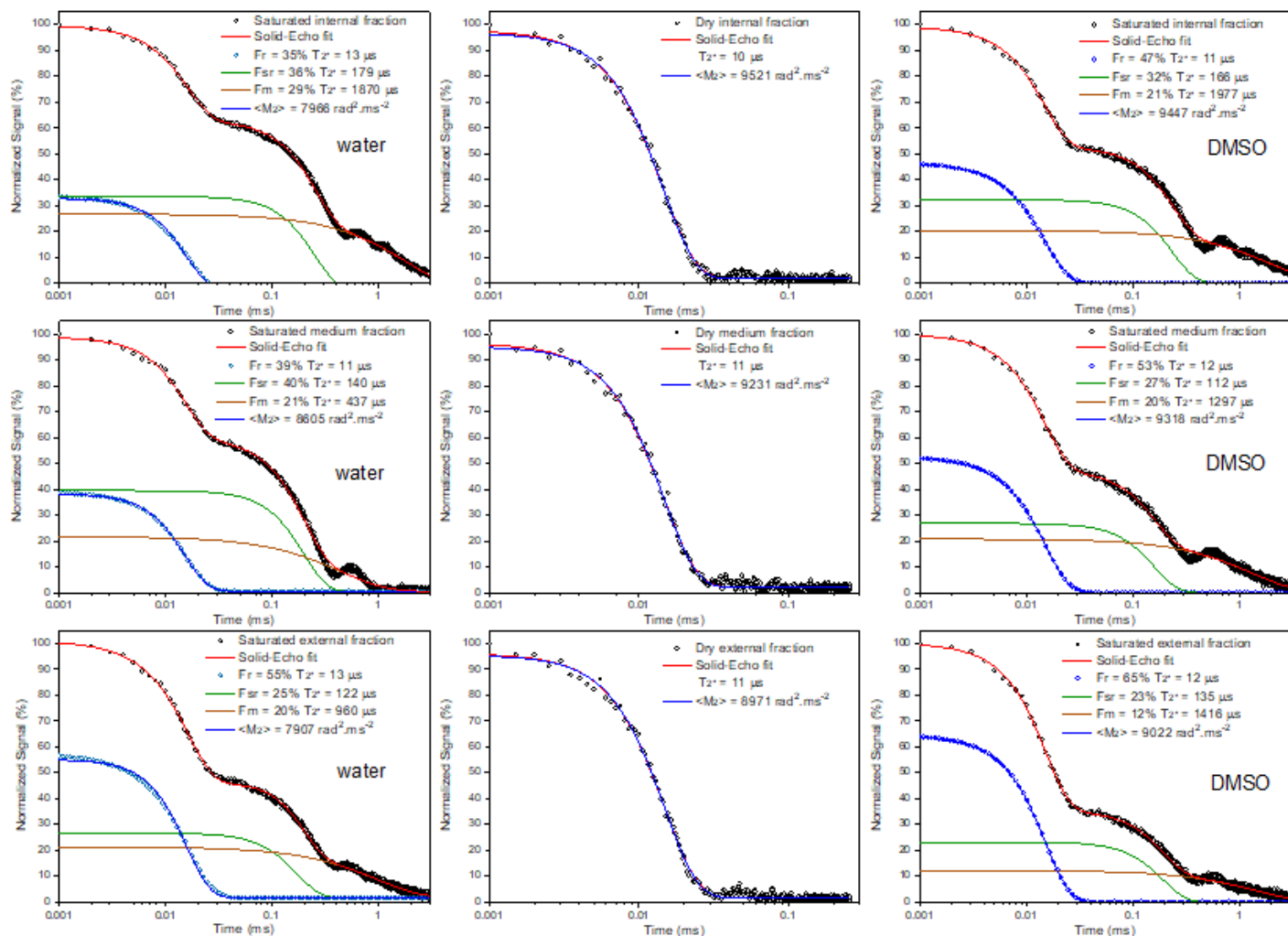


Figure 6

Obtained SEPS NMR signal intensities fitted to Equation 1 (signal components) and Equation 2 (M_2 values) for the samples saturated with distilled H_2O (left) and DMSO (right). The middle column refers to the dry bamboo samples.

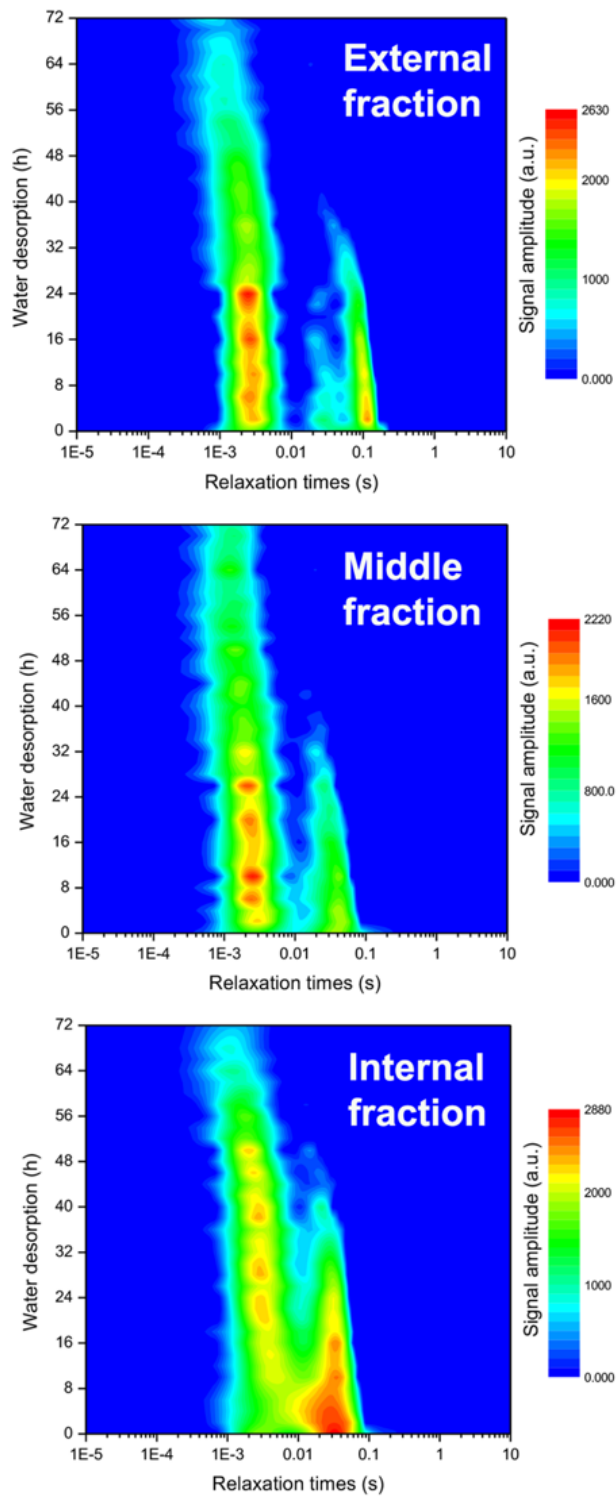


Figure 7

Surface contour plots of the 72h long water desorption process are monitored in real time via the relaxation of water molecules.

Supplementary Files

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