



***Molinia caerulea* alters forest *Quercus petraea* seedling growth through reduced mycorrhization**

Marine Fernandez, Philippe Malagoli, Lucie Vincenot, Antoine Vernay, Thierry Ameglio, Philippe Balandier

► To cite this version:

Marine Fernandez, Philippe Malagoli, Lucie Vincenot, Antoine Vernay, Thierry Ameglio, et al.. *Molinia caerulea* alters forest *Quercus petraea* seedling growth through reduced mycorrhization. *AoB Plants*, 2022, 15 (2), plac043. <10.1093/aobpla/plac043>. <hal-03977500>

HAL Id: hal-03977500

<https://hal.science/hal-03977500v1>

Submitted on 28 Feb 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons CC BY 4.0 - Attribution - International License

Studies

Molinia caerulea alters forest *Quercus petraea* seedling growth through reduced mycorrhization

Marine Fernandez¹, Philippe Malagoli^{1,*}, Lucie Vincenot², Antoine Vernay³, Thierry Améglio¹ and Philippe Balandier¹

¹Université Clermont Auvergne, INRAE, PIAF, F-63000 Clermont-Ferrand, France

²Normandie Univ, UNIROUEN, Laboratoire ECODIV USC INRAE 1499, 76000 Rouen, France

³Univ Lyon, Université Claude Bernard Lyon 1, CNRS, ENTPE, UMR 5023 LEHNA, F-69622 Villeurbanne, France

*Corresponding author's e-mail address: philippe.malagoli@uca.fr

Populations & Communities. Chief Editor: Jean Burns

Associate Editor: Adam Martin

Abstract

Oak regeneration is jeopardized by purple moor grass, a well-known competitive perennial grass in the temperate forests of Western Europe. Below-ground interactions regarding resource acquisition and interference have been demonstrated and have led to new questions about the negative impact of purple moor grass on ectomycorrhizal colonization. The objective was to examine the effects of moor grass on root system size and ectomycorrhization rate of oak seedlings as well as consequences on nitrogen (N) content in oak and soil. Oak seedlings and moor grass tufts were planted together or separately in pots under semi-controlled conditions (irrigated and natural light) and harvested 1 year after planting. Biomass, N content in shoot and root in oak and moor grass as well as number of lateral roots and ectomycorrhizal rate in oak were measured. Biomass in both oak shoot and root was reduced when planting with moor grass. Concurrently, oak lateral roots number and ectomycorrhization rate decreased, along with a reduction in N content in mixed-grown oak. An interference mechanism of moor grass is affecting oak seedlings performance through reduction in oak lateral roots number and its ectomycorrhization, observed in conjunction with a lower growth and N content in oak. By altering both oak roots and mycorrhizas, moor grass appears to be a species with a high allelopathic potential. More broadly, these results show the complexity of interspecific interactions that involve various ecological processes involving the soil microbial community and need to be explored *in situ*.

Keywords: Ectomycorrhization rate; fungal symbionts; interaction; lateral root; pot experiment.

Introduction

Competition from common perennial grasses has long been known to restrict tree seedling growth and survival during regeneration, whether natural or artificial (i.e. plantation) (Davis *et al.* 1998). Grass species can rapidly colonize large volumes of soil with their fast-growing fasciculate root systems (Freschet *et al.* 2017) and their efficient nutrient uptake (Coll *et al.* 2004; Picon-Cochard *et al.* 2006). Colonization by grasses depletes resources in forest soil, a process termed competition by resource exploitation (Bleasdale 1960). Competition for soil resources such as water and inorganic nitrogen (N) can stunt tree seedling growth and increase mortality rate (Gordon *et al.* 1989; Coll *et al.* 2003; Balandier *et al.* 2006), thus jeopardizing forest regeneration.

Competition detrimental to tree seedlings can also be mediated by interference. Interference results in the negative effect of a plant A on a plant B not by affecting the quantity of a resource, but by affecting the ability of plant B to access that non-limiting resource (Holdridge *et al.* 2016). For instance, tree seedlings root systems are found to be smaller in biomass

and length due to the presence of graminoids (Harmer and Robertson 2003; Collet *et al.* 2006) and it has been demonstrated that, allelochemicals exuded by graminoids can cause neighbour trees growth inhibition (Fernandez *et al.* 2021). Such negative effects of graminoids on tree seedling root systems can affect water and nutrient uptake ability. This is the case in oak seedlings exhibiting a decreased concentration in both shoot and root resulting in a reduced N content when grown with annual and perennial graminoids like *Stipa pulchra*, *Avena barbata* or *Deschampsia cespitosa* (Welker *et al.* 1991; Cheng and Bledsoe 2004; Vernay *et al.* 2018a, b).

Below-ground competition by interference can also involve alteration of interactions of tree seedling roots with other organisms such as symbiotic fungi. Many studies have demonstrated that mycorrhizal symbiosis can favour tree seedling establishment, growth and productivity (Lu *et al.* 1998; Simard and Durall 2004; Javaid 2007; Richard *et al.* 2009; Van Der Heijden and Horton 2009; Walker and Mallik 2009; Bonfante and Genre 2010). The role of mycorrhizae in N uptake by tree seedlings is well-investigated at physiological and

Received: 23 May 2022; Editorial decision: 12 August 2022; Accepted: 26 September 2022

© The Author(s) 2022. Published by Oxford University Press on behalf of the Annals of Botany Company.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by/4.0/>), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

molecular levels (Chalot and Brun 1998; Govindarajulu *et al.* 2005; Chalot and Plassard 2011). In natural ecosystems (e.g. low anthropogenic impacts forests), mycorrhizae can provide up to 80 % of tree seedling N requirements (Van Der Heijden *et al.* 2008). In this context, many studies have demonstrated that grass can exert an inhibitory effect on mycorrhizal growth and association with tree seedlings, particularly for conifer species (Handley 1963; Robinson 1972; Inderjit and Mallik 1999; Inderjit and Callaway 2003; Mallik 2003; Mallik *et al.* 2016).

For instance, Nilsson *et al.* (1993) showed both root system growth, mycorrhizal fungus mycelium diameter (*Paxillus involutus*) and N uptake in *Pinus sylvestris* were affected by the Ericaceae *Empetrum hermaphroditum*. However, molecules involved in this interaction and their targets were not identified. Moreover, considering that evergreen conifer and deciduous trees have different functional traits involved in soil resource capture (root extension rate, root mass fraction, for instance) and use (N use efficiency [NUE], N content, for instance) (Tomlinson *et al.* 2019), it appears necessary to increase the knowledge of mycorrhizae-mediated interactions between grasses and deciduous tree seedlings.

The nature of the mycorrhization process may also differ according to plant ecological strategies: acquisitive species, that promote fast growth, resource acquisition and use, and biomass turnover (Reich 2014; Fernandez *et al.* 2022), are more likely to be colonized by arbuscular mycorrhizae than by ectomycorrhizae (Craig *et al.* 2018). Thus, the inhibitory effect of grass may not harm the two functional groups of trees, evergreen and deciduous, in the same way. Additionally, mycorrhizae can also negatively impact plant growth when the cost of symbiosis outweighs the benefits (Johnson *et al.* 1997; Jones and Smith 2004). Näsholm *et al.* (2013) demonstrated that positive feedback between decreased soil N availability and increased mycorrhizal N immobilization through increased below-ground carbon allocation may exacerbate N depletion in soils.

Such a negative effect of ectomycorrhizal association was also supported by Alberton *et al.* (2007), showing competition for N increases under elevated CO₂. Other studies also explained that arbuscular mycorrhizal (AM) fungi are more likely to affect trees seedlings establishment than ectomycorrhizal fungi (EMF), especially in context of competition with grasses, due to common AM network existence between tree and grass (Alberton *et al.* 2007; Zangaro *et al.* 2018). The mycorrhization process therefore directly or indirectly, positively or negatively, affects plant dispersal and competition that shape plant populations and communities (Tedersoo *et al.* 2020).

Negative effects of a herbaceous plant on mycorrhization of a deciduous tree seedling were demonstrated by Timbal *et al.* (1990): They demonstrated neighbouring moor grass, *Molinia caerulea*, reduced red oak (*Quercus rubra*) growth 3- to 5-fold compared to sole-grown oak seedlings. They concomitantly found shifts in mycorrhizal biodiversity: biomass and/or proportion of efficient fungi (i.e. those involved in higher nutrient uptake ability) such as *Laccaria* sp. fell, while less efficient fungi such as *Cenococcum* sp. became more abundant. In some circumstances of low water or nutrient resources, the competition of *Molinia* on *Quercus*, especially *Quercus petraea*, can be problematic, compromising oak regeneration in Western Europe (Vernay *et al.* 2016). Our previous work has demonstrated strong competition for soil N in

favour of moor grass, but nothing is known about interfering interactions targeting mycorrhizae and more broadly on the complexity of bidirectional interactions between plants.

For many years, work on plants interactions has mainly focused on competition (Grime 1974; Tilman 1990), generally described as unidirectional, i.e. negative effect of plant A on plant B without considering the effect of plant B on plant A. Callaway (1995) points out that facilitative interactions have been less studied because they would be 'interesting but not convincing' in plant communities. However, facilitation has gained interest since ecologists have shown that it can become the primary interaction when stress increases to some extent (Bertness and Callaway 1994; Michalet *et al.* 2006). According to the stress-gradient hypothesis, competition may also turn into facilitation when environmental conditions change (Pugnaire and Luque 2001; Maestre *et al.* 2009). Facilitation can also occur simultaneously with resource competition or allelopathy (Callaway 1995; Bronstein 2009; Schöb *et al.* 2014a, b) underlining the interest of considering interactions as a bidirectional process (Schöb *et al.* 2015). Recent studies showed that sessile oak (*Q. petraea*) improved grasses growth, evidencing an antagonistic interaction where reduced oak growth was concomitant with improved grass development (Vernay *et al.* 2018a; Fernandez *et al.* 2020). In this context we aimed at identifying the effect of moor grass on oak seedling mycorrhization and its effects on oak N uptake and biomass.

We investigated the interactions between *M. caerulea* (purple moor grass) and *Q. petraea* (sessile oak). *Molinia caerulea* is a grass with one of the strongest soil N uptake abilities (Heil and Bruggink 1987; Aerts and Berendse 1988; Persson and Nasholm 2001; Vernay *et al.* 2016). Sessile oak is a common species widespread in European temperate forests and is economically important (Eaton *et al.* 2016). We sought to determine whether competition between oak seedlings and moor grass involved not only exploitation of resources such as N, but also interference mechanisms affecting oak root system size and reducing ectomycorrhizal colonization of oak. We specifically hypothesized that moor grass (i) has a detrimental effect on the number of oak seedling lateral roots, and (ii) depresses ectomycorrhizal colonization of roots, resulting in (iii) a decrease in N uptake ability.

Materials and Methods

Plant material and experimental design

The experiment was conducted in pots under outdoor conditions in Clermont-Ferrand (Auvergne, France, 45°45'N 3°07'E, altitude 394 m a.s.l.) from April 2018 to May 2019 (mean temperature, $T_{\text{mean}} = 12.9^{\circ}\text{C}$, annual rainfall = 546 mm). An experiment in pots rather than under natural conditions was chosen to avoid confounding effects such as water availability. In April 2018, 12 one-year-old bare-root sessile oak seedlings and 12 moor grass tufts were planted in plastic pots, either separately or together. Oak seedlings were sourced from a local nursery (Pépinières Naudet, Leuglay, France). They weighed 32 ± 8.2 g (mean $\pm$ SE) (fresh), and measured 52 ± 5.8 cm in height, and 6.8 ± 0.9 mm in diameter. Moor grass (fresh weight 2.2 ± 0.9 g) was collected in a local forest at Paray-le-Frésil (Auvergne, France; 46°39'N 3°36'E); shoots and roots were cut at 6 cm each with 2.04 ± 0.97 g fresh weight per lot.

A total of six 10-L pots and twelve (grouped in pairs) 5-L pots were filled with soil (typical luvisol-redoxisol pseudogley, sandy loam) collected in the same forest as the moor grass tufts. Natural forest soil was used to preserve natural composition of EMF community. Two treatments based on root system separation or interaction were set up: (i) two separate 5-L pots containing either one oak seedling or one moor grass tuft were placed side-by-side, precluding all root interactions ('sole-grown' treatment) and (ii) one oak and one moor grass tuft were placed in a shared 10-L pot to allow full below-ground interactions through root and hyphae contacts ('mixed-grown' treatment). Each treatment was replicated six times with a random spatial pot arrangement. To avoid interaction with water availability, pots were irrigated to field capacity throughout the experiment. To ensure the correct water amount was delivered, soil water content was continuously measured with TDR probes with a target soil water content maintained between 15 and 30 % [see [Supporting Information—Figure S1](#) and [Appendix S1](#)]. No fertilizer was added to the pots during the experiment.

Plant harvesting, lateral roots and ectomycorrhizal oak count

Plants were harvested in May 2019, after 13 months in pots to allow full mycorrhizal root colonization by fungi as well as interactions between plants. Shoots (stem + leaves together) and roots were collected for each plants. They were then dried at 60 °C for 48 h to measure shoot and root dry biomasses.

Immediately after root system harvest, approximately one-tenth of each oak total root system (fine and coarse roots) was stored in plastic bags at –20 °C for subsequent observation of lateral roots and occurrence of ectomycorrhizal root tips. For each treatment, 12 microscope slides were prepared, each slide harbouring ten 1-cm fragments of random primary fine roots (diameter < 2 mm) cut with a cutter was placed with water under a coverslip (20 fragments per root system of mixed-grown oak and 30 fragments per root system of sole-grown oak). The total of these 120 primary roots per treatment was observed with a binocular magnifier and Leica LAS Core® software with ×40 magnification. For each fine root fragment, the number of fine lateral roots and the number of ectomycorrhizal root tips were counted. Fungal species were not identified morphologically, but root tips were considered as ectomycorrhizal based on the distinctive presence of a hyphal sheath (the ectomycorrhiza mantle) surrounded with extramatrical hyphae [see [Supporting Information—Photo S1](#)]. The rate of ectomycorrhization was then calculated as the number of ectomycorrhizal root tips/total number of observed root tips.

Molecular identification of oak EMF

Ectomycorrhizal root tips were investigated with molecular barcoding when oak root systems were developed enough to apply the following procedure, that is, for six sole-grown oaks and only four mixed-grown oaks (insufficient root system with little or no branching with mycorrhizae). Under binocular magnifier and Leica LAS Core® software with ×40 magnification, 10 ectomycorrhizal root tips per oak root system were excised and stored at –20 °C until DNA extraction. For each excised ectomycorrhizal root tip, total DNA was extracted using DNeasy 96 Plant Pro Kit (Qiagen) following the supplier's protocol. PCR amplification of locus

ITS was performed using the primers ITS1-F ([Gardes and Bruns 1993](#)) and ITS4 ([White *et al.* 1990](#)) following PCR conditions described in [Vincenot *et al.* \(2012\)](#). Amplicons were purified using PCR DNA and Gel Band Purification Kit (GFX™), and Sanger-sequenced on ABI 3730 XL (Life Technologies/ThermoFisher). Nucleotide sequences were visually inspected, then aligned and paired to retrieve contig sequences in MEGAX ([Kumar *et al.* 2018](#)). A total of 87 samples were sent for Sanger sequencing: 57 sequences of ectomycorrhizal roots of sole-grown oaks and 30 sequences of ectomycorrhizal roots of mixed-grown oaks. These sequences are available in GenBank database under accession numbers ON391357–ON391414. Eventually, taxonomic identity of fungi in ectomycorrhizal root tips was retrieved by submitting contig sequences to BLAST search in fungi molecular database UNITE ([Nilsson *et al.* 2019](#)).

Plant N content

After drying at 60 °C for at least 48 h, shoots and roots were weighed and ground to a fine powder. Total N content (% of dry weight) was then determined with an elemental analyser (vario ISOTOPE cube, Elementar, Hanau, Germany) in line with a gas isotope ratio mass spectrometer (IsoPrime 100, Isoprime Ltd, Cheadle, UK) at the analytic platform SilvaTech.

Statistics

Statistical analysis was performed using R software (Version 3.4.1). Data were means of $n = 6$ plants for dry weight in each treatment, and $n = 120$ pseudo-replicates ($n = 20$ fragments per root system of the total six mixed-grown oak and $n = 30$ fragments per root system of the total four sole-grown oak) for lateral root tips and ectomycorrhizal root tip counts. All variables were tested for normality and homoscedasticity, using the Shapiro–Wilk and Levene tests. Data analysed using ANOVA. Data of lateral root tips and ectomycorrhizal root tip counts were analysed using linear models (lmer function), where treatment was the predictor variable and oak identity was a random effect.

Results

Mean total oak dry weight increased by 3.2-fold between the planting (April 2018) and the harvest (May 2019) for sole-grown oaks, compared with only 1.3-fold for mixed-grown oaks with moor grass ([Fig. 1A](#); see [Supporting Information—Table S1](#)). Both shoot (stem + leaves) and root biomass showed 2.5- and 2.2-fold decreases, respectively, when mixed-grown with moor grass ($P < 0.001$ and $P = 0.011$, respectively; [Fig. 1B](#); see [Supporting Information—Table S2](#)). Whether sole-grown or grown in mix, moor grass grew much faster than oak: between April 2018 and May 2019, mean moor grass dry weight increased over 150 times when sole-grown, compared with over 260 times when mixed-grown with oak ([Fig. 1A](#); see [Supporting Information—Table S1](#)). Conversely to oak, moor grass exhibited a higher (1.7-fold increase) mean dry weight when mixed-grown than when sole-grown for both shoot and root ($P = 0.005$ and $P = 0.052$, respectively; [Fig. 1B](#); see [Supporting Information—Table S2](#)).

Sole-grown oak roots displayed a mean number of 16.7 fine lateral roots per cm ([Fig. 2A](#); see [Supporting Information—Table S3](#)), with an average of 6.6 ectomycorrhizal root tips per cm ([Fig. 2B](#); see [Supporting Information—Table S4](#)), thus

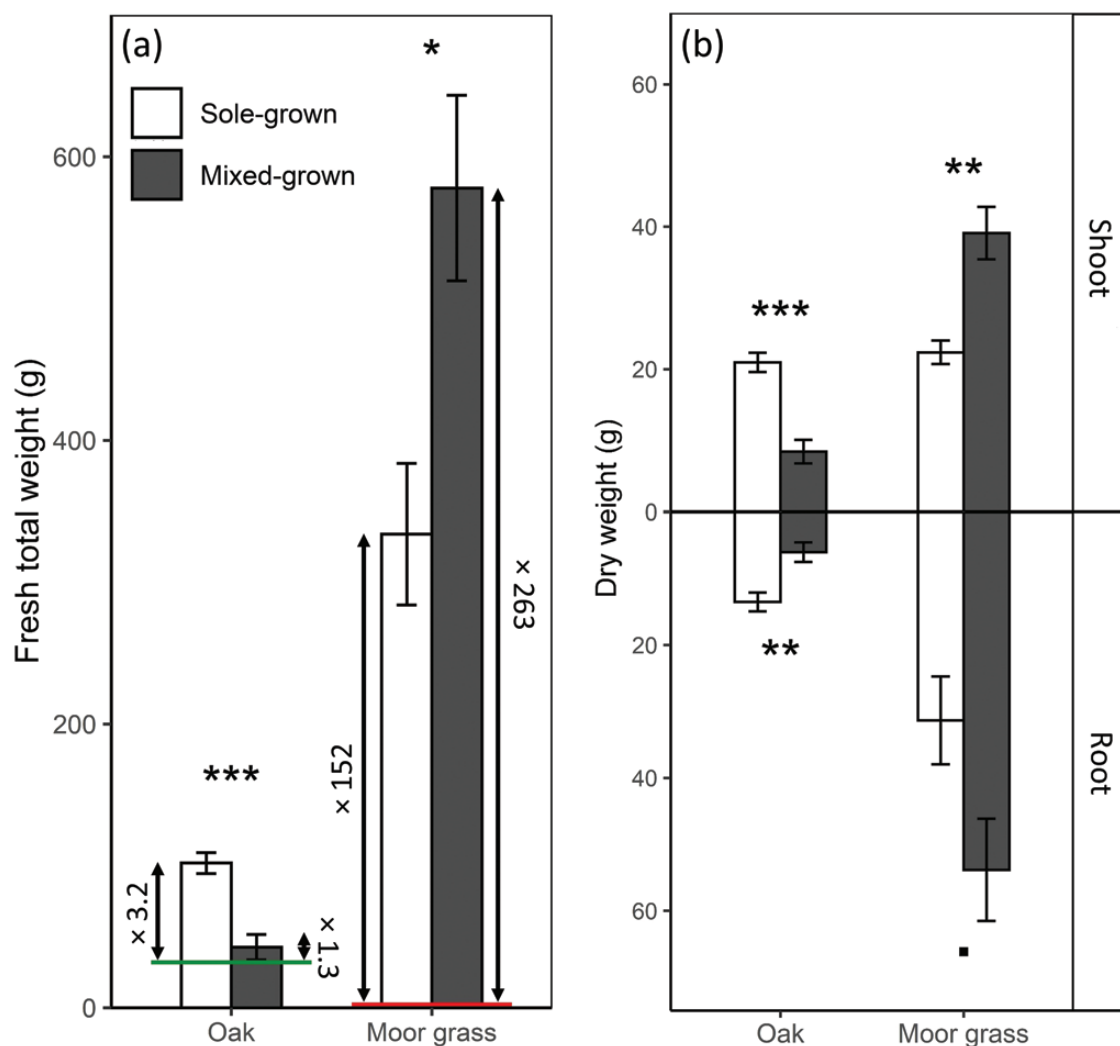


Figure 1. (A) Fresh total weight of oak and moor grass when sole-grown (white) or mixed-grown (grey). The two horizontal lines indicate the total fresh weight at the beginning of experiment for oak and moor grass, respectively. The numbers indicate the increase in biomass between the beginning of the experiment and the harvest. (B) Dry weight of oak and moor grass shoots and roots. Values are reported as means $\pm$ SE. *, **, *** correspond to $P < 0.05$, 0.01 and 0.001 , respectively, for Student's t -test for each organ, $n = 6$.

about 40 % of root tips having formed ectomycorrhizal association. When mixed-grown with moor grass, both the numbers of oak fine lateral roots ($11.2 \cdot \text{cm}^{-1}$) and ectomycorrhizal root tips ($2.8 \cdot \text{cm}^{-1}$) were markedly reduced ($P < 0.001$ for both root tips and ectomycorrhizal root tips), leading to a 2.4-fold decrease of oak ectomycorrhization rate.

Molecular typing of ectomycorrhizal root tips allowed the identification of 35 sequences among the 57 sequences in sole-grown oaks and 16 sequences among the 30 sequences in mixed-grown oaks, the rest remaining as 'unidentified taxa' after BLAST due to poor-quality sequence. Six different taxa were identified in sole-grown versus five in mixed-grown treatment (Fig. 3). In sole-grown, Ascomycota was more abundant (36.8 %) than Basidiomycota (24.6 %) on the contrary to mixed-grown (23.3 % and 30 % of Ascomycota and Basidiomycota, respectively). *Sphaerospora brunnea* (Ascomycota) and *Thelophora terrestris* (Basidiomycota) were present in both treatments, with a higher proportion of *S. brunnea* (21.1 %) than *T. terrestris* (17.5 %) in sole-grown treatment. Conversely, in mixed-grown the proportion of *S. brunnea* (13.3 %) was lower than of *T. terrestris* (20 %).

Mycorrhizal fungi of the orders Helotiales and Pezizales were also present in both treatments (7.0 % and 6.7 % in sole-grown and mixed-grown, respectively, for Helotiales; 8.8 % and 3.3 % in sole-grown and mixed-grown, respectively, for Pezizales), yet their families could not be identified. *Laccaria proxima* and *Hebeloma vaccinum* were also detected in sole-grown treatment only, in lower proportions (3.5 % for each of these). In mixed-grown, 10 % of Sebaciales were identified, but none in sole-grown. Unidentified sequences represent 38.6 % in sole-grown and 46.7 % in mixed-grown.

Along with the decrease in oak biomass when mixed-grown, there was a decrease in oak shoots N content when mixed-grown moor grass ($P = 0.014$; from 1.2 % of dry shoot weight in sole-grown to 0.9 % in mixed-grown oak; Fig. 4A; see Supporting Information—Table S5) but N content in oak root was not changed in mixed-grown ($P = 0.69$; 0.57 % and 0.54 % of dry root weight when sole-grown vs. mixed-grown, respectively). Nitrogen content in soil was statistically similar between pots containing sole-oak and pots containing mixed-grown ($P = 0.340$), but significantly higher in pots containing sole-moor grass ($P = 0.014$ and $P < 0.001$ for sole-oak vs.

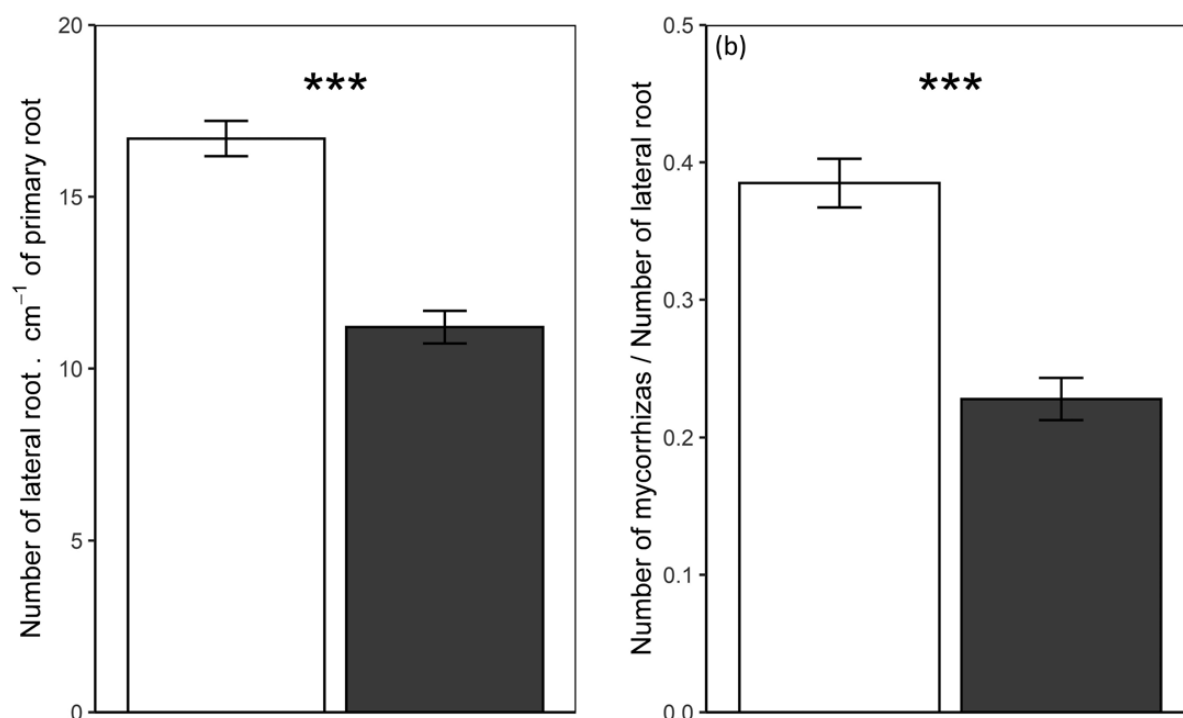


Figure 2. Number of lateral roots (A) and ectomycorrhization rate, i.e. ratio of number of mycorrhizal root tips to number of lateral roots (B) when sole-grown (white) or mixed-grown (grey). Values are reported as means $\pm$ SE. *** corresponds to $P < 0.001$ for Student's t -test for each organ, $n = 120$.

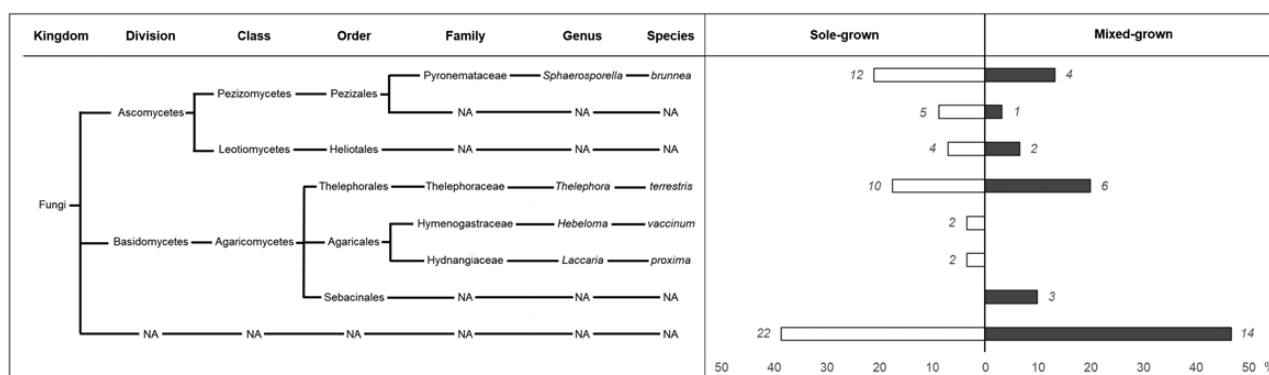


Figure 3. Proportion in percentage of oak root mycorrhizas in sole-grown (white) and mixed-grown (grey) for each taxon. Italic number next bar indicated total number of mycorrhizas sequences. The results of BLAST analyses assigning taxa to each sequence are shown on the left. NA means that the BLAST result did not give more details about the taxon. The last line represents the sequences that did not give any BLAST result.

sole-moor grass and mixed-species vs. sole-moor grass, respectively) (Fig. 4B; see [Supporting Information—Tables S6 and S7](#)).

Discussion

Moor grass competition impairs oak root ability to forage for N

Presence of moor grass in the same pot as oak seedlings resulted in reduced growth of oak compared to sole-grown seedlings, as shown by a lower biomass production. Also, oak root branching (number of lateral fine roots/fine root length) lowered when mixed-grown with moor grass. These results are consistent with those of other studies reporting the negative effect of grass species on the number of lateral roots, leading to decreased tree seedling growth (Schroth

1999; Harmer and Robertson 2003; Schaller *et al.* 2003; Collet and Chenost 2006). Root foraging ability associated with root system architecture, and particularly branching, plays a critical role in nutrient uptake, particularly of N and phosphorus (Bar-Tal *et al.* 1997; Postma *et al.* 2014; Shahzad and Amtmann 2017; Duque and Villordon 2019). Profuse branching of root systems increases both soil exchange surfaces and the volume of soil that can be explored (Fitter *et al.* 1991). Some studies have demonstrated that a localized supply of nutrients increases lateral root density and elongation only where the nutrient levels are high (Hackett 1972; Drew and Saker 1975; Granato and Raper 1989; Hodge 2004), yet a homogenous nutrient deficiency can stimulate lateral root formation to increase soil exploitation (Shahzad and Amtmann 2017). Conversely, others have observed an inhibitory effect of high nutrient concentration on root

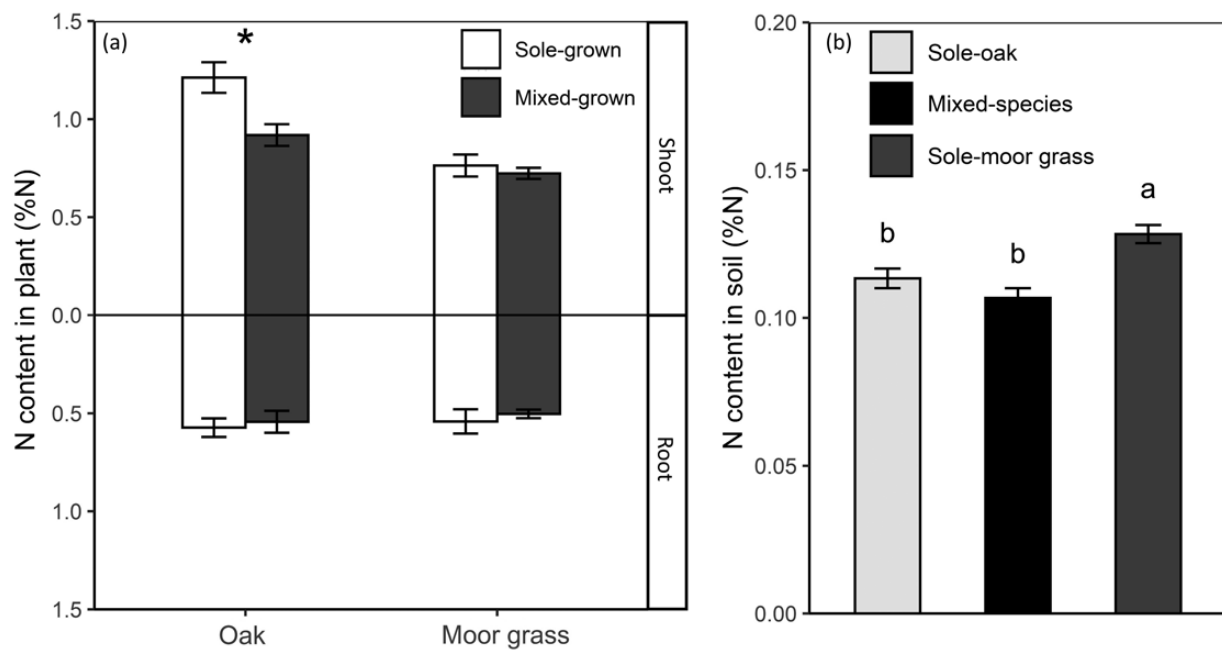


Figure 4. Nitrogen content in plant (A) shoots and roots when sole-grown (white) or mixed-grown (grey) and in soil (B) in sole-oak pot (light grey), sole-moor grass pot and (dark grey) in mixed-species pot (black). Values are reported as means $\pm$ SE. *correspond to $P < 0.05$, for Student's t -test for each organ, $n = 6$.

growth (Zhang and Forde 1998; Zhang *et al.* 1999; Linkohr *et al.* 2002; Celis-Arámburo *et al.* 2011; Gruber *et al.* 2013).

In our experiment, N availability in soil probably does not explain lateral root inhibition, as N content values were similar in sole- and mixed-grown treatments (N content was 0.13 ± 0.003 % and 0.11 ± 0.003 % in sole-grown and mixed-grown, respectively, without statistically significant difference). However, numerous other factors have been identified as inhibitors of lateral root formation and growth, including phytohormones (Fukaki and Tasaka 2009; Shkolnik-Inbar and Bar-Zvi 2010; Lewis *et al.* 2011). In the specific context of a mixed-grown treatment, shoot and root extracts of some species, generally grass and crops, can have an allelopathic effect on lateral root formation and length in neighbours (Horsley 1977; Pardales *et al.* 1992; Chon *et al.* 2002; Amoo *et al.* 2008; Hossain *et al.* 2016). Ahmed *et al.* (2007) reported such an inhibitory effect was much more pronounced in root and lateral root development than for shoot or seed germination. This suggests inhibition of oak root system development by moor grass could play a key role in the interaction between the two species.

Moor grass depletes oak root ectomycorrhization

Moor grass also reduced ectomycorrhizal association rate with oak roots, as the number of ectomycorrhizal root tips was 1.9 times lower when mixed-grown with moor grass. To our knowledge, such a reduction of association rate with EMF by moor grass has not been reported yet.

Regarding mechanisms that can inhibit mycorrhizal colonization, most studies focused on vesicular AM symbiosis (Abbott and Gazey 1994; Koide and Schreiner 2003; Garg and Chandel 2010). As for lateral root initiation and development, edaphic parameters can inhibit root association with EMF, such as phosphorus enrichment,

low soil temperature or phytohormones (Graham *et al.* 1982; el Ghachtouli *et al.* 1996; Javaid 2008; Kobae *et al.* 2016). Allelochemical exudation by roots may also be involved in the inhibition of both endomycorrhizae and ectomycorrhizae, but only a few studies have addressed that mechanism, mostly testing for allelopathy using leaf extracts at various concentrations (Olsen *et al.* 1971; Nilsson 1994; Javaid 2007, 2008). In this line, Nilsson *et al.* (1993) suggested that a decrease in ectomycorrhizal symbiosis of *P. sylvestris* seedlings was probably due to a direct negative effect of *E. hermaphroditum* on hyphal diameter. Pellissier (1994) identified phenolic allelochemicals in forest floor humus involved in conifers (*Picea abies* and *Picea mariana*) regeneration failure, possibly because these compounds can directly reduce root growth and ectomycorrhizal formation (Mallik 1987, 1998, 2003; Zhu and Mallik 1994; Inderjit and Mallik 1996).

An allelopathic activity of moor grass has been suggested in a few studies (Becker and Lévy 1982; Timbal *et al.* 1990), but no allelopathic compound has been clearly identified so far (Fernandez *et al.* 2021). For instance, Timbal *et al.* (1990) showed no impact of moor grass on fungal taxonomic richness but a change of species community composition of EMF in *Q. rubra* seedlings roots, suggesting the necessity of studying composition of *Molinia* roots exudates. Moor grass roots and shoots might contain phenolic compounds that can be released into the soil through litter decomposition, leaching and exudation, with possible inhibitory effects on oak lateral root initiation, mycorrhizal symbiosis establishment or functioning, or directly on fungal hyphae development. Allelopathic compounds of moor grass may also act as an environmental filter on the structure of ectomycorrhizal communities associated to oak roots (Becker and Lévy 1982; Timbal *et al.* 1990; Fernandez *et al.* 2021).

Number of oak EMF sequences decreases when mixed-grown with moor grass

Analyses of molecular identification of EMF confirmed oaks mixed-grown with moor grass harboured lower numbers of sequenced mycorrhizal taxa than sole-grown oaks. However, caution should be exercised in interpreting the effect of moor grass on oak mycorrhizal biodiversity because 38.6 % and 46.7 % of sequences could not be identified in sole-grown and mixed-grown, respectively. Unidentified fungal sequences represent 17.3 % in sole-grown and 40.7 % in mixed-grown. Such a difference can be explained by a more necrotic root system that was observed in mixed-grown during harvest, and thus this could lead to lower quality ectomycorrhizal samples. Because oaks mixed-grown with moor grass had a lower root system biomass and a lower mycorrhization rate than sole-grown oaks, it was then more difficult to harvest mycorrhizae and extract DNA.

Moreover, Timbal *et al.* (1990) demonstrated a change in EMF community composition but not in abundance. Voříšková *et al.*'s (2014) study on *Q. petraea* showed that Basidiomycota was the dominant division, in line with our results only in mixed-grown oak. Also, in temperate forest, Basidiomycota are commonly more represented than Ascomycota (Venturella *et al.* 2011; Weber *et al.* 2013), especially under *Quercus* spp. (Morris *et al.* 2008; Smith *et al.* 2009; Toju *et al.* 2013a, b). Nevertheless, He *et al.* (2016) measured a higher proportion of Ascomycota (64.6 %) than Basidiomycota (26.6 %) on *Quercus mongolica* root system, in accordance with our study in sole-grown treatment. Weber *et al.* (2013) demonstrated that the proportion of Ascomycota in bulk soil increases with soil depth with a corresponding decrease in Basidiomycota. Here, we harvested mycorrhizae from the entire root system, not just from the surface, which possibly partly explains a higher proportion of Ascomycota than Basidiomycota in sole-grown treatment in comparison with other studies where roots were harvested in the surface soil only (0–15 cm). Other drivers may explain an unexpected proportion of Ascomycota in ectomycorrhizal fungal community, such as seedlings or trees origin and age, soil properties and environmental conditions (Morris *et al.* 2008; Wu *et al.* 2013; Lamit *et al.* 2016). In the studies cited above, all samples were taken from mature trees located in forests, whereas we worked with oak nursery seedlings in a pot experiment.

Sebacinales were only found in mixed-grown oak roots, but possibly this taxon was also present among the unidentified sequences in sole-grown oaks. Sebacinales are commonly found on the roots of many dicot and monocot plant species (Weiß *et al.* 2016), including oaks (Richard *et al.* 2011; Oberwinkler *et al.* 2013), and participate to common mycorrhizal networks that can allow nutrients exchanges amongst host plants (Selosse *et al.* 2017). Potentially shared Sebacinales raise the hypothesis of C transfer between oak and moor grass through shared mycorrhizal symbiotic mycelia, which would need further isotopic investigation to be verified. *Hebeloma vaccinum* and *L. proxima* were only found in sole-grown oak and also commonly form ectomycorrhizae with *Quercus* sp. (Leski *et al.* 2010; Otsing and Tedersoo 2015). We demonstrated the strong negative effect of moor grass on relative numbers of retrieved EMF sequences, underlying the importance of mycorrhizae identification to describe interactions between plants. Our results suggest possible allelochemical compounds of moor grass may filter oak EMF community

composition and impair ectomycorrhizal establishment in oak root system, limiting oak growth (Fernandez *et al.* 2021). The specific intensity of moor grass negative effects on EMF taxa can also be discussed. Relative numbers in sequenced EMF community were lower on mixed-grown oaks than on sole-grown oaks for EMF various taxa (Pezizales 2.6 times lower; *S. brunnea*, 1.6 times lower; Helotiales 1.1 time), suggesting a fungal-specific variation of intensity of moor grass negative effect. Such shifts in the EMF taxa relative numbers of EMF community were consistent with other studies demonstrating disproportional impact of allelochemicals on mycorrhizal fungi (Roche *et al.* 2021).

Oaks N allocation: the conservative strategy

Along with a decrease in shoot dry weight, the significantly lower N content in oak shoot for mixed-grown strongly suggests a reduced ability to take soil N up, associated with the decrease in lateral roots and ectomycorrhization as discussed above. In sole-grown treatment, mean N content in oak shoot was 1.21 ± 0.08 %, in line with previous findings (Espelta *et al.* 2005), but when oak was mixed-grown with moor grass, mean N content was only 0.92 ± 0.05 %.

Nitrogen content in oak roots was similar in the two treatments (Fig. 4B), although dry weight was lower when mixed-grown, indicating N allocation towards root in oaks with moor grass was larger than in sole-grown oaks. These results are consistent with previous findings highlighting a modification of N allocation in forest tree seedlings in the presence of grasses (Welker *et al.* 1991; Coll *et al.* 2004). Nitrogen allocation to roots did not correlate with investment in lateral root development, prospection and resource capture, so it strongly suggests a strategy for N storage (i.e. conservative). According to Grime (1974), moor grass, through its strong competitive strategy, induces stress in oak seedling resulting in intensification of N conservative strategy and a lower ability to compete for N soil. Vernay *et al.* (2018b) also described modification of N allocation in oak seedling when mixed-grown with *D. cespitosa*, whereas fine root dry weight was constant. This confirms a different conservative strategy of N economy due to the perturbation caused by neighbouring grass. One possibility is that limiting root growth and biomass will maintain sufficient N for plant survival.

Nitrogen content in soil was unexpectedly higher in pot containing sole-grown moor grass compared to pot containing sole-grown oaks or pot with the mixed-grown association. This result was not consistent with other study demonstrating that grass significantly decrease nitrate amount—but not ammonium—n soil compared to oak seedling (Vernay *et al.* 2018a). Although N content was higher in soil of sole-grown moor grass compared to mixed-grown, N content was statistically similar in both moor grass shoot and root. Difference of N content between shoot and root did not corroborate difference of biomass which was much higher in mixed-grown. It questions the effect of the presence of a neighbour on the plant NUE (Congreves *et al.* 2021): do oak seedlings favour moor grass NUE? Beyond N allocation and use, competition for soil N between oak and moor grass refers to theories of interspecific competition regarding resource use and availability. According to Tilman (1990) R* rule, R* (i.e. the lowest minimum resource threshold tolerated by a given species) of moor grass is supposed to be lower than oak R* suggesting that its competitive potential is greater as soil N

availability decreases. However, contrary to Tilman's theory as well as game theory (Mcnicke and Dybzinski 2013), competitive potential for soil resource of moor grass was not expressed by higher root biomass, but by higher shoot biomass. These suggestions also question the tragedy of the commons indicating that inter-plant competition for shared space should lead to increased root biomass and forage but decreased yield (Gersani *et al.* 2001). On the contrary, our study demonstrates that in a context of interspecific competition, a Poaceae can show a small increase in its root biomass but a clear increase in its aerial biomass. The challenge in fitting our results with existing ecological theories probably stems from the fact that the oak–molinia interaction is antagonistic, which is still poorly documented.

Oak increased moor grass biomass: the antagonistic interaction

Higher biomass production of moor grass when mixed-grown with oak observed here is consistent with previous results demonstrating facilitative effects of oak seedlings on grasses (Vernay *et al.* 2018a) due to rapid and significant N transfer from oak to grass (Fernandez *et al.* 2020). In the complex framework of plant interactions, our results raise the following questions: (i) Is the competitive potential of the moor grass enhanced in the presence of oak seedlings because of intensification for N competition according to Tilman's R^* theory? (ii) Does the competitive potential of the moor grass increase the stress in oak seedlings resulting in a decrease of N competition according to Grime's theory? And then, (iii) can we really talk about antagonistic interaction or modification of competitive potential of moor grass in the presence of oak seedlings? Our results perfectly corroborate Callaway's (1995) statement: 'the overall effect of one species on another may be the product of multiple, complex interactions'. Interactions between these two species constitute a double jeopardy for oak, which is stunted by moor grass and has been also shown here to facilitate moor grass biomass.

Consequences of the interference interactions in natural ecosystems

Our experiment needs to be replicated *in situ*, as growth in pots can modify root architecture compared with naturally regenerated oak seedlings (Tsakalidimi *et al.* 2009), and impacts on ectomycorrhizal rate could be altered due to reduced space in the pot. The role of soil bacteria modulating N provision in the rhizosphere could be studied to gain a better understanding of the mechanisms by which lateral root formation and ectomycorrhization are inhibited. Then, in order to better characterize the interactions and question the ecological theories on plant competition, it would be interesting to measure the amount of nutrients (N, C, P, K) in the soil throughout the experiment. In general, analyses of plant, soil nutrients and mycorrhizal content would help clarify the processes and bio-actors targeted by the moor grass.

Furthermore, a common AM network can enhance inter- or intraspecific transfer of allelochemicals directly to the root system of the target species in pot experiment (Barto *et al.* 2011). Several oak species appear as dual host trees that can associate both with ectomycorrhizal and AM fungi (Dickie *et al.* 2004; He *et al.* 2006; Toju *et al.* 2013a, b). The latter being typically connected to herbaceous host plants, potential

allelopathic effects of moor grass may be relayed by mycorrhizal symbionts shared with oak seedlings. This hypothesis could be further investigated by searching for shared AM fungal species and/or genotypes in moor grass and oak root systems. Also, the impacts of oak mycorrhizal communities' alteration by moor grass on N uptake decrease would deserve further investigation. Eventually, our findings suggesting a strong allelopathic effect of moor grass on oak seedlings call for the identification of phenolic compounds present in moor grass root exudates.

In oak–moor grass stands, it has been clearly demonstrated that thinning induces a large influx of light into the understory favouring the development of the moor grass and hinders the regeneration of the oak (Vernay *et al.* 2016). However, the understory vegetation often appears to be essential to the forest renewal process, by promoting nutrient turnover and thus produce more fertile soils than bare soils (Watt 1919; Bobiec *et al.* 2018). Forest management must therefore consider the balance between thinning intensity and the consequences on the interactions between understory species and seedlings.

Supporting Information

The following additional information is available in the online version of this article—

Photo S1. Oak mycorrhizal roots observed with a binocular magnifying glass (×40).

Figure S1. Evolution of volumetric soil water content (%) in the pot (mean ± SD). The horizontal red line indicates 40 % of soil water holding capacity, i.e. the threshold below which plants can begin to experience a drought (Vicca *et al.* 2012). [see **Supporting Information—Appendix S1**].

Vicca, S., *et al.* (2012). "Urgent need for a common metric to make precipitation manipulation experiments comparable." *New Phytologist* 195(3): 518–522.

Appendix S1. Evolution of soil water content in the pots.

To avoid interaction with confounding factors in the experiment such as water availability, pots were irrigated to field capacity throughout the experiment. To ensure the correct water amount was delivered, volumetric soil water content was continuously measured with TDR probes in a previous experiment (2017–18), allowing to test the optimal water supply for this experiment [see **Supporting Information—Fig. S1**]. The hydric characteristics of the soil were determined by establishing a curve between volumetric soil water content and matrix potential and gave 22.5 % for field capacity and 9.4 % for permanent wilting point. **Supporting Information—Figure S1** shows that most of time pots were at field capacity and experienced only slight droughty periods (values below the red line, 40 % of soil water holding capacity, Vicca *et al.* 2012), too short to influence results reported in that study. In particular at the end of the experiment, we stopped irrigation 2 weeks before plant harvest to facilitate root extraction from the soil. Exactly the same water supply was added in 2018–19 experiment.

Tables S1–S7. Statistical analyses.

Acknowledgements

The authors thank André Marquier, Christophe Serre, Pascal Walser and Brigitte Saint-Joanis for their valuable help in processing plant material.

Contributions by the Authors

M.F. conceptualized, designed and performed the experiments, collected and analyzed data, wrote and edited the original version. P.M. conceptualized, designed the experiments, analyzed data and edited the original version. L.V. designed and performed the experiments, analyzed data, edited the original version. A.V. conceptualized, edited the original version. P.B. conceptualized, designed the experiments, analyzed data and edited the original version. T.A. conceptualized and edited the original version.

Conflict of Interest Statement

None declared.

Data Availability

The data sets generated during and/or analysed for the present study are available from the corresponding author on reasonable request. Sequences produced for molecular identification of fungal mycorrhizal symbionts are deposited in GenBank under accession numbers ON391357–ON391414.

Sources of Funding

This experiment was conducted with funds from the University of Clermont Auvergne and Urban MycoServe project. Marine Fernandez PhD work was supported by a Grant of the French Ministry of Research.

Literature Cited

- Abbott, L.K., Gazey, C., 1994. An ecological view of the formation of VA mycorrhizas. *Plant Soil* 159, 69–78. <https://doi.org/10.1007/BF00000096>
- Aerts, R., Berendse, F., 1988. The effect of increased nutrient availability on vegetation dynamics in wet heathlands. *Vegetatio* 76, 63–69. <https://doi.org/10.1007/BF00047389>
- Ahmed, R., Uddin, M.B., Khan, M.A.S.A., Mukul, S.A., Hossain, M.K., 2007. Allelopathic effects of *Lantana camara* on germination and growth behavior of some agricultural crops in Bangladesh. *Journal of Forestry Research (Harbin)* 18, 301–304. <https://doi.org/10.1007/s11676-007-0060-6>
- Alberton, O., Kuyper, T.W., Gorissen, A., 2007. Competition for nitrogen between *Pinus sylvestris* and ectomycorrhizal fungi generates potential for negative feedback under elevated CO₂. *Plant and Soil* 296, 159–172. <https://doi.org/10.1007/s11104-007-9306-5>
- Amoo, S.O., Ojo, A.U., van Staden, J., 2008. Allelopathic potential of *Tetrapleura tetraptera* leaf extracts on early seedling growth of five agricultural crops. *South African Journal of Botany* 74, 149–152. <https://doi.org/10.1016/j.sajb.2007.08.010>
- Balandier, P., Collet, C., Miller, J.H., Reynolds, P.E., Zedaker, S.M., 2006. Designing forest vegetation management strategies based on the mechanisms and dynamics of crop tree competition by neighbouring vegetation. *Forestry* 79, 3–27. <https://doi.org/10.1093/forestry/cpi056>
- Bar-Tal, A., Ganmore-Neumann, R., Ben-Hayyim, G., 1997. Root architecture effects on nutrient uptake, in: Altman, A., Waisel, Y. (Eds.), *Basic Life Sciences*. Springer US, Boston, MA, pp. 39–45. https://doi.org/10.1007/978-1-4615-5403-5_5
- Barto, E.K., Hilker, M., Müller, F., Mohny, B.K., Weidenhamer, J.D., Rillig, M.C., 2011. The fungal fast lane: common mycorrhizal networks extend bioactive zones of allelochemicals in soils. *PLoS One* 6, 1–7. <https://doi.org/10.1371/journal.pone.0027195>
- Becker, M., Lévy, G., 1982. Le dépérissement du chêne en forêt de Tronçais Les cause ecologiques. *Annales des sciences forestières* 39, 439–444.
- Bertness, M.D., Callaway, R.M., 1994. Positive interactions in communities. *Tree* 9, 191–193. <https://doi.org/10.1201/9780203738559>
- Bleasdale, J.K.A., 1960. Studies on plant competition., in: Harper, J.L. (Ed.), *Biology of Weeds*. Oxford, England: Blackwell Science Pub, pp. 133–142.
- Bobiec, A., Reif, A., Öllerer, K., 2018. Seeing the oakscape beyond the forest: a landscape approach to the oak regeneration in Europe. *Landscape Ecology*. <https://doi.org/10.1007/s10980-018-0619-y>
- Bonfante, P., Genre, A., 2010. Mechanisms underlying beneficial plant-fungus interactions in mycorrhizal symbiosis. *Nature Communications*. <https://doi.org/10.1038/ncomms1046>
- Bronstein, J.L., 2009. The evolution of facilitation and mutualism. *Journal of Ecology* 97, 1160–1170. <https://doi.org/10.1111/j.1365-2745.2009.01566.x>
- Callaway, R.M., 1995. Positive interactions among plants. *The Botanical Review* 61, 306–349. <https://doi.org/10.1007/BF02912621>
- Celis-Arámburo, T. de J., Carrillo-Pech, M., Castro-Concha, L.A., Miranda-Ham, M. de L., Martínez-Estévez, M., Echevarría-Machado, I., 2011. Exogenous nitrate induces root branching and inhibits primary root growth in *Capsicum chinense* Jacq. *Plant Physiology and Biochemistry* 49, 1456–1464. <https://doi.org/10.1016/j.plaphy.2011.09.003>
- Chalot, M., Brun, A., 1998. Physiology of organic nitrogen acquisition by ectomycorrhizal fungi and ectomycorrhizas. *FEMS Microbiology Reviews* 22, 21–44. <https://doi.org/10.1111/j.1574-6976.1998.tb00359.x>
- Chalot, M., Plassard, C., 2011. Ectomycorrhiza and nitrogen provision to the host tree, in: Polacco, J.C., Todd, C.D. (Eds.), *Ecological Aspects of Nitrogen Metabolism in Plants*. Wiley-Blackwell, England, pp. 69–94.
- Cheng, X., Bledsoe, C.S., 2004. Competition for inorganic and organic N by blue oak (*Quercus douglasii*) seedlings, an annual grass, and soil microorganisms in a pot study. *Soil Biology and Biochemistry* 36, 135–144. <https://doi.org/10.1016/j.soilbio.2003.08.024>
- Chon, S.-U., Choi, S.-K., Jung, S., Jang, H.-G., Pyo, B.-S., Kim, S.-M., 2002. Effects of alfalfa leaf extracts and phenolic allelochemicals on early seedling growth and root morphology of alfalfa and barnyard grass. *Crop Protection* 21, 1077–1082.
- Coll, L., Balandier, P., Picon-Cochard, C., 2004. Morphological and physiological responses of beech (*Fagus sylvatica*) seedlings to grass-induced belowground competition. *Tree Physiology* 24, 45–54.
- Coll, L., Balandier, P., Picon-Cochard, C., Prévosto, B., Curt, T., 2003. Competition for water between beech seedlings and surrounding vegetation in different light and vegetation composition conditions. *Annals of Forest Science* 60, 593–600. <https://doi.org/10.1051/forest:2003051>
- Collet, C., Chenost, C., 2006. Using competition and light estimates to predict diameter and height growth of naturally regenerated beech seedlings growing under changing canopy conditions. *Forestry* 79, 489–502. <https://doi.org/10.1093/forestry/cpl033>
- Collet, C., Löf, M., Pagès, L., 2006. Root system development of oak seedlings analysed using an architectural model. Effects of competition with grass. *Plant Soil* 279, 367–383. <https://doi.org/10.1007/s11104-005-2419-9>
- Congreves, K.A., Otchere, O., Ferland, D., Farzadfar, S., Williams, S., Arcand, M.M., 2021. Nitrogen use efficiency definitions of today and tomorrow. *Frontiers in Plant Science* 12. <https://doi.org/10.3389/fpls.2021.637108>
- Craig, M.E., Turner, B.L., Liang, C., Clay, K., Johnson, D.J., Phillips, R.P., 2018. Tree mycorrhizal type predicts within-site variability in the storage and distribution of soil organic matter. *Global Change Biology* 24, 3317–3330. <https://doi.org/10.1111/gcb.14132>

- Davis, M.A., Wragg, K.J., Reich, P.B., 1998. Competition between tree seedlings and herbaceous vegetation: support for a theory of resource supply and demand. *Journal of Ecology* 86, 652–661. <https://doi.org/10.1046/j.1365-2745.1998.00087.x>
- Dickie, I.A., Guza, R.C., Krazewski, S.E., Reich, P.B., 2004. Shared ectomycorrhizal fungi a herbaceous between perennial (*Helianthemum bicknellii*) and oak (*Quercus*) seedlings. *New Phytologist* 164, 375–382. <https://doi.org/10.1111/j.1469-8137.2004.01177.x>
- Drew, M.C., Saker, L.R., 1975. Nutrient supply and the growth of the seminal root system in Barley. *Journal of Experimental Botany* 26, 79–90. <https://doi.org/10.1093/jxb/26.1.79>
- Duque, L.O., Villordon, A., 2019. Root branching and nutrient efficiency: status and way forward in root and tuber crops. *Frontiers in Plant Science* 10, 1–8. <https://doi.org/10.3389/fpls.2019.00237>
- Eaton, E., Caudullo, G., Oliveira, S., de Rigo, D., 2016. *Quercus robur* and *Quercus petraea* in Europe: distribution, habitat, usage and threats. *European Journal of Forest Tree Species* 23, 160–163. <https://doi.org/10.15421/011515>
- el Ghachtouli, N., Martin-Tanguy, J., Paynot, M., Gianinazzi, S., 1996. First report of the inhibition of arbuscular mycorrhizal infection of *Pisum sativum* by specific and irreversible inhibition of polyamine biosynthesis or by gibberellic acid treatment. *FEBS Letters* 385, 189–192. [https://doi.org/10.1016/0014-5793\(96\)00379-1](https://doi.org/10.1016/0014-5793(96)00379-1)
- Espelta, J.M., Cortés, P., Mangirón, M., Retana, J., 2005. Differences in biomass partitioning, leaf nitrogen content, and water use efficiency ($\delta^{13}\text{C}$) result in similar performance of seedlings of two Mediterranean oaks with contrasting leaf habit. *Écoscience* 12, 447–454. <https://doi.org/10.2980/1195-6860-12-4-447.1>
- Fernandez, M., Malagoli, P., Vernay, A., Améglio, T., Balandier, P., 2020. Below-ground nitrogen transfer from oak seedlings facilitates *Molinia* growth: ^{15}N pulse-chase labelling. *Plant Soil* 449, 343–356. <https://doi.org/10.1007/s11040-020-04473-9>
- Fernandez, M., Malagoli, P., Gallet, C., Fernandez, C., Vernay, A., Améglio, T., Balandier, P., 2021. Investigating the role of root exudates in the interaction between oak seedlings and purple moor grass in temperate forest. *Forest Ecology Management* 491, 1–9. <https://doi.org/10.1016/j.foreco.2021.119175>
- Fernandez, M., Vernay, A., Henneron, L., Adamik, L., Malagoli, P., Balandier, P., 2022. Plant N economics and the extended phenotype: Integrating the functional traits of plants and associated soil biota into plant-plant interactions. *Journal of Ecology* 00, 1–18. <https://doi.org/10.1111/1365-2745.13934>
- Fitter, A.H., Stickland, T.R., Harvey, M.L., Wilson, G.W., 1991. Architectural analysis of plant root systems 1. Architectural correlates of exploitation efficiency. *New Phytol* 118, 375–382.
- Freschet, G.T., Valverde-Barrantes, Oscar J., Tucker, Caroline M., Craine, Joseph M., McCormack, M. Luke, Vielle, Cyrille, Fort, Florian, Blackwood, Christopher B., Urban-Mead, Katherine R., Iversen, Colleen M., Bonis, Anne, Comas, Louise H., Cornelissen, Johannes H. C., Dong, Ming, Guo, Dali, Hobbie, Sarah E., Holdaway, Robert J., Kembel, Steven W., Makita, Naoki, Onipchenko, Vladimir G., Picon-Cochard, Catherine, Reich, Peter B., de la Riva, Enrique G., Smith, Stuart W., Soudzilovskaia, Nadejda A., Tjoelker, Mark G., Wardle, David A., Roumet, Catherine, (2017). “Climate, soil and plant functional types as drivers of global fine-root trait variation.” *Journal of Ecology* 105(5): 1182–1196.
- Fukaki, H., Tasaka, M., 2009. Hormone interactions during lateral root formation. *Plant Molecular Biology*. <https://doi.org/10.1007/s11103-008-9417-2>
- Gardes, M., Bruns, T.D., 1993. ITS primers with enhanced specificity for basidiomycetes-application to the identification of mycorrhizae and rusts. *Molecular Ecology*.
- Garg, N., Chandel, S., 2010. Arbuscular mycorrhizal networks: process and functions. A review. *Agron Sustain Dev* 30, 581–599. <https://doi.org/10.1051/agro/2009054i>
- Gersani, M., Brown, J.S., O'Brien, E.E., Maina, G.M., Abramsky, Z., 2001. Tragedy of the commons as a result of root competition. *Journal of Ecology* 89, 660–669.
- Gordon, D.R., Welker, J.M., Menke, J.W., Rice, K.J., 1989. Competition for soil water between annual plants and blue oak (*Quercus douglasii*) seedlings. *Oecologia* 79, 533–541.
- Govindarajulu, M., Pfeffer, P.E., Jin, H., Abubaker, J., Douds, D.D., Allen, J.W., Bücking, H., Lammers, P.J., Shachar-Hill, Y., 2005. Nitrogen transfer in the arbuscular mycorrhizal symbiosis. *Nature* 435, 819–823. <https://doi.org/10.1038/nature03610>
- Graham, J.H., Leonard, R.T., Menge, J.A., 1982. Interaction of light intensity and soil temperature with phosphorus inhibition of vesicular-arbuscular mycorrhiza formation. *New Phytologist* 91, 683–690.
- Granato, T.C., Raper, C.D., 1989. Proliferation of maize (*Zea mays* L.) roots in response to localized supply of nitrate. *Journal of Experimental Botany* 40, 263–275. <https://doi.org/10.1093/jxb/40.2.263>
- Grime, J.P., 1974. Vegetation classification by reference to strategies. *Nature* 250, 26–31. <https://doi.org/10.1038/252497a0>
- Gruber, B.D., Giehl, R.F.H., Friedel, S., von Wieren, N., 2013. Plasticity of the *Arabidopsis* root system under nutrient deficiencies. *Plant Physiology* 163, 161–179. <https://doi.org/10.1104/pp.113.218453>
- Hackett, C., 1972. A Method of applying nutrients locally to roots under controlled conditions, and some morphological effects of locally applied nitrate on the branching of wheat roots. *Australian Journal of Biological Sciences* 25, 1169–1180. <https://doi.org/10.1071/BI9721169>
- Handley, W.R.C., 1963. Mycorrhizal associations and *Calluna* heathland afforestation. *Forestry Commission Bulletin* 36, 1–70.
- Harmer, R., Robertson, M., 2003. Seedling root growth of six broad-leaved tree species grown in competition with grass under irrigated nursery conditions. *Annals of Forest Science* 60, 601–608. <https://doi.org/10.1051/forest:2003052>
- He, F., Yang, B., Wang, H., Yan, Q., Cao, Y., He, X., 2016. Changes in composition and diversity of fungal communities along *Quercus mongolica* forests developments in Northeast China. *Applied Soil Ecology* 100, 162–171. <https://doi.org/10.1016/j.apsoil.2015.12.014>
- He, X., Bledsoe, C.S., Zasoski, R.J., Southworth, D., Horwath, W.R., 2006. Rapid nitrogen transfer from ectomycorrhizal pines to adjacent ectomycorrhizal and arbuscular mycorrhizal plants in a California oak woodland. *New Phytologist* 170, 143–151. <https://doi.org/10.1111/j.1469-8137.2006.01648.x>
- Heil, G.W., Bruggink, M., 1987. Competition for nutrients between *Calluna vulgaris* (L.) Hull and *Molinia caerulea* (L.) Moench. *Oecologia* 73, 105–107. <https://doi.org/10.1007/BF00376984>
- Hodge, A., 2004. The plastic plant: root responses to heterogeneous supplies of nutrients. *New Phytologist* 162, 9–24. <https://doi.org/10.1111/j.1469-8137.2004.01015.x>
- Holdridge, E.M., Cuellar-Gempeler, C., terHorst, C.P., 2016. A shift from exploitation to interference competition with increasing density affects population and community dynamics. *Ecology and Evolution* 6, 5333–5341. <https://doi.org/10.1002/ece3.2284>
- Horsley, S.B., 1977. Allelopathic inhibition of black cherry by fern, grass, goldenrod and aster. *Canadian Journal of Forest Research* 7, 205–216.
- Hossain, M.K., Anwar, S., Nandi, R., 2016. Allelopathic effects of *Mikania cordata* on forest and agricultural crops in Bangladesh. *Journal of Forestry Research (Harbin)* 27, 155–159. <https://doi.org/10.1007/s11676-015-0161-6>
- Inderjit, Callaway, R.M., 2003. Experimental designs for the study of allelopathy. *Plant Soil* 256, 1–11. <https://doi.org/10.1023/A:1026242418333>
- Inderjit, Mallik, A.U., 1999. Nutrient status of black spruce (*Picea mariana* [Mill.] BSP) forest soils dominated by *Kalmia angustifolia* L. *Acta Oecologica* 20, 87–92. [https://doi.org/10.1016/S1146-609X\(99\)80020-4](https://doi.org/10.1016/S1146-609X(99)80020-4)
- Inderjit, Mallik, A.U., 1996. The nature of interference potential of *Kalmia angustifolia*. *Canadian Journal of Forest Research* 26, 1899–1904.

- Javaid, A., 2008. Allelopathy in mycorrhizal symbiosis in the poaceae family. *Allelopathy Journal* 21, 207–218. <https://doi.org/citeulike-article-id:5726070>
- Javaid, A., 2007. Allelopathic interactions in mycorrhizal associations. *Allelopathy Journal* 20, 29–42.
- Johnson, N.C., Graham, J.H., Smith, F.A., 1997. Functioning of mycorrhizal associations along the mutualism-parasitism continuum. *New Phytologist* 135, 575–586. <https://doi.org/10.1046/j.1469-8137.1997.00729.x>
- Jones, M.D., Smith, S.E., 2004. Exploring functional definitions of mycorrhizas: Are mycorrhizas always mutualisms? *Canadian Journal of Botany* 82, 1089–1109. <https://doi.org/10.1139/B04-110>
- Kobae, Y., Ohmori, Y., Saito, C., Yano, K., Ohtomo, R., Fujiwara, T., 2016. Phosphate treatment strongly inhibits new arbuscule development but not the maintenance of arbuscule in mycorrhizal rice roots. *Plant Physiology* 171, 566–579. <https://doi.org/10.1104/pp.16.00127>
- Koide, R.T., Schreiner, R.P., 2003. Regulation of the vesicular-arbuscular mycorrhizal symbiosis. *Annual Review of Plant Physiology and Plant Molecular Biology* 43, 557–581. <https://doi.org/10.1146/annurev.pp.43.060192.003013>
- Kumar, S., Stecher, G., Li, M., Knyaz, C., Tamura, K., 2018. MEGA X: Molecular evolutionary genetics analysis across computing platforms. *Molecular Biology and Evolution* 35, 1547–1549. <https://doi.org/10.1093/molbev/msy096>
- Lamit, L.J., Holeski, L.M., Flores-Rentería, L., Whitham, T.G., Gehring, C.A., 2016. Tree genotype influences ectomycorrhizal fungal community structure: Ecological and evolutionary implications. *Fungal Ecology* 24, 124–134. <https://doi.org/10.1016/j.funeco.2016.05.013>
- Leski, T., Pietras, M., Rudawska, M., 2010. Ectomycorrhizal fungal communities of pedunculate and sessile oak seedlings from bare-root forest nurseries. *Mycorrhiza* 20, 179–190. <https://doi.org/10.1007/s00572-009-0278-6>
- Lewis, D.R., Negi, S., Sukumar, P., Muday, G.K., 2011. Ethylene inhibits lateral root development, increases IAA transport and expression of PIN3 and PIN7 auxin efflux carriers. *Development* 138, 3485–3495. <https://doi.org/10.1242/dev.065102>
- Linkohr, B.I., Williamson, L.C., Fitter, A.H., Leyser, H.M.O., 2002. Nitrate and phosphate availability and distribution have different effects on root system architecture of *Arabidopsis*. *Plant Journal* 29, 751–760. <https://doi.org/10.1046/j.1365-313X.2002.01251.x>
- Lu, X., Malajczuk, N., Dell, B., 1998. Mycorrhiza formation and growth of *Eucalyptus globulus* seedlings inoculated with spores of various ectomycorrhizal fungi. *Mycorrhiza* 8, 81–86. <https://doi.org/10.1007/s005720050216>
- Maestre, F.T., Callaway, R.M., Valladares, F., Lortie, C.J., 2009. Refining the stress-gradient hypothesis for competition and facilitation in plant communities. *Journal of Ecology* 97, 199–205. <https://doi.org/10.1111/j.1365-2745.2008.01476.x>
- Mallik, A., 1998. Allelopathy and competition in coniferous forests. *Environmental Forest Science*. 54, 309–315. https://doi.org/10.1007/978-94-011-5324-9_33
- Mallik, A.U., 2003. Conifer regeneration problems in boreal and temperate forests with ericaceous understory: role of disturbance, seed-bed limitation, and keystone species change. *Critical Reviews in Plant Sciences* 22, 341–366.
- Mallik, A.U., 1987. Allelopathic potential of *Kalmia angustifolia* to black spruce (*Picea mariana*). *Forest Ecology and Management* 20, 43–51. [https://doi.org/10.1016/0378-1127\(87\)90149-6](https://doi.org/10.1016/0378-1127(87)90149-6)
- Mallik, A.U., Biswas, S.R., Collier, L.C.S., 2016. Belowground interactions between *Kalmia angustifolia* and *Picea mariana*: roles of competition, root exudates and ectomycorrhizal association. *Plant Soil* 403, 471–483. <https://doi.org/10.1007/s11104-016-2819-z>
- McNickle, G.G., Dyzinski, R., 2013. Game theory and plant ecology. *Ecology Letters* 16, 545–555. <https://doi.org/10.1111/ele.12071>
- Michalet, R., Brooker, R.W., Cavieres, L.A., Kikvidze, Z., Lortie, C.J., Pugnaire, F.I., Valiente-Banuet, A., Callaway, R.M., 2006. Do biotic interactions shape both sides of the humped-back model of species richness in plant communities? *Ecology Letters* 9, 767–773. <https://doi.org/10.1111/j.1461-0248.2006.00935.x>
- Morris, M.H., Smith, M.E., Rizzo, D.M., Rejmánek, M., Bledsoe, C.S., 2008. Contrasting ectomycorrhizal fungal communities on the roots of co-occurring oaks (*Quercus* spp.) in a California woodland. *New Phytologist* 178, 167–176. <https://doi.org/10.1111/j.1469-8137.2007.02348.x>
- Näsholm, T., Högberg, P., Franklin, O., Metcalfe, D., Keel, S.G., Campbell, C., Hurry, V., Linder, S., Högberg, M.N., 2013. Are ectomycorrhizal fungi alleviating or aggravating nitrogen limitation of tree growth in boreal forests? *New Phytologist* 198, 214–221. <https://doi.org/10.1111/nph.12139>
- Nilsson, M.C., 1994. Separation of allelopathy and resource competition by the boreal dwarf shrub *Empetrum hermaphroditum* Hagerup. *Oecologia* 98, 1–7. <https://doi.org/10.1007/BF00326083>
- Nilsson, M.-C., Högberg, P., Zackrisson, O., Fengyou, W., 1993. Allelopathic effects by *Empetrum hermaphroditum* on development and nitrogen uptake by roots and mycorrhizae of *Pinus silvestris*. *Canadian Journal of Botany* 71, 620–628. <https://doi.org/10.1139/b93-071>
- Nilsson, R.H., Larsson, K.H., Taylor, A.F.S., Bengtsson-Palme, J., Jeppesen, T.S., Schigel, D., Kennedy, P., Picard, K., Glöckner, F.O., Tedersoo, L., Saar, I., Kõljalg, U., Abarenkov, K., 2019. The UNITE database for molecular identification of fungi: Handling dark taxa and parallel taxonomic classifications. *Nucleic Acids Research* 47, D259–D264. <https://doi.org/10.1093/nar/gky1022>
- Oberwinkler, F., Riess, K., Bauer, R., Selosse, M.A., Weiß, M., Garnica, S., Zuccaro, A., 2013. Enigmatic Sebaciniales. *Mycological Progress* 12, 1–27. <https://doi.org/10.1007/s11557-012-0880-4>
- Olsen, R.A., Odham, G., Lindeberg, G., 1971. Aromatic substances in leaves of *Populus tremula* as inhibitors of mycorrhizal fungi. *Physiologia Plantarum* 25, 122–129. <https://doi.org/10.1111/j.1399-3054.1971.tb01100.x>
- Otsing, E., Tedersoo, L., 2015. Temporal dynamics of ectomycorrhizal fungi and persistence of *Tuber melanosporum* in inoculated *Quercus robur* seedlings in North Europe. *Mycorrhiza* 25, 61–66. <https://doi.org/10.1007/s00572-014-0591-6>
- Pardales, J.R., Kono, Y., Yamauchi, A., Iijima, M., 1992. Seminal root growth in sorghum (*Sorghum bicolor*) under allelopathic influences from residues of taro (*Colocasia esculenta*). *Annals of Botany* 69, 493–496. <https://doi.org/10.1093/oxfordjournals.aob.a088377>
- Pellissier, F., 1994. Effect of phenolic compounds in humus on the natural regeneration of spruce. *Phytochemistry* 36, 865–867. [https://doi.org/10.1016/S0031-9422\(00\)90452-7](https://doi.org/10.1016/S0031-9422(00)90452-7)
- Persson, J., Näsholm, T., 2001. Amino acid uptake: a widespread ability among boreal forest plants. *Ecology Letters* 4, 434–438. <https://doi.org/10.1046/j.1461-0248.2001.00260.x>
- Picon-Cochard, C., Coll, L., Balandier, P., 2006. The role of below-ground competition during early stages of secondary succession: the case of 3-year-old Scots pine (*Pinus sylvestris* L.) seedlings in an abandoned grassland. *Oecologia* 148, 373–383. <https://doi.org/10.1007/s00442-006-0379-2>
- Postma, J.A., Dathe, A., Lynch, J.P., 2014. The optimal lateral root branching density for maize depends on nitrogen and phosphorus availability. *Plant Physiology* 166, 590–602. <https://doi.org/10.1104/pp.113.233916>
- Pugnaire, F.I., Luque, M.T., 2001. Changes in plant interactions along a gradient of environmental stress. *Oikos* 93, 42–49.
- Reich, P.B., 2014. The world-wide “fast-slow” plant economics spectrum: A traits manifesto. *Journal of Ecology* 102, 275–301. <https://doi.org/10.1111/1365-2745.12211>
- Richard, F., Roy, M., Shahin, O., Sthultz, C., Duchemin, M., Joffre, R., Selosse, M.A., 2011. Ectomycorrhizal communities in a Mediterranean forest ecosystem dominated by *Quercus ilex*: Seasonal dynamics and response to drought in the surface organic horizon. *Annals of Forest Science* 68, 57–68. <https://doi.org/10.1007/s13595-010-0007-5>

- Richard, F., Selosse, M.A., Gardes, M., 2009. Facilitated establishment of *Quercus ilex* in shrub-dominated communities within a Mediterranean ecosystem: Do mycorrhizal partners matter? *FEMS Microbiology Ecology* 68, 14–24. <https://doi.org/10.1111/j.1574-6941.2009.00646.x>
- Robinson, R.K., 1972. The production by roots of *Calluna Vulgaris* of a factor inhibitory to growth of some mycorrhizal fungi. *The Journal of Ecology* 60, 219–224. <https://doi.org/10.2307/2258051>
- Roche, M.D., Pearse, I.S., Bialic-Murphy, L., Kivlin, S.N., Sofaer, H.R., Kalisz, S., 2021. Negative effects of an allelopathic invader on AM fungal plant species drive community-level responses. *Ecology* 102, 1–12. <https://doi.org/10.1002/ecy.3201>
- Schaller, M., Schroth, G., Beer, J., Jiménez, F., 2003. Root interactions between young *Eucalyptus deglupta* trees and competitive grass species in contour strips. *Forest Ecology and Management* 179, 429–440. [https://doi.org/10.1016/S0378-1127\(02\)00534-0](https://doi.org/10.1016/S0378-1127(02)00534-0)
- Schöb, C., Callaway, R.M., Anthelme, F., Brooker, R.W., Cavieres, L.A., Kikvidze, Z., Lortie, C.J., Michalet, R., Pugnaire, F.I., Xiao, S., Cranston, B.H., García, M.-C., Hupp, N.R., Llambí, L.D., Lingua, E., Reid, A.M., Zhao, L. and Butterfield, B.J. (2014a), The context dependence of beneficiary feedback effects on benefactors in plant facilitation. *New Phytologist* 204: 386–396. <https://doi.org/10.1111/nph.12908>
- Schöb, C., Michalet, R., Cavieres, L.A., Pugnaire, F.I., Brooker, R.W., Butterfield, B.J., Cook, B.J., Kikvidze, Z., Lortie, C.J., Xiao, S., Hayek, P. al, Bagousse-pinguet, Y., Anya, M., Anthelme, F., Cranston, B.H., Garc, M., 2014b. A global analysis of bidirectional interactions in alpine plant communities shows facilitators experiencing strong reciprocal fitness costs. *New Phytologist* 202, 95–105. <https://doi.org/10.1111/nph.12641>
- Schroth, G., 1999. A review of belowground interactions in agroforestry, focussing on mechanisms and management options. *Agroforestry Systems* 43, 5–34. https://doi.org/10.1007/978-94-017-0679-7_1
- Selosse, M.A., Bocayuva, M.F., Kasuya, M.C.M., Courty, P.E., 2017. Mixotrophy in mycorrhizal plants: Extracting carbon from mycorrhizal networks, in: Martin, F. (Ed.), *Molecular Mycorrhizal Symbiosis*. John Wiley, Hoboken, pp. 451–471. <https://doi.org/10.1002/9781118951446.ch25>
- Shahzad, Z., Amtmann, A., 2017. Food for thought: how nutrients regulate root system architecture. *Current Opinion in Plant Biology* 39, 80–87. <https://doi.org/10.1016/j.pbi.2017.06.008>
- Shkolnik-Inbar, D., Bar-Zvi, D., 2010. ABI4 mediates abscisic acid and cytokinin inhibition of lateral root formation by reducing polar auxin transport in *Arabidopsis*. *Plant Cell* 22, 3560–3573. <https://doi.org/10.1105/tpc.110.074641>
- Simard, S.W., Durall, D.M., 2004. Mycorrhizal networks: a review of their extent, function, and importance. *Canadian Journal of Botany* 82, 1140–1165. <https://doi.org/10.1139/b04-116>
- Smith, M.E., Douhan, G.W., Fremier, A.K., Rizzo, D.M., 2009. Are true multihost fungi the exception or the rule? Dominant ectomycorrhizal fungi on *Pinus sabiniana* differ from those on cooccurring *Quercus* species. *New Phytologist* 182, 287–290. <https://doi.org/10.1111/j.1469-8137.2009.02808.x>
- Tedersoo, L., Bahram, M., Zobel, M., 2020. How mycorrhizal associations drive plant population and community biology. *Science* (1979) 367, 1–9. <https://doi.org/10.1126/science.aba1223>
- Tilman, D., 1990. Constraints and tradeoffs: toward a predictive theory of competition and succession. *Oikos* 58, 3–15. <https://doi.org/10.2307/3565355>
- Timbal, J., Gelpe, J., Garbaye, J., Courrier, G., 1990. Étude préliminaire sur l'effet dépressif de la molinie (*Molinia caerulea*) sur la croissance et l'état mycorrhizien de semis de chêne rouge (*Quercus rubra*). *Annales of Forest Science* 21, 643–649.
- Toju, H., Yamamoto, S., Sato, H., Tanabe, A.S., 2013a. Sharing of diverse mycorrhizal and root-endophytic fungi among plant species in an oak-dominated cool-temperate forest. *PLoS One* 8, 1–13. <https://doi.org/10.1371/journal.pone.0078248>
- Toju, H., Yamamoto, S., Sato, H., Tanabe, A.S., Gilbert, G.S., Kadowaki, K., 2013b. Community composition of root-associated fungi in a *Quercus*-dominated temperate forest: “codominance” of mycorrhizal and root-endophytic fungi. *Ecology and Evolution* 3, 1281–1293. <https://doi.org/10.1002/ece3.546>
- Tomlinson, K.W., Sterck, F.J., Barbosa, E.R.M., de Bie, S., Prins, H.H.T., van Langevelde, F., 2019. Seedling growth of savanna tree species from three continents under grass coition and nutrient limitation in a greenhouse experiment. *Journal of Ecology* 107, 1051–1066. <https://doi.org/10.1111/1365-2745.13085>
- Tsakalimi, M., Tsitsoni, T., Ganatsas, P., Zagas, T., 2009. A comparison of root architecture and shoot morphology between naturally regenerated and container-grown seedlings of *Quercus ilex*. *Plant Soil* 324, 103–113. <https://doi.org/10.1007/s1104-009-9974-4>
- Van Der Heijden, M.G.A., Bardgett, R.D., Van Straalen, N.M., 2008. The unseen majority: Soil microbes as drivers of plant diversity and productivity in terrestrial ecosystems. *Ecology Letters* 11, 651. <https://doi.org/10.1111/j.1461-0248.2008.01199.x>
- Van Der Heijden, M.G.A., Horton, T.R., 2009. Socialism in soil? the importance of mycorrhizal fungal networks for facilitation in natural ecosystems. *Journal of Ecology* 97, 1139–1150. <https://doi.org/10.1111/j.1365-2745.2009.01570.x>
- Venturella, G., Altobelli, E., Bernicchia, A., di Piazza, S.D., Donnini, D., Gargano, M.L., Gorjón, S.P., Granito, V.M., Lantieri, A., Lunghini, D., Montemartini, A., Padovan, F., Pavarino, M., Pecoraro, L., Perini, C., Rana, G., Ripa, C., Salerni, E., Savino, E., Tomei, P.E., Vizzini, A., Zambonelli, A., Zotti, M., 2011. Fungal biodiversity and in situ conservation in Italy. *Plant Biosystems* 145, 950–957. <https://doi.org/10.1080/11263504.2011.633115>
- Vernay, A., Balandier, P., Guinard, L., Améglio, T., Malagoli, P., 2016. Photosynthesis capacity of *Quercus petraea* (Matt.) saplings is affected by *Molinia caerulea* (L.) under high irradiance. *Forest Ecology and Management* 376, 107–117. <https://doi.org/10.1016/j.foreco.2016.05.045>
- Vernay, A., Malagoli, P., Fernandez, M., Perot, T., Améglio, T., Balandier, P., 2018a. Improved *Deschampsia cespitosa* growth by nitrogen fertilization jeopardizes *Quercus petraea* regeneration through intensification of competition. *Basic and Applied Ecology* 31, 21–32. <https://doi.org/10.1016/j.baae.2018.06.002>
- Vernay, A., Malagoli, P., Fernandez, M., Perot, T., Améglio, T., Balandier, P., 2018b. Carry-over benefit of high internal N pool on growth and function of oak seedlings (*Quercus petraea*) competing with *Deschampsia cespitosa*. *Forest Ecology and Management* 419–420, 130–138. <https://doi.org/10.1016/j.foreco.2018.03.039>
- Vicca, S., Gilgen, A. K., Camino Serrano, M., Dreesen, F. E., Dukes, J. S., Estiarte, M., Gray, S. B., Guidolotti, G., Hoepfner, S. S., Leakey, A. D. B., Ogaya, R., Ort, D. R., Ostrogovic, M. Z., Rambal, S., Sardans, J., Schmitt, M., Siebers, M., van der Linden, L., van Straaten, O., Granier, A., 2012. Urgent need for a common metric to make precipitation manipulation experiments comparable. *New Phytologist* 195: 518–522.
- Vincenot, L., Nara, K., Sthultz, C., Labbé, J., Dubois, M.P., Tedersoo, L., Martin, F., Selosse, M.A., 2012. Extensive gene flow over Europe and possible speciation over Eurasia in the ectomycorrhizal basidiomycete *Laccaria amethystina* complex. *Molecular Ecology* 21, 281–299. <https://doi.org/10.1111/j.1365-294X.2011.05392.x>
- Voříšková, J., Brabcová, V., Cajthaml, T., Baldrian, P., 2014. Seasonal dynamics of fungal communities in a temperate oak forest soil. *New Phytologist* 201, 269–278. <https://doi.org/10.1111/nph.12481>
- Walker, G.R., Mallik, A.U., 2009. Black spruce reforestation in *Kalmia* heath: seedling response to forest floor mixing and mycorrhizal inoculation with *Paxillus involutus*. *Canadian Journal of Forest Research* 39, 2007–2020. <https://doi.org/10.1139/x09-115>
- Watt, A.S., 1919. On the causes of failure of natural regeneration in British oakwoods. *Journal of Ecology* 7, 173–203.
- Weber, C.F., Vilgalys, R., Kuske, C.R., 2013. Changes in fungal community composition in response to elevated atmospheric CO₂ and

- nitrogen fertilization varies with soil horizon. *Frontiers in Microbiology* 4, 1–14. <https://doi.org/10.3389/fmicb.2013.00078>
- Weiß, M., Waller, F., Zuccaro, A., Selosse, M.A., 2016. Sebaciales - one thousand and one interactions with land plants. *New Phytologist* 211, 20–40. <https://doi.org/10.1111/nph.13977>
- Welker, J.M., Gordon, D.R., Rice, K.J., 1991. Capture and allocation of nitrogen by *Quercus douglasii* seedlings in competition with annual and perennial grasses. *Oecologia* 87, 459–466. <https://doi.org/10.1007/BF00320407>
- White, T.J., Bruns, T., Lee, S., Taylor, J., 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics, in: Innis, M., Gelfand, D., Sninsky, J., White, T. (Eds.), *PCR Protocols: A Guide to Methods and Applications*. Academic Press., San Diego, pp. 315–322. <https://doi.org/10.1016/b978-0-12-372180-8.50042-1>
- Wu, Y.T., Wubet, T., Trogisch, S., Both, S., Scholten, T., Bruehlheide, H., Buscot, F., 2013. Forest age and plant species composition determine the soil fungal community composition in a Chinese subtropical forest. *PLoS One* 8, 1–12. <https://doi.org/10.1371/journal.pone.0066829>
- Zangaro, W., Lescano, L.E.A.M., Matsuura, E.M., Rondina, A.B.L., Nogueira, M.A., 2018. Interactions between arbuscular mycorrhizal fungi and exotic grasses differentially affect the establishment of seedlings of early- and late-successional woody species. *Applied Soil Ecology* 124, 394–406. <https://doi.org/10.1016/j.apsoil.2017.12.003>
- Zhang, H., Forde, B.G., 1998. An *Arabidopsis* MADS box gene that controls nutrient-induced changes in root architecture. *Science* (1979) 279, 407–409. <https://doi.org/10.1126/science.279.5349.407>
- Zhang, H., Jennings, A., Barlow, P.W., Forde, B.G., 1999. Dual pathways for regulation of root branching by nitrate. *Proceedings of the National Academy of Sciences* 96, 6529–6534.
- Zhu, H., Mallik, A.U., 1994. Interactions between *Kalmia* and black spruce: isolation and identification of allelopathic compounds. *Journal of Chemical Ecology* 20, 407–421. <https://doi.org/10.1007/BF02064447>