



**INTERNATIONAL JOURNAL OF
PHARMACEUTICAL SCIENCES**
[ISSN: 0975-4725; CODEN(USA): IJPS00]
Journal Homepage: <https://www.ijpsjournal.com>



Review Article

Formulation and Evaluation of Herbal Antibacterial Cream using *Tridax procumbens* Leaf Extract

Lina Nagpure*, Prachi Dashwant, Shraddha Pisal, Avadhut Dukandar

Department of Pharmaceutics, Ashokrao Mane College of Pharmacy, Peth Vadgaon, Shivaji University, Maharashtra, India 416112

ARTICLE INFO

Published: 27 May 2026

Keywords:

Asteraceae family,
phytochemical
characterisation, vitro
antibacterial assays,
staphylococcus aureus,
assessment etc

DOI:

10.5281/zenodo.20404726

ABSTRACT

Tridax procumbens, commonly referred to as coat buttons or tridax daisy, is a widely distributed plant in the Asteraceae family with a rich history in traditional Ayurvedic medicine. Over the last two decades, extensive scientific research has proven its medicinal properties, particularly its strong antibacterial action, on phytochemical characterisation, in vitro antibacterial assays, cream formulation design, physicochemical criteria for assessment, stability testing, and improved nano-dispense methods. The evidence consistently shows that ethanolic, methanolic, and aqueous leaf extracts of *T. procumbens* have broad-spectrum antibacterial activity against clinically significant pathogens such as *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Klebsiella pneumonia*, *Bacillus subtilis*, and *Bacillus cereus*.

INTRODUCTION

The worldwide propagation of antibiotic-resistant bacteria has produced an urgent demand for new antimicrobial medicines derived from natural sources. Medicinal plants have long been the core of conventional healthcare systems in Asia, Africa, and Latin America, and contemporary pharmacological research supports their therapeutic claims. Among these, *Tridax procumbens* (L.), a member of the family Asteraceae, stands out as a plant with outstanding medicinal value that is currently inadequately

utilised in mainstream pharmaceutical development.

T. procumbens is a prostrate, branching annual herb that grows as a weed throughout tropical and subtropical climates such as India, West Africa, Southeast Asia, and the Americas. Despite its weed designation, it has been used in folk medicine studies as an anticoagulant, wound healer, antiseptic, antifungal agent, insect repellent, and hepatoprotective treatments. It has been prescribed by Indian Ayurvedic practitioners as an alternative for "Beringia," a well-known hair

*Corresponding Author: Lina Nagpure

Address: Ashokrao Mane College of Pharmacy, Peth Vadgaon, Shivaji University, Maharashtra, India 416112.

Email ✉: linanagpure1804@gmail.com

Relevant conflicts of interest/financial disclosures: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.



growth booster. The plant's leaves, stems, blossoms, and roots have all been studied for their therapeutic properties, with the leaves receiving the most scientific interest.

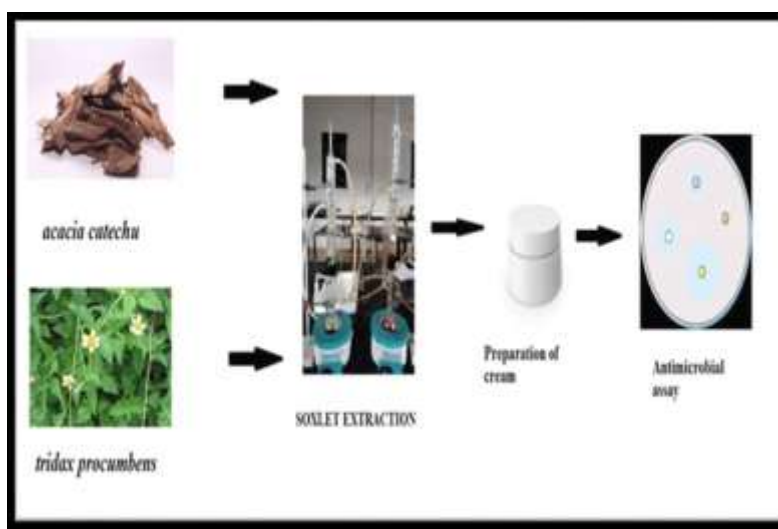


Fig No-1

The development of herbal antibacterial creams represents a tremendous opportunity to convert *T. procumbens*' raw antimicrobial potential into a standardised, patient-acceptable topical dose form. Creams, whether oil-in-water (O/W) or water-in-oil (W/O) emulsions, have several advantages, including ease of application, good skin penetration, cosmetic acceptability, and the potential to transport both hydrophilic and lipophilic active components. Such formulations are evaluated using a series of physicochemical tests to assure safety, stability, and medicinal efficacy.

This study critically examines 30 published research publications on the phytochemistry, pharmacology, formulation, and efficacy of *T. procumbens*-based herbal antibacterial creams and closely related topical treatments. The papers are organised thematically to produce a cohesive narrative that includes botanical background, phytochemical profiling, antibacterial mechanisms, formulation tactics, assessment parameters, and future directions.

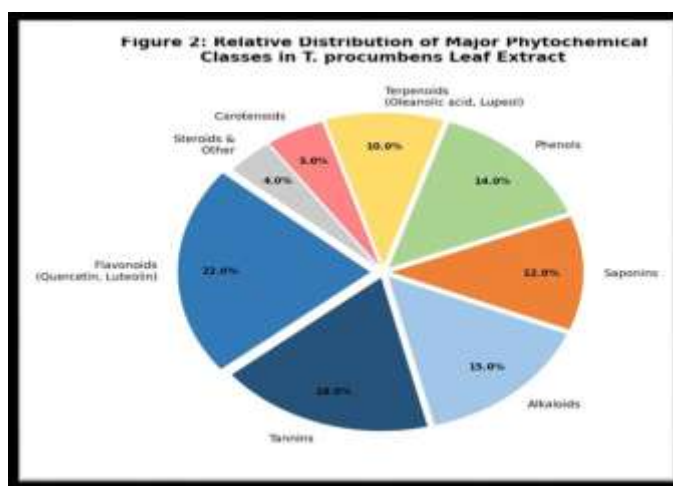


Fig No-2: Relative Distribution of Major Phytochemical Classes in *T. procumbens* Leaf Extract

In the world, India is well-known for its indigenous valuable flora and fauna with excellent therapeutic potential. India is the world's largest source of luxuriant flora and vast variety of medicinally beneficial plants, which play an important role in people's health. Medicinal plant use is regarded a "living tradition" in many underdeveloped countries, including India. The reliance on plants is a significant component of Indian cultural heritage, which is represented in practices and lifestyles throughout the country.

India has a rich history of historically valued medicine. Ayurveda, Unani, Siddha, Amechi, and Homoeopathy are among the various medicinal systems practiced in India. Many people use a huge number of plants for disease therapy encompassing both human and animal ailments. The plants employed in treatments are referred to as "valuable" medicinal plants. It is no surprise that one-fourth of the world's population, i.e. 1.42 billion people, use medicinal plants as home remedies. According to World Health Organization (WHO) data, in nations with limited resources, approximately 80% of people consume traditional medicines to maintain mental health and vigour. According to one estimate throughout the entire world, 20,000 to 35,000 species have been used as medications, pharmaceuticals, cosmetics, and nutraceuticals. [1]

Botanical Profile and Ethnomedicinal Background

Taxonomy and Morphology

The plant is designated as Kingdom Plantae, Family Asteraceae, Genus *Tridax*, and Species *procumbens*. Morphologically, it is a prostrate herb with opposite, lanceolate leaves with serrated sides and little yellow ray flowers. The plant's geographical distribution includes India, West Africa, Australia, and tropical America, as well as

its traditional usage in many cultures, such as use as a poultice for wounds, diarrhoea management, and skin infection treatment.

T. procumbens is a semi-prostrate and annual creeper herb with branching, sparsely hairy stems that root at nodes and reach heights of 30-50 cm. Leaf structures are single, opposite, serrate or dentate, sharp, fleshy, pubescent, exstipulate, lanceolate to ovate in shape with 3-7 cm long, irregularly serrated margin with wedge-shaped base, slightly petioles, and hairy on both surfaces. The leaves are dorsiventral, having a single layer of epidermis on both surfaces and a thick cuticle. The upper epidermis has a single layer of multicellular covering trichomes, whereas the lower epidermis has a single layer of elongated cells that are densely packed. The crystals of calcium can be seen in vessels in the xylem. Vascular bundles are concentric in form. Meristeeel is composed of a single, centrally situated collateral vascular bundle surrounded by some parenchymatous cells.

Flowers are tubular in design, yellow in colour, and have hairs on the capitulum part of the inflorescence [4, 13, 14]. This plant produces two types of flowers: ray florets and disc florets with basal placentar development [13].



Fig No-3: *Tridax procumbens* plant

Fruit is a rigid achene with stiff hairs and a feathery, plume-like white pappus on one end that aids in aerial distribution. The heads are diverse, with long peduncles that can reach heights of up to 2 feet. The ray florets are female, with a ligulate corolla, trifold, and always pale-yellow in colour.

T. procumbens seeds germinate at warmer temperatures (35/25 and 30/20 °C) with 58-78% light. These are particularly susceptible to salt concentration and water stress [15]. Gametes have 36 (diploid) and 18 (haploid) chromosomes [8]. Propagation occurs through spreading stems and seed production [4].



Fig No-4: Botanical specimen

Botanical Description

Kingdom: Plantae | Sub-kingdom: Tracheobionta |
Division: Magnoliophyta | Class: Magnoliopsida |
Sub-class: Asteridae | Order: Asterales | Family:
Asteraceae | Genus: *Tridax* | Species: *procumbens*

Vernacular Names

English: Coat Buttons and Tridax Daisy | Hindi:
Ghamra | Sanskrit: Jayanti Veda | Marathi: Dagadi
Pala | Telugu: Gaddi Chemanthi | Tamil: Thata
poodu | Malayalam: Chiravanak | Spanish: Cadillp
Chisaca | French: Herbe Caille | Chinese:
Kotobukigiku (Satish et al., 2012)

Phytochemical Characterization

Qualitative and Quantitative Phytochemical Screening

Saponins, steroids, diterpenes, alkaloids, flavonoids, phenols, and tannins were found in the leaves of *Tridax procumbens*. The methodical use of conventional colour reactions and precipitation assays resulted in a credible phytochemical fingerprint for the plant. The study emphasised that the co-presence of flavonoids and tannins is especially relevant for antibacterial activity, as both chemical groups are known to damage bacterial cell membranes and precipitate proteins.

A quantitative phytochemical and antioxidant analysis revealed that *T. procumbens* ethanolic extracts had significantly higher total phenolic content (TPC) and total flavonoid content (TFC) than many other commonly used medicinal plants. High TPC values correlate with substantial DPPH radical scavenging activity, and because oxidative stress plays a role in bacterial pathogenesis, the extract's antioxidant and antibacterial dual action is clinically relevant.

HPLC analysis was used to isolate and identify quercetin and rutin from *T. procumbens* leaves. Quercetin, a flavonoid aglycone, has been extensively tested for antibacterial action via mechanisms such as DNA gyrase inhibition, cytoplasmic membrane disruption, and suppression of fatty acid production. Rutin, a glycoside derivative, enhances anti-inflammatory action. [7]

A systematic and extensive review, which included 101 references and screened 839 papers, identified over 138 chemical compounds isolated or characterised from *T. procumbens*. These include luteolin, quercetin, isquercetin, glucoluteolin, carotenoids, beta-sitosterol,

oleanolic acid, lupeol, fumaric acid, radixin, betaine, pentacyclic triterpenes, alkyl esters, and fatty acids. [8]

Advanced Phytochemical Profiling

Cutting-edge work using ultrasonic-assisted extraction, ultra-performance liquid chromatography, and quadrupole time-of-flight mass spectrometry achieved remarkable phytochemical resolution. The study carefully assessed the impact of various solvent polarity on phytochemical yield and discovered a diverse matrix of tannins, flavonoids, phenolic acids, and terpenoids. This work presents a thorough,

reproducible extraction procedure for industrial-scale cream formulation. [9]

Individual fractions of ethyl acetate extract were separated and evaluated for biological activity using column chromatography. The F4-5 fraction demonstrated the highest antibacterial inhibition against *E. coli*, *S. aureus*, *B. subtilis*, and *Proteus mirabilis*, whereas different fractions had the highest antioxidant or xanthine oxidase inhibitory activity. This bioactivity-guided fractionation approach is essential for determining the exact active fractions that are most suited for cream formulation. [10]

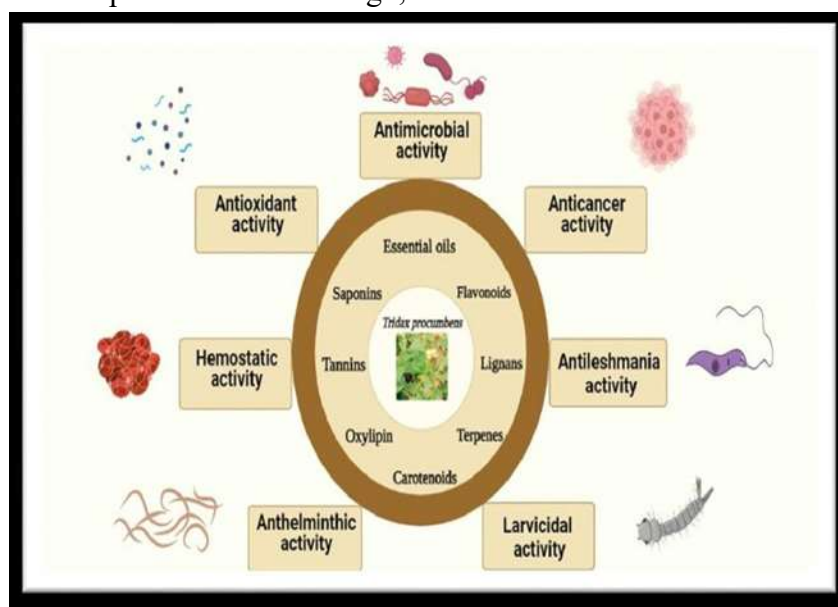


Fig No-5: Pharmacological activities of *T. procumbens*

In Vitro Antibacterial Studies

Agar Diffusion and Minimum Inhibitory Concentration Studies

The agar well diffusion method was used to determine the phytochemical content and antibacterial activity of *T. procumbens* leaf extracts against three gram-positive and three gram-negative bacterial species. The chloroform extract had the strongest antibacterial action, producing results comparable to the standard

medication ampicillin. The MIC for ethanol and methanol extracts against examined species ranged from 1.96 to 19.5 mg/mL, with *S. aureus* being the most susceptible. [11]

A systematic examination of acetone and methanolic extracts of all plant parts (stem, root, leaf, flower, and whole plant) against *E. coli*, *Klebsiella pneumoniae*, *Proteus vulgaris* (gram-negative), and *B. subtilis* and *S. aureus* (gram-positive) revealed that ethanol and methanolic extracts had broad-spectrum efficacy against all

test organisms. Root extracts in chloroform and petroleum ether exhibited no efficacy against Gram-negative bacteria. [12]

In a study aimed specifically at bovine mastitis-causing *Staphylococcus aureus*, both aqueous and methanolic leaf extracts of *T. procumbens*

exhibited considerable in vitro antibacterial activity. This study is relevant since *S. aureus* is the most prevalent skin pathogen found in topical antibacterial cream applications, demonstrating the clinical utility of *T. procumbens*-based formulations for skin infections. [13]

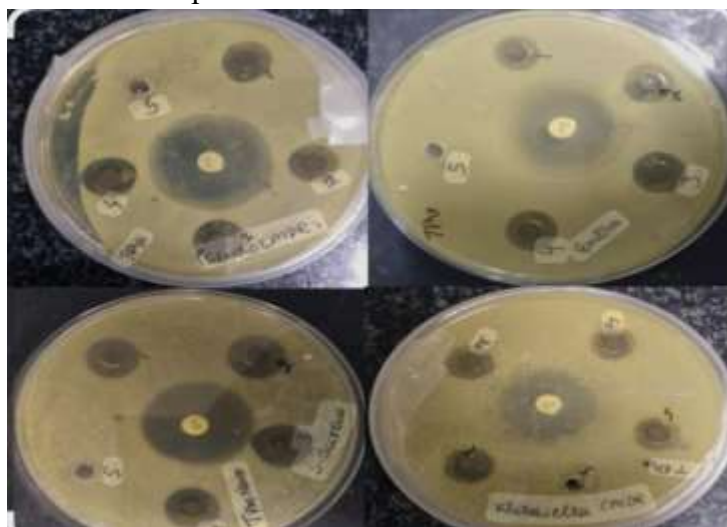


Fig No-6: Antibacterial assay results

Mechanisms of Antibacterial Action

T. procumbens' bactericidal action against clinically relevant infections such as *S. aureus* and *P. aeruginosa* has been attributed to mechanisms involving cell membrane rupture and bioactive chemical intercalation inside bacterial nucleic acids. The flavonoid-rich fractions appear to hinder DNA gyrase and topoisomerase IV, enzymes required for bacterial DNA replication. Tannins denature bacterial membrane proteins through protein precipitation, resulting in cytoplasmic leakage and cell death.

A novel study combined ethanolic *T. procumbens* extract and chitosan to produce a composite with improved antibacterial efficacy. The composite was tested for sensitivity against *E. coli*, *S. aureus*, *P. aeruginosa*, *K. pneumoniae*, *A. niger*, *M. canis*, and *C. acremonium* using agar well diffusion and double serial dilution (MIC/MBC determination). The chitosan-extract combination demonstrated substantially higher zones of inhibition than either drug alone, indicating synergistic interaction. [14]

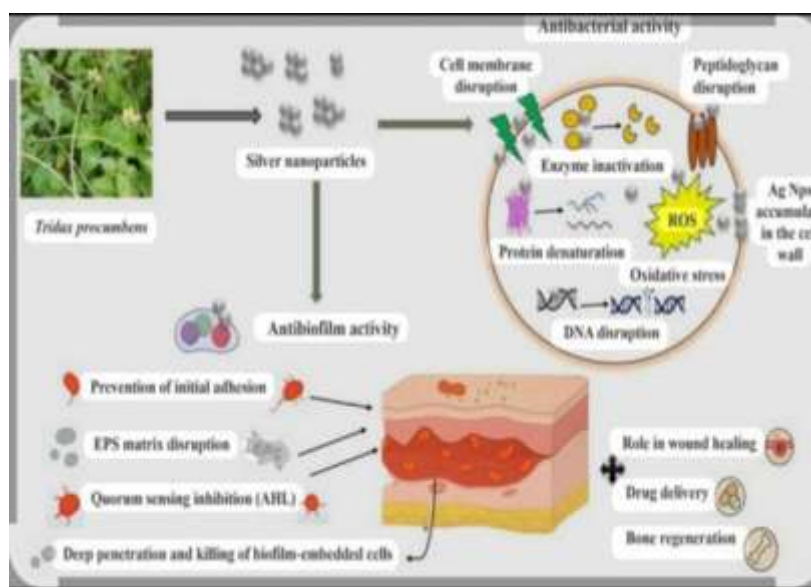


Fig No-7: Antibacterial mechanism of silver nanoparticles

Formulation of Herbal Antibacterial Creams

Formulation Design and Excipient Selection

Preparation of the cream followed a systematic procedure involving both oil and water phases (Table 1). Initially, the oil phase was prepared by combining 5 gm of Liquid Paraffin and 1.6 gm of White Beeswax, which were gently heated using a double boiler or microwave until the wax melted completely and a homogeneous mixture was obtained. The temperature during this step was maintained between 70 and 75 °C, and occasional stirring ensured complete melting and uniformity.

Concurrently, the water phase was prepared by dissolving 0.08 gm of Borax in 3.3 ml of Purified Water, also heated to approximately 70 °C, ensuring the borax dissolved completely. [18]

Once both phases reached the appropriate temperature, the heated water phase was slowly poured into the oil phase with continuous stirring to initiate emulsification. The mixture was stirred vigorously until a stable emulsion began to form. This blending step was crucial for achieving the proper consistency and stability of the final product.

After the emulsion was formed, the mixture was allowed to cool to less than 40 °C. During the cooling process, the cream began to thicken, developing the desired semi-solid texture. Once the temperature dropped sufficiently, preservatives and Rose Water were added to the cooled cream, followed by thorough stirring to ensure uniform distribution of all components. The result was a stable and homogeneous cream formulation ready for further evaluation and use. [19]

Table No. 1: Formula for Cream Base

Ingredients	Quantity (for 10 gm)	Role of Ingredients
Liquid Paraffin	5 gm	Moisturizer
White Beeswax	1.6 gm	Penetration Enhancer
Borax	0.08 gm	Stabilizer
Methyl Paraben	0.018 gm	Preservative
Propyl Paraben	0.002 gm	Preservative
Rose Water	q.s	Perfume
Purified Water	3.3 ml	Vehicle

Table No. 2: Cream Concentrations

Cream	Concentration of Acacia catechu	Concentration of Tridax procumbens
1st	1%	1%
2nd	2%	2%
3rd	3%	3%



4th	3%	-
5th	-	3%

Evaluation Parameters for Herbal Antibacterial Creams

Physicochemical Evaluation

The evaluation of a cream involves assessing its organoleptic properties, physical characteristics, and other attributes that determine its effectiveness and appeal to consumers. Further evaluation has been conducted on higher concentration of cream by incorporating 3% of Acacia catechu and Tridax procumbens.

Organoleptic evaluation is a crucial aspect of assessing the quality and acceptability of a cream formulation through sensory parameters such as appearance, colour, odour, and texture. The appearance of the cream is initially examined visually to ensure uniformity and the absence of any phase separation or irregularities; a well-formulated cream should appear smooth and consistent without visible separation of oil and water phases. The colour is also visually inspected to confirm consistency; it should typically be clear, white, or slightly off-white, depending on the ingredients used. The odour is assessed by smelling the cream to detect any unpleasant or undesirable scents. Finally, the texture is evaluated by applying a small quantity of the cream to the skin or rubbing it between the fingers.

pH Determination

The pH of the prepared formulation Tridax procumbens phospholipid complex (phytosome) gel was determined to be between 6 and 7.5, which falls within the typical pH range of the skin. Specifically, the pH was found to be 7.35. Hence, the formulated Tridax procumbens phospholipid

complex (phytosome) gel is within the skin pH range.

Determination of Spreadability

Spreadability is a key factor in patient adherence, as it ensures even distribution of the Tridax procumbens phytosomal gel on the skin. The spreading coefficient values for the formulated gel indicate that it spreads easily on the skin's surface. The spreadability test reveals a value of 27.5, which indicates excellent spreading properties.

Determination of Viscosity

The viscosity of the Tridax procumbens phospholipid complex (phytosome) gel was found to be 7100 cps at 5 rpm, indicating that it has a semi-solid texture, which is appropriate for topical use. The high viscosity at low shear rates suggests shear-thinning behavior, which is essential for ease of application and better skin retention.

FTIR Analysis

FTIR analysis was performed using a Bruker-Alpha II. The range of wavenumbers used to capture the spectrum was 4000 to 500 cm^{-1} . The FTIR spectra of the Tridax procumbens extract, Tridax procumbens phospholipid complex (phytosome) and Tridax procumbens phytosomal gel were analysed along with the values of their principal peaks.

Washability

The cream was easily removed by washing with water, indicating good washability. No oily residue or stickiness remained on the skin after washing. The formulation showed satisfactory cleansing properties.

Evaluation of Cream Antimicrobial Assay



Table No. 3: Evaluation Parameters

Evaluation Parameters	Observation
Appearance	Clear
Odour	Characteristic
pH	5.5
State	Semisolid
Consistency	Smooth
Spreadability	Spreadable
Washability	Washable
Phase Separation	No phase separation
Homogeneity	Homogeneous

**Fig 8 & 9: Antimicrobial assay against E. coli and Bacillus subtilis**

DISCUSSION

The antibacterial, antifungal, anti-inflammatory, antioxidant, and wound healing properties of *T. procumbens* translate into a multi-target topical product that may outperform single-mechanism synthetic antibiotics, particularly against biofilm-forming and multi-drug-resistant pathogens. A significant strength of *T. procumbens*-based cream research is the consistency of findings across geographically diverse research groups operating in India, Nigeria, Indonesia, Thailand, and other countries. This geographical validation mitigates concerns about chemotype variability and suggests that the core antibacterial phytoconstituent profile is robustly maintained across growing conditions.

Several research gaps emerge from this review. First, while in vitro antibacterial activity is

extensively documented, in vivo clinical studies on human subjects remain scarce. The transition from laboratory data to clinical proof-of-concept is essential for regulatory acceptance. Second, formal toxicological profiling — including acute oral toxicity, dermal sensitization, and repeated-dose toxicity — is incompletely documented. Third, pharmacokinetic data on skin penetration and percutaneous absorption of *T. procumbens* phytoconstituents from cream matrices is largely absent, making it difficult to correlate in vitro release with in vivo therapeutic outcomes. Fourth, standardization of raw material (leaf extract) in terms of marker compound content (e.g., quercetin percentage) is inconsistently applied, making cross-study comparisons challenging.

The integration of nanotechnology addresses several of these limitations. Silver nanoparticle

formulations have demonstrated enhanced skin permeation, improved bioavailability, and superior antibacterial activity in animal wound models, providing a pathway toward rigorous in vivo validation. Phytosome complexation improves the oral bioavailability of polar phytoconstituents and is increasingly being applied to topical systems. From a regulatory perspective, the extensive traditional use history of *T. procumbens* across multiple cultures provides a safety precedent that can support Generally Recognized as Safe (GRAS) designation for cosmetic-grade preparations. [26]

CONCLUSIONS AND FUTURE DIRECTIONS

Tridax procumbens leaf extract possesses scientifically validated antibacterial properties attributable to a rich and synergistically active phytochemical profile. Herbal antibacterial creams formulated with *T. procumbens* leaf extract consistently exhibit acceptable physicochemical properties, satisfactory in vitro antibacterial activity, dermal safety, and promising in vivo wound healing efficacy. The plant therefore merits serious consideration as a lead candidate for commercial herbal antibacterial cream development.

Future research should prioritize the following directions:

1. Randomized controlled clinical trials comparing *T. procumbens* cream against standard topical antibiotics (mupirocin, fusidic acid) in patients with superficial skin infections to generate evidence-grade I clinical data.
2. Formal standardization protocols establishing minimum quercetin and luteolin content in leaf extracts to ensure batch-to-batch consistency in antibacterial activity.

3. Comprehensive dermal safety assessment including repeated insult patch test (RIPT), phototoxicity, and sensitization studies in human volunteers.

4. Pharmacokinetic skin penetration studies using Franz diffusion cells with pig skin or human cadaver skin to characterize percutaneous absorption of key phytoconstituents from O/W cream matrices.

5. Scale-up formulation development with industrial stability testing (12 months under ICH Q1A) to support regulatory dossier preparation.

6. Exploration of combination nanotechnology platforms — such as AgNP-loaded phytosomes in cream matrices — to achieve superior bioavailability, prolonged release, and enhanced antibacterial potency against MDR pathogens.

In conclusion, *T. procumbens* represents a compelling convergence of traditional wisdom and modern pharmacological evidence — a plant whose time for mainstream pharmaceutical development has arrived. With appropriate investment in clinical research and formulation standardization, *T. procumbens*-based antibacterial cream has the potential to become a significant addition to the global dermatological product portfolio. [29]

REFERENCES

1. Gubbiveeranna V, Nagaraju A, Bharathi DR, Chandrashekar K. Ethnomedical, phytochemical constituents and pharmacological activities of Tridax procumbens: a review. Asian Pac J Trop Biomed. 2016.
2. Amutha R, Geetha RV, Anitha R. Tridax procumbens (coat button) – a gift of nature: an overview. In: Pharmacological Benefits of



- Natural Products. 1st ed. Chapter 12. 2019. p.193-212.
3. Ikewuchi CJ, Ikewuchi CC, Igboh NM. Chemical profile of *Tridax procumbens* Linn. *Pak J Nutr.* 2009;8(5):548-50.
4. Ikewuchi JC, Ikewuchi CC. Comparative study of the mineral element composition of some common Nigerian medicinal plants. *Pac J Sci Technol.* 2009;10:362.
5. Sawant L, Godghate AG. Qualitative phytochemical screening of rhizomes of *Curcuma longa* Linn. *Int J Sci Environ Technol.* 2013;2(3):634-641.
6. Singh P, Jain K, Khare S, Shrivastav P. Evaluation of phytochemical and antioxidant activity of *Tridax procumbens* extract. *Pharm Biosci J.* 2017;5:41-7.
7. Pande PS, Mishra MN. Isolation and identification of quercetin and rutin from leaves of *Tridax procumbens* Linn. by HPLC analysis. *Int J Chem Phys Sci.* 2015;4:65-8.
8. Khadse CD, Kakde RB. Phytochemistry and pharmacological aspects of *Tridax procumbens* (L.): A systematic and comprehensive review. *J Ethnopharmacol Phytochem. ScienceDirect.* 2021.
9. Custodio-Mendoza JA, et al. Ultrasonic assisted extraction and phytochemical profiling of *Tridax procumbens* using UPLC-QTOF-MS/MS. *Research Square.* 2025 [Preprint].
10. Ninlachkar S, et al. Antihyperuricemia, antioxidant, and antibacterial activities of *Tridax procumbens* L. *MDPI Molecules / PMC.* 2019.
11. Christudas S, Kulathivel TM, Agastian P. Phytochemical and antibacterial studies of leaves of *Tridax procumbens* L. *Asian Pac J Trop Biomed.* 2012;2(1):S159-61.
12. Phytochemical screening and in vitro antimicrobial activity of *Tridax procumbens* L. [Multiple authors]. *Academia.* 2015.
13. Dhanabalan R. In vitro phytochemical screening and antibacterial activity of aqueous and methanolic leaf extracts of *Tridax procumbens* against bovine mastitis isolated *Staphylococcus aureus*. *Ethnobot Leaflets.* 2008;12:1090-5.
14. Pai C, Kulkarni U, Borde M, Murali S, Mrudula P, Deshmukh Y. Antibacterial activity of *Tridax procumbens* with special reference to nosocomial pathogens. *Br J Pharm Res.* 2011;1(4):164-73.
15. In vitro antibacterial, antioxidant potentials and cytotoxic activity of the leaves of *Tridax procumbens*. *PMC.* 2020.
16. Custodio-Mendoza JA, et al. Mechanism of antibacterial action — UPLC-QTOF analysis. *Research Square.* 2025 [Preprint].
17. Antimicrobial efficacy of chitosan-*Tridax procumbens* extract composite on selected wound surface microorganisms. *ResearchGate.* 2025.
18. Development and evaluation of antimicrobial cream with *Acacia catechu* and *Tridax procumbens*. *PC Biochem Res.* 2025.
19. Gund AL, Jawale SS, Walunj K, et al. Formulation and evaluation of polyherbal cream containing extracts of *Tridax procumbens*, *Ficus religiosa*, *Azadirachta indica* and *Curcuma longa*. *Int J Sci Res Sci Technol.* 2022.
20. Design and formulation of *Tridax procumbens*-based polyherbal cream for wound healing potential. *Scholars Res Libr.*
21. Sonawane GB, Patil NP, Pendharkar MD, et al. Review article on *Tridax procumbens* herbal antibacterial cream. *STM Journals.* 2026.
22. Nimbekar TP, Meshram AS, Farooqui MS. Formulation and evaluation of ointment containing natural wound healing activity of *Tridax procumbens*. *JETIR.* 2019;6(5).



23. Development of wound healing ointment formulation containing active extracts of *Tridax procumbens*, *Calendula officinalis*, *Murraya koenigii*, and *Aloe barbadensis*. *Int J Pharm Phytopharmacol Res (EIJPPR)*.
24. Formulation and evaluation of a *Tridax procumbens*-loaded phospholipid complex (phytosome) gel for wound healing and antimicrobial activities. *RSC Pharmaceutics*. 2025/2026.
25. General physicochemical evaluation parameters for herbal creams. [Multiple authors — synthesis of evaluation standards from literature corpus].
26. Study of antimicrobial and hair regrowth activity of *Tridax procumbens* for scalp disorder. *Int J Sci Res Sci Technol*. 2020.
27. Ethnomedicinal, phytochemical constituents and pharmacological activities of *Tridax procumbens* — comprehensive review. *ResearchGate/Academia*.
28. Kubendiran M, et al. Green synthesized silver nanoparticles using *Tridax procumbens* for topical application: excision wound model and histopathological studies. *Pharmaceutics*. 2021;13(11):1754. MDPI.
29. Muthukumar S, Nandini V, Elumalai S, et al. Enhancement of cell migration and wound healing by nano-herb ointment formulated with biosurfactant, silver nanoparticles and *Tridax procumbens*. *Front Microbiol*. 2023.
30. Bharathi S, Ramesh B, Kumaran S, et al. In vitro preparation of biosurfactant based herbal-nano topical ointment from *Tridax procumbens* infused oil using gelatin stabilized silver nanoparticle and its efficacy on fibroblastic cell lines. *Appl Nanosci*. 2021.

HOW TO CITE: Lina Nagpure, Prachi Dashwant, Shraddha Pisal, Avadhut Dukandar, Formulation and Evaluation of Herbal Antibacterial Cream using *Tridax procumbens* Leaf Extract, *Int. J. of Pharm. Sci.*, 2026, Vol 4, Issue 5, 7108-7119. <https://doi.org/10.5281/zenodo.20404726>

