

Influence of *Potamogeton crispus* Harvesting on Phosphorus Composition of Lake Yimeng

Li-zhi Wang (✉ wanglizhi@lyu.edu.cn)

Linyi University <https://orcid.org/0000-0002-8613-1444>

Xiyuan Wu

Linyi University

Bing Dong

Linyi University

Hongli Song

Linyi University

Juan An

Linyi University

Yuanzhi Wu

Linyi University

Yun Wang

Linyi University

Bao Li

Linyi University

Qianjin Liu

Linyi University

Research Article

Keywords: submerged macrophytes, phosphorus composition, environmental factors, harvesting, Lake Yimeng, *Potamogeton crispus*

Posted Date: June 1st, 2022

DOI: <https://doi.org/10.21203/rs.3.rs-1592007/v1>

License: © ⓘ This work is licensed under a Creative Commons Attribution 4.0 International License. [Read Full License](#)

Abstract

Harvesting is an important method used to control the overproduction of *Potamogeton crispus* in lakes. A three-year comparative field study was performed in a eutrophic lake (harvested area) and its connected lake (non-harvested area) to determine the effects of harvesting on the phosphorus (P) composition and environmental factors in the water and sediment. Results revealed that harvesting significantly reduced the dissolved total P (DTP) and dissolved organic P (DOP) and increased the alkaline phosphatase activity (APA) and particulate P (PP) in the water. No significant differences were detected in the water total P (TP), soluble reactive P (SRP), chlorophyll-a (Chl-a), pH, and dissolved oxygen (DO) between the harvested and non-harvested areas. Sediment TP and organic P (OP) were significantly reduced in the harvested area. Harvesting changed the P composition in the water. In the non-harvested area, P was mainly formed by DOP (40%) in the water body, while in the harvested area, PP was the main water component (47%). Harvesting increased the proportion of inorganic P (IP) in the sediment and decreased the proportion of OP. In the water, the IP to TP ratio in the non-harvested and harvested areas were 58.26% and 63.51%, respectively. Our results showed that harvesting changed the P composition in the water and sediment. In the harvesting of submerged vegetation, our results can serve as a reference for the management of vegetation-rich lakes.

Introduction

Submerged macrophytes as primary producers play important roles in maintaining the structure, function, and biodiversity of lake ecosystems (Nyieku et al. 2020). Submerged macrophytes secrete allelochemicals, inhibit the propagation of algae, and absorb nutrients in the overlying water and sediment, which has important ecological value in controlling lake eutrophication (Sarvala et al. 2020). The restoration, reconstruction, and transformation of aquatic plants have become important methods in the ecological regulation and endogenous pollution load control of shallow lakes (Chorus et al. 2020). However, when a large number of submerged macrophytes decompose, the residue still exists in the water and decomposes as well, which releases nitrogen (N), phosphorus (P), and other raw elements into the overlying water, thereby leading to secondary pollution (Malaviya et al. 2020). Harvesting is an important method used for the overgrowth of submerged macrophytes in lakes. Harvesting submerged macrophytes is conducive not only to the recovery and growth of plants, which increases biodiversity, but also to improving the stability of the community to continuously purify the water. It can also reduce the nutrient load in a lake by harvesting plants or transferring nutrients from the lake. Additionally, harvesting can prevent the negative effects of excessive submerged macrophyte growth. For submerged macrophytes whose biomass is mainly concentrated in the upper layer or surface of the water body, harvesting can alleviate excessive biomass concentration. Therefore, it is of great importance to study the effects of harvesting on aquatic environments.

Harvesting directly affects the N and P cycles in the water. Currently, due to excess N, P, and other nutrients, submerged macrophytes overgrow and bloom in many lakes (Franceschini et al. 2020). As the seasons change, the decomposition of many submerged macrophyte residues causes serious secondary pollution, which poses a great threat to the safety of aquatic ecosystems (Wang et al. 2018). Decomposition and death are necessary stages in the life history of submerged macrophytes. The decomposition and release of nutrients are important ecological processes that play important roles in the biogeochemical cycles of N, P, and other nutrients in lake ecosystems. In the field of lake ecology, the investigation of these processes is currently a popular research area (Shilla et al. 2006). In the process of natural succession and seasonal change, if aquatic plants are not harvested, then their litter will disperse into the water and sediment, and their decomposition products (organic matter, N, P, and other nutrients) will participate in the biogeochemical cycles of lake nutrients again (Bianchi & Findlay 1991). Therefore, the decomposition of aquatic plants is the key link between material cycling and energy flow. This process will reduce the transparency of the water body and increase the contents and ratios of organic matter, N, P, and other pollutants in the water body and sediment, which will thereby result in secondary pollution (Battle & Mihuc 2000).

The P status in lakes is an important factor of submerged macrophytes that affects eutrophication. The relationship between eutrophication and limiting nutrients (mostly P) differs between lakes with and without submerged vegetation. Therefore, it is of great importance to explore the effects of harvesting on the P cycle in the water body to assist the management of vegetative lakes (Overbeek et al. 2019).

Potamogeton crispus is a submerged macrophyte widely distributed throughout China that is often used in the ecological restoration of eutrophic lakes (Hao et al. 2018). Most accomplishments in efforts to restore eutrophic lakes have been attributed to the success of aquatic macro-vegetation (Wang et al. 2017). The viable growth temperature of *P. crispus* is 10–20°C, and it stops growing when temperatures are > 24°C. *P. crispus* germinates in autumn and grows during the winter; large reproductive growth occurs in April and May, and plants finally decompose and die in the summer. It has a rapid growth rate, can tolerate high nutrient-rich environments, and grows well in contaminated water bodies (Leoni et al. 2016).

Lake Yimeng was formed in 1997 when a rubber dam (1,135 m) was built across the Yi River, capturing 12 million m³ of water. In recent decades, the lake has exhibited dense canopy-forming populations of *P. crispus*, which covers nearly 90% of the lake during spring and summer (Wang et al. 2018). Different types of management strategies have been implemented to reduce *P. crispus* blooms, including harvesting during the summer. Almost 100% of the present *P. crispus* was harvested in 2017 in the Yi River part of the lake; however, in the Beng River part of the

lake, *P. crispus* was not harvested. Therefore, we selected these two areas (harvested and non-harvested) to study the effects of harvesting on P composition in the water (Fig. 1).

The aim of this work was to determine how harvesting submerged macrophytes (*P. crispus*) affects the P composition and environmental factors in the water. We compared the P in the water and sediment, chlorophyll-a (Chl-a), and environmental factors between the harvested and non-harvested areas of the lake for three years. The relationship between Chl-a and P in the water and environmental factors are also discussed.

Materials And Methods

Study area and sampling

For several years, *P. crispus* has bloomed in Lake Yimeng, which is a eutrophic lake. The Beng River part of the lake (non-harvested *P. crispus* area) is connected to the Yi River part of the lake (35°2'40"–35°7'11"N; 118°22'11"–118°23'12"E). *P. crispus* was harvested in 2017. Three corresponding sampling sites were selected in the non-harvested and harvested areas (Fig. 1). Beng River was completely covered by *P. crispus* in its growing season with a mean biomass of 1,025 g·m⁻². Although the Beng River is connected to the Yi River, there is little water exchange. During the rainy season from July to August, water flows from the Beng River to the Yi River. The Secchi disc depth varies between 20 cm and 60 cm in the Yi River and between 50 cm and 145 cm in the Beng River.

Mixed water samples from the upper layer of each sampling site (0, 0.5, and 1 m) were collected using a Grasp sampler (Grasp BC-2300, Beijing, China) every month. Afterwards, water samples were stored in a refrigerator at 4°C for further analysis. For total P (TP) analysis, water samples were autoclaved at 121°C for 30 min after the addition of K₂S₂O₈. Then, 10% ascorbic acid was added and the samples were measured using the molybdenum blue spectrophotometric method (Sommers 1977). A continuous flow analyzer (Flowsys III, Systea Company, Italy) was used to determine the P concentration. The same method was used for the soluble reactive P (SRP) determination, except the water samples were filtered through a 0.45 µm cellulose acetate membrane and not autoclaved. Dissolved total P (DTP) was measured using the same method as TP, except the water samples were filtered through a 0.45 µm cellulose acetate membrane. The difference between the TP and DTP was defined as the sum of the particulate P (PP) fraction. The difference between the DTP and SRP was defined as the sum of the dissolved organic P (DOP) fraction. The detection limit for the SRP and DTP concentrations in the overlying water was 1 µg·L⁻¹. All materials used for the analyses were purchased from the Shanghai N&D Co., Ltd. (Shanghai, China). For all samples, triplicates were analyzed and the data are expressed as the mean. The P concentration was determined according to previously described methods Goulden and Brooksbank (1975).

Two liters were collected for measuring the physical and chemical indices and plankton analysis. Phytoplankton samples (1 L) for quantitative analysis were preserved with Lugol's solution and concentrated to 30 mL after one week of sedimentation.

Sediment samples from each sampling site were collected at the same time when mixed water samples were collected. Sediment samples were collected using a self-made hand-driven polyethylene corer (patent no. ZL201420135437.1), immediately transported to the lab in sealed plastic bags placed in iceboxes, and then freeze-dried and sieved (< 2 mm). All samples were stored at 0–4°C for further analysis.

Sediment P fractionation of the TP, inorganic P (IP), and organic P (OP) was conducted based on the Standards Measurements and Testing (SMT) protocol, which consists of three extraction procedures applied to 0.2 g aliquots of sediment samples. The SMT protocol is a common approach used for studying the P fractions of lake sediment (Ruban et al. 2001). Extractions (16 h) used 1 mol·L⁻¹ HCl and were performed to remove the IP. The residues from the extractions were placed in a porcelain crucible and calcined in a furnace for 3 h at 450°C. The residues were extracted (16 h) again using 1 mol·L⁻¹ HCl to remove P associated with the organic matter of the sediment (OP). To obtain the TP, a simple extraction (16 h) using 3.5 mol·L⁻¹ HCl was performed after the calcination of a separate sample for 3 h at 450°C. For all cases, phosphate was determined in the extracts via a spectrophotometric procedure. Measurements were conducted at 886.0 nm.

The extracellular alkaline phosphatase activity (APA) in the water samples was determined following previously described methods Gage and Gorham (1985). Water samples were filtered through 0.45 µm Millipore filters to determine the Chl-a content using an extraction of 90% acetone (García-Nieto et al. 2020).

The pH (PHSJ-4A; Lei-ci, Shanghai, China) and dissolved oxygen (DO) in the overlying water (YSI 5750; USA) were measured before sampling in each site.

Statistical analysis

Correlation analyses were performed using SPSS v25.0 (IBM Corp., Chicago, IL, USA). Differences were considered statistically significant at $P < 0.05$ and $P < 0.01$. Tukey's post-hoc test after two-way ANOVA was used to determine the differences between the harvested and non-harvested areas of the water and sediment indices of different sections. The relationship between the water chemical and physical parameters of the two areas were assessed using Pearson correlation analyses. The results are presented as the average of three replicates. The default Z-scores in

SPSS were adopted for data standardization. Principal component analysis (PCA) was used for data dimensions, and the extraction method was used for principal components.

Results

Monthly variations in water P composition and physical parameters

In the two water bodies, the TP concentration was the lowest during the winter and highest during the summer (Fig. 2a). From April to June 3, TP rapidly increased and peaked in the summer (from May to August, TP ranged from $0.20 \text{ mg}\cdot\text{L}^{-1}$ to $0.25 \text{ mg}\cdot\text{L}^{-1}$). From January to March, TP remained low ($0.04\text{--}0.06 \text{ mg}\cdot\text{L}^{-1}$) in the non-harvested area for three years. In the harvested area, the TP remained relatively low in 2017 and showed its first peak in April ($0.17 \text{ mg}\cdot\text{L}^{-1}$). From May to October, the TP gradually increased and showed its second peak in October ($0.20 \text{ mg}\cdot\text{L}^{-1}$). From 2018 to 2019, the TP trend of the harvested area changed in the same way as the non-harvested area, but the TP concentration was relatively low in the summer and high in the winter.

In the non-harvested area, DTP exhibited roughly the same trend as TP and peaked in the summer (from May to August, the DTP ranged from $0.12 \text{ mg}\cdot\text{L}^{-1}$ to $0.18 \text{ mg}\cdot\text{L}^{-1}$). In the harvested area, DTP exhibited an increasing trend, with the highest concentration detected in August 2019 ($0.11 \text{ mg}\cdot\text{L}^{-1}$). From winter to summer, DTP gradually increased and plateaued when compared to the non-harvested area (Fig. 2b). The change trend in PP was not obvious in either water body but seemed to increase in the summer and decrease in the winter (Fig. 2c).

In the non-harvested area, SRP increased rapidly from May to July in 2017 ($0.31\text{--}0.10 \text{ mg}\cdot\text{L}^{-1}$) and increased again from May to June in 2018 and 2019. After it peaked, the SRP decreased gradually (Fig. 2d). In the harvested area, the SRP showed its first peak in April ($0.05 \text{ mg}\cdot\text{L}^{-1}$) and its second peak in November ($0.07 \text{ mg}\cdot\text{L}^{-1}$) in 2017; it also peaked in August ($0.07 \text{ mg}\cdot\text{L}^{-1}$) and July ($0.08 \text{ mg}\cdot\text{L}^{-1}$) in 2018 and 2019, respectively. The SRP peaks in the harvested area lagged behind those in the non-harvested area. Generally, TP, DTP, and SRP exhibited increasing trends in the harvested area but decreasing trends in the non-harvested area from 2017 to 2019.

DOP increased and was maintained at a high level ($0.05\text{--}0.13 \text{ mg}\cdot\text{L}^{-1}$) from May to September in the non-harvested area and was higher than in the harvested area. DOP was maintained at a low level and changed slightly in the harvested area (Fig. 2e). TP, DTP, PP, SRP, and DOP increased in the non-harvested area and were consistent with the senescence period of *P. crispus* (Fig. 2j).

Changes in the Chl-a content exhibited the same trend as TP, DTP, and SRP. The Chl-a content had a positive relationship with the TP, DTP, and SRP concentrations in the non-harvested and harvested areas. After combining the data from the two water bodies, this relationship was still observed (Fig. 2h). The Chl-a content was significantly higher in the harvested area when compared to the non-harvested area during the growth period of *P. crispus*. On the seasonal scale, the Chl-a content was the lowest during the winter and highest during the summer. In April, the Chl-a content sharply increased when *P. crispus* began to decompose.

The total APA was maintained at a high level ($46.02\text{--}55.01 \text{ nmol}\cdot\text{L}^{-1}\cdot\text{min}^{-1}$) from July to September in the non-harvested area. In the harvested area, the total APA was maintained at a relatively high level from April to December ($42.01\text{--}55.03 \text{ nmol}\cdot\text{L}^{-1}\cdot\text{min}^{-1}$). In the two water bodies, the total APA was the lowest during the winter and highest during the summer (Fig. 2g). The pH and DO increased during the growth period of *P. crispus* from March to May and decreased during the decomposition period in the non-harvested area. In the harvested area, the pH was the lowest during the winter and highest during the summer, while the DO seemed to exhibit the opposite trend (Fig. 2h, i).

Monthly variations in the sediment P fractions

The sediment of different P fraction concentrations exhibited increasing trends in the two water bodies. The sediment TP increased during the decomposition period of *P. crispus* from June to October in the non-harvested area. In the harvested area, the sediment TP changed slightly and exhibited an increasing trend during the summer alongside a small amount of *P. crispus* decomposition (Fig. 3a). The IP roughly exhibited the same change trend as the sediment TP in the two water bodies. The IP was higher from June to October in the non-harvested area when compared to the harvested area in 2019 (Fig. 3b). In the non-harvested area, the OP increased during the decomposition of *P. crispus*. The OP exhibited an increasing trend after 2018. Most of the time, the OP in the non-harvested area was higher than in the harvested area. In the harvested area, the OP was maintained at a relatively stable level (Fig. 3c).

The sediment P in the two areas was mainly formed by IP. The average percentages of the IP in the non-harvested and harvested areas were 58.26% and 63.51%, respectively. The OP percentage in the non-harvested area was higher than in the harvested area. The average percentages of the OP in the non-harvested and harvested areas were 41.74% and 36.49%, respectively. Thus, *P. crispus* harvesting clearly increased the IP proportion in the sediment.

Comparison of the mean water and sediment indices between the harvested and non-harvested areas

No significant differences were detected in the water TP and SRP between the non-harvested and harvested areas (Table 1). The DTP and DOP were significantly lower in the harvested area when compared to the non-harvested area ($P < 0.05$). The PP exhibited the opposite trend ($P < 0.05$). The Chl-a, pH, and DO did not significantly differ between the non-harvested and harvested areas. The APA was significantly higher in the harvested area ($P < 0.05$).

Table 1
Tukey's post hoc analysis of the two-way ANOVA test between Harvest area and *P. crispus* area of water and sediment chemical and physical parameters (means $\pm$ SD)

	water										sediment		
	TP	DTP	PP	SRP	DOP	Chl a	APA	Biomass	pH	DO	TP	IP	OP
	mg·L ⁻¹	mg·L ⁻¹	mg·L ⁻¹	mg·L ⁻¹	mg·L ⁻¹	mg·L ⁻¹	nmol·L ⁻¹ ·min ⁻¹	g		mg·L ⁻¹	mg·g ⁻¹	mg·g ⁻¹	mg·g ⁻¹
Harvest area	0.13 $\pm$ 0.04	0.07 $\pm$ 0.02	0.06 $\pm$ 0.02	0.05 $\pm$ 0.02	0.02 $\pm$ 0.01	0.15 $\pm$ 0.08	42.36 $\pm$ 7.96	104.26 $\pm$ 118.27	8.17 $\pm$ 0.33	8.2 $\pm$ 0.57	0.09 $\pm$ 0.01	0.06 $\pm$ 0.01	0.03 $\pm$ 0.01
<i>P. crispus</i> area	0.14 $\pm$ 0.07	0.10 $\pm$ 0.05	0.04 $\pm$ 0.03	0.04 $\pm$ 0.02	0.06 $\pm$ 0.04	0.13 $\pm$ 0.11	30.14 $\pm$ 12.35	405.72 $\pm$ 258.31	8.2 $\pm$ 0.39	8.66 $\pm$ 1.8	0.10 $\pm$ 0.02	0.06 $\pm$ 0.02	0.04 $\pm$ 0.01
F value	1.18	10.94	6.73	3.59	37.86	0.57	24.90	40.54	0.19	2.10	7.27	0.24	36.07
P value	0.28	0.00	0.01	0.06	0.00	0.45	0.00	0.00	0.67	0.15	0.01	0.63	0.00

In the sediment (Table 1), the TP and OP were significantly lower in the harvested area when compared to the non-harvested area ($P < 0.05$). The IP did not significantly differ between the non-harvested and harvested areas.

Relationship between the Chl-a, environmental factors, and P composition in the water based on PCA

To study the relationship between Chl-a and water chemical and physical parameters, PCA was conducted for data reduction purposes. The indices related to Chl-a were reduced in the two areas. The two main factors (F_h and F_p) were extracted in the two study areas (F_h in the harvested area, F_p in the non-harvested area). The coefficient matrix is shown in Table 2. Factor scores were calculated according to the following formulas:

Table 2
PCA score coefficient matrix of harvest area and *P. crispus* area

harvest area		<i>P. crispus</i> area	
items	F_h	items	F_p
TP	0.22	TP	0.26
DTP	0.20	DTP	0.25
PP	0.19	PP	0.21
SRP	0.20	SRP	0.20
DO	-0.15	DOP	0.22
pH	0.19		

$$F_h = 0.22Z_{TP} + 0.20Z_{DTP} + 0.19Z_{PP} + 0.20Z_{SRP} - 0.15Z_{DO} + 0.19Z_{pH}$$

$$F_p = 0.26Z_{TP} + 0.25Z_{DTP} + 0.21Z_{PP} + 0.20Z_{SRP} + 0.22Z_{DOP}$$

where Z_{TP} , Z_{DTP} , Z_{PP} , Z_{SRP} , Z_{DOP} , Z_{DO} , and Z_{pH} are the z-standardized data of the TP, DTP, PP, SRP, DOP, DO, and pH, respectively.

Since PCA was based on z-standardized data, the correlation analysis of the Chl-a content was also based on the z-standardized data. The Z-standardized score of the Chl-a content positively correlated with F_h and F_p (Fig. 4).

The relationship between the Chl-a content, environmental factors, and P concentration in the water is expressed as follows:

1. Harvested area:

$$Z_{Chla} = 0.17Z_{TP} + 0.16Z_{DTP} + 0.14Z_{PP} + 0.16Z_{SRP} - 0.12Z_{DO} + 0.15Z_{pH} + 0.09$$

(2) Non-harvested area:

$$Z_{Chla} = 0.27Z_{TP} + 0.26Z_{DTP} + 0.22Z_{PP} + 0.21Z_{SRP} + 0.23Z_{DOP} - 0.09$$

Comprehensive analysis revealed that the Chl-a content in the water body was affected by the contents of different P forms in the water body and environmental factors in the harvested area. However, the Chl-a content in the water body was only affected by the content of various P forms in the water body of the non-harvested area. Clearly, the harvesting of *P. crispus* changed the influencing factors in the water body on the Chl-a content.

Discussion

Effects of harvesting on P composition in the water

In this field investigation, we compared changes triggered by the harvesting of the submerged macrophyte, *P. crispus*. Our results suggested that harvesting *P. crispus* significantly decreased the P concentration in terms of DTP, PP, and DOP in the water column; TP did not significantly decrease. Harvesting *P. crispus* increased SRP but not significantly. Fluctuations of different P fractions in the water of the non-harvested area were larger than in the harvested area. In the non-harvested area, the P in the water was mainly formed by DOP (40%), which was due to the growth and decomposition of *P. crispus* (Fig. 5), especially its decomposition. Our results are consistent with the findings of Cao et al. (2018), who reported that *P. crispus* released a large amount of DOP after decomposition. In the harvested area, PP was the main component of P in the water (47%). The water body was greatly disturbed by wind and waves in the absence of *P. crispus*, thereby increasing the PP content (Fig. 5). The percentage of the SRP was 40% in the absence of *P. crispus*. PP is unstably bound to P (Zhu et al. 2013). Changes in environmental regulations lead to desorption, reduction, or dissociation reactions of unstably bound P on the surface of some particles, which are converted to SRP and other P fractions (Song et al. 2017). Therefore, high percentages of the SRP and PP increase the risk of eutrophication.

TP decreased after April 2017, where the TP decreased the TP of other P fractions in the harvested area. However, different P fractions increased in the non-harvested area at the same time, indicating that *P. crispus* harvesting reduced the P concentration in the water. The TP and other P fractions increased and reached their second peak in October 2017, which may be due to the decomposition of algae and residual aquatic plants. The release of P in the sediment increases the P in the water body. Changes in environmental factors lead to the release of P in the sediment. The pH and DO significantly decreased after *P. crispus* harvesting; a low pH and DO also leads to P release. The data from 2018 and 2019 showed that the P concentration in the water recovered the increasing trend during the summer and decreasing trend during the winter after one year of harvesting. Collectively, these findings show that *P. crispus* harvesting can reduce the P concentration within a short period of time.

P is a key element that limits the primary productivity in the water body, as evidenced by the significant positive relationship detected between the P concentration and Chl-a in many freshwater lakes (Hudson et al. 2000). The mean SRP concentration in the non-harvested area was significantly lower than in the harvested area, suggesting that the presence of *P. crispus* decreased the bioavailable P in the water column. The DOP concentration in the non-harvested area increased and exceeded the amount in the harvested area when submerged macrophytes decomposed, indicating that OP was released during the decomposition process of *P. crispus* in the lake. Consistently, submerged macrophytes increased the DOP concentration during the decomposition process in a lake (Wang et al. 2018). The decomposition of *P. crispus* is a major source of bioavailable P and decomposable dissolved organism matter (DOM) for the phytoplankton community in a lake, suggesting that macrophyte decomposition potentially stimulates phytoplankton production (Granéli & Solander 1988). The decomposition of *P. crispus* increased the turnover rate of the DOP, indicating more bioavailable DOP mediated by alkaline phosphatase (Cao et al. 2018).

Effects of harvesting on environmental factors in the water

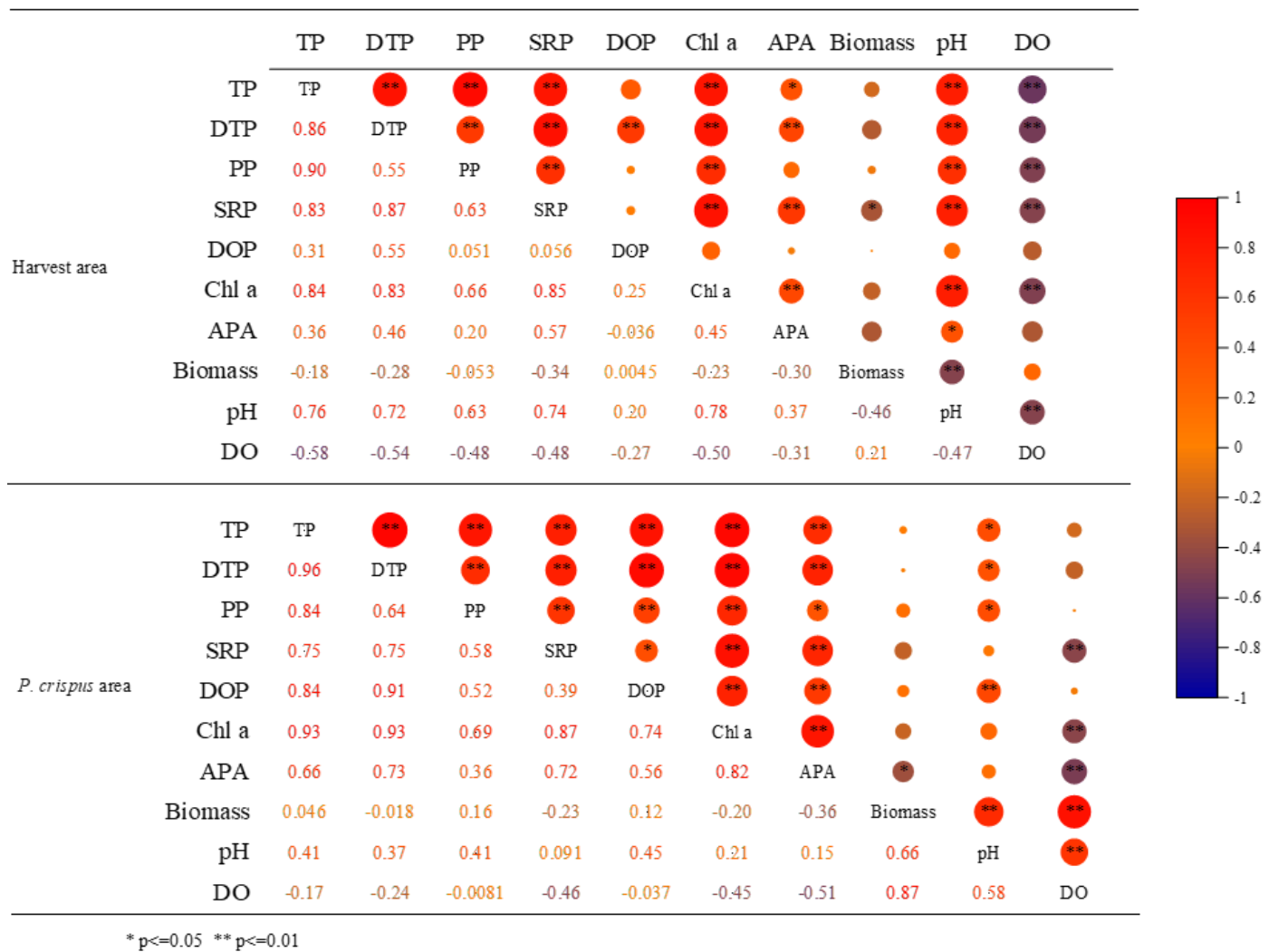
Alkaline phosphatase is a type of phosphate hydrolase with wide specificity that can catalyze the hydrolysis reactions of all phosphate esters and transfer reactions of phosphate groups. It directly participates in P metabolism, provides P for the rapid growth of plankton in the water body, and plays an important role in the transformation of P in aquatic ecosystems (Gross et al. 2007). APA is increased during the summer and

decreased during the winter in the two areas of this study. The ability of phytoplankton to excrete extracellular phosphatase that catalyzes DOP to overcome P deficiency is a possible explanatory reason (Ding et al. 2016). For many lakes, most of the TP in the overlying water is in the form of OP, in which, > 70% is particulate OP in the suspension, while the rest is dissolved OP in colloidal form. However, the dissolved inorganic orthophosphate content that directly enters the cell body and is absorbed and utilized by phytoplankton only accounts for 5%, which changes quickly due to its direct utilization (Brovini et al. 2021). In the absence of orthophosphate in the water, alkaline phosphate can be induced in algae and bacteria as an inducer for hydrolyzing monophosphate. Submerged macrophytes can reduce the maximum reaction rate of alkaline phosphatase in overlying water (Song et al. 2006). Kalinowska (1997) found that in lakes dominated by submerged macrophytes, the decomposition of OP by phosphatase was almost negligible. The reason may be that more enzymes were inactivated due to the complexation of humus. *P. crispus* changed the composition of alkaline phosphatase in the water, thereby reducing the maximum enzymatic reaction rate. A previous study demonstrated that the total APA in an Indian pond system was proportional to the TP and total bacteria in the water (Barik et al. 2015). Additionally, the resuspension of sediment led to a higher P content in the particles, which promoted the reaction rate of alkaline phosphate. Zhou and Dodds (1995) showed that the APAs associated with different-sized particles had different kinetic characteristics. A similar phenomenon was observed in Lake Donghu and Lake Nanhu in China (Cao et al. 2005).

Phytoplankton are important producers in river ecosystems and play important roles in the material circulation and energy transformation processes (Rydberg et al. 2020). Chl-a is a comprehensive performance that reflects physical and chemical factors and biological indicators in the water body; it is also restricted by environmental factors (Chaffin et al. 2018). Therefore, Chl-a can be used to characterize the growth state of phytoplankton in the water.

The Chl-a content had significant positive relationships with the TP, DTP, PP, SRP, and pH, but a negative relationship with the DO in the harvested area, while in the non-harvested area, it had significant positive relationships with the TP, DTP, PP, DOP, and SRP (Table 3). The Chl-a content in the non-harvested area was mainly affected by the P content of the water body, while in the harvested area, Chl-a was affected by the P content and environmental factors in the water body.

Table 3 Pearson's correlation coefficients describing the relationships between water chemical and physical parameters in the two studied areas



Effects of harvesting on the sediment P fractions

The migration and transformation of P by *P. crispus* remains controversial. One notion is that *P. crispus* activates the P in the sediment, wherein *P. crispus* mainly absorbs the P in the sediment during its growth period, then releases it into the water body during its decomposition period (Jarvie et al. 2020). Therefore, *P. crispus* serves a P pump role and activates the P in the sediment into the water, which intensifies eutrophication. Another explanation is that *P. crispus* metabolizes easily-released P into stable P. *P. crispus* releases P into the water body during the decomposition period for a relatively short period of time (Wang et al. 2020, Wang et al. 2018). After a month, the P in the water body returns to a relatively low level. The growth cycle of *P. crispus* absorbs easily released forms of P in the sediment and then converts them into difficult-to-release forms of P, which are finally deposited in the sediment for permanent storage. *P. crispus* can also cause P adsorption in the sediment (Hao et al. 2018). This increased P capacity indicates that the final migration direction of P is to the sediment.

In this study, the TP, IP, and OP contents in the sediment of the non-harvested area exhibited upward trends, indicating that the migration direction of P in the water body was to the sediment after a long adjustment period by *P. crispus*. The OP percentage in the non-harvested area was higher than in the harvested area, indicating that under the action of *P. crispus*, P forms in the sediment migrate in the direction of OP.

Phytoplankton respond accordingly to P composition variations. A cyanobacteria bloom occurred when *P. crispus* decomposed. Freshwater cyanobacteria blooms are expected to be a major environmental problem in the urban lake and Three Gorge Reservoir due to excessive nutrient release from the sediment (Li et al. 2011).

Endogenous P in the sediment is mainly derived from the precipitation of PP in lakes but dissolved P (after adsorption by mineral particles) and OP in organisms. The migration and transformation process of P at the sediment-water interface is a key process of the P cycle in eutrophic lakes and is affected by many factors, including sediment properties, the interface environment, and biological characteristics (Mi et al. 2008).

The classical P cycle theory suggests that P will be adsorbed by iron (hydrogen) oxides or precipitate FePO_4 and that the release of P is mainly controlled by DO and the redox potential (Song et al. 2017). In recent years, scholars have conducted several studies on the migration and transformation mechanism of P at the sediment-water interface and gained a certain understanding of the influence mechanism of environmental and biological factors (Chattopadhyay et al. 1995, Tarasova et al. 2018, Xu et al. 2017). For example, DO will affect the decomposition rate of organic matter and occurrence of P, pH will affect the combination of P and metals, such as Fe and Al, and temperature will affect the activity of microorganisms and decomposition rate of organic matter, thereby affecting the migration and transformation of P.

In this study, *P. crispus* released a large amount of organic matter into the water and sediment during the decomposition process, thereby changing environmental factors in the water body. The pH and DO increased during the growth period of *P. crispus* and decreased during the decomposition period in the non-harvested area. The increase in the TP content in the sediment may be related to the input of external P from the water body. *P. crispus* fixes external P and migrates into the sediment during the growth and decomposition processes. The increased P content in the sediment in the non-harvested area may be the result of the combined effects of these environmental factors.

Conclusion

During the three years of field observations, we found that *P. crispus* harvesting significantly reduced DTP and DOP and increased APA and PP in the water. No significant differences were detected in the water TP, SRP, Chl-a, pH, or DO between the non-harvested and harvested areas. The sediment TP and OP significantly decreased in the harvested area. No significant difference was detected in the IP between the non-harvested and harvested areas.

The sediment P in the two areas was mainly formed by IP. The average percentages of IP in the non-harvested and harvested areas were 58.26% and 63.51%, respectively. The OP percentage in the non-harvested area was higher than in the harvested area.

Different forms of P decreased during the growing season, while all P forms increased during the decomposition period of *P. crispus*. The growth and decomposition of *P. crispus* changed the composition of P in the water body. In the non-harvested area, P in the water was mainly formed by DOP (40%), while PP was the main component in the harvested area (47%).

The harvesting of *P. crispus* changed the factors in the water body affecting the Chl-a content. The Chl-a content in the water was affected by different forms of P and environmental factors in the harvested area. However, the Chl-a content in the water was only affected by P in the non-harvested area.

Declarations

Authors' contributions

All authors contributed to the study conception and design. Material preparation, data collection, and analysis were performed by Lizhi Wang, Xiyuan Wu, BinDong and Hongli Song. The first draft of the manuscript was written by Lizhi Wang, and the data analysis was performed by Juan An, Yuanzhi Wu, Yun Wang. Bao Li and Qianjin Liu commented on previous versions of the manuscript. All authors read and approved the final manuscript.

Funding

This work was supported by the National Natural Science Foundation of China (41977067, 32071630, 41303061 and 41601283); the Natural Science Foundation of Shandong Province, China (ZR2021MD045, ZR2021MD003 and ZR2020MD102).

Data availability

The datasets used during the current study are available from the corresponding author on reasonable request.

Compliance with ethical standards

Competing interests The authors declare that they have no conflict of interest.

Ethical approval Not applicable.

Consent to participate Not applicable.

Consent to publish Not applicable

References

1. Barik SK, Purushothaman CS, Mohanty AN (2015) Phosphatase activity with reference to bacteria and phosphorus in tropical freshwater aquaculture pond systems. *Aquac Res* 32:819–832
2. Battle JM, Mihuc TB (2000) Decomposition dynamics of aquatic macrophytes in the lower Atchafalaya, a large floodplain river. *Hydrobiologia* 418:123–136
3. Bianchi TS, Findlay S (1991) Decomposition of Hudson estuary macrophytes: Photosynthetic pigment transformations and decay constants. *Estuaries* 14:65–73
4. Brovini EM, Cardoso SJ, Quadra GR, Vilas-Boas JA, Paranaíba JR, Pereira RdO, Mendonça RF (2021) Glyphosate concentrations in global freshwaters: are aquatic organisms at risk? *Environ Sci Pollut Res* 28:60635–60648
5. Cao X, Štrojsová A, Znachor P, Zapomělová E, Liu G, Vrba J, Zhou Y (2005) Detection of extracellular phosphatases in natural spring phytoplankton of a shallow eutrophic lake (Donghu, China). *Eur J Phycol* 40:251–258
6. Cao XY, Wan LL, Xiao J, Chen XY, Zhou YY, Wang ZC, Song CL (2018) Environmental effects by introducing *Potamogeton crispus* to recover a eutrophic Lake. *Sci Total Environ* 621:360–367
7. Chaffin JD, Kane DD, Stanislawczyk K, Parker EM (2018) Accuracy of data buoys for measurement of cyanobacteria, chlorophyll, and turbidity in a large lake (Lake Erie, North America): implications for estimation of cyanobacterial bloom parameters from water quality sonde measurements. *Environ Sci Pollut Res* 25:25175–25189
8. Chattopadhyay M, Mukherjee D, Bhattacharya SK, Lahiri SC (1995) Studies on soil parameters of sediments from the River Ganga phosphorus mobilization, interactions with sediment and the phosphorus cycle. *Environmentalist* 15:211–219
9. Chorus I, Köhler A, Beulker C, Fastner J, van de Weyer K, Hegewald T, Hupfer M (2020) Decades needed for ecosystem components to respond to a sharp and drastic phosphorus load reduction. *Hydrobiologia* 847:4621–4651
10. Ding YQ, Qin BQ, Xu H, Wang XD (2016) Effects of sediment and turbulence on alkaline phosphatase activity and photosynthetic activity of phytoplankton in the shallow hyper-eutrophic Lake Taihu, China. *Environ Sci Pollut Res* 23:16183–16193
11. Franceschini MC, Murphy KJ, Moore I, Kennedy MP, Martínez FS, Willems F, De Wysiecki ML, Sickingabula H (2020) Impacts on freshwater macrophytes produced by small invertebrate herbivores: Afrotropical and Neotropical wetlands compared. *Hydrobiologia* 847:3931–3950
12. Gage MA, Gorham E (1985) Alkaline phosphatase activity and cellular phosphorus as an index of the phosphorus status of phytoplankton in Minnesota lakes. *Freshw Biol* 15:227–233
13. García-Nieto PJ, García-Gonzalo E, Alonso Fernández JR, Díaz Muñoz C (2020) A New Predictive Model for Evaluating Chlorophyll-a Concentration in Tanes Reservoir by Using a Gaussian Process Regression. *Water Resour Manage* 34:4921–4941
14. Goulden PD, Brooksbank P (1975) The determination of total phosphate in natural waters. *Anal Chim Acta* 80:183–187
15. Granéli W, Solander D (1988) Influence of aquatic macrophytes on phosphorus cycling in lakes. *Hydrobiologia* 170:245–266
16. Gross EM, Hilt S, Lombardo P, Mulderij G (2007) Searching for allelopathic effects of submerged macrophytes on phytoplankton—state of the art and open questions. *Hydrobiologia* 584:77–88
17. Hao B, Roejkjaer AF, Wu H, Cao Y, Jeppesen E, Li W (2018) Responses of primary producers in shallow lakes to elevated temperature: a mesocosm experiment during the growing season of *Potamogeton crispus*. *Aquat Sci* 80:34–44
18. Hudson JJ, Taylor WD, Schindler DW (2000) Phosphate concentrations in lakes. *Nature* 406:54–56
19. Jarvie H, Pallet D, Shafer S, Macrae M, Bowes M, Farrand P, Warwick A, King S, Williams R, Armstrong L, Nicholls D, Lord W, Rylett D, Roberts C, Fisher N (2020) Biogeochemical and Climate Drivers of Wetland Phosphorus and Nitrogen Release: Implications for Nutrient Legacies and Eutrophication Risk. *J Environ Qual* 49:1703–1716
20. Kalinowska K (1997) Eutrophication processes in a shallow, macrophyte dominated lake – alkaline-phosphatase activity in Lake Łuknajno (Poland). *Hydrobiologia* 342:395–399
21. Leoni B, Marti CL, Forasacco E, Mattavelli M, Soler V, Fumagalli P, Imberger J, Rezzonico S, Garibaldi L (2016) The contribution of *Potamogeton crispus* to the phosphorus budget of an urban shallow lake: Lake Monger, Western Australia. *Limnology* 17:175–182
22. Li J, Peng FL, Ding DB, Zhang SB, Li DL, Ting Z (2011) Characteristics of the phytoplankton community and bioaccumulation of heavy metals during algal blooms in Xiangjiang River (Hunan, China). *Sci China Life Sci* 54:931–938
23. Malaviya P, Singh A, Anderson TA (2020) Aquatic phytoremediation strategies for chromium removal. *Reviews in Environmental Science and Bio/Technology* 19:897–944
24. Mi WJ, Zhu DW, Zhou YY, Zhou HD, Yang TW, Hamilton DP (2008) Influence of *Potamogeton crispus* growth on nutrients in the sediment and water of Lake Tangxunhu. *Hydrobiologia* 603:139–146
25. Nyieku FE, Essandoh HMK, Armah FA, Awuah E (2020) Joint influence of hydraulic load and hydraulic retention time on oilfields wastewater contaminant removal dynamics in free water surface flow constructed wetland. *SN Appl Sci* 2:2180–2191

26. Overbeek CC, van der Geest HG, van Loon EE, Admiraal W (2019) Decomposition of Standing Litter Biomass in Newly Constructed Wetlands Associated with Direct Effects of Sediment and Water Characteristics and the Composition and Activity of the Decomposer Community Using *Phragmites australis* as a Single Standard Substrate. *Wetlands* 39:113–125
27. Ruban V, López-Sánchez JF, Pardo P, Rauret G, Muntau H, Quevauviller P (2001) Harmonized protocol and certified reference material for the determination of extractable contents of phosphorus in freshwater sediments – A synthesis of recent works. *Fresenius J Anal Chem* 370:224–228
28. Rydberg J, Cooke CA, Tolu J, Wolfe AP, Vinebrooke RD (2020) An assessment of chlorophyll preservation in lake sediments using multiple analytical techniques applied to the annually laminated lake sediments of Nylandssjön. *J Paleolimnol* 64:379–388
29. Sarvala J, Helminen H, Heikkilä J (2020) Invasive submerged macrophytes complicate management of a shallow boreal lake: a 42-year history of monitoring and restoration attempts in Littoistenjärvi. SW Finland *Hydrobiologia* 847:4575–4599
30. Shilla D, Asaeda T, Fujino T, Sanderson B (2006) Decomposition of dominant submerged macrophytes: implications for nutrient release in Myall Lake, NSW, Australia. *Wetlands Ecol Manage* 14:427–433
31. Sommers LE (1977) Chemical Analysis of Ecological Materials. *J Environ Qual* 41:494
32. Song CL, Cao XY, Li JQ, Li QM, Chen GY, Zhou YY (2006) Contributions of phosphatase and microbial activity to internal phosphorus loading and their relation to lake eutrophication. *Sci China Ser D* 49:102–113
33. Song K, Winters C, Xenopoulos MA, Marsalek J, Frost PC (2017) Phosphorus cycling in urban aquatic ecosystems: connecting biological processes and water chemistry to sediment P fractions in urban stormwater management ponds. *Biogeochemistry* 132:203–212
34. Tarasova NP, Makarova AS, Vasileva EG, Shlyakhov PI, Zanin AA (2018) Evaluation of the Phosphorus Load on Freshwater Bodies of Subjects of the Russian Federation: Modeling of the Migration of Phosphorus and Its Compounds among Environmental Components. *Dokl Earth Sci* 480:818–822
35. Wang J, Song Y, Xue Y (2020) Influence Scope of *Potamogeton crispus* Population on Lake Waterquality during the Growth Period in Xuanwu Lake, China. *J Water Chem Technol* 42:491–497
36. Wang JQ, Song YZ, Wang GX (2017) Causes of large *Potamogeton crispus* L. population increase in Xuanwu Lake. *Environ Sci Pollut Res* 24:5144–5151
37. Wang LZ, Liu QJ, Hu CW, Liang RJ, Qiu JC, Wang Y (2018) Phosphorus release during decomposition of the submerged macrophyte *Potamogeton crispus*. *Limnology* 19:355–366
38. Xu W, Li R, Liu S, Ning Z, Jiang Z (2017) The phosphorus cycle in the Sanggou Bay. *Acta Oceanol Sin* 36:90–100
39. Zhou Y, Dodds W (1995) Kinetics of size-fractionated and dissolved alkaline phosphate in a farm pond. *Arch Hydrobiol* 134:93–102
40. Zhu YR, Zhang RY, Wu FC, Qu XX, Xie FZ, Fu ZY (2013) Phosphorus fractions and bioavailability in relation to particle size characteristics in sediments from Lake Hongfeng, Southwest China. *Environ Earth Sci* 68:1041–1052

Figures

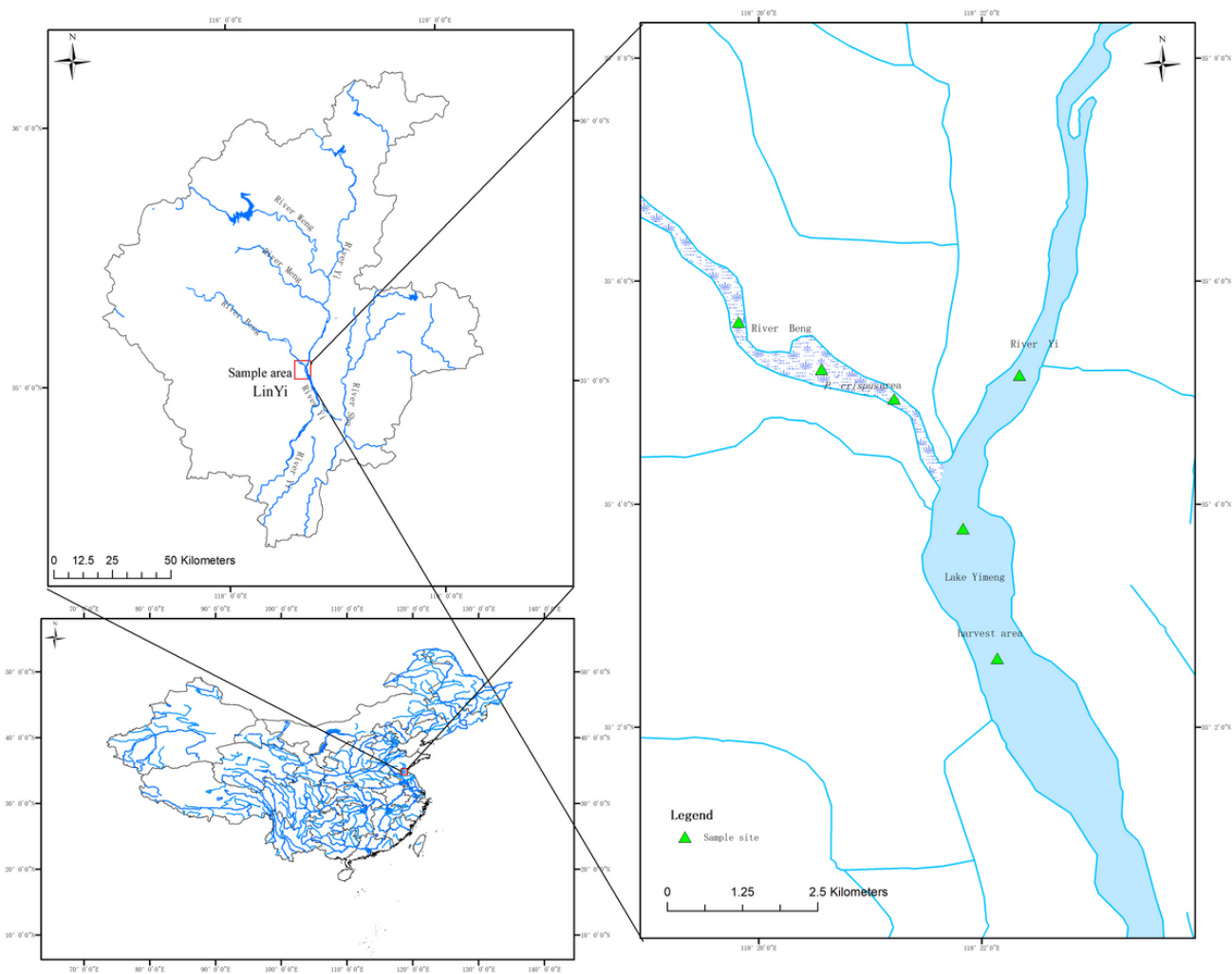


Figure 1

Location of studied area and its sampling sites.

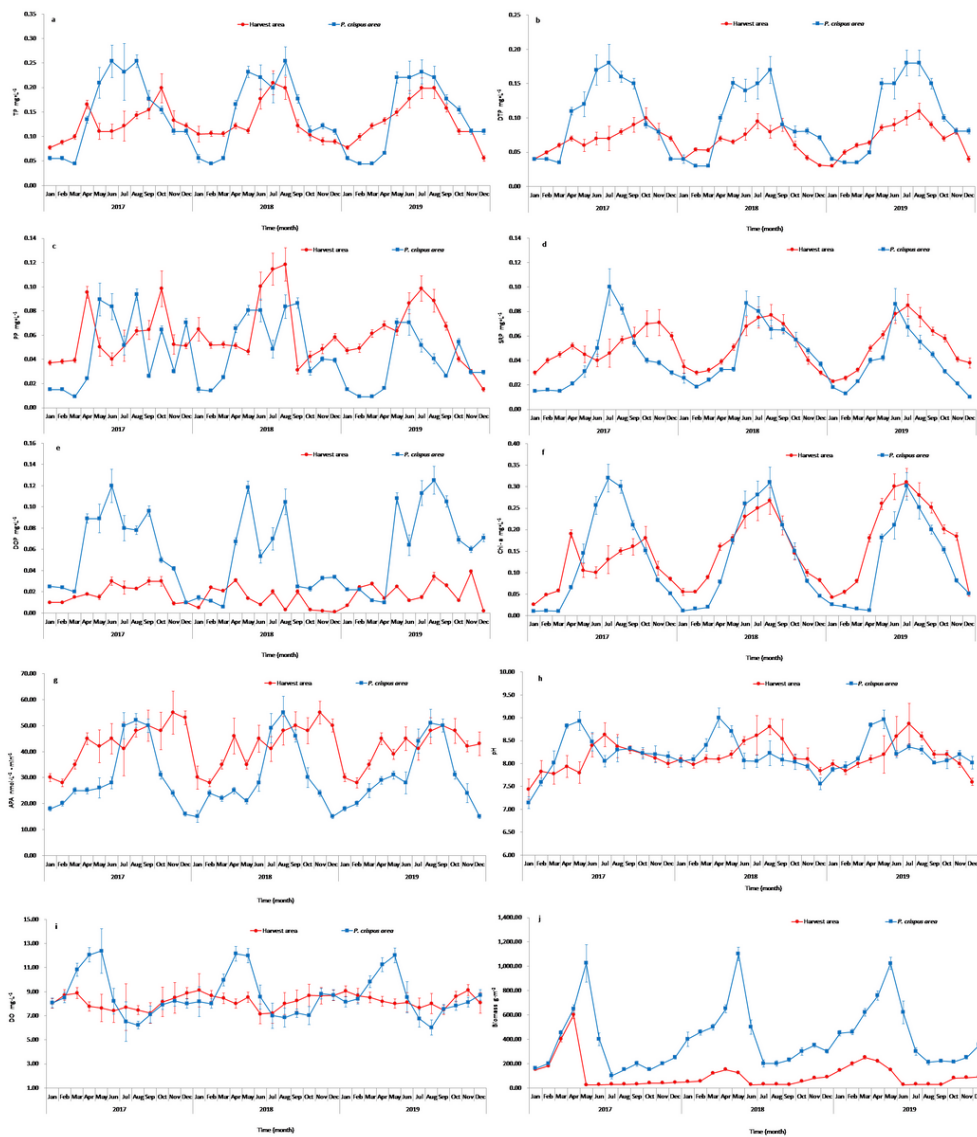


Figure 2

Monthly variations of water chemical and physical parameters (mean $\pm$ SD) in different areas of Yimeng Lake from January 2017 to October 2019. a-e water different P fraction concentration in different lake areas; f water chl-a concentration in different lake areas; g water APA concentration in different lake areas; h-i physical parameters in different lake areas. j biomass in different lake areas.



Figure 3

Monthly variations of sediment P fractions (mean $\pm$ SD) in different areas of Yimeng Lake from January 2017 to October 2019. a-c different P fractions in sediment. d percentage of IP and OP in the harvest area, e percentage of IP and OP in the *P. crispus* area.

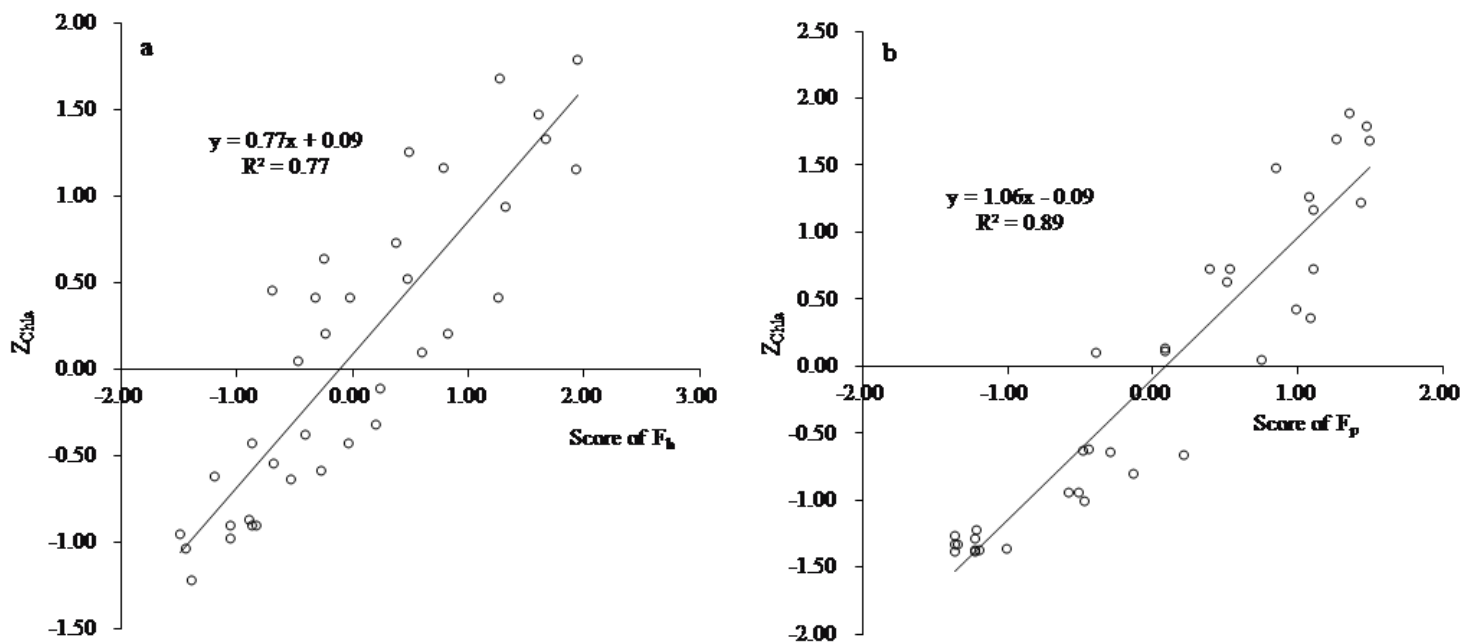


Figure 4

The relationship between Z score of chl a and score of F_h and F_p

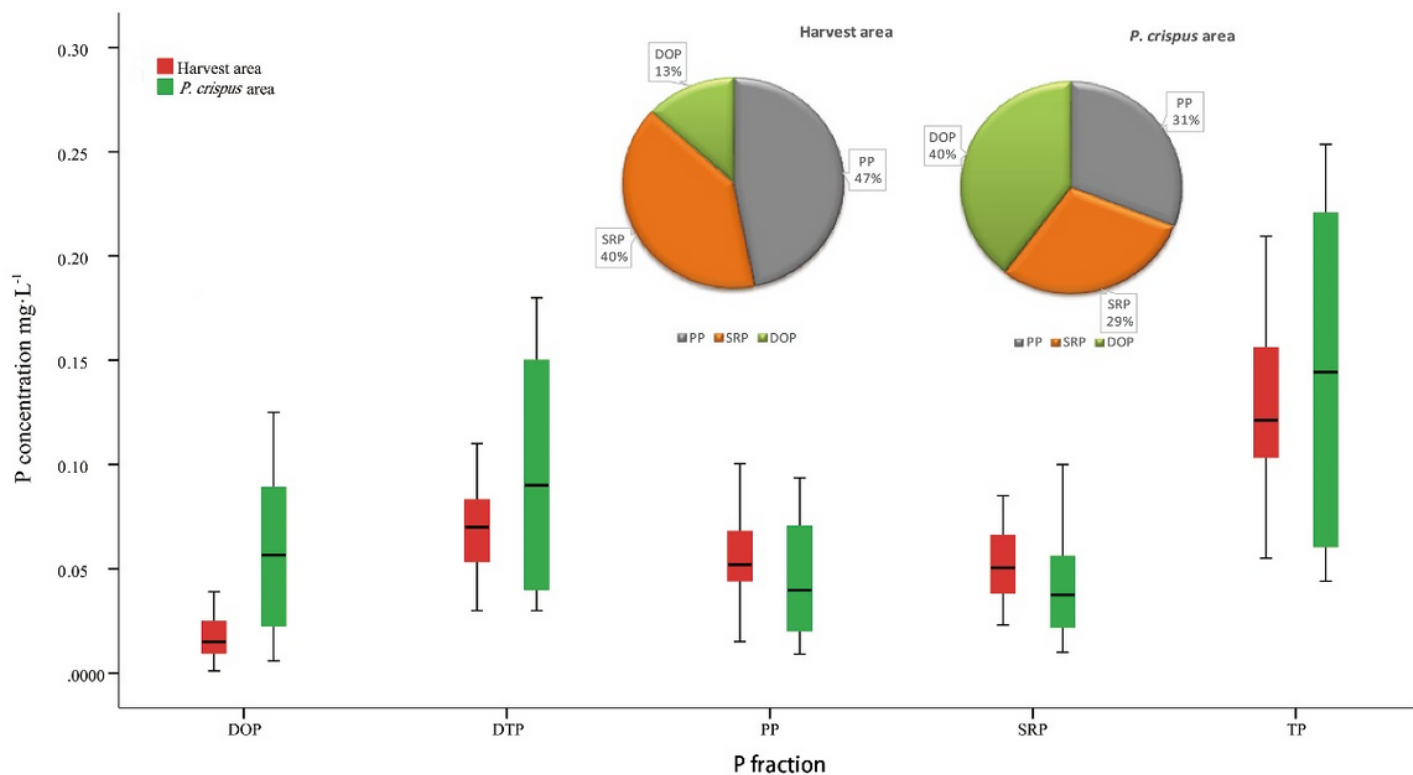


Figure 5

Box plots of different P fractions and P structure in water in different areas of Yimeng Lake