



OPEN Impact of dry-off and lyophilized *Aloe arborescens* supplementation on plasma metabolome of dairy COWS

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Positive effects have been observed as a result of *Aloe arborescens* supplementation in the dry-off phase in dairy cows. Metabolomic approaches can provide additional information about animal physiology. Thus, we characterized plasma metabolome around dry-off in 12 cows supplemented (AL) or not (CTR) with 10 g/d of lyophilized *A. arborescens* with an untargeted metabolomic approach. Overall, 1658 mass features were annotated. Regardless of treatment, multivariate statistics discriminated samples taken before and after dry-off. Overall, 490 metabolites were different between late lactation and early dry period, of which 237 were shared between AL and CTR. The most discriminant compounds (pentosidine and luteolin 7-*O*-glucoside) were related to the more fibrous diet. Pathway analysis indicated that pyrimidine and glycerophospholipid metabolisms were down-accumulated, suggesting reduced rumen microbial activity and liver load. Samples from AL were discriminated from CTR either the day of dry-off or 7 days after. At dry-off, aloin and emodin were the most discriminant metabolites, indicating that *Aloe's* bioactive compounds were absorbed. Seven days later, 534 compounds were different between groups, and emodin was among the most impacted. Pathway analysis highlighted that glycerophospholipid, pyrimidine, and folate metabolisms were affected. These results might indicate that *Aloe* has positive effects on liver function and a modulatory effect on rumen fermentation.

The dry-off represents a stressful event for dairy cows, encompassing an abrupt milking cessation, diet and group changes, and consequent physiological alterations^{1,2}. In fact, the dry-off causes impairments of metabolism, liver function, and antioxidant system, and also triggers an inflammatory response^{3–5}. Recently, these changes have been intensively investigated^{5–7}, but a lot of work is still required to decipher all the pathways involved. Dry-off is a transitional phase that takes place during the last weeks of gestation and is characterized by biological variations related to the changes in diet and hormonal and metabolic settings⁵. In contrast, the transition period in dairy cows represents the three weeks before and after calving^{8,9} and is characterized by sudden and dramatic changes including alterations in metabolism, inflammatory response, oxidative stress, and a reduction in immune competence¹⁰. At dry-off, all these phenomena are present^{4,6,11}, even though their magnitude is usually lower.

To cope with similar drastic physiological changes to those occurring at dry-off, the administration of nutraceuticals during the transition period has shown positive results in regulating immune responses and metabolism¹². However, literature addressing the application of these products at dry-off is scarce. *Aloe* spp. are among the most used plants in traditional medicine for their therapeutic properties, such as wound healing, anti-inflammatory, antioxidant, antitumor, antimicrobial, and immunomodulatory effects¹³. The most relevant active compounds of *Aloe* are anthraquinones and their glycosides, which are present in the leaf green rind, and complex carbohydrates, which are detectable in the parenchyma¹⁴. The biological activities of *Aloe* are likely related to a synergic action of several compounds¹⁴, but the two main components are aloin, a C-glycoside derivative of a 1,8-dihydroxyanthraquinone, and an acemannan (a typical D-isomer mucopolysaccharide of *Aloe* leaves)¹⁵. The use of *Aloe arborescens* has been evaluated in dairy cows. Aloin appears to be effectively absorbed, since aloin is detectable in the blood of dairy cows 2 h after oral administration of 200 g/d of whole leaves *A.*

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arborescens homogenate, without any adverse effect on feed intake and digestibility¹⁶. Moreover, *A. arborescens* supplementation during transition period showed a positive impact on lipid mobilization, liver and kidney function, and mitigated the inflammatory response at calving¹⁷. Recently, we tested the use of *A. arborescens* as a lyophilized powder at dry-off¹⁸. The lyophilization of the homogenate can facilitate the storage, preservation, and administration of the product, which are impractical on commercial farms. Cows that received *A. arborescens* had a slightly different rumen fermentation pattern and improved liver function at dry-off¹⁸. Moreover, they had a reduced inflammatory response later at calving and higher milk production in the first weeks of lactation. However, the exact mode of action of *A. arborescens* has yet to be fully elucidated.

Omics approaches have been recently used by researchers to gain a better understanding of both cow physiology^{19–21} and different feeding strategies^{22,23}. Particularly, metabolomics is a high-throughput technique that allows for the concurrent determination of myriad metabolites in several biological samples²⁴, consisting of untargeted and targeted approaches. Targeted metabolomics is used to quantitatively detect metabolites in metabolic pathways of interest, whereas untargeted metabolomics examines metabolite differences between control and experimental groups, which also plays an important role in marker screening for a certain disease or treatment¹⁹. Of interest, metabolomics has been used to study the transition period^{25–27} and phytogetic compound action in animal models²⁸. Moreover, blood metabolomics has been extensively applied in the last few years to identify markers of several diseases^{29–32}. However, to the best of our knowledge, studies correlating blood metabolomics to the dry-off period are still lacking.

Therefore, in this study, we used an untargeted metabolomic approach to analyze the plasma metabolite profile. The first aim was to assess whether *A. arborescens* metabolites could be detected in plasma with this technique. Then, according to previous results, we hypothesized that dry-off (performed without antibiotic treatment) would alter the plasma metabolome and lyophilized *A. arborescens* Mill. supplementation could partially improve the physiological response in this phase.

Results and discussion

To the best of our knowledge, a metabolomic approach has not yet been applied to study the dry-off phase in dairy cows. Nevertheless, both targeted and untargeted metabolomic approaches have been mainly used to explore the periparturient period in dairy cows and the supplementation with different products^{25,33}. In particular, untargeted metabolomics can provide new insights into dairy cow physiology in critical phases of the lactation cycle. Therefore, in the present study, we used untargeted metabolomics, based on liquid chromatography coupled with high-resolution mass spectrometry and multivariate statistical modeling, to characterize the biochemical changes occurring in plasma in response to either dry-off or *A. arborescens* supplementation.

Plasma metabolome profile. The untargeted metabolomic analysis resulted in the identification of 1657 mass features, that were annotated according to a Level 2 of confidence, including both putative annotation and MS/MS confirmation against a comprehensive ad-hoc database of mass spectra (Bovine Metabolome Database³⁴). A comprehensive list reporting each annotated metabolite together with its composite mass spectrum, total annotation score, and relative abundance value can be found in Supplementary Table S1. Under our experimental conditions, a wide variety of chemical entities could be recorded, and the most represented classes of compounds detected are reported in Fig. 1. The high number of blood metabolites obtained by the untargeted metabolomic approach denotes the complexity of the biofluid investigated. Overall, amino acids and peptides

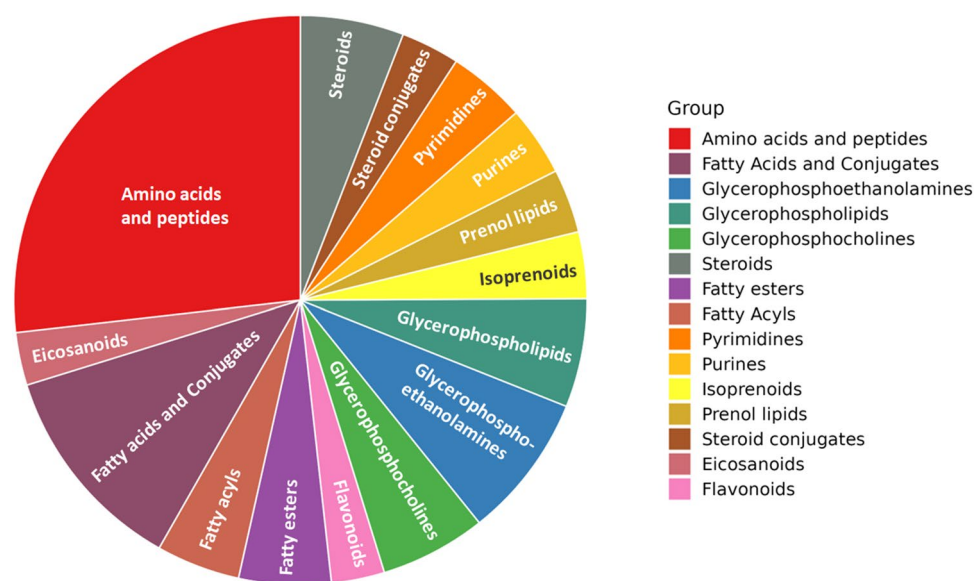


Figure 1. Pie chart showing the chemical classes annotated by untargeted metabolomics in the different plasma samples considering the classes of metabolites containing at least two entries.

were by far the most numerically represented class (170 compounds), followed by several sub-classes of lipids and derivatives (such as fatty acids, glycerophosphoethanolamines, glycerophospholipids, glycerophosphocholines, steroids, fatty esters, and fatty acyls). Among the annotated amino acids, we found all those classified as aromatic (i.e. phenylalanine, tryptophan, and tyrosine), together with proline, histidine, lysine, serine, cysteine, glutamine, leucine, methionine, arginine, and glutamic acid (Supplementary Table S1). Of interest, several peptides characterized the dataset, with a great abundance of dipeptides. Also, some metabolites associated with protein mobilization in cattle, such as 3-methylhistidine³⁵, creatine, and creatinine were detected. Lipids and lipid metabolism are key aspects of the physiology of transition cows^{36,37} but, to some extent, have been studied also in relation to late lactation and dry-off^{38–40}. Additionally, metabolomics allowed the identification of feed-derived compounds (such as isoprenoids and flavonoids), together with purine and pyrimidine derivatives (cumulatively accounting for 53 compounds), prenol lipids (including terpenoids), and some endocrine mediators (i.e. eicosanoids). Among feed-derived compounds, we found hippuric acid. Hippuric acid is formed in the liver to detoxify benzoic acid (a phenolic acid) in the reaction with glycine and can represent an indicator of plant digestion⁴¹. Furthermore, looking at some nucleic acid intermediates, uric acid was observed in each plasma sample. The latter is produced from the oxidation of xanthine via xanthine dehydrogenase in the intestinal mucosa, liver, and blood⁴¹. Taken together, the preliminary information obtained by exploring the plasma metabolome profile was coherent with the complex phenomena under investigation, justifying further data reduction and extrapolations to identify the key biomarkers involved.

Effect of dry-off. Regardless of dietary treatment, the unsupervised principal components analysis (PCA) discriminated the samples taken at the beginning of dry period from those collected in late lactation (Fig. 2). The first principal component explained around 17% of the total variance, thus suggesting the presence of great variability among samples. The dry-off is a moment of intense physiological and metabolic modifications^{5–7}, during which milking is stopped. To firstly depress milk synthesis stimuli and then adapt to the lower demand of the mammary gland, the energy content of the diet is reduced, increasing the fiber fraction. Consistently, the heatmap resulting from the unsupervised hierarchical cluster analysis (Fig. 3) showed a strong clustering of samples taken after dry-off (i.e. 7 DFD). Metabolomic profiles in AL and CTR at 7 DFD clustered together, and they were markedly different from those observed in late lactation (i.e. –7 or 0 DFD). Results obtained with this omics approach confirmed previous observations made with other techniques highlighting a different overall metabolic state between lactating and nonlactating periods.

To identify the metabolites that were mostly affected by the dry-off event, supervised OPLS-DA models were applied separately to AL and CTR groups, comparing samples before dry-off with those collected after milking cessation (7 DFD; Supplementary Fig. S1). This technique allows us to identify the most discriminant compounds (i.e. the plasma metabolites driving the discrimination according to the cluster type) between lactation and early dry period. OPLS-DA is widely used in metabolomic research for identifying those biomarker compounds maximizing group separation²³. In both treatment groups, the score plots separated samples taken before dry-off from those collected in early dry period, confirming the results of the unsupervised PCA. This outcome was also supported by the validation parameters, being the goodness-of-fit (R^2Y) = 0.99 and the

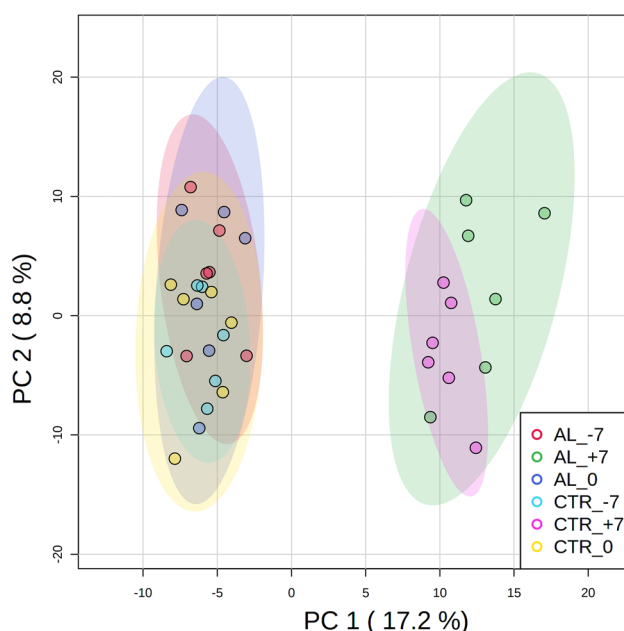


Figure 2. Principal component analysis carried out on plasma samples collected at –7, 0, and 7 days from dry-off (DFD) in dairy cows receiving 10 g/d of lyophilized *Aloe arborescens* Mill. (AL) from –7 to 7 DFD or in the control group (CTR).

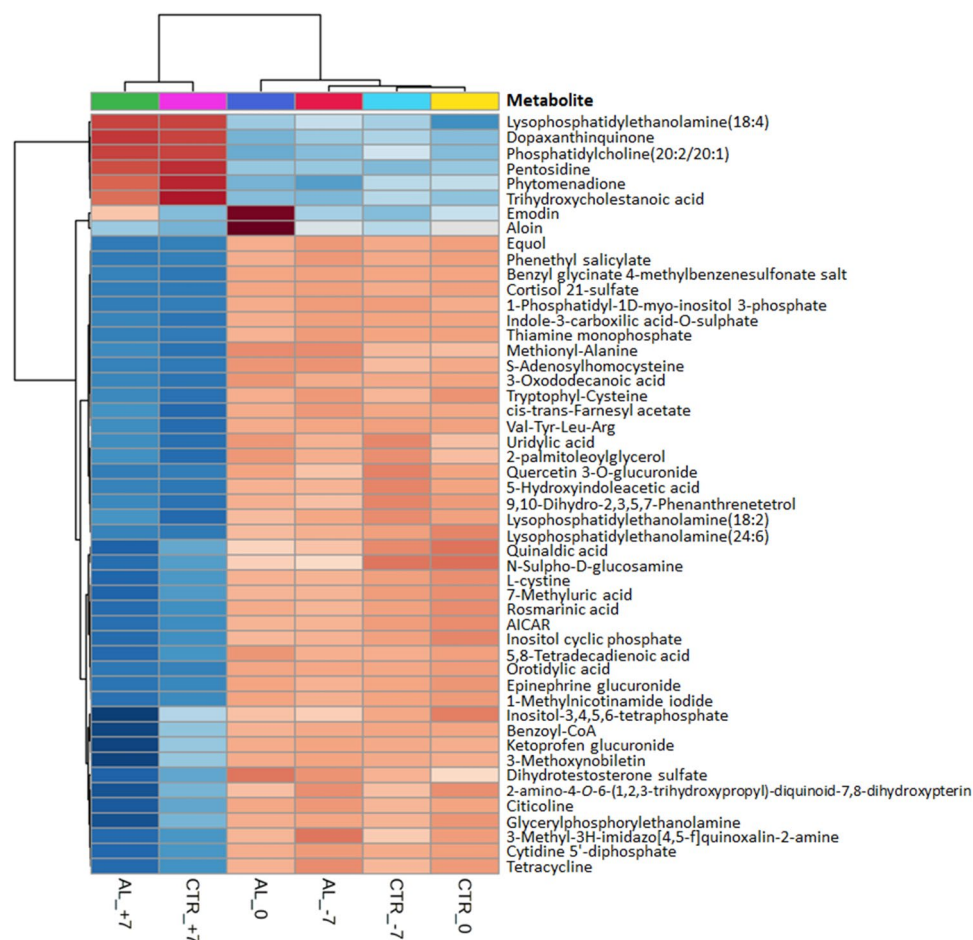


Figure 3. Heatmap with hierarchical cluster analysis showing the longitudinal changes in the plasma metabolome profile across sampling day (−7, 0, and 7 days from dry-off) and dietary treatment (lyophilized *A. arborescens* supplementation [AL] or control diet [CTR]). Groups at each sampling day are represented in the columns. The top 50 metabolites were ranked by the *P*-value of one-way ANOVA and represented in the lines. Data were auto-scaled by features.

goodness-of-prediction (Q^2) = 0.92. The discriminant compounds were extrapolated by VIP selection method (Table 1), and 490 compounds showed the highest prediction ability (VIP score ≥ 1). Among those, 127 were exclusively differentially expressed in AL, 126 only in CTR, and 237 were shared between the groups. Looking at the common discriminant compounds, the most represented and enriched classes were amino acids and peptides (20 compounds, with a great abundance of dipeptides), steroids (9 compounds, including some metabolites of progesterone and 17- β -estradiol), pyrimidines (8 compounds, including orotidylic acid, thiamine, and dihydrouracil), glycerophosphoethanolamines (7 compounds), and indoles (including some tryptophan derivatives, such as 5-methoxytryptophan and tryptophanol). Interestingly, among the most discriminant compounds in both models pentosidine and luteolin 7-*O*-glucoside were present. The first one is an advanced glycation end product, linked to the Maillard reaction. The up-accumulation of this compound after dry-off might be related to dietary intake, due to Maillard reaction happening during the sun drying of the hay fed⁴². However, it is usually rapidly excreted by glomerular filtration in healthy subjects⁴³. Thus, alongside the latter, another possible explanation could be represented by a transient impairment of kidney function in this phase. The latter could be supported by the higher plasma concentration of creatinine after dry-off^{5,44}, since circulating creatinine is related to the kidneys' ability to remove it from the bloodstream⁴⁵. Moreover, advanced glycation end products are recognized as pro-inflammatory mediators, their formation is related to that of reactive oxygen metabolites, and can be formed as a result of oxidative stress and along inflammatory pathways⁴⁶. Oxidative stress and inflammation can happen as a result of dry-off⁵. Luteolin 7-*O*-glucoside, instead, is found in grass hay⁴⁷. Its increase was related to the hay-based diet fed after dry-off, which drastically changed from the lactation diet, which had lower hay inclusion and higher amount of silages and concentrates.

From the pathway analysis conducted on each group comparing samples before dry-off with those collected at 7 DFD, there were 2 shared pathways. The most important was the pyrimidine metabolism pathway ($P < 0.01$). Pyrimidine metabolites were mostly down-accumulated, likely indicating a diminished rumen microbial N production. Since pyrimidines are nucleic acid constituents and the amount of DNA and RNA synthesized in the

Compound	VIP score	cvSE ^a	LogFC ^b
Citicoline	4.78	0.73	− 9.33
Epinephrine glucuronide	4.51	1.27	− 8.81
Benzoyl-CoA	4.39	0.93	− 8.27
Dopaxanthin quinone	4.37	1.09	9.09
Pentosidine	4.29	1.26	8.62
Phosphatidylinositol 3-phosphate	4.29	1.88	− 8.03
Tetracycline	4.05	0.87	− 7.16
Indole-3-carboxylate-O-sulphate	3.94	0.88	− 6.48
S-Adenosylhomocysteine	3.92	1.34	− 6.77
Benzyl glycinate 4-methylbenzenesulfonate	3.80	0.70	− 5.88
Phytomenadione	3.72	1.11	6.17
Luteolin 7-O-glucoside	3.70	1.57	6.75
Quetiapine	3.56	1.97	− 5.76
Lysophosphatidylethanolamine(18:4)	3.47	1.22	5.79
Maltohexaose	3.44	2.50	6.20
(3 α ,7 α ,12 α)-Trihydroxy-5 β -cholestanoic acid	3.43	1.29	5.40
Cytidine 5'-diphosphate	3.37	0.97	− 4.98
Triacylglycerol(18:3/22:6/22:2)	3.36	1.32	5.48
6-Succinoaminopurine	3.34	1.15	6.05
Tryptophyl-cysteine	3.18	1.25	− 4.20

Table 1. Most discriminant compounds (according to variable importance in projection [VIP]) between plasma samples collected before (−7 and 0 days from dry-off [DFD]) or after dry-off (7 DFD). ^acvSE, cross-validation standard error. ^bLogFC, logarithm of Fold-Change values.

rumen depends largely on bacterial growth^{48,49}, we hypothesized that the reduced rumen fermentation intensity after dry-off⁵⁰ could account for this result.

Liver function is transiently impaired after dry-off⁵. In the present study, this was supported also by the down-accumulation of compounds taking part in the glycerophospholipid metabolism pathway ($P < 0.01$). Glycerophospholipids are involved in triglyceride export by the liver, playing a key role in very low-density lipoprotein (VLDL) hepatic synthesis. Therefore, the mild increase in plasma nonesterified fatty acids (NEFA) usually observed immediately after dry-off^{5,6,18} might be interpreted as a decreased liver ability to remove NEFA from the bloodstream rather than excessive fat mobilization. Glycerophospholipid down-accumulation after dry-off might also be linked to a reduced liver load driven by the lower intake, which makes less necessary VLDL synthesis.

Effect of Aloe supplementation. Previous research showed the beneficial effects of *A. arborescens* supplementation during transition¹⁷ and dry-off periods¹⁸. Particularly, *A. arborescens* homogenate supplementation during transition period resulted in lower plasma concentrations of NEFA, β -hydroxybutyrate (as markers of reduced lipid mobilization), myeloperoxidase, and ceruloplasmin, and higher concentrations of cholesterol, retinol, tocopherol, and paraoxonase (as markers of mitigated inflammatory response at calving), also paired with lower plasma concentrations of creatinine and bilirubin (indicating an improved kidney and liver function)¹⁷. At dry-off (in the same cows of this study), lyophilized *A. arborescens* supplementation led to higher plasma concentrations of paraoxonase and glucose, and lower urea¹⁸, suggesting a better liver function. Paraoxonase is a liver enzyme that is released into the bloodstream and its decrease can be interpreted as a liver impairment indicator⁵¹. Similarly, glucose supply heavily relies on liver endogenous production, particularly during transition period⁵².

Consistently with those results, alongside the sampling day effect, the heatmap reported in Fig. 3 also shows the clustering of samples according to the *Aloe* treatment. There was a clear grouping between AL and CTR at 7 DFD, and samples taken at 0 DFD in AL were markedly different from their counterparts in CTR, with aloin and emodin being the most discriminant compounds. In our previous paper¹⁸, we showed positive effects of *A. arborescens* supplementation. The annotation of these compounds in the plasma of supplemented cows biologically supported those results showing that *Aloe* bioactive compounds were effectively absorbed. Emodin is a natural anthraquinone derivative with a wide spectrum of pharmacological properties, including anticancer, hepatoprotective, anti-inflammatory, antioxidant, and antimicrobial activities⁵³ and it represents, together with aloin, one of the main secondary phenolic metabolites of *Aloe*⁵⁴. It takes its origin from the conversion of barbaloin (a mixture of two diastereomers, aloin A and aloin B) by intestinal bacteria. Previous research suggested that aloin can be metabolized into aloe-emodin in the intestine and then absorbed into the bloodstream⁵⁵. However, Bani et al.¹⁶ successfully detected aloin, but not emodin, in plasma of dairy cows supplemented with 200 g/d of *A. arborescens* homogenate. This discrepancy between these two studies could be related to the different analytic techniques used, mainly in terms of FWHM resolution (liquid chromatography coupled to triple quadrupole mass spectrometry vs ultra-high-pressure liquid chromatography coupled with orbitrap mass spectrometry), or to the different

forms of supplementation (lyophilized vs homogenate). High mass resolution is particularly important for all types of experiments involving complex mixtures, such as samples generated from a biological matrix (such as plasma, milk, and others), since these latter potentially contain a significant number of background (matrix) ions in addition to the possible analytes of interest. In such cases, high mass resolution makes the difference between detecting analyte molecules at low concentration and not detecting them due to the masking effect of isobaric matrix interferences^{56,57}. Then, considering that samples taken at 7 DFD were different from those before dry-off and that a tendency toward separation between AL and CTR was observed at 7 DFD (Supplementary Fig. S1), we focused our analysis on the early dry period when, also in our previous paper¹⁸, the largest differences were noted. Another OPLS-DA model was applied to the data, considering three clusters: samples taken before dry-off and samples taken at 7 DFD either in AL or CTR (Fig. 4). The plot showed a clear separation between the clusters ($R^2Y = 0.98$ and $Q^2 = 0.55$). As far as the VIP scores are concerned, 534 compounds showed the highest prediction ability (VIP score ≥ 1) thus successfully discriminating AL from CTR. Amino acids and peptides were the most represented compounds (54 metabolites), followed by fatty acids and conjugates (21 metabolites), pyrimidines, steroids conjugates, and fatty acyls (13 metabolites each), and isoprenoids, steroids, glycerophospholipids, and glycerophosphoethanolamines (12 metabolites each).

Of interest, the most discriminating compound according to the VIP algorithm between AL and CTR at 7 DFD was emodin (Table 2), consistent with the previous discussion. In this case, aloin was not among those compounds with the highest prediction ability. The reason might be related to the different rumen environment which could have resulted in higher degradation of aloin or a higher conversion into emodin, the latter potentially involving a higher deglycosylation activity at the intestinal level by hydrolyzing enzymes. It is also important to highlight that, in this work, the database used for the annotation does not contain structural information regarding glucuronides/sulfates forms of anthraquinones, which are reported to play an important role in aloin/emodin absorption following the activity of phase II enzymes⁵⁸. Therefore, up to date, limited information is available on anthraquinone bioavailability, and further studies are required to confirm our findings. The second most discriminant ability was outlined for glycylphosphorylethanolamine. It is a membrane breakdown product resulting from the cleavage of the lipid group from glycerophosphoethanolamine fatty acids (i.e. phosphatidylethanolamine). Its lower relative accumulation in AL ($\log FC = -0.95$) was likely related to the aforementioned better liver function observed in those cows, as supported by the higher plasma paraoxonase concentration¹⁸.

The compound with the highest fold change was a compound of the class of phosphatidylcholines ($\log FC = 5.57$). In cattle, a decrease in the abundance of lipids related to phosphatidylcholine metabolism was associated with increased severity of fatty liver⁵⁹. Phosphatidylcholine is involved in VLDL synthesis and helps prevent fatty liver, exporting triglycerides from the hepatocytes as VLDL^{60,61}. This might help explain the anti-hyperlipidemic role of *Aloe* previously reported¹⁷ and the improved liver function that we observed¹⁸.

Then, we investigated the differentially expressed pathways (Table 3). Among those that resulted significant, glycerophospholipid, pyrimidine, folate, nicotinate, and nicotinamide metabolisms were the most impacted. Together, these pathways suggest that liver and rumen metabolism might have been key regulators of the *Aloe* effects described previously^{17,18}. Relative plasma concentration of compounds taking part in the glycerophospholipid metabolism (mostly phosphatidylethanolamine and phosphatidylcholine) was reduced. To the best of our knowledge, information about glycerophospholipid metabolism at dry-off is lacking. Nevertheless, it has

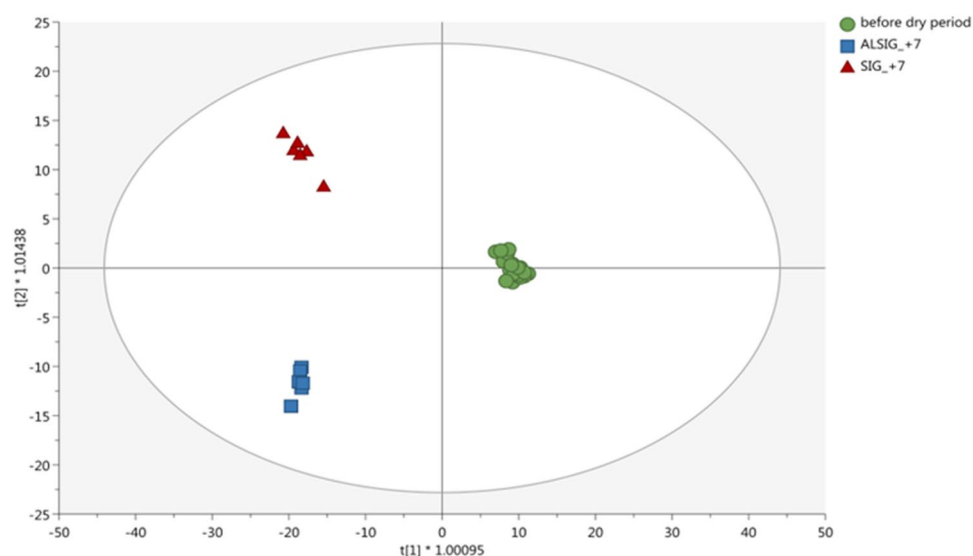


Figure 4. Orthogonal projection to latent structures (OPLS) discriminant analysis (DA) considering the cluster type as class discrimination parameter. Plasma samples were collected at -7 , 0 , and 7 days from dry-off (DFD) in dairy cows receiving 10 g/d of lyophilized *Aloe arborescens* Mill. (AL) from -7 to 7 days relative to dry-off or in the control group (CTR). Samples taken before dry-off (-7 and 0 DFD) were considered as a reference. The comparison between samples collected at 7 DFD in AL and CTR was further discussed.

Metabolite	VIP score	cvSE ^a	LogFC ^b
Emodin	2.32	0.81	3.25
Glycerolphosphorylethanolamine	2.27	0.95	− 0.95
3-Methoxynobletin	2.26	0.93	− 0.93
Ketoprofen glucuronide	2.26	0.93	− 0.93
Succinylaminoimidazole carboxamide ribotide	2.24	0.97	− 0.95
Geranylgeranylethanolamine	2.07	0.66	1.73
4-t-octylphenol	2.06	1.16	− 1.02
Cinnamyl alcohol	2.02	0.76	2.98
Cytidine-diphosphate diacylglycerol (18:0/22:3)	2.02	1.23	0.32
Ethenodeoxyadenosine	2.00	1.01	− 1.09
Sinapic acid	1.99	1.17	0.22
Quinaldic acid	1.99	1.07	− 0.22
Flavin Mononucleotide	1.99	1.08	− 1.01
L-isoleucyl-L-proline	1.98	1.16	− 0.30
D-Xylono-1,5-lactone	1.96	0.96	− 0.79
D-Myo-inositol 3,4,5,6-tetrakisphosphate	1.96	1.37	− 1.82
Triacylglycerol(20:3n6/15:0/O-18:0)	1.91	1.34	0.29
Alpha-N-Phenylacetyl-L-glutamine	1.90	0.90	− 0.62
Vanillylmandelic acid	1.88	1.06	− 1.06
Tricosanoic acid	1.85	1.30	− 0.27

Table 2. Most discriminant compounds (according to variable importance in projection [VIP]) between plasma samples collected 7 days from dry-off (DFD) in dairy cows receiving 10 g/d of lyophilized *Aloe arborescens* Mill. (AL) considering −7 to 7 days from dry-off or in the control group (CTR). ^acvSE, cross-validation standard error. ^bLogFC, logarithm of Fold-Change values.

Pathway name	Metabolites impacted ^a	P-value	Impact
Glycerophospholipid metabolism	8/36	< 0.01	0.52
Pyrimidine metabolism	8/38	< 0.01	0.24
One carbon pool by folate	3/9	0.02	0.91
Purine metabolism	9/66	0.02	0.20
Nicotinate and nicotinamide metabolism	8/13	0.05	0.43

Table 3. Significantly impacted pathways by *Aloe* supplementation (AL; 10 g/d of lyophilized *A. arborescens* Mill. from −7 to 7 days from dry-off [DFD]) compared with control diet (CTR) in dairy cows at 7 DFD. ^aNumber of metabolites impacted by *Aloe* supplementation out of the total number of metabolites in the considered pathway.

been reported that, during transition period, blood glycerophospholipid decrease can be interpreted as a marker of fatty liver³⁶ but was also associated with a desired healthy metabolic phenotype²⁵. In the present experiment, cows supplemented with *A. arborescens* at dry-off seemed to have improved liver function. Thus, the down-accumulation of glycerophospholipids might result from reduced liver load in a phase when the liver has to cope with several adaptations⁵. Alternatively, glycerophospholipid down-accumulation might have been driven by the lower availability of methyl donors, discussed thereafter,

Pyrimidine relative concentrations were mostly reduced in AL cows compared with CTR. As mentioned before, pyrimidines in the bloodstream are mainly of rumen origin, being constituents of microbial DNA and RNA^{23,48,49}. Despite the lack of data after dry-off, the day of dry-off rumen fluid volatile fatty acid concentration tended to be lower in AL cows¹⁸. Together, these results can indicate that *Aloe* has a modulatory effect on ruminal fermentation. Although in vitro trials did not show any adverse effect of *Aloe* incubation on rumen fermentation parameters^{16,62}, in vivo results might be different. We previously hypothesized that a different epithelial absorption rate might also be implied, as suggested by the higher daily rumination time observed¹⁸. Looking at changes in pyrimidines, we would support the modulatory effect hypothesis rather than the increased absorption, but further studies are needed to disentangle this matter. It may also be hypothesized that *Aloe* could reduce microbial turnover through some effect on protozoa⁶³.

Finally, folate metabolites were mostly down-accumulated. Folate cycle is part of the one-carbon metabolism^{12,64}. The decrease in plasma concentrations of these compounds might suggest an increased consumption of methyl donors in those cows^{60,65}. Therefore, it may be possible to speculate that *Aloe* action was related to some effect on DNA methylation pattern, supporting the mitigated inflammatory response observed

two months later at calving¹⁸. These hypotheses agree with previous findings associating cow condition at dry-off and calving⁶⁶ and might help explain why priming dairy cows long before calving has beneficial effects⁹.

Conclusions

The dry-off confirmed to be a phase of intense modifications in the metabolism of dairy cows, with deep alterations in metabolism and liver function. Of interest, metabolic pathways involved in pyrimidines and glycerophospholipid metabolisms were impacted by the transition to dry period. At the same time, supplementing lyophilized *A. arborescens* seemed to improve the adaptation to dry period at two levels: at the rumen level and tuning some metabolic pathways. In this study, using an untargeted metabolomic approach, we demonstrated that aloin, the main *Aloe* bioactive compound, is absorbed by cows and metabolized to emodin, supporting previous observations and confirming the positive effect of *Aloe* supplementation on liver function in this phase.

Methods

Animal management and experimental design. The research was carried out at the Università Cattolica del Sacro Cuore dairy barn (Cezzo, San Bonico, Piacenza, Italy) in accordance with animal welfare guidelines (including ARRIVE guidelines) and Italian laws on animal experimentation and ethics (Italian Health Ministry Authorization N 444/2019-PR in agreement with D. Lgs. n. 26, 04/03/2014). This work was part of a larger study and detailed information about animals and experimental design, as well as results in terms of performance, milk, rumen fluid, and plasma analyses have been reported elsewhere¹⁸. Briefly, 23 Holstein dairy cows with milk somatic cell count less than 200,000 cells/mL and without intramammary infections due to major pathogens (i.e. *Staphylococcus aureus*, *Streptococcus uberis*, *Enterobacter* spp., *Escherichia coli*, *Trueperella pyogenes*, *Serratia marcescens*) were included in this study. During lactation, cows were housed in a free-stall pen, received lactation diet, and were milked twice daily (5:00 and 17:00 h). About 55 days before expected calving, cows were abruptly dried off with the infusion of internal teat sealant (Noroseal, Norbrook Laboratories Limited, UK) after the last milking. Then, they were moved to a dry pen and received ad libitum grass hay only for 7 days, after which dry period diet was administered. Cows received 10 g/d of lyophilized *A. arborescens* Mill. whole leaf homogenate supplemented to their diet from -7 to 7 days from dry-off (DFD) (AL; n = 11) or none for the control group (CTR; n = 12). Groups were balanced for parity, previous lactation length, and somatic cell history. Before total mixed ration (TMR) distribution, feed bunk was cleaned, all cows were restrained in the headlocks, and each dose of lyophilized *Aloe* was mixed with 1 kg of lactation TMR and fed to AL cows, whereas CTR cows received 1 kg of lactation TMR only. During the whole trial, an operator checked that cows ate all the supplemented TMR.

Blood samples. Blood samples were collected from the jugular vein into heparinized tubes before the morning feeding on -7, 0, and 7 DFD. Tubes were immediately cooled in an ice-water bath. Afterwards, whole blood was centrifuged at 3500×g for 15 min at 4 °C, and plasma was stored at -20 °C for subsequent assays. A subset of 6 cows/group was randomly selected and samples from these cows underwent metabolomic analysis, as described below.

The extraction of metabolites from plasma samples was carried out as previously reported by Luo et al.²⁷, with few modifications. Briefly, seventy-two plasma samples were slowly thawed at 4 °C, and from each sample, a 200-μL aliquot was taken and added to 800 μL of pre-cooled methanol/acetonitrile solution (1:1, vol/vol). Samples were then vortex mixed and maintained at -20 °C for 60 min, followed by centrifugation at 14,000×g and 4 °C for 20 min. The resulting supernatants were finally filtered through 0.22-μm cellulose syringe filters in ultra-high-pressure liquid chromatography (UHPLC) vials until further untargeted metabolomic profiling.

Untargeted metabolomic analysis. The untargeted UHPLC-HRMS analysis was done using a Q Exactive™ Focus Hybrid Quadrupole-Orbitrap Mass Spectrometer (Thermo Scientific, Waltham, MA, USA) coupled to a Vanquish UHPLC pump, equipped with heated electrospray ionization (HESI)-II probe (Thermo Scientific, USA). A water-acetonitrile (both LC-MS grade, from Sigma-Aldrich, Milan, Italy) gradient elution (6–94% acetonitrile in 35 min) was done for the chromatographic separation, using 0.1% formic acid as phase modifier, exploiting an Agilent Zorbax Eclipse Plus C18 column (50×2.1 mm, 1.8 μm). The flow rate was 200 μL/min, considering an injection volume of 6 μL. The untargeted analysis consisted of a full scan in the 100–1200 m/z range, using a positive ionization mode and setting a mass resolution of 70,000 at m/z 200. Automatic gain control (AGC) target and maximum injection time were 1×10⁶ and 200 ms, respectively. Under our instrumental conditions, pooled quality control samples were randomly injected and analyzed in a data-dependent (Top N = 3, under a stepped normalized collisional energy) MS/MS mode, with full scan mass resolution reduced to 17,500 at m/z 200, AGC target of 1×10⁵, maximum injection time of 100 ms, and isolation window of 1.0 m/z. The HESI parameters are reported elsewhere²³. The mass spectrometer was calibrated using Pierce positive ion calibration solution (Thermo Fisher Scientific, San Jose, CA, USA) before the UHPLC-HRMS run.

The collected raw data were further processed using the software MS-DIAL (version 4.70) for the automatic peak finding, LOWESS normalization, and annotation via spectral matching against the comprehensive Bovine Metabolome Database³⁴ (last accessed date: 15/11/2022). The MS and MS/MS tolerance for peak centroiding was set to 0.01 and 0.05 Da, respectively, considering features with a minimum peak height of 10,000 cps²³. Therefore, the identification step was based on mass accuracy, isotopic pattern, and spectral matching. The selected identification criteria were used to calculate a total identification score. The total identification score cut-off was > 50%, considering the most common source adducts. Additionally, a gap filling (using the peak finder algorithm) was done to fill in missing peaks, setting a 5-ppm tolerance for m/z values.

Statistical analysis. The metabolomics-based dataset was further elaborated for multivariate statistical modeling using two different software, namely MetaboAnalyst⁶⁷ and SIMCA 13 (Umetrics, Malmo, Sweden)²³. Data were log-transformed, and Pareto scaled. After normalization, both unsupervised and supervised multivariate statistics were carried out. The unsupervised approach was based on both PCA and hierarchical cluster analysis, while the orthogonal projections to latent structures discriminant analysis (OPLS-DA) was considered as a supervised tool. The OPLS-DA model validation parameters (goodness-of-fit R^2Y together with goodness-of-prediction Q^2Y) were recorded, considering a Q^2Y prediction ability of >0.5 as the acceptability threshold²³, and excluding model overfitting by permutation testing ($N > 100$). The discriminant potential of each plasma metabolite was then calculated according to the variable selection method variable importance in projection (VIP), using as the minimum significant threshold a VIP score ≥ 1 . Finally, the software MetaboAnalyst was used initially to inspect those metabolic pathways mostly represented by the metabolites annotated (using as pathway library: *Bos taurus*, KEGG), and then to provide a metabolite set enrichment analysis based on those significant classes of metabolites/metabolic pathways revealed by multivariate statistics.

Ethics declaration. The study was approved by the Università Cattolica del Sacro Cuore Animal Welfare Committee (OPBA) and authorized by the Italian Health Ministry (Authorization N 444/2019-PR in agreement with D. Lgs. n. 26, 04/03/2014).

Data availability

The dataset used and analyzed during the current study is available in Supplementary Table S1.

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References

- Vilar, M. J. & Rajala-Schultz, P. J. Dry-off and dairy cow udder health and welfare: Effects of different milk cessation methods. *Vet. J.* **262**, 105503 (2020).
- Zobel, G., Weary, D. M., Leslie, K. E. & von Keyserlingk, M. A. G. Invited review: Cessation of lactation: Effects on animal welfare. *J. Dairy Sci.* **98**, 8263–8277 (2015).
- Bertulat, S. *et al.* Measurement of fecal glucocorticoid metabolites and evaluation of udder characteristics to estimate stress after sudden dry-off in dairy cows with different milk yields. *J. Dairy Sci.* **96**, 3774–3787 (2013).
- Cattaneo, L. *et al.* Gene network expression of whole blood leukocytes in dairy cows with different milk yield at dry-off. *PLoS ONE* **16**, e0260745 (2021).
- Mezzetti, M., Minuti, A., Piccioli-Cappelli, F. & Trevisi, E. Inflammatory status and metabolic changes at dry-off in high-yield dairy cows. *Ital. J. Anim. Sci.* **19**, 51–65 (2020).
- Putman, A. K., Brown, J. L., Gandy, J. C., Wisnieski, L. & Sordillo, L. M. Changes in biomarkers of nutrient metabolism, inflammation, and oxidative stress in dairy cows during the transition into the early dry period. *J. Dairy Sci.* **101**, 9350–9359 (2018).
- Cattaneo, L. *et al.* Drying-off cows with low somatic cell count with or without antibiotic therapy: A pilot study addressing the effects on immunometabolism and performance in the subsequent lactation. *Livest. Sci.* **254**, 104740 (2021).
- Drackley, J. K. Biology of dairy cows during the transition period: The final frontier?. *J. Dairy Sci.* **82**, 2259–2273 (1999).
- Mezzetti, M. *et al.* The transition period updated: A review of the new insights into the adaptation of dairy cows to the new lactation. *Dairy* **2**, 617–636 (2021).
- Trevisi, E. & Minuti, A. Assessment of the innate immune response in the periparturient cow. *Res. Vet. Sci.* **116**, 47–54 (2018).
- Mezzetti, M., Bionaz, M. & Trevisi, E. Interaction between inflammation and metabolism in periparturient dairy cows. *J. Anim. Sci.* **98**, S155–S174 (2020).
- Lopreato, V. *et al.* Role of nutraceuticals during the transition period of dairy cows: A review. *J. Anim. Sci. Biotechnol.* **11**, 96 (2020).
- Singab, A.-N.B., El-Hefnawy, H. M., Esmat, A., Gad, H. A. & Nazeem, J. A. A systemic review on *Aloe arborescens* pharmacological profile: Biological activities and pilot clinical trials. *Phyther. Res.* **29**, 1858–1867 (2015).
- Hamman, J. Composition and applications of Aloe vera LEAF GEL. *Molecules* **13**, 1599–1616 (2008).
- Lucini, L., Pellizzoni, M. & Molinari, G. P. Anthraquinones and β -polysaccharides content and distribution in Aloe plants grown under different light intensities. *Biochem. Syst. Ecol.* **51**, 264–268 (2013).
- Bani, P. *et al.* Administration of *Aloe arborescens* homogenate to cattle: Interaction with rumen fermentation and gut absorption of aloin. *Ital. J. Anim. Sci.* **15**, 233–240 (2016).
- Mezzetti, M., Minuti, A., Bionaz, M., Piccioli-Cappelli, F. & Trevisi, E. Effects of *Aloe arborescens* whole plant Homogenate on lipid metabolism, inflammatory conditions and liver function of dairy cows during the transition period. *Animals* **10**, 917 (2020).
- Cattaneo, L., Piccioli-Cappelli, F., Minuti, A. & Trevisi, E. Drying-off dairy cows without antibiotic therapy and orally supplemented with lyophilized *Aloe arborescens*: Effects on rumen activity, immunometabolic profile, and milk yield. *J. Anim. Physiol. Anim. Nutr. (Berl)*. <https://doi.org/10.1111/jpn.13777> (2022).
- Cecilian, F., Lecchi, C., Urh, C. & Sauerwein, H. Proteomics and metabolomics characterizing the pathophysiology of adaptive reactions to the metabolic challenges during the transition from late pregnancy to early lactation in dairy cows. *J. Proteomics* **178**, 92–106 (2018).
- Yanibada, B. *et al.* Inhibition of enteric methanogenesis in dairy cows induces changes in plasma metabolome highlighting metabolic shifts and potential markers of emission. *Sci. Rep.* **10**, 15591 (2020).
- Lisuzzo, A. *et al.* Differences in the serum metabolome profile of dairy cows according to the BHB concentration revealed by proton nuclear magnetic resonance spectroscopy (1H-NMR). *Sci. Rep.* **12**, 2525 (2022).
- Pacífico, C., Stauder, A., Reisinger, N., Schwartz-Zimmermann, H. E. & Zebeli, Q. Distinct serum metabolomic signatures of multiparous and primiparous dairy cows switched from a moderate to high-grain diet during early lactation. *Metabolomics* **16**, 96 (2020).
- Rocchetti, G. *et al.* Milk metabolome reveals pyrimidine and its degradation products as the discriminant markers of different corn silage-based nutritional strategies. *J. Dairy Sci.* **105**, 8650–8663 (2022).
- Suh, J. H. Critical review: Metabolomics in dairy science—Evaluation of milk and milk product quality. *Food Res. Int.* **154**, 110984 (2022).
- Kenéz, Á., Dänicke, S., Rolle-Kampczyk, U., von Bergen, M. & Huber, K. A metabolomics approach to characterize phenotypes of metabolic transition from late pregnancy to early lactation in dairy cows. *Metabolomics* **12**, 165 (2016).
- Ghaffari, M. H. *et al.* Metabolomics meets machine learning: Longitudinal metabolite profiling in serum of normal versus over-conditioned cows and pathway analysis. *J. Dairy Sci.* **102**, 11561–11585 (2019).

27. Luo, Z. Z. *et al.* Plasma metabolite changes in dairy cows during parturition identified using untargeted metabolomics. *J. Dairy Sci.* **102**, 4639–4650 (2019).
28. Humer, E. *et al.* Supplementing phytogenic compounds or autolyzed yeast modulates ruminal biogenic amines and plasma metabolome in dry cows experiencing subacute ruminal acidosis. *J. Dairy Sci.* **101**, 9559–9574 (2018).
29. Zhang, G. *et al.* Metabotyping reveals distinct metabolic alterations in ketotic cows and identifies early predictive serum biomarkers for the risk of disease. *Metabolomics* **13**, 43 (2017).
30. Dervishi, E., Zhang, G., Mandal, R., Wishart, D. S. & Ametaj, B. N. 043 Metabolomics uncovers serum biomarkers that can predict the risk of retained placenta in transition dairy cows. *J. Anim. Sci.* **95**, 21–22 (2017).
31. Dervishi, E. *et al.* GC–MS metabolomics identifies metabolite alterations that precede subclinical mastitis in the blood of transition dairy cows. *J. Proteome Res.* **16**, 433–446 (2017).
32. Zhang, G. *et al.* Dairy cows affected by ketosis show alterations in innate immunity and lipid and carbohydrate metabolism during the dry off period and postpartum. *Res. Vet. Sci.* **107**, 246–256 (2016).
33. Xu, W. *et al.* Effects of dietary branched-chain amino acid supplementation on serum and milk metabolome profiles in dairy cows during early lactation. *J. Dairy Sci.* <https://doi.org/10.3168/jds.2022-21892> (2022).
34. Foroutan, A. *et al.* The bovine metabolome. *Metabolites* **10**, 233 (2020).
35. Houweling, M., van der Drift, S. G. A., Jorritsma, R. & Tielens, A. G. M. Technical note: Quantification of plasma 1- and 3-methylhistidine in dairy cows by high-performance liquid chromatography–tandem mass spectrometry. *J. Dairy Sci.* **95**, 3125–3130 (2012).
36. Imhasly, S. *et al.* Blood plasma lipidome profile of dairy cows during the transition period. *BMC Vet. Res.* **11**, 252 (2015).
37. Rico, J. E. *et al.* Characterization of the plasma lipidome in dairy cattle transitioning from gestation to lactation: Identifying novel biomarkers of metabolic impairment. *Metabolites* **11**, 290 (2021).
38. Putman, A. K., Brown, J. L., Gandy, J. C., Abuelo, A. & Sordillo, L. M. Oxy lipid profiles of dairy cattle vary throughout the transition into early mammary gland involution. *J. Dairy Sci.* **102**, 2481–2491 (2019).
39. Javadi, A. *et al.* Effects of acute intravenous lipopolysaccharide administration on the plasma lipidome and metabolome in lactating Holstein cows experiencing hyperlipidemia. *Metabolomics* **18**, 75 (2022).
40. Fan, Y. *et al.* Identification of milk fat metabolism-related pathways of the bovine mammary gland during mid and late lactation and functional verification of the ACSL4 gene. *Genes (Basel)* **11**, 1357 (2020).
41. Prah, M. C. *et al.* Hepatic urea, creatinine and uric acid metabolism in dairy cows with divergent milk urea concentrations. *Sci. Rep.* **12**, 17593 (2022).
42. van Soest, P. J. & Mason, V. C. The influence of the Maillard reaction upon the nutritive value of fibrous feeds. *Anim. Feed Sci. Technol.* **32**, 45–53 (1991).
43. Miyata, T. *et al.* Renal catabolism of advanced glycation end products: The fate of pentosidine. *Kidney Int.* **53**, 416–422 (1998).
44. Premi, M. *et al.* Changes of plasma analytes reflecting metabolic adaptation to the different stages of the lactation cycle in healthy multiparous Holstein dairy cows raised in high-welfare conditions. *Animals* **11**, 1714 (2021).
45. Bauer, J. H., Brooks, C. S. & Burch, R. N. Clinical appraisal of creatinine clearance as a measurement of glomerular filtration rate. *Am. J. Kidney Dis.* **2**, 337–346 (1982).
46. Valle, E. *et al.* Association of the glycoxidative stress marker pentosidine with equine laminitis. *Vet. J.* **196**, 445–450 (2013).
47. Besle, J. M. *et al.* Ultraviolet-absorbing compounds in milk are related to forage polyphenols. *J. Dairy Sci.* **93**, 2846–2856 (2010).
48. Fujihara, T. & Shem, M. N. Metabolism of microbial nitrogen in ruminants with special reference to nucleic acids. *Anim. Sci. J.* **82**, 198–208 (2011).
49. Stentoft, C. *et al.* Absorption and intermediary metabolism of purines and pyrimidines in lactating dairy cows. *Br. J. Nutr.* **113**, 560–573 (2015).
50. Odensten, M. O., Chilliard, Y. & Holtenius, K. Effects of two different feeding strategies during dry-off on metabolism in high-yielding dairy cows. *J. Dairy Sci.* **88**, 2072–2082 (2005).
51. Bionaz, M. *et al.* Plasma paraoxonase, health, inflammatory conditions, and liver function in transition dairy cows. *J. Dairy Sci.* **90**, 1740–1750 (2007).
52. Drackley, J. K., Overton, T. R. & Douglas, G. N. Adaptations of glucose and long-chain fatty acid metabolism in liver of dairy cows during the periparturient period. *J. Dairy Sci.* **84**, E100–E112 (2001).
53. Dong, X. *et al.* Aloe-emodin: A review of its pharmacology, toxicity, and pharmacokinetics. *Phyther. Res.* **34**, 270–281 (2020).
54. Fanali, S. *et al.* Analysis of Aloe-based phytotherapeutic products by using nano-LC-MS. *J. Sep. Sci.* **33**, 2663–2670 (2010).
55. Frolidi, G., Baronchelli, F., Marin, E. & Grison, M. Antiglycation activity and HT-29 cellular uptake of Aloe-emodin, aloin, and *Aloe arborescens* leaf extracts. *Molecules* **24**, 2128 (2019).
56. Rocchetti, G. & O'Callaghan, T. F. Application of metabolomics to assess milk quality and traceability. *Curr. Opin. Food Sci.* **40**, 168–178 (2021).
57. Crutchfield, C. A. & Clarke, W. Present and future applications of high resolution mass spectrometry in the clinic. *Discoveries* **2**, e17 (2014).
58. Park, M.-Y., Kwon, H.-J. & Sung, M.-K. Intestinal absorption of aloin, aloe-emodin, and aloesin; A comparative study using two in vitro absorption models. *Nutr. Res. Pract.* **3**, 9 (2009).
59. Gerspach, C. *et al.* Altered plasma lipidome profile of dairy cows with fatty liver disease. *Res. Vet. Sci.* **110**, 47–59 (2017).
60. Palombo, V. *et al.* Unique adaptations in neonatal hepatic transcriptome, nutrient signaling, and one-carbon metabolism in response to feeding ethyl cellulose rumen-protected methionine during late-gestation in Holstein cows. *BMC Genomics* **22**, 280 (2021).
61. Piepenbrink, M. S. & Overton, T. R. Liver metabolism and production of cows fed increasing amounts of rumen-protected choline during the periparturient period. *J. Dairy Sci.* **86**, 1722–1733 (2003).
62. Calabrò, S. *et al.* Effect of *Aloe arborescens* on in vitro rumen fermentations. *J. Nutr. Ecol. Food Res.* **1**, 117–123 (2013).
63. Vasta, V. *et al.* Invited review: Plant polyphenols and rumen microbiota responsible for fatty acid biohydrogenation, fiber digestion, and methane emission: Experimental evidence and methodological approaches. *J. Dairy Sci.* **102**, 3781–3804 (2019).
64. McFadden, J. W. *et al.* Symposium review: One-carbon metabolism and methyl donor nutrition in the dairy cow. *J. Dairy Sci.* <https://doi.org/10.3168/jds.2019-17319> (2020).
65. Bucktrout, R. E. *et al.* One-carbon, carnitine, and glutathione metabolism-related biomarkers in periparturient Holstein cows are altered by prepartal body condition. *J. Dairy Sci.* **104**, 3403–3417 (2021).
66. Cattaneo, L., Lopreato, V., Piccoli-Cappelli, F., Trevisi, E. & Minuti, A. Plasma albumin-to-globulin ratio before dry-off as a possible index of inflammatory status and performance in the subsequent lactation in dairy cows. *J. Dairy Sci.* **104**, 8228–8242 (2021).
67. Pang, Z. *et al.* Using MetaboAnalyst 5.0 for LC–HRMS spectra processing, multi-omics integration and covariate adjustment of global metabolomics data. *Nat. Protoc.* **17**, 1735–1761 (2022).

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Author contributions

F.P.C. and E.T. conceptualized the study and acquired the financial support; L.C., F.P.C., and A.M. executed the study; G.R. and S.Z. analyzed the samples; L.C., G.R., and S.Z. analyzed the data and prepared the data visualization; L.C. wrote the initial draft; G.R., A.M., and E.T. revised the draft. All authors read and approved the final manuscript.

Competing interests

The authors declare no competing interests.

Additional information

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