

1 **Effects of planting density on the growth and photosynthetic characteristics of *Alternanthera***
2 ***philoxeroides* under different nutrient conditions**

3

4 Ke Li¹, Hao Chen¹, Haijie Zhang², Miansong Huang³, Quan Quan⁴, Tong Wang⁵, Jian Liu^{1*}

5

6 ¹ Institute of Environmental Research, Shandong University, Jinan 250100, China

7 ² State Key Laboratory of Biocontrol, School of Life Sciences, Sun Yat-sen University, Guangzhou 5

8 10275, China

9 ³ Ningxia Capital Sponge City Construction & development Co., Ltd, Guyuan, 756000, China

10 ⁴ State Key Laboratory of Eco-hydraulics in Northwest Arid Region of China, Xi'an University

11 Technology, Xi'an, 710048, China

12 ⁵ Institute of Ecology and Biodiversity, School of Life Sciences, Shandong University, Jinan 250100,

13 China

14

15

16

17 *Corresponding author

18 E-mail: ecology@sdu.edu.cn (JL)

19

20

21

22

Abstract

Density and nutrient level are important factors that might affect the growth of invasive plants. To reveal the effects of plant density on the performance of invasive plant *Alternanthera philoxeroides* under different nutrient conditions, a greenhouse experiment was conducted in which *A. philoxeroides* was planted at three densities (low, medium and high) under three nutrient levels (low, medium and high). The results showed that both planting density and nutrient levels had significant effects on the growth of the plant. The biomass of individual plant and all plants in one pot under medium nutrient level were the highest while the photosynthetic rate and total chlorophyll content were the highest at the high nutrient level. Under different nutrient levels, the photosynthetic rate was the highest at medium planting density. The biomass of single plant decreased with the increase of population density, while the total biomass in the whole pot increased with the increase of density. These characteristics might contribute to the invasion of *A. philoxeroides* and help the plant to form monodominant community.

keywords: *Alternanthera philoxeroides*; Biomass; Nutrients; Population density; Photosynthesis.

Introduction

Biological invasion has far reaching impacts on ecological functions and the ecological diversity of native environments [1-5]. The invasive species have high fecundity, and once the invasion succeeds, it is easy to form the predominant population. The success of biological invasion is not only determined by the physiological and genetic characteristics of invasive species, but also by the factors such as population density and nutrient levels [6-10].

The nutrient is one of the important factors for plant growth. Previous studies have found that traits associated with the resource used determine the invasion success of several invasive species [11, 12]. Invasive plants generally show stronger photosynthesis and growth when invading habitats with richer nutrients [12-15]. Therefore, the nutrient level is also associated with the invasion success [9]. The research on the influence of nutrient on invasive plants plays an important role in understanding the successful plant invasion.

The competition among plants is also an important factor affecting the spatial distribution, dynamics and species diversity, promoting the succession of communities [16]. Invasive plants can replace local plants through interspecific competition, accomplishing the process of successful invasion [17]. However, when invasive plants spread to a certain extent, there is a degree of intraspecific competition. In order to adapt to this change, plants tend to change their own characteristics, such as plant height, biomass of branches and physiological characteristics of leaves, etc [18, 19]. And this phenotypic plasticity allows invaders to allocate more nutrients than their native counterparts to increase biomass [20, 21]. Several studies found that the physiological indexes of invasive plant under high density are less than those under low population density [8, 22]. Plant density determines the competitiveness of aquatic clonal plants

in complex habitats [23]. Therefore, we can analyze the growth mechanism by studying the morphological changes of plant. At the same time, the effect of population density on invasive plants is one of the core problems in the study of invasion ecology at present [24].

Aquatic species might have a fast response to nutrient enrichment, increasing their biomass rapidly, which is particularly true in the aquatic invasive species [9, 25-27]. Therefore, aquatic invasive species affects the productivity and management of land and water resources worldwide [28]. *A. philoxeroides* is one of these aquatic invasive plants. It is a clone weed that is native to South America and it is a stoloniferous and rhizomatous perennial herbaceous plant [28-30], which propagates clonally and expands rapidly in both aquatic and terrestrial habitats [31-34]. *A. philoxeroides* has experienced an invasion history of more than 80 years in China and become one of the most harmful invasive species [34]. At present, there are many researches focused on the physiological characteristics of *A. philoxeroides*, its response to natural environmental factors, and biological and chemical control [10, 35-37]. Many studies have shown that if the plant has a high photosynthetic rate, and it usually grows and propagate rapidly [38, 39]. Moreover, the invasive capacity of invasive plants is closely related to the ability of photosynthesis, they often enhance its invasive ability by increasing the photosynthesis of ramets [40, 41]. Thus, studying the photosynthetic capacity of *A. philoxeroides* is essential for understanding the potential of invasive species and to develop appropriate control strategies.

Our previous studies have compared the growth of *A. philoxeroides* with native plants [10]. So in the present study, we conducted a greenhouse experiment which combined nutrient levels with planting density to explore their effects on the growth and photosynthetic characteristic of *A. philoxeroides*. We asked the following questions: (1) Will the increase of planting density

inhibit the growth of *A. philoxeroides* under various nutrient levels? (2) How do the nutrient level and planting density affect the invasion of *A. philoxeroides*.

91

92 **Materials and methods**

93 **Ethics statement**

94 We collected plant material for our study with the official permission of the Environmental
95 Protection Bureau of Weishan County, and the Management Committee of the Weishan Lake
96 Constructed Wetland Park. We did not collect endangered or protected species.

97

98 **Plant material and experimental design**

99 The *A. philoxeroides* seedlings were collected from Weishan Lake Wetland Park, Shandong
100 province, in July 2017. The seedlings were cultured a week in the greenhouse of Fanggan
101 Research Station of Shandong University (36°26'N, 117°27'E). The method of sand culture was
102 used in the experiment, and the river sand was washed thoroughly before planting. The length of
103 the selected stem was about 11cm, and then was transplanted into a pot (h: 23.5cm; d: 22cm)
104 with 7 kg sand. Set up two factors in our experiment: the nutrient level of the sand and planting
105 density of *A. philoxeroides*. The nutrient treatments consisted of three levels (low, medium and
106 high; labeled as A, B and C) and three kinds of planting density (low, medium and high; 1, 2 and 4
107 seedlings per pot, respectively). In total, there were nine treatment combinations, each dealing
108 with five replicates. The nutrient levels and planting density settings in the experiment are
109 shown in Table 1.

110 **Table 1. Nutrient levels and planting density of *A. philoxeroides* at different treatment.**

Nutrient levels	TN	Amount of chemical	planting density	planting number
	(g kg ⁻¹ sand)	fertilizer added		(plant pot ⁻¹)
		(g kg ⁻¹ sand)		
A	0.5	2.5	1	1
			2	2
			4	4
B	0.7	3.5	1	1
			2	2
			4	4
C	2.1	10.5	1	1
			2	2
			4	4

111 *A, B, and C: low, medium and high nutrient levels.

112 In the experiment, the compound fertilizer with the ratio of nitrogen and phosphorus (N: P
113 = 4: 1) similar to Nansi Lake was used as the nutrient source. According to our previous
114 experiments, the medium nutrient gradient in this experiment is the most suitable for the
115 growth of *A. philoxeroides* [37]. Water 200 mL every day, in order to ensure the normal growth
116 of *A. philoxeroides*, and would not lead to the loss of fertilizers in the pot. At the time of
117 fertilization, grounding the fertilizer into powder, then half dissolved in 200 mL of water each
118 time, added for two days, to prevent once adding cause damage to plants. At the time of
119 experiment, the low nutrient level was added the fertilizer only in the first week; medium and
120 high nutrient levels were added every two weeks during the experiment. The time of the

experiment was from the July 24, 2017 to September 20, 2017.

Determination of photosynthetic rate and light response curve

The photosynthetic characteristics of each group were measured by a Portable Photosynthesis System (*LI-COR 6800*, USA) using PAR of 1000 $\mu\text{molm}^{-2}\text{s}^{-1}$. Leaves were measured under ambient CO_2 concentration [$385 \mu\text{molmol}^{-1}$] [42]. The light response curves were measured under the PAR of 1600, 1200, 1000, 800, 600, 400, 200, 100, and 0 $\mu\text{molm}^{-2}\text{s}^{-1}$ [42]. The fitting of the light response curve adopts a non - right - angle hyperbolic model, and the model formula is:

$$Pn = \frac{\alpha I + Pn_{\max} - \sqrt{(\alpha I + Pn_{\max})^2 - 4\theta\alpha I Pn_{\max}}}{2\theta} - Rd$$

In this formula, α is the apparent quantum rate; I is the light quantum flux density; $Pn_{\max}$ is the maximum photosynthetic rate; Rd is the dark respiration rate; θ is the angle parameter that reflects the degree of bending of the light response curve, and the range of values is $0 \leq \theta \leq 1$ [43]. According to the light response curve, we calculated the light compensation point (LCP) and the light saturation point (LSP).

Determination of chlorophyll content

In each treatment, using a volume fraction of 95% ethanol extract chlorophyll, then the spectrophotometer was used to measure the absorbance values at 649nm and 665nm wavelength [44]. The concentration of chlorophyll a, chlorophyll b and total chlorophyll were calculated according to the formula:

$$Ca = 13.95A_{665} - 6.88A_{649}$$

$$Cb = 24.96A_{649} - 7.32A_{665}$$

$$Cr = Ca + Cb = 18.08A_{649} - 6.63A_{665}$$

In the formula, Ca, Cb and Cr represent the concentrations of chlorophyll a, chlorophyll b and total chlorophyll respectively, [mg cm⁻²]; A₆₄₉ and A₆₆₅ represent the absorbance at the wavelengths of 649 nm and 665 nm respectively [45].

Determination of specific leaf area (SLA) of *A. philoxeroides*

Before harvest, randomly selected 20 mature leaves of each treatment, wiped clean and flattened on the scanner. The determination of leaf area using Photoshop software.

Leaf area = the percentage of leaf pixels / background pixels · background paper area [46-48].

SLA = leaf area / leaf dry weight.

Determination of morphological indexes of *A. philoxeroides*

We measured the length of stolon and recorded the number of internode before harvest.

The internode length of *A. philoxeroides* under different treatments was calculated.

Determination of biomass index of *A. philoxeroides*

At the end of the experiment, the roots of *A. philoxeroides* were washed thoroughly, and the leaves, stems and roots of each treatment were respectively put into the envelopes, numbered and then dried up to constant weight in an oven which the temperature was 80 °C, recorded the dry weight of each part.

Statistical analysis

The date of different variables, such as biomass, photosynthetic rate, SLA and total chlorophyll content was analyzed by two-way ANOVA with SPSS 21.0 software (Table 2). The significance test in all tests was performed at a level of $P < 0.05$. Use Origin 8.5 software to draw charts.

Results

Table 2. The two-way ANOVA of the effects of the independent variable nutrient level and plant density, and their combination on studied parameters of species *A. philoxeroides* in the experiment.

Indices	Source	df	F	p
Single leaf biomass	Density	2	56.564	<0.001
	Nutrient	2	114.140	<0.001
	Nutrient $\times$ Density	4	7.131	<0.001
Single biomass	Density	2	201.859	<0.001
	Nutrient	2	97.273	<0.001
	Nutrient $\times$ Density	4	17.055	<0.001
Total leaf biomass	Density	2	25.660	<0.001
	Nutrient	2	108.667	<0.001
	Nutrient $\times$ Density	4	5.568	0.001
Total biomass	Density	2	28.790	<0.001
	Nutrient	2	68.308	<0.001
	Nutrient $\times$ Density	4	3.485	0.017

Internode length	Density	2	77.514	<0.001
	Nutrient	2	4.017	0.027
	Nutrient $\times$ Density	4	2.251	0.083
SLA	Density	2	49.876	<0.001
	Nutrient	2	195.844	<0.001
	Nutrient $\times$ Density	4	1.751	0.160
Photosynthetic rate	Density	2	35.230	<0.001
	Nutrient	2	42.370	<0.001
	Nutrient $\times$ Density	4	3.346	0.020
Total chlorophyll content	Density	2	10.451	<0.001
	Nutrient	2	156.668	<0.001
	Nutrient $\times$ Density	4	0.502	0.735

Effects of density on Plant Biomass Index and Morphology Index under different nutrient gradients

Results showed that the interaction between nutrient level and plant density significantly affect the leaf biomass of per plant, single plant biomass, total leaf biomass and total biomass of a whole pot (Table 2).

Under three nutrient levels, the leaf biomass of single plant and the biomass of single plant all decreased significantly with the increase of planting density (Table 3). Among various nutrient levels, under three planting density, the reduction rate of leaf dry weight per plant was 47.9%, 53.9%, 62.5%; the reduction rate of biomass per plant was 57.2%, 64.4%, 63.4% (Table 3). At

different nutrient levels, the difference in the leaf biomass of per plant between low density (1) and medium density (2) was slightly greater than the difference between medium density (2) and high density (4) (Table 3). At every nutrient levels, there was a significant difference in the biomass of single plant among all three planting densities (Table 3). In low nutrient level (A), the difference between medium planting density (2) and high planting density (4) is slightly larger than that between low planting density (1) and medium planting density (2) (Table 3). Under medium nutrient (B) and high nutrient (C), the gap between low density (1) and medium planting density (2) is slightly larger than medium planting density (2) and high planting density (4) (Table 3).

At the three nutrient levels, the total leaf biomass and the total biomass of a pot all increased significantly with the increase of planting density (Table 3). To total leaf biomass of *A. philoxeroides*, there was no significant difference between low planting density (1) and medium planting density (2) under the treatment of low (A) and medium (B) nutrient levels and the difference among the three planting densities was not significant under the high nutrient level (C) treatment (Table 3). For total biomass of a pot, at low nutrient level (A), the difference between medium planting density (2) and high planting density (3) is significantly smaller than that between low planting density (1) and medium planting density (2), besides there is no significant difference between low planting density (1) and medium planting density (2) at medium (B) and high (C) nutrient levels (Table 3).

According to the two-way ANOVA analysis (Table 2), there were have obvious interaction between nutrient level and plant density on internode length of *A. philoxeroides*.

With the increase of planting density, the average internode length of *A. philoxeroides*

decreased in varying degrees, and the internode length was the longest at the treatment of low planting density (1) (Table 3). At the treatment of three nutrient levels, the average internode length of medium nutrient level (B) was the highest, is 5.6 cm (Table 3).

Table 3. Different nutrient levels and different planting densities of *A. philoxeroides* biomass and morphological index.

Treatments	Single leaf biomass (g)	Total leaf biomass (g)	Single biomass (g)	Total biomass (g)	Internode length (cm)
A1	2.90±0.56d	2.90±0.56e	22.83±3.35b	22.83±3.35d	5.66±0.28ab
A2	1.97±0.43e	3.94±0.88e	19.32±0.91c	38.64±1.83b	5.65±0.35b
A4	1.51±0.33e	5.80±1.30d	9.76±2.06de	39.03±8.25b	4.29±0.20d
B1	11.61±1.05a	11.61±1.05bc	40.52±3.03a	40.52±3.03b	6.60±0.61a
B2	7.21±2.13bc	14.42±4.25b	22.08±3.18b	44.16±6.36b	5.56±0.19c
B4	5.35±0.92c	21.38±3.66a	14.41±2.14cd	57.65±8.56a	4.65±0.50cd
C1	7.49±1.53b	7.49±1.53cd	21.38±2.60b	21.38±2.60d	6.05±0.40ab
C2	5.06±0.63c	10.11±1.26c	11.91±2.13d	23.83±4.26d	5.57±0.27c
C4	2.81±0.77de	11.25±3.07bc	7.83±0.91e	31.30±3.64c	4.55±0.06d

*A, B, and C: low, medium and high nutrient levels; 1, 2, and 4: the number of seedlings per pot.

Values are presented as means ±SD. Different letters indicate significant differences (p<0.05)

Effects of plant density on SLA of *A. philoxeroides* under different nutrient levels

According to the two-way ANOVA analysis (Table 2), there is no interaction between effects of nutrient level and plant density on SLA of *A. philoxeroides*.

The SLA of *A. philoxeroides* increased significantly with the increase of nutrient level (Fig 1). Different planting density had certain effects on the SLA of *A. philoxeroides*, which was the highest under medium planting density (2) and the lowest under high planting density (4), and both have significant differences (Fig 1).

Fig 1. The SLA of *A. philoxeroides* at different nutrient level and plant density. A, B, and C: low, medium and high nutrient levels; 1, 2, and 4: the seedlings per pot. Values are presented as means $\pm$ SD. Different letters indicate significant differences ($p < 0.05$).

Effects of density on photosynthetic rate and total chlorophyll content of *A. philoxeroides* under different nutrient levels.

We found that nutrient levels and planting densities on the photosynthetic rate of *A. philoxeroides* has a obvious interaction, and for the total chlorophyll content, they had no significant interaction to it (Table 2).

The analysis of photosynthetic rate and total chlorophyll content of *A. philoxeroides* under different treatments showed that the change trend of the two indexes are basically the same, and both of them increased with the increasing nutrient level (Fig 2). The photosynthetic rate of medium planting density (2) of the three nutrient levels were the highest and had significant differences, and they were the lowest under the treatment of high density (3) (Fig 2-A). The total chlorophyll content of the medium nutrient (B) and high nutrient (C) had obvious differences (Fig 2-B). The total chlorophyll content of *A. philoxeroides* reached the maximum under high nutrient

level (C), and for the planting density, when the planting density was medium (2), the total chlorophyll content of *A. philoxeroides* reached the maximum (Fig 2-B).

Fig 2. The photosynthetic rate and chlorophyll content of *A. philoxeroides* at different nutrient levels and plant densities. A, B, and C: low, medium and high nutrient levels; 1, 2, and 4: the number of seedlings per pot. Values are presented as means $\pm$ SD. Different letters indicate significant differences ($p < 0.05$).

Effects of density on light response curve of *A. philoxeroides* under different nutrient levels.

The data were analyzed and fitted by SPSS software. The fitting coefficients (R^2) of the light response curve of each group were all greater than 0.9, which showed that the curve fitting degree was better and the photosynthetic characteristic of *A. philoxeroides* could be more accurately reflected.

With the increase of the nutrient level, the maximum photosynthetic rate (Pn_{max}) of *A. philoxeroides* increased in different degrees, so the Pn_{max} at the high nutrient level (C) were the largest (Table 4). Among different planting density the Pn_{max} of *A. philoxeroides* were the largest at the treatments of medium planting density (2), and the Pn_{max} of *A. philoxeroides* was minimal under the high planting density (4) (Table 4). Besides, under the same nutrient level treatment, there were significant differences among the three different planting densities on the Pn_{max} of *A. philoxeroides* (Table 4).

The light compensation point (LCP) and light saturation point (LSP) of *A. philoxeroides* under different nutrient treatments were the biggest when planting density was medium (2), and when the planting density was high level (4), the value of LCP and LSP were the smallest (Table 4). At the same planting density, both LCP and LSP increased with increasing nutrient levels, among them, there was a larger gap between medium nutrient (B) and high nutrient (C) (Table 4). Under

the same nutrient level, at the difference of the value of LCP and LSP between medium planting density (2) and high planting density (4) was larger than that between medium planting density (2) and low planting density (1) (Table 4).

Table 4. Light response curve parameters of *A. philoxeroides* at different nutrient levels and planting densities.

Treatments	Pn_{max}	LCP	LSP
A1	$4.79 \pm 0.30g$	$51.54 \pm 4.60fg$	$250.21 \pm 14.90e$
A2	$6.08 \pm 0.28f$	$56.39 \pm 4.46f$	$283.08 \pm 9.79d$
A4	$3.77 \pm 0.25h$	$40.68 \pm 3.92h$	$205.30 \pm 16.82f$
B1	$7.55 \pm 0.34e$	$65.48 \pm 3.67e$	$307.15 \pm 8.56c$
B2	$8.34 \pm 0.50d$	$72.2 \pm 2.951d$	$341.17 \pm 13.12b$
B4	$6.25 \pm 0.18f$	$45.86 \pm 1.42g$	$238.56 \pm 4.46e$
C1	$11.31 \pm 0.43b$	$91.21 \pm 4.79b$	$377.10 \pm 30.20ab$
C2	$13.40 \pm 0.56a$	$117.73 \pm 2.47a$	$392.55 \pm 6.47a$
C4	$10.00 \pm 0.82c$	$84.09 \pm 4.33c$	$355.47 \pm 21.62b$

*A, B, and C: low, medium and high nutrient levels; 1, 2, and 4: the number of seedlings per pot. Values are presented as means $\pm$ SD. Different letters indicate significant differences ($p < 0.05$)

Discussion

In our experiment, nutrient and planting density had significant interaction effects on the biomass accumulation of *A. philoxeroides*. Under the same planting density, the biomass accumulation of *A. philoxeroides* could be promoted by increasing nutrients [49-51]. And our

previous studies concluded that compared with native plants, the *A. philoxeroides* had better environmental adaptability and higher biomass and ratio of leaf area under different nutrient conditions [10, 37]. At the treatment of medium nutrient level, the whole pot biomass of *A. philoxeroides* among three different planting densities was the highest, indicating that too high nutrient levels could be harmful for *A. philoxeroides* at the initial stage. At medium nutrient level, the increase of planting density had the most obvious effects on the single plant biomass of *A. philoxeroides* and the effects will be weakened at low nutrient. Therefore, we conclude that under appropriate nutrient level, the effects of planting density on the growth of *A. philoxeroides* were nutrient dependent. Some studies have shown that at same nutrient condition, the biomass of invasive plants increased more than that of native plants [52]. So in our experiments, under the same nutrient level, although the biomass of single plant was reduced with the increase of planting density, the total biomass of the plant in whole pot was increased, especially at low nutrient. That would enhance the invasive ability of *A. philoxeroides*.

In addition, according to the leaf biomass of *A. philoxeroides*, we analyzed the SLA of *A. philoxeroides*. In our study, with the increase of nutrient level, the SLA of *A. philoxeroides* increased, which is consistent with the studies on SLA of other plants [53-55]. Similarly, the increase of planting density has different influences on the SLA of *A. philoxeroides*. In this study, the SLA of *A. philoxeroides* was the highest under medium planting density treatment. SLA is one of the important plant leaf traits and closely related to plant growth and survival strategy. Its value can reflect the ability of plant leaves to intercept light and self-protection in bright light [56]. It is closely related to the photosynthesis and respiration of plants [57]. We therefore concluded that at the medium planting density, the leaf of *A. philoxeroides* had a higher net

photosynthetic rate. So we then analyzed the photosynthetic rate of rate of each treatments to confirm our conclusion.

In this study, photosynthetic rate had increase with the increasing nutrient levels that is consistent with previous studies [58-60]. Among the three kinds of plant density treatment, the photosynthetic rate in medium planting density was the highest, and the difference with high planting density was obvious. Chlorophyll is the main pigment of photosynthesis in plants, which reflects the size of photosynthesis in plants [44]. By this experiment, we found that the content of the total chlorophyll of *A. philoxeroides* had the same trend, indicating that under medium planting density, plants can capture resources better [61].

In order to better understand the effects of planting density on the photosynthetic characteristics of *A. philoxeroides* under different nutrient conditions, we studied the light response curve parameters of *A. philoxeroides*. Photosynthetic parameters, such as maximum photosynthetic rate (Pn_{max}), light compensation point (LCP) and light saturation point (LSP), are important scientific basis for rapid growth of plants [62-64]. Light saturation point (LSP) can reflected the adaptability of plants to strong light; the lower the light compensation point (LCP) is, the better the normal photosynthesis is under the weak light and the maximum net photosynthetic rate (Pn_{max}) reflects the utilization ability of *A. philoxeroides* to strong light under different treatments. In our study, among the three kinds of nutrient, the Pn_{max} , LCP and LSP of the *A. philoxeroides* seedlings in medium planting density were the largest, indicating that under this treatment, *A. philoxeroides* had the strongest utilization ability and adaptability to glare, and under high planting density, *A. philoxeroides* has better capability to utilize weak light. Moreover, the photosynthetic parameters of the plant increased with the increasing nutrient levels. That is

similar to previous studies [65, 66]. It shows that the increase of nutrient will enhance the utilization ability of *A. philoxeroides* to strong light. This suggests that at higher nutrient levels, higher light intensities are required to produce more biomass, may be that is why the *A. philoxeroides* has higher photosynthetic rate but the biomass is lower than the medium nutrient level. Under the same nutrient level, the increase of planting density resulted in the decrease of Pn_{max} , LCP and LSP and there were significant differences among the three planting densities. The results suggested that the increase of planting density decreased the Pn_{max} of *A. philoxeroides* and its ability to use strong light and its adaptability. Under the low nutrient level, there was no obvious difference in the LCP between the medium and low planting density, which indicated that under the low nutrient level, the high planting density *A. philoxeroides* had more obvious photosynthetic ability at low light. But at the medium and high nutrient levels, the planting density had more obvious influence on the LCP of *A. philoxeroides*, and the effects became more obvious with the increase of nutrient level.

In conclusion, our study showed that at the three nutrient levels, the SLA, photosynthetic rate and total chlorophyll content of *A. philoxeroides* at medium planting density were the highest. What's more, although the biomass of single plant, SLA, photosynthetic rate and the content of Chlorophyll reduced with the increase of planting density, the biomass of whole pot tended to increase. These attributes may increase the competitive dominance of *A. philoxeroides* and could help the *A. philoxeroides* population develop into a monodominant community.

References

1. Pyšek P, Richardson DM. Invasive species, environmental change and management, and health. Annual Review of Environment & Resources. 2010;35(1).

- 341 2. Vilà M, Weiner J. Are invasive plant species better competitors than native plant species?
342 – evidence from pair - wise experiments. *Oikos*. 2004;105(2):229-38.
- 343 3. Vicente JR, Pereira HM, Randin CF, Gonçalves J, Lomba A, Alves P, et al. Environment and
344 dispersal paths override life strategies and residence time in determining regional patterns of invasion
345 by alien plants. *Perspectives in Plant Ecology Evolution & Systematics*. 2014;16(1):1-10.
- 346 4. Oduor AMO, Leimu R, Kleunen M. Invasive plant species are locally adapted just as frequently
347 and at least as strongly as native plant species. *Journal of Ecology*. 2016;104(4):957-68.
- 348 5. Seebens H, Blackburn TM, Dyer EE, Genovesi P, Hulme PE, Jeschke JM, et al. No saturation in the
349 accumulation of alien species worldwide. *Nature Communications*. 2017;8:14435.
- 350 6. Crawley MJ. *The Structure of Plant Communities*: Blackwell Publishing Ltd.; 2003. 475-531 p.
- 351 7. Forsman A. Effects of genotypic and phenotypic variation on establishment are important for
352 conservation, invasion, and infection biology. *Proceedings of the National Academy of Sciences of the*
353 *United States of America*. 2013;111(1):302.
- 354 8. Li SL, Vasemägi A, Ramula S. Genetic variation facilitates seedling establishment but not
355 population growth rate of a perennial invader. *Annals of Botany*. 2016;117(1).
- 356 9. Sardans J, Bartrons M, Margalef O, Gargallo-Garriga A, Janssens IA, Ciais P, et al. Plant invasion is
357 associated with higher plant-soil nutrient concentrations in nutrient-poor environments. *Glob Chang*
358 *Biol*. 2017;23(3):1282-91.
- 359 10. Zhang H, Chang R, Guo X, Liang X, Wang R, Liu J. Shifts in growth and competitive dominance of
360 the invasive plant *Alternanthera philoxeroides* under different nitrogen and phosphorus supply.
361 *Environmental & Experimental Botany*. 2016;135:118-25.
- 362 11. Daehler CC. Performance Comparisons of Co-Occurring Native and Alien Invasive Plants:
363 Implications for Conservation and Restoration. *Annual Review of Ecology Evolution & Systematics*.
364 2003;34(1):183-211.
- 365 12. González AL, Kominoski JS, Danger M, Ishida S, Iwai N, Rubach A. Can ecological stoichiometry
366 help explain patterns of biological invasions? *Oikos*. 2010;119(5):779–90.
- 367 13. Davis MA, Grime JP, Thompson K. Fluctuating Resources in Plant Communities: A General Theory
368 of Invasibility. *Journal of Ecology*. 2000;88(3):528-34.
- 369 14. Eva S, Christoph K, Peterj E, Hansjörg D. Influence of light and nutrient conditions on seedling
370 growth of native and invasive trees in the Seychelles. *Biological Invasions*. 2009;11(8):1941-54.
- 371 15. Quan G, Mao D, Zhang J, Xie J. Effects of nutrient level on plant growth and biomass allocation of
372 invasive *Chromolaena odorata*. *Ecological Science*. 2015.
- 373 16. Xiang X, Ganlin WU, Duan R, Yan Y, Zhang X. Intraspecific and interspecific competition of *Pinus*
374 *dabeshanensis*. *Acta Ecologica Sinica*. 2015;35(2).
- 375 17. Chittka L, Schürkens S. Successful invasion of a floral market. *Nature*. 2001;411(6838):653.
- 376 18. Schooler S, Baron Z, Julien M. Effect of simulated and actual herbivory on alligator weed,
377 *Alternanthera philoxeroides*, growth and reproduction. *Biological Control*. 2006;36(1):74-9.
- 378 19. Liu LM, Song, H., Liu, H. The effect of water, light and density on the growth of the plant.
379 *Environmental Protection Science*. 2014;40(4):29-35.
- 380 20. Drenovsky RE, Martin CE, Falasco MR, James JJ. Variation in resource acquisition and utilization
381 traits between native and invasive perennial forbs. *American Journal of Botany*. 2008;95(6):681-7.
- 382 21. Funk JL. Differences in Plasticity between Invasive and Native Plants from a Low Resource
383 Environment. *Journal of Ecology*. 2008;96(6):1162-73.
- 384 22. S.Y. Q, Chang EZ, Dong JJ, Guo TT. Competitive effect between invasive plant *Galinsoga parviflora*

385 and *Trifolium repens*. Guangdong Agricultural Sciences. 2014;41(1):141-5.

386 23. Cipollini DF, Bergelson J. Plant density and nutrient availability constrain constitutive and
387 wound-induced expression of trypsin inhibitors in *Brassica napus*. Journal of Chemical Ecology.
388 2001;27(3):593-610.

389 24. Fraver S, D'Amato AW, Bradford JB, Jonsson BG, Jönsson M, Esseen PA. Tree growth and
390 competition in an old - growth *Picea abies* forest of boreal Sweden: influence of tree spatial
391 patterning. Journal of Vegetation Science. 2014;25(2):374-85.

392 25. Butzler JM. THE ROLE OF NUTRIENT VARIABILITY IN AQUATIC ECOSYSTEMS. 2002.

393 26. Smith SDP. The roles of nitrogen and phosphorus in regulating the dominance of floating and
394 submerged aquatic plants in a field mesocosm experiment. Aquatic Botany. 2014;112:1-9.

395 27. Zhao H, Yang W, Xia L, Qiao Y, Xiao Y, Cheng X, et al. Nitrogen-Enriched Eutrophication Promotes
396 the Invasion of *Spartina alterniflora* in Coastal China. CLEAN - Soil, Air, Water. 2015;43(2):244-50.

397 28. Wang B, Li W, Wang J. Genetic diversity of *Alternanthera philoxeroides* in China. Aquatic Botany.
398 2005;81(3):277-83.

399 29. Li J, Ye WH. Genetic diversity of alligator weed ecotypes is not the reason for their different
400 responses to biological control. Aquatic Botany. 2006;85(2):155-8.

401 30. Lian Jin G, Tao W. Impact of invasion of exotic plant *Alternanthera philoxeroides* on interspecies
402 association and stability of native plant community. Chinese Journal of Eco-Agriculture.
403 2009;17(5):851-6.

404 31. Kolar, Cynthia S, Lodge, David M. Progress in invasion biology: predicting invaders. Trends in
405 Ecology & Evolution. 2001;16(4):199-204.

406 32. Zhang B, Jin, Y.G., Huai, H.Y., Shi, H.Y. Anatomical structure of hollow lotus leaf blade in two
407 habitats. Journal of weeds. 2001(4):6-7.

408 33. Liu J, Dong M, Miao SL, Li ZY, Song MH, Wang RQ. Invasive alien plants in China: role of clonality
409 and geographical origin. Biological Invasions. 2006;8(7):1461-70.

410 34. Pan X, Geng Y, Sosa A, Zhang W, Li B, Chen J. Invasive *Alternanthera philoxeroides*: biology,
411 ecology and management. Acta Phytotaxonomica Sinica. 2007;45(6):884-900.

412 35. Weng BQ, Lin S, Wang YX. Discussion on adaptability and invasion mechanisms of *Alternanthera*
413 *philoxeroides* in China. Acta Ecologica Sinica. 2006;26(7):2373-81.

414 36. Cao YS, Xiao YA, Zhou B, Wen-Jie HU. A THE PHENOTYPIC PLASTICITY OF *ALTERNANTHERA*
415 *PHILOXEROIDES* TO DIFFERENT WATER HABITATS. Journal of Jiangangshan University. 2012.

416 37. Chang RY, Wang RQ, Zhang YR, Liu J. Effects of N:P Ratio and Nutrient Level on the Competition
417 between Invasive *Alternanthera philoxeroides* and Native *Oenanthe javanica*. Advanced Materials
418 Research. 2012;534:337-42.

419 38. McDowell SCL. Photosynthetic Characteristics of Invasive and Noninvasive Species of *Rubus*
420 (Rosaceae). American Journal of Botany. 2002;89(9):1431-8.

421 39. Penuelas J, Sardans J, Llusà J, Owen SM, Carnicer J, Giambelluca TW, et al. Faster returns on 'leaf
422 economics' and different biogeochemical niche in invasive compared with native plant species. Global
423 Change Biology. 2010;16(8):2171-85.

424 40. Pearcy RW, Tumosa N, Williams K. Relationships between growth, photosynthesis and
425 competitive interactions for a C 3 and C 4 plant. Oecologia. 1981;48(3):371.

426 41. Liu J, He WM, Zhang SM, Liu FH, Dong M, Wang RQ. Effects of clonal integration on
427 photosynthesis of the invasive clonal plant *Alternanthera philoxeroides*. Photosynthetica.
428 2008;46(2):299.

- 429 42. Zhu JW. Physiological and biochemical characteristics of hollow lotus seed grass under high
430 manganese stress and the study of glyphosate tolerance: Zhejiang University; 2008.
- 431 43. Liang WB, Nie DL, Si-Zheng WU, Bai WF, Shen SZ. Photosynthetic light response curves of
432 *Macropanax rosthornii* and their model fitting. Nonwood Forest Research. 2014.
- 433 44. Chen QZ, Tang N, Zhang BJ, Wang LK, Yang P. Chromium-induced Photosynthetic Physiological
434 Parameters in *Alternanthera philoxeroides*. Hubei Agricultural Sciences. 2015.
- 435 45. Guo C, Wei X, Yun-Feng LI, Zhao FM. Physiological Characteristics of Osmotic Adjustment and
436 Content of Chlorophyll of H8 and H10 Carrying DNA Segments of *Alternanthera philoxeroides*.
437 Southwest China Journal of Agricultural Sciences. 2014;27(2):573-7.
- 438 46. Xiao Q, Ye, W.J., Zhu, Z., Chen, Y., Zheng, H.L. A simple method for measuring leaf area using
439 digital camera and Photoshop software. Chinese Journal of Ecology. 2005;24(6):711-4.
- 440 47. Chen WX, Huang, J.J. Comparative study on the method of measuring leaf area of two kinds of
441 plants. Jilin agricultural. 2010(10):50-1.
- 442 48. Cui SG, Qin, J.H. Image processing method for the determination of leaf area of rape. Hubei
443 Agricultural Sciences. 2017;56(14):2756-7.
- 444 49. Maron JL, Connors PG. A native nitrogen-fixing shrub facilitates weed invasion. *Oecologia*.
445 1996;105(3):302-12.
- 446 50. Burns JH. A comparison of invasive and non - invasive dayflowers (Commelinaceae) across
447 experimental nutrient and water gradients. *Diversity & Distributions*. 2004;10(5 - 6):387-97.
- 448 51. Maestre FT, Reynolds JF. Amount or Pattern? Grassland Responses to the Heterogeneity and
449 Availability of Two Key Resources. *Ecology*. 2007;88(2):501-11.
- 450 52. Lapointe BE, Bedford BJ. Stormwater nutrient inputs favor growth of non-native macroalgae
451 (Rhodophyta) on O'ahu, Hawaiian Islands. *Harmful Algae*. 2011;10(3):310-8.
- 452 53. Arendonk JJCMV, Niemann GJ, Boon JJ, Lambers H. Effects of nitrogen supply on the anatomy
453 and chemical composition of leaves of four grass species belonging to the genus *Poa*, as determined
454 by image - processing analysis and pyrolysis - mass spectrometry. *Plant Cell & Environment*.
455 1997;20(7):881-97.
- 456 54. Meziane D, Shipley B. Interacting determinants of specific leaf area in 22 herbaceous species:
457 effects of irradiance and nutrient availability. *Plant Cell & Environment*. 1999;22(5):447-59.
- 458 55. Niinemets UKK. Leaf structure vs. nutrient relationships vary with soil conditions in temperate
459 shrubs and trees. *Acta Oecologica*. 2003;24(4):209-19.
- 460 56. Zhang L, Luo T. Advances in ecological studies on leaf lifespan and associated leaf traits. *Acta*
461 *Phytoecologica Sinica*. 2004;28(6):844-52.
- 462 57. BenomarLahcen, DesRochersAnnie, Larocqueguy R. Changes in specific leaf area and
463 photosynthetic nitrogen-use efficiency associated with physiological acclimation of two hybrid poplar
464 clones to intraclonal competition. *Canadian Journal of Forest Research*. 2011;41(7):1465-76.
- 465 58. Van KM, Weber E, Fischer M. A meta-analysis of trait differences between invasive and
466 non-invasive plant species. *Ecology Letters*. 2010;13(2):235-45.
- 467 59. Feng YL, Auge H, Ebeling SK. Invasive *Buddleja davidii* Allocates More Nitrogen to Its
468 Photosynthetic Machinery than Five Native Woody Species. *Oecologia*. 2007;153(3):501-10.
- 469 60. Zhang W, Xiao H, Yin Z, Zeng X, Huang M, Feng Y, et al. Effects of simulated nitrogen deposition
470 on photosynthetic characteristics of the invasive plant *Mikania micrantha*. *Ecology & Environmental*
471 *Sciences*. 2013;22(12):1859-66.
- 472 61. Han LH, Liu, C., Wang, J.J. Effect of nitrogen fertilizer on the content of pigment in purple stem of

473 different population. Hubei Agricultural Sciences. 2012;51(3):475-7.
 474 62. Sharp RE, Matthews MA, Boyer JS. Kok Effect and the Quantum Yield of Photosynthesis: Light
 475 Partially Inhibits Dark Respiration. Plant Physiology. 1984;75(1):95-101.
 476 63. Awada T, Radoglou K, Fotelli MN, Constantinidou HI. Ecophysiology of seedlings of three
 477 Mediterranean pine species in contrasting light regimes. Tree Physiology. 2003;23(1):33-41.
 478 64. Hui Z. Photosynthetic characteristics comparison between an invasive plant, *Lantana camara*
 479 L., and associated species. Acta Ecologica Sinica. 2009;29(5):2701-9.
 480 65. Jiao JY, Yin CY, Chen K. Effects of soil water and nitrogen supply on the photosynthetic
 481 characteristics of *Jatropha curcas* seedlings. Chinese Journal of Plant Ecology. 2011;35(1):91-9.
 482 66. Wang S, Han, X.R., Zhan, X.M., Yang, J.F., Liu, Y.F., Wang, Y., Li, N. The comparative study on
 483 fitting light response curve model of photosynthesis of maize under different nitrogen fertilizer levels.
 484 Journal of Plant Nutrition & Fertilizer. 2014;20(6):1403-12.
 485



