



OPEN ACCESS

EDITED BY

Diaa Abd El Moneim,
Arish University, Egypt

REVIEWED BY

Kalaichelvi Kalaignan,
Tamil Nadu Agricultural University, India
Manish Singh,
Govind Ballabh Pant National Institute of
Himalayan Environment and Sustainable
Development, India

*CORRESPONDENCE

Seham M. Al Raish
✉ seham.alraish@uaeu.ac.ae

RECEIVED 21 December 2025

REVISED 11 April 2026

ACCEPTED 20 April 2026

PUBLISHED 15 May 2026

CITATION

Abu-Elsaoud AM, Aldawsari EO,
Alotaibi RN, Aldayel Aa, Alarefi ANA,
Aldossari RMS and Al Raish SM (2026)
Comparative study on the effect of
elevated levels of carbon dioxide on
growth and photosynthesis of three
selected plant species.
Front. Plant Sci. 17:1772837.
doi: 10.3389/fpls.2026.1772837

COPYRIGHT

© 2026 Abu-Elsaoud, Aldawsari, Alotaibi,
Aldayel, Alarefi, Aldossari and Al Raish. This
is an open-access article distributed under
the terms of the [Creative Commons
Attribution License \(CC BY\)](#). The use,
distribution or reproduction in other
forums is permitted, provided the
original author(s) and the copyright
owner(s) are credited and that the
original publication in this journal is
cited, in accordance with accepted
academic practice. No use, distribution
or reproduction is permitted which does
not comply with these terms.

Comparative study on the effect of elevated levels of carbon dioxide on growth and photosynthesis of three selected plant species

Abdelghafar M. Abu-Elsaoud¹, Eman Obaid Aldawsari¹,
Reem Nigr Alotaibi¹, Abeer abdulrahman Aldayel¹,
Alya Nasser Abdulrahman Alarefi¹,
Renad Mohammed Salem Aldossari¹ and Seham M. Al Raish^{2*}

¹Department of Biology, College of Science, Imam Mohammad Ibn Saud Islamic University (IMSIU), Riyadh, Saudi Arabia, ²Country Department of Biology, College of Science, United Arab Emirates University, Al Ain, United Arab Emirates

The aim of this study was to evaluate the growth and physiological responses of two C3 species (*Capsicum annuum* and *Mentha × piperita*) and one C4 species (*Gomphrena globosa*) under elevated CO₂ conditions. Plants were exposed to three CO₂ concentrations—~400 parts per million (ppm) (control), ~2,900 ppm, and ~5,400 ppm—in a controlled greenhouse experiment. Elevated CO₂ significantly enhanced plant height, leaf number, bud formation, shoot and root fresh weight, total biomass, chlorophyll content, and relative leaf nitrogen index across all species. The magnitude of response differed among species, with *C. annuum* showing the strongest overall growth stimulation at 5,400 ppm. Multivariate analyses further confirmed that CO₂ concentration significantly influenced physiological parameters. These findings demonstrate species-specific responses of C3 and C4 plants to high CO₂ enrichment and highlight potential implications for crop productivity under changing atmospheric conditions.

KEYWORDS

Elevated CO₂, growth, photosynthesis, C3, C4, *Capsicum annuum*, *Mentha × piperita*, *Gomphrena globosa*

1 Introduction

Atmospheric carbon dioxide (CO₂) concentrations have increased substantially due to anthropogenic activities and are projected to continue to rise throughout this century, with significant implications for global climate and agricultural productivity (Sage, 1994; Cassia et al., 2018; Shishegaran et al., 2020; Wang et al., 2022; Sendall et al., 2024; Jiang et al., 2025). Because CO₂ is the primary substrate for photosynthesis, rising atmospheric CO₂ directly influences plant carbon assimilation, biomass production, and ecosystem carbon cycling (Sage, 1994; Keenan et al., 2022). In addition to their ecological and agronomic importance, plant species, particularly those with medicinal properties, are critical components of global healthcare systems, with growing evidence of widespread reliance on plant-based remedies

and strong associations among knowledge, perceptions, and patterns of use (Almasri et al., 2025; Bedir et al., 2025; Al Raish et al., 2025, 2026; Al Raish, 2026). Global photosynthesis represents the largest CO₂ flux in the carbon cycle, and even modest changes in photosynthetic performance can substantially alter carbon balance and crop productivity (Keenan et al., 2022; Baslam and Sanz-Saez, 2023). Although elevated CO₂ generally enhances net photosynthesis in C3 species, long-term exposure often induces acclimation responses characterized by reductions in maximum carboxylation capacity (V_{cmax}), ribulose-1,5-bisphosphate carboxylase/oxygenase (RuBisCO) content, and leaf nitrogen concentration (Ainsworth and Long, 2021; Ancin et al., 2024). These acclimatory adjustments complicate predictions of future crop performance under elevated CO₂ scenarios.

Plants utilize two primary photosynthetic pathways, C3 and C4, which differ fundamentally in their carbon fixation mechanisms and responses to atmospheric CO₂ enrichment (Raines, 2011; Cheng and He, 2019; Verhage, 2021; Chen et al., 2023; Mukundan et al., 2024). In C3 species, CO₂ is directly fixed by RuBisCO, a process accompanied by photorespiration that reduces carbon-use efficiency under ambient CO₂ (Raines, 2011; Mukundan et al., 2024). Elevated CO₂ suppresses photorespiration in C3 plants, typically enhancing photosynthetic carbon gain and biomass accumulation (Bravdo and Canvin, 1979; Zheng et al., 2019; Busch, 2020). In contrast, C4 plants possess a CO₂-concentrating mechanism that minimizes photorespiration under current atmospheric conditions, resulting in comparatively smaller stimulation under elevated CO₂ (Verhage, 2021; Mukundan et al., 2024). Large-scale syntheses and recent experimental studies confirm that C3 crops generally exhibit stronger growth and photosynthetic stimulation than C4 species under elevated CO₂, although responses vary across environmental conditions and developmental stages (Reich et al., 2018; Kaur et al., 2025).

Beyond steady-state photosynthesis, elevated CO₂ also alters stomatal conductance, biochemical capacity, and the balance between diffusional and biochemical limitations (Ancin et al., 2024; Tang et al., 2024). Recent work demonstrates that elevated CO₂ can shift the dominant limitation of photosynthesis from stomatal to biochemical constraints during induction, depending on species-specific stomatal morphology and resource allocation patterns (Tang et al., 2024; Zhang et al., 2024). Moreover, interactions between elevated CO₂ and environmental factors such as temperature, drought, and nutrient availability further influence RuBisCO regulation, electron transport capacity, and nitrogen dynamics (Wang et al., 2022; Ancin et al., 2024). These findings underscore the importance of evaluating species-specific physiological plasticity under controlled CO₂ enrichment conditions.

Although numerous studies have examined the effects of elevated CO₂ on individual crop species, direct comparative evaluations of C3 and C4 plants under identical controlled high-CO₂ exposure remain limited. Such comparisons are critical for understanding pathway-dependent physiological responses, nitrogen-related adjustments, and biomass allocation patterns that may influence future crop selection and adaptation strategies.

In this study, two C3 species (*Capsicum annuum* and *Mentha × piperita*) and one C4 species (*Gomphrena globosa*) were selected to investigate differential physiological responses under controlled CO₂

enrichment. These species were chosen based on both physiological and practical considerations: *C. annuum* and *M. piperita* are economically important C3 crops, while *G. globosa* represents a C4 photosynthetic pathway. This combination enables direct comparison of growth, chlorophyll dynamics, and relative nitrogen status between photosynthetic types under strong CO₂ enrichment conditions.

The selected plant species were chosen based on both physiological and practical considerations. *C. annuum* (pepper) and *Mentha × piperita* (peppermint) are economically important C3 crops widely cultivated for food and medicinal uses. *G. globosa*, a representative C4 species, was included to allow direct comparison between C3 and C4 photosynthetic pathways under identical CO₂ enrichment conditions. This combination enables evaluation of pathway-dependent growth and physiological responses relevant to agricultural adaptation under rising atmospheric CO₂.

The objectives of this study were to:

1. evaluate the growth responses of two C3 species (*C. annuum* and *Mentha × piperita*) and one C4 species (*G. globosa*) under elevated CO₂ concentrations [400, 2,900, and 5,400 parts per million (ppm)];
2. assess changes in chlorophyll content, relative leaf nitrogen index, and biomass allocation under high CO₂ exposure; and
3. compare species-specific physiological responses between C3 and C4 photosynthetic pathways under controlled CO₂ enrichment conditions.

The selection of plant species in this study was deliberate to enable a comparative evaluation of different photosynthetic pathways under elevated CO₂ conditions. *C. annuum* and *Mentha × piperita* are C3 species, while *G. globosa* follows the C4 photosynthetic pathway. Since C3 plants generally exhibit stronger stimulation of photosynthesis under elevated CO₂ due to reduced photorespiration, whereas C4 plants already possess a CO₂-concentrating mechanism, comparing these species provides insight into pathway-specific growth and physiological responses under high CO₂ environments.

Although the stimulatory effects of elevated CO₂ on plant growth have been widely reported, comparative studies integrating C3 and C4 species under identical experimental conditions and high CO₂ exposure remain limited. The novelty of the present study lies in the simultaneous evaluation of two C3 crops and one C4 species using a unified experimental system, combined with comprehensive physiological, biochemical, and multivariate analyses. This approach allows a clearer interpretation of pathway-specific responses to elevated CO₂ enrichment.

2 Materials and methods

2.1 Experimental design

This study was conducted to evaluate the effects of elevated CO₂ concentrations on the growth and photosynthesis of three selected

plant species: *G. globosa*, *C. annuum*, and *M. piperita*. Each species was subjected to three different CO₂ treatment levels over a specified period to assess the influence of elevated CO₂ on various growth and physiological parameters (Figure 1).

Plants were grown in 7-cm-diameter pots filled with a uniform mixture of sterilized soil and peat moss (3:1 ratio). The soil had a pH of 6.5 and was supplemented with a balanced slow-release fertilizer (NPK 15-15-15) to ensure adequate nutrient supply throughout the experiment. Watering was conducted uniformly with tap water as needed to maintain optimal soil moisture levels across all experimental units. The experiment was carried out in a naturally lit greenhouse with a controlled temperature of 25 °C (± 2 °C), and plants were exposed to a 12-h photo period of natural sunlight filtered through transparent containers. A total of 45 plants of each species were divided into three treatment groups based on the CO₂ concentration they were exposed to:

Group I (Control): Plants were exposed to ambient CO₂ levels (~400 ppm), which were continuously monitored and maintained under natural greenhouse conditions throughout the experiment.

Group II: ~2,900 ppm.

Group III: ~5,400 ppm.

Each treatment group contained five replicates per species ($n = 5$ per species per treatment), totaling 15 plants per treatment and 45 plants per species. The methodology was implemented following the procedures described by Shenglei and Ferris (2006) and Akhlaq et al. (2023).

CO₂ enrichment was achieved using dry ice (solid CO₂) to simulate elevated atmospheric CO₂ levels. Transparent plastic containers were used to enclose each treatment group, ensuring controlled CO₂ concentrations while allowing sufficient light penetration. Dry ice was placed in small, perforated containers within each sealed chamber to gradually release CO₂ through sublimation. The amount of dry ice required for each treatment was calculated based on the container volume and the desired CO₂ concentration. For Groups II and III, dry ice quantities were calculated to achieve the target final chamber concentrations of approximately 2,900 and 5,400 ppm, respectively.

CO₂ levels in all treatment groups, including the ambient control, were continuously monitored using a Multifunction ELIKLIV Portable Air Quality Detector (LORD Electrical, Shenzhen Yong An Cun Technology Co., Ltd, China) to ensure stability. To regulate CO₂ release and prevent oxygen depletion, small vents were incorporated into the containers to allow excess gas to escape without significant CO₂ loss. Dry ice was replenished every 24 h to maintain consistent CO₂ concentrations, as the sublimation rate varied with temperature and container size.

All CO₂ values reported in this study refer exclusively to the final monitored concentrations inside the experimental chambers.

Although the applied CO₂ concentrations exceed projected near-future atmospheric levels, the purpose of this study was to examine short-term physiological and growth responses under strong CO₂ enrichment conditions in a controlled greenhouse system. Such enrichment levels are commonly used in experimental

plant physiology to amplify metabolic responses and allow clearer differentiation between C3 and C4 species.

2.2 Growth parameters

Growth and physiological parameters were measured at 3, 9, and 14 days after treatment (DAT) intervals throughout the experiment. The parameters assessed included the following:

1. Plant height (cm)
2. Number of leaves
3. Number of buds
4. Total leaf chlorophyll content (estimated using an AtLeaf device, FT Green LLC)
5. Relative leaf nitrogen index (estimated using the AtLeaf chlorophyll meter)
6. Root length (cm)
7. Shoot and root fresh weight (FW)

The AtLeaf device provided a non-destructive estimate of relative chlorophyll content for continuous monitoring throughout the study (Supplementary Figures 1-4).

Leaf nitrogen status was estimated using the AtLeaf chlorophyll meter (FT Green LLC, USA). The device provides relative index readings based on leaf chlorophyll measurements that are correlated with nitrogen status. The values reported in this study represent AtLeaf index units rather than direct chemical nitrogen concentration.

The present study focused exclusively on physiological and growth responses to elevated CO₂ and did not include molecular analyses such as gene expression or enzymatic activity assays.

2.3 Statistical analysis

All collected data, including plant height, number of leaves, number of buds, chlorophyll content, relative leaf nitrogen index, root length, and shoot and root FW, were recorded in a spreadsheet for statistical analysis. Data normality was verified using the Shapiro–Wilk and Kolmogorov–Smirnov tests. Since the data followed a parametric distribution, results were presented as mean $\pm$ standard error of the mean (SEM).

A one-way analysis of variance (ANOVA) was performed to determine the effects of CO₂ concentration on each parameter for all plant species. A Tukey's HSD *post hoc* test was conducted to compare group differences. Statistical significance was determined at $p < 0.05$, $p < 0.01$, and $p < 0.001$. All analyses used IBM-SPSS software (Version 30.0 for Mac OS).

3 Results

Elevated CO₂ significantly improved several physiological and biochemical traits, including plant height, leaf number, bud formation, shoot and root fresh weight, total biomass, chlorophyll content, relative leaf nitrogen index, and biomass allocation (shoot/root ratio).

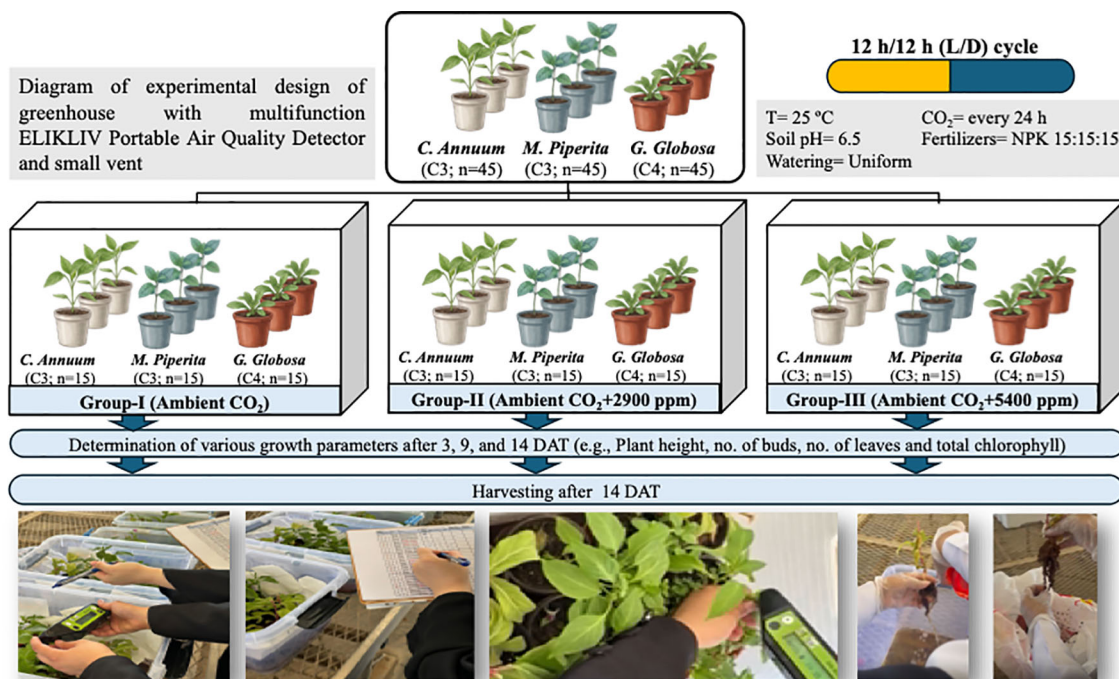


FIGURE 1

Experimental greenhouse setup and CO₂ treatment design. The controlled environment chamber was equipped with a multifunction portable air quality detector (ELIKLIV) and a ventilation system to maintain target CO₂ concentrations. A total of 45 plants per species (*Gomphrena globosa*, *Capsicum annuum*, and *Mentha × piperita*) were distributed equally across three CO₂ treatment groups: Group I — ambient control (~400 ppm), Group II — elevated CO₂ (~2,900 ppm), and Group III — highly elevated CO₂ (~5,400 ppm). Representative photographs of the experimental conditions and plant materials are shown.

3.1 Plant height

The plant height of *M. piperita* in Group I measured at 3, 9, and 14 DAT exhibited an average $\pm$ standard deviation (SD) of 8.7 ± 0.6 , 11.0 ± 1.0 , and 11.7 ± 1.5 cm, respectively. In Group II, the corresponding values were 14.2 ± 2.3 , 16.0 ± 1.0 , and 19.7 ± 0.6 cm, while in Group III, plant height increased to 16.8 ± 2.0 , 19.7 ± 0.6 , and 21.7 ± 1.5 cm at the same time points (Table 1, Figure 2).

For *C. annuum*, plant height in Group I at 3, 9, and 14 DAT was recorded as 13.7 ± 1.5 , 14.0 ± 1.0 , and 15.3 ± 1.2 cm, respectively. In Group II, plant height increased to 15.0 ± 1.0 , 17.7 ± 0.6 , and 19.0 ± 1.0 cm, while in Group III, it reached 20.7 ± 1.2 , 22.0 ± 3.5 , and 25.7 ± 0.6 cm over the same period.

Moreover, for *G. globosa*, plant height showed a progressive increase across all groups over time. At 3 DAT, Group I and Group II recorded identical values of 9.7 ± 1.2 cm, whereas Group III was slightly lower at 9.3 ± 0.6 cm. By 9 DAT, plant height increased modestly in all groups, reaching 11.7 ± 0.6 cm in Group I, 11.0 ± 1.0 cm in Group II, and 11.3 ± 1.2 cm in Group III. The greatest increase was observed at 14 DAT, with Group III attaining the highest value (16.3 ± 0.6 cm), followed by Group II (14.3 ± 1.2 cm) and Group I (13.0 ± 1.0 cm), indicating that Group III promoted the strongest growth response over time.

A two-way ANOVA revealed a highly significant effect on the total number of plant leaves, influenced by plant species ($p < 0.001$), treatment groups ($p < 0.001$), and the interaction between species and treatment groups ($p < 0.001$).

3.2 Number of leaves

The number of leaves in *M. piperita* at 3, 9, and 14 DAT in Group I exhibited an average ($\pm$ SD) of 10.7 ± 1.2 , 12.3 ± 0.6 , and 15.0 ± 1.0 , respectively. In Group II, the corresponding values were 12.3 ± 1.5 , 13.7 ± 1.5 , and 15.3 ± 0.6 , while in Group III, the number of leaves increased to 11.7 ± 0.6 , 13.7 ± 0.6 , and 17.3 ± 1.2 (Table 2, Figure 3).

Similarly, for *C. annuum*, the number of leaves at 3, 9, and 14 DAT in Group I was recorded as 8.3 ± 0.6 , 9.7 ± 0.6 , and 15.3 ± 0.6 , respectively. In Group II, the number of leaves increased to 8.3 ± 0.6 , 12.7 ± 1.2 , and 18.7 ± 1.2 , while in Group III, it reached 8.0 ± 1.0 , 15.3 ± 1.2 , and 21.7 ± 1.5 over the same period.

For *G. globosa*, the number of leaves in Group I at 3, 9, and 14 DAT showed an average ($\pm$ SD) of 7.3 ± 0.6 , 10.3 ± 0.6 , and 10.7 ± 0.6 , respectively. In Group II, the values increased to 9.0 ± 1.0 , 11.0 ± 1.0 , and 15.3 ± 1.5 , while in Group III, the corresponding values were 13.7 ± 0.6 , 15.0 ± 1.0 , and 17.3 ± 0.6 .

A two-way ANOVA revealed a highly significant effect on the total number of plant leaves, influenced by plant species ($p < 0.001$), treatment groups ($p < 0.001$), and the interaction between species and treatment groups ($p < 0.001$).

3.3 Number of buds

The number of buds in *M. piperita* at 3, 9, and 14 DAT in Group I exhibited an average ($\pm$ SD) of 11.3 ± 1.2 , 14.0 ± 2.0 , and 15.3 ± 0.6 , respectively. In Group II, the corresponding values were $12.7 \pm$

TABLE 1 The plant height (cm) under varying CO₂ concentrations presented as mean and standard deviation.

Plant species	Treatment group	Plant height		
		3 DAT	9 DAT	14 DAT
<i>Mentha × piperita</i>	Group I	8.7 ± 0.6 g	11.0 ± 1.0 f	11.7 ± 1.5 e
	Group II	14.2 ± 2.3 d	16.0 ± 1.0 c	19.7 ± 0.6 b
	Group III	16.8 ± 2.0 c	19.7 ± 0.6 b	21.7 ± 1.5 a
<i>Capsicum annuum</i>	Group I	13.7 ± 1.5 g	14.0 ± 1.0 g	15.3 ± 1.2 f
	Group II	15.0 ± 1.0 e	17.7 ± 0.6 d	19.0 ± 1.0 cd
	Group III	20.7 ± 1.2 c	22.0 ± 3.5 b	25.7 ± 0.6 a
<i>Gomphrena globosa</i>	Group I	9.7 ± 1.2 e	11.7 ± 0.6 d	13.0 ± 1.0 c
	Group II	9.7 ± 1.2 e	11.0 ± 1.0 d	14.3 ± 1.2 b
	Group III	9.3 ± 0.6 e	11.3 ± 1.2 d	16.3 ± 0.6 a
Two-way repeated-measures ANOVA				
Plant species		<0.001***		
Treatment		<0.001***		
Time		<0.001***		
Plant × Treatment × Time		<0.001***		

Significance levels are indicated as $p < 0.05$ (*), $p < 0.01$ (**), and $p < 0.001$ (***), while “ns” denotes non-significant differences ($p > 0.05$). Different letters (a, b) indicate statistically significant differences according to Tukey’s HSD test at $p \leq 0.05$. Abbreviations: ANOVA, analysis of variance; DAT, days after treatment. CO₂ treatments: Group I (Control, ~400 ppm CO₂), Group II (Elevated CO₂, ~2,900 ppm CO₂), and Group III (Highly Elevated CO₂, ~5,400 ppm CO₂).

1.2, 15.0 ± 1.0, and 18.3 ± 0.6, while in Group III, the number of buds increased to 12.7 ± 1.2, 19.3 ± 1.2, and 22.7 ± 0.6 (Figure 4).

Similarly, for *C. annuum*, the number of buds at 3, 9, and 14 DAT in Group I was recorded as 10.3 ± 1.5, 12.7 ± 1.2, and 18.3 ± 0.6, respectively. In Group II, the values increased to 9.0 ± 1.0, 15.0 ± 1.0, and 21.3 ± 1.5, while in Group III, they reached 10.3 ± 1.5, 16.0 ± 1.7, and 27.3 ± 2.3.

For *G. globosa*, the number of buds in Group I at 3, 9, and 14 DAT showed an average (± SD) of 12.3 ± 0.6, 15.3 ± 1.2, and 18.3 ± 2.5, respectively. In Group II, the values were 10.7 ± 1.2, 13.7 ± 1.5, and 19.7 ± 1.5, while in Group III, the corresponding values were 11.0 ± 1.0, 15.3 ± 2.5, and 21.0 ± 1.0 (Figure 2).

A two-way ANOVA revealed a highly significant effect of treatment groups ($p < 0.001$) and the interaction between treatment groups and species ($p < 0.001$) on the total number of buds. However, the difference in bud numbers among plant species was not statistically significant.

3.4 Total leaf chlorophyll content

The chlorophyll content in *M. piperita* at 3, 9, and 14 DAT in Group I showed an average (± SD) of 35.5 ± 0.5, 36.7 ± 1.5, and 37.7 ± 1.2, respectively. In Group II, the corresponding values were 39.7 ± 2.5, 43.2 ± 0.6, and 47.2 ± 0.3, while in Group III, chlorophyll content increased to 43.7 ± 0.6, 46.9 ± 2.5, and 48.5 ± 0.6 (Figure 5).

Similarly, in *C. annuum*, total leaf chlorophyll content at 3, 9, and 14 DAT in Group I was recorded as 34.9 ± 0.6, 38.7 ± 1.2, and 40.8 ± 2.3, respectively. In Group II, the values increased to 39.9 ± 0.8, 42.6 ± 0.6, and 47.4 ± 0.4, while in Group III, they reached 47.1 ± 0.5, 49.2 ± 0.3, and 55.1 ± 1.5.

For *G. globosa*, chlorophyll content at 3, 9, and 14 DAT in Group I showed an average (± SD) of 22.4 ± 0.8, 25.3 ± 4.0, and 31.8 ± 1.4, respectively. In Group II, the values were 27.7 ± 1.9, 32.9 ± 6.1, and 36.9 ± 1.5, while in Group III, the corresponding values increased to 34.9 ± 3.5, 37.7 ± 2.0, and 40.3 ± 1.6.

A two-way ANOVA indicated a highly significant effect of plant species ($p < 0.001$), time of observation ($p < 0.001$), treatment groups ($p < 0.001$), and the interaction between treatment groups and species ($p < 0.001$) on total leaf chlorophyll content.

3.5 Relative leaf nitrogen index

The relative leaf nitrogen index in *M. piperita* exhibited a progressive increase across treatment groups. In Group I, the values at 3, 9, and 14 DAT were 94.9 ± 1.4, 98.2 ± 4.3, and 101.0 ± 3.3, respectively. In Group II, the corresponding values were 106.7 ± 7.1, 116.7 ± 1.7, and 127.9 ± 0.8. Group III showed the highest nitrogen content, with 118.0 ± 1.6, 127.1 ± 6.9, and 131.5 ± 1.7 at 3, 9, and 14 DAT, respectively.

Similarly, in *C. annuum*, relative leaf nitrogen index in Group I was recorded as 93.2 ± 1.7, 104.0 ± 3.4, and 109.8 ± 6.4 at 3, 9, and 14 DAT. In Group II, the values increased to 107.3 ± 2.2, 115.1 ± 1.7, and 128.5 ± 1.0. Group III exhibited the highest nitrogen content, with values of 127.6 ± 1.3, 133.6 ± 0.7, and 150.2 ± 4.3 at the respective time points.

For *G. globosa*, the relative leaf nitrogen index in Group I was 57.8 ± 2.3, 66.1 ± 11.3, and 84.6 ± 4.1 at 3, 9, and 14 DAT. In Group II, the recorded values were 73.0 ± 5.5, 87.6 ± 17.1, and 98.9 ± 4.2. Group III exhibited the highest nitrogen content, with values of 93.1 ± 9.9, 101.0 ± 5.7, and 108.4 ± 4.4 at the respective time points (Figure 2).

A two-way ANOVA indicated a highly significant effect of plant species ($p < 0.001$), time of observation ($p < 0.001$), treatment groups ($p < 0.001$), and the interaction between treatment groups and species ($p < 0.001$) on total relative leaf nitrogen index.

3.6 Root length

The root length after 14 DAT at harvesting, in *M. piperita* in Group I, Group II, and Group III, showed an average (± SD) of 11.3 ± 0.6 cm, 10.3 ± 0.6 cm, and 11.0 ± 1.7 cm. Similarly in *C. annuum* in Group I, Group II, and Group III, the root length showed an

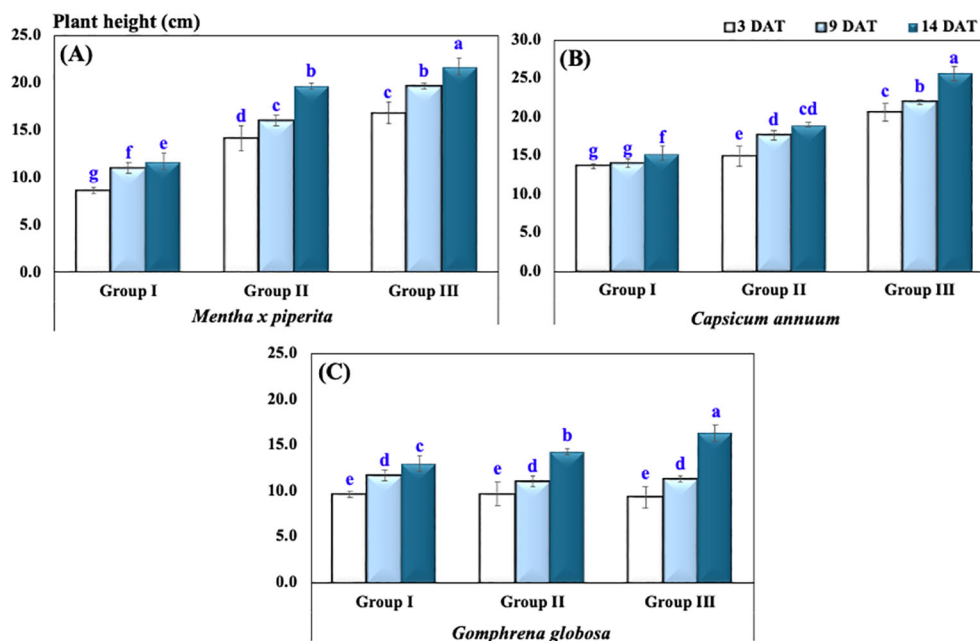


FIGURE 2

Plant height under varying CO₂ concentrations in (A) *Mentha x piperita*, (B) *Capsicum annuum*, (C) *Gomphrena globosa*. Different letters indicate statistically significant differences based on Tukey's HSD test at $p \leq 0.05$. Treatments included Group I (Control, ~400 ppm CO₂), Group II (Elevated CO₂, ~2,900 ppm), and Group III (Highly Elevated CO₂, ~5,400 ppm). DAT, Days after treatment.

average ($\pm$ SD) of 13.3 ± 0.6 cm, 12.3 ± 1.2 cm, and 12.0 ± 1.0 cm. *G. globosa* in Group I, Group II, and Group III showed an average ($\pm$ SD) of 11.0 ± 1.0 cm, 12.7 ± 0.6 cm, and 11.0 ± 1.0 cm.

3.7 Shoot fresh weight

The shoot FW of the studied plant species was recorded at 14 DAT, corresponding to the harvesting stage, and is presented as mean $\pm$ SD (Table 3, Figure 6).

For *M. piperita*, shoot FW in Group I was 7.0 ± 2.6 g, while in Group II and Group III treatment groups, the values increased to 9.3 ± 1.2 g and 11.0 ± 1.0 g, respectively. In *C. annuum*, shoot FW in Group I was measured at 3.7 ± 0.6 g, 5.0 ± 1.0 g in Group II, and 7.7 ± 0.6 g in Group III. Similarly, in *G. globosa*, shoot FW was 3.0 ± 1.0 g in Group I, while the treatment in Group II and Group III showed higher values of 7.7 ± 1.5 g and 9.7 ± 0.6 g, respectively.

3.8 Root fresh weight

The root FW of the studied plant species was measured at 14 DAT, corresponding to the harvesting stage, and is presented as mean $\pm$ SD (Table 4, Figure 7). For *M. piperita*, the root FW in Group I was 3.3 ± 0.6 g, while the treatment groups in Group II and Group III exhibited increased values of 5.3 ± 0.6 g and 6.3 ± 0.6 g, respectively. In *C. annuum*, root FW was recorded as 2.3 ± 0.6 g in Group I, whereas Group II and Group III treatments resulted in higher values of 3.3 ± 0.6 g and 4.6 ± 0.7 g, respectively. Similarly, for *G. globosa*, root FW in Group I was 2.0 ± 0.2 g, with significant

increases observed in Group II (3.7 ± 0.8 g) and Group III (5.2 ± 0.8 g) treatment groups.

3.9 Plant fresh weight

The plant FW of the studied species was measured at 14 DAT, corresponding to the harvesting stage and is presented as mean $\pm$ SD (Table 5, Figure 8). For *M. piperita*, plant FW in Group I was 10.3 ± 2.1 g, while Group II and Group III treatment groups exhibited higher values of 14.7 ± 1.5 g and 17.3 ± 1.5 g, respectively. In *C. annuum*, plant FW Group I was recorded as 6.0 ± 1.0 g, whereas Group II and Group III treatments resulted in increased values of 8.3 ± 0.6 g and 12.3 ± 1.2 g, respectively. Similarly, for *G. globosa*, plant FW in Group I was 5.0 ± 1.0 g, with notable increases observed in Group II (11.3 ± 2.3 g) and Group III (14.8 ± 0.3 g) treatment groups.

3.10 Shoot/root ratio

The shoot-to-root ratio (shoot/root ratio) was assessed at 14 DAT, corresponding to the harvesting stage, and is presented as mean $\pm$ SD (Table 6). For *M. piperita*, the shoot/root ratio in Group I was 2.2 ± 1.1 , while Group II and Group III treatment groups exhibited lower values of 1.8 ± 0.2 and 1.7 ± 0.1 , respectively. In *C. annuum*, the shoot/root ratio in Group I was recorded as 1.6 ± 0.3 , with similar values observed in Group II (1.6 ± 0.5) and Group III (1.7 ± 0.2) treatment groups. For *G. globosa*, the shoot/root ratio in Group I was 1.5 ± 0.5 , whereas in Group II, it resulted in an

TABLE 2 The number of leaves under varying CO₂ concentrations presented as mean and standard deviation.

Plant species	Treatment group	No. of leaves		
		3 DAT	9 DAT	14 DAT
<i>Mentha × piperita</i>	Group I	10.7 ± 1.2 f	12.3 ± 0.6 e	15.0 ± 1.0 b
	Group II	12.3 ± 1.5 d	13.7 ± 1.5 c	15.3 ± 0.6 b
	Group III	11.7 ± 0.6 d	13.7 ± 0.6 c	17.3 ± 1.2 a
<i>Capsicum annuum</i>	Group I	8.3 ± 0.6 f	9.7 ± 0.6 e	15.3 ± 0.6 c
	Group II	8.3 ± 0.6 e	12.7 ± 1.2 d	18.7 ± 1.2 b
	Group III	8.0 ± 1.0 e	15.3 ± 1.2 c	21.7 ± 1.5 a
<i>Gomphrena globosa</i>	Group I	7.3 ± 0.6 f	10.3 ± 0.6 d	10.7 ± 0.6 d
	Group II	9.0 ± 1.0 e	11.0 ± 1.0 d	15.3 ± 1.5 b
	Group III	13.7 ± 0.6 c	15.0 ± 1.0 b	17.3 ± 0.6 a
Two-way repeated-measures ANOVA				
Plant species		<0.001***		
Treatment		<0.001***		
Time		<0.001***		
Plant × Treatment × Time		<0.001***		

Significance levels are indicated as $p < 0.05$ (*), $p < 0.01$ (**), and $p < 0.001$ (***), while “ns” denotes non-significant differences ($p > 0.05$). Different letters (a, b) indicate statistically significant differences according to Tukey’s HSD test at $p \leq 0.05$. Abbreviations: ANOVA, analysis of variance; DAT, days after treatment. CO₂ treatments: Group I (Control, ~400 ppm CO₂), Group II (Elevated CO₂, ~2,900 ppm CO₂), and Group III (Highly Elevated CO₂, ~5,400 ppm CO₂).

increased ratio of 2.1 ± 0.2 , followed by a slight decrease in Group III (1.9 ± 0.4).

3.11 Correlation analysis of CO₂ levels and plant physiological responses

The blue–red heatmap presented in Figure 9 illustrates the correlation patterns among various study variables. In this heatmap, blue signifies a positive correlation, red represents a negative correlation, and white indicates no correlation. Statistically significant correlations are highlighted using gray boxes. The analysis reveals a strong inverse relationship between plant presence and CO₂ levels, suggesting that the presence of plants significantly reduces CO₂ concentration. Furthermore, indoor environments exhibited higher environmental factors (EFs) and noise levels compared to outdoor conditions. Additionally, a notable positive correlation was observed between CO₂ levels and chlorophyll content, indicating that increased CO₂ concentrations may influence photosynthetic pigment accumulation.

3.12 Canonical correspondence analysis of CO₂ effects on plant growth

The canonical correspondence analysis (CCA) presented in Figure 10 illustrates the relationships between dependent and independent variables. In this multivariate analysis, arrows represent both the strength and direction of correlations, providing insights into how different variables respond to changes in CO₂ levels and observation time. Independent variables, such as CO₂ concentration and time of observation, are depicted by green arrows, with their length indicating the strength of their influence. The analysis demonstrates that CO₂ concentration and observation time significantly impact chlorophyll content, total nitrogen, and plant growth parameters, highlighting the role of elevated CO₂ in modulating physiological and biochemical processes in plants.

All measurements were conducted under standardized greenhouse conditions to ensure experimental reproducibility.

4 Discussion

The results of this study indicate that elevated CO₂ levels significantly enhanced multiple physiological and biochemical traits, including plant height, leaf number, bud formation, biomass accumulation, chlorophyll content, and relative leaf nitrogen index in *C. annuum*, *G. globosa*, and *M. piperita*. At the highest CO₂ concentration (5,400 ppm, Group III), *C. annuum* exhibited the most substantial growth response, displaying the most significant increase in plant height, number of buds, leaf chlorophyll content, and leaf nitrogen content, while also experiencing the most pronounced reduction in root length. It ranked second in leaf number and root weight increase but showed the smallest growth in shoot weight.

In contrast, *G. globosa* demonstrated the highest increase in leaf number, shoot weight, and root weight, with root length remaining unchanged. However, it exhibited the lowest response in all other measured parameters. *M. piperita* displayed a moderate response across most parameters under 5,400 ppm CO₂, except for root weight, which showed the least increase among the three species. Overall, all three species experienced enhanced growth compared to Group I (ambient CO₂ at 400 ppm), confirming that elevated CO₂ positively affects plant growth parameters.

Elevated CO₂ consistently enhanced growth-related parameters across the studied species, including plant height, leaf number, bud formation, shoot and root fresh weight, and total biomass. These findings are consistent with previous reports demonstrating that CO₂ enrichment stimulates carbon assimilation and biomass accumulation in diverse crop species (Idso et al., 1987; Thompson et al., 2017; Yadav et al., 2019; Lamichaney et al., 2021; Mndela et al., 2022; Kaur et al., 2023). However, the magnitude of growth stimulation differed among species, with *C. annuum* exhibiting the strongest overall response at 5,400 ppm. Such variation likely

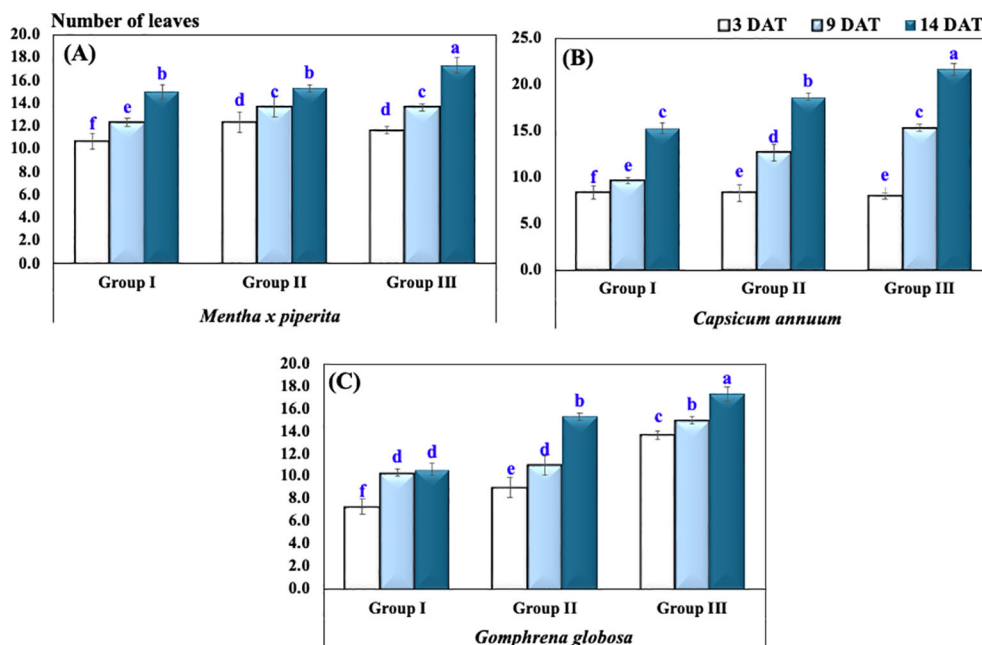


FIGURE 3

Number of leaves of plants under different CO₂ concentrations in (A) *Mentha x piperita*, (B) *Capsicum annum*, (C) *Gomphrena globosa*. Different letters indicate significant differences based on Tukey's HSD test at $p \leq 0.05$. Treatments included Group I (Control, ~400 ppm CO₂), Group II (Elevated CO₂, ~2,900 ppm), and Group III (Highly Elevated CO₂, ~5,400 ppm). DAT, Days after treatment.

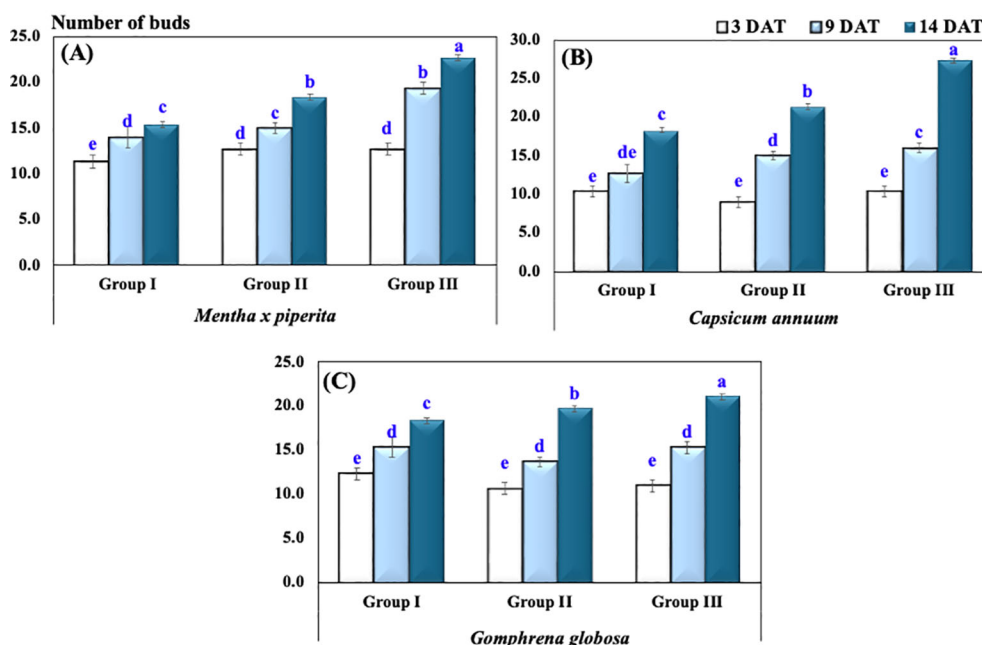


FIGURE 4

Number of buds of plants under varying CO₂ concentrations in (A) *Mentha x piperita*, (B) *Capsicum annum*, (C) *Gomphrena globosa*. Different letters indicate significant differences based on Tukey's HSD test at $p \leq 0.05$. Treatments included Group I (Control, ~400 ppm CO₂), Group II (Elevated CO₂, ~2,900 ppm), and Group III (Highly Elevated CO₂, ~5,400 ppm). DAT, Days after treatment.

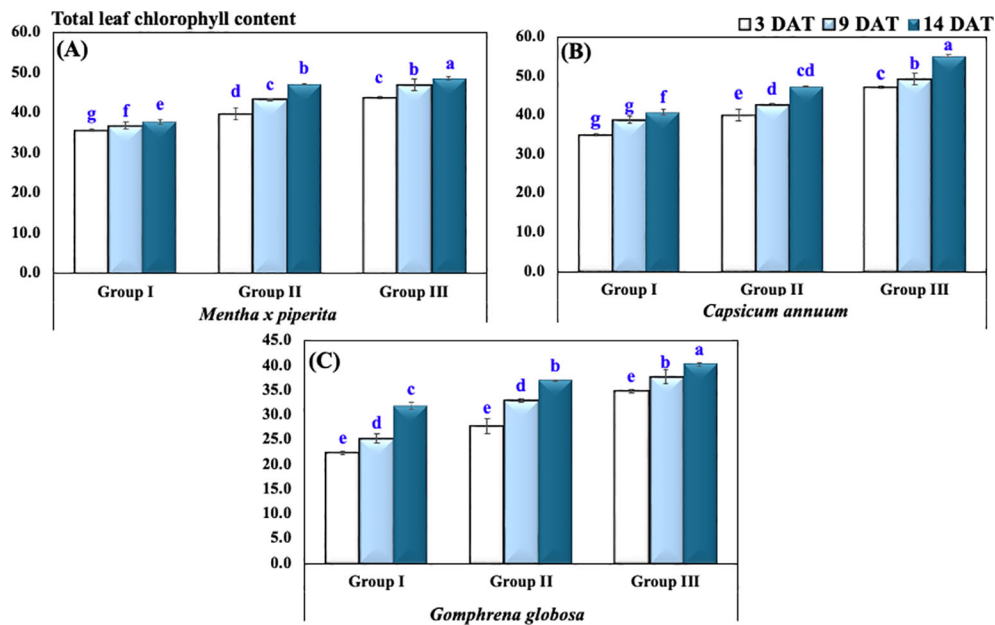


FIGURE 5

Total leaf chlorophyll content of plants under varying CO₂ concentrations in (A) *Mentha x piperita*, (B) *Capsicum annum*, (C) *Gomphrena globosa*. Different letters indicate significant differences based on Tukey's HSD test at $p \leq 0.05$. Treatments included Group I (Control, ~400 ppm CO₂), Group II (Elevated CO₂, ~2,900 ppm), and Group III (Highly Elevated CO₂, ~5,400 ppm). DAT, Days after treatment.

reflects differences in photosynthetic pathway characteristics, resource allocation strategies, and physiological plasticity under high CO₂ conditions.

Nutrient availability may further modulate plant responses to elevated CO₂. Previous studies have shown that phosphorus limitation can reduce the stimulatory effects of CO₂ enrichment on biomass accumulation (Ellsworth et al., 2017; Jiang et al., 2020; Li et al., 2023). Although nutrient levels were controlled in the present study, interactions between CO₂ enrichment and nutrient dynamics

remain important factors influencing long-term plant performance.

Regarding chlorophyll content, CO₂ enrichment is crucial in modulating photosynthetic pigments. For instance, a study on wheat cultivars under drought stress demonstrated that elevated CO₂ influences leaf photosynthesis and chlorophyll fluorescence parameters. These findings suggest that CO₂ enrichment can alter chlorophyll content and photosynthetic efficiency, although the extent of these effects varies depending on plant species and environmental conditions (Yang et al., 2023). Although photosynthetic rate, stomatal conductance, and transpiration were not directly quantified in the present study, the observed increases in chlorophyll content and biomass under elevated CO₂ are consistent with enhanced carbon assimilation. Elevated CO₂ typically increases photosynthetic rate in C3 species by reducing photorespiration, while simultaneously decreasing stomatal conductance and transpiration, thereby improving water-use efficiency (Bravdo and Canvin, 1979; Engineer et al., 2016; Xu et al., 2016; Hatfield and Dold, 2019; Cao et al., 2022; Wei et al., 2022; Haghpanah et al., 2024). In C4 species, which already possess a CO₂-concentrating mechanism, the stimulation of photosynthesis is generally less pronounced. Therefore, the growth enhancement observed in this study likely reflects improved carbon fixation efficiency and altered stomatal regulation under high CO₂ conditions.

Our study demonstrated that elevated CO₂ significantly enhanced chlorophyll and leaf nitrogen content, suggesting improved photosynthetic capacity. These findings are consistent with previous studies that reported increased photosynthetic rates and higher chlorophyll content under CO₂ enrichment (Cai et al., 2021; Zhang et al., 2022). However, other research has documented decreased relative leaf nitrogen index under similar conditions (Laza et al., 2021). Additionally, Wang et al. (2023) found that elevated CO₂

TABLE 3 The number of buds under varying CO₂ concentrations presented as mean and standard deviation.

Treatment group	Shoot fresh weight		
	<i>Mentha x piperita</i>	<i>Capsicum annum</i>	<i>Gomphrena globosa</i>
Group I	7.0 ± 2.6 c	3.7 ± 0.6 b	3.0 ± 1.0 c
Group II	9.3 ± 1.2 b	5.0 ± 1.0 b	7.7 ± 1.5 b
Group III	11.0 ± 1.0 a	7.7 ± 0.6 a	9.7 ± 0.6 a
Two-way repeated-measures ANOVA			
Plant species	<0.001***		
Treatment	<0.001***		
Plant × Treatment × Time	<0.001***		

Significance levels: $p < 0.05$ (*), $p < 0.01$ (**), $p < 0.001$ (***); "ns" indicates non-significant differences ($p > 0.05$). Different letters (a, b) denote significant differences according to Tukey's HSD test at $p \leq 0.05$. Abbreviations: ANOVA, analysis of variance; DAT, days after treatment. CO₂ treatments: Group I (Control, ~400 ppm CO₂), Group II (Elevated CO₂, ~2,900 ppm CO₂), and Group III (Highly Elevated CO₂, ~5,400 ppm CO₂).

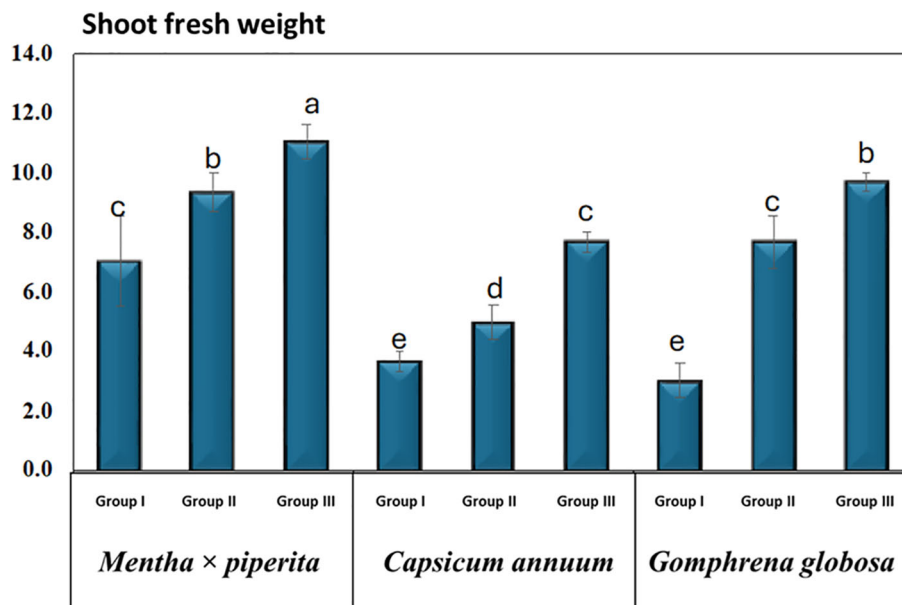


FIGURE 6

Shoot fresh weight under varying CO₂ concentrations. Different letters indicate significant differences based on Tukey's HSD test at $p \leq 0.05$.

increases nitrogen concentrations in shoots while simultaneously reducing nitrogen levels in roots (Wang et al., 2023). Similarly, our study showed an overall increase in total relative leaf nitrogen index under elevated CO₂ conditions (Wang et al., 2023).

The observed differences in relative leaf nitrogen index between C3 (*C. annuum* and *M. piperita*) and C4 (*G. globosa*) species may be partially explained by their contrasting photorespiratory pathways. In C3 plants, elevated CO₂ reduces photorespiration, thereby improving nitrogen-use efficiency and potentially enhancing nitrogen accumulation in leaves. In contrast, C4 plants possess a CO₂-

concentrating mechanism that minimizes photorespiration even under ambient conditions, resulting in relatively smaller changes in nitrogen assimilation under elevated CO₂. This physiological distinction may explain the comparatively moderate nitrogen response observed in *G. globosa*. The contrasting responses observed between the C3 and C4 species under 2,900 and 5,400 ppm further support the concept that C3 plants benefit more strongly from elevated CO₂ due to reduced photorespiration, whereas C4 plants exhibit comparatively moderate stimulation because of their inherent CO₂-concentrating mechanism. From an agricultural perspective, these findings suggest that C3 crops may exhibit stronger nitrogen-related growth stimulation under elevated CO₂, whereas C4 crops may maintain more stable nitrogen metabolism due to their inherently efficient carbon fixation pathway.

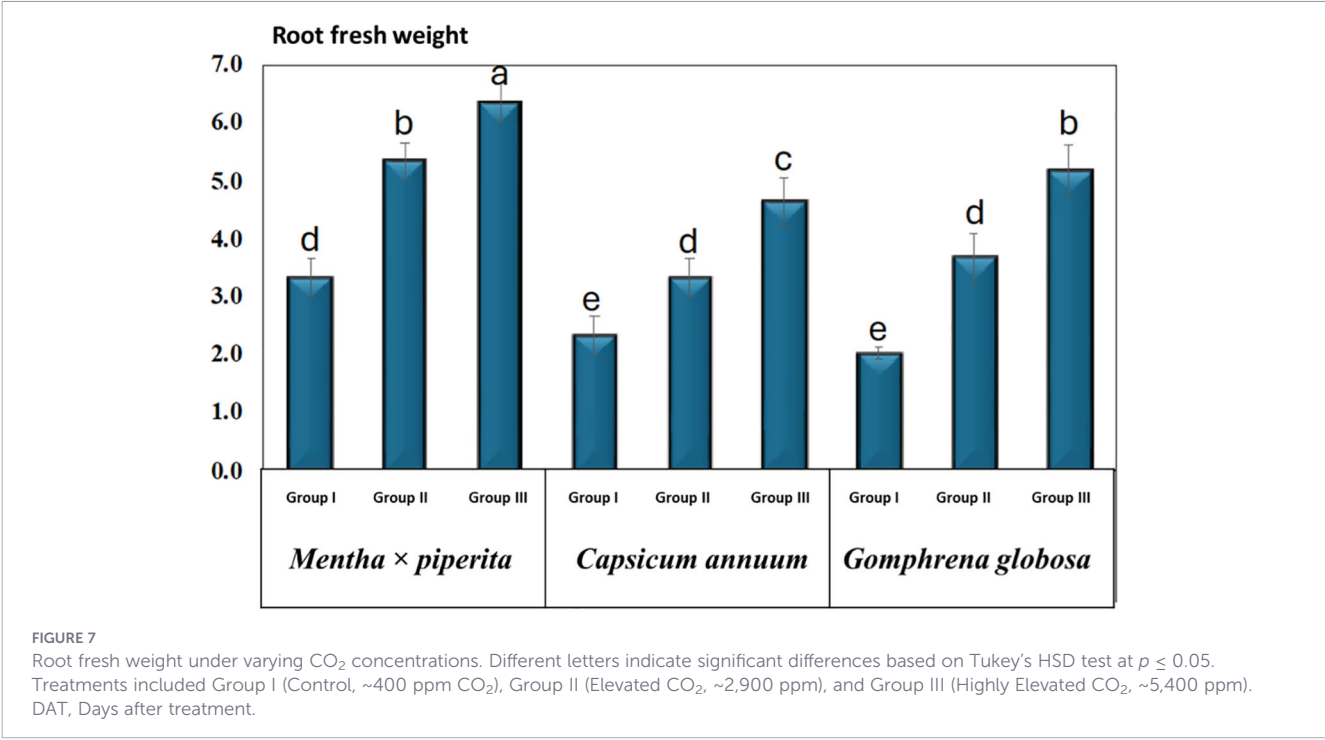
The positive effects of CO₂ enrichment on biomass accumulation have been widely documented. For instance, previous studies reported that elevated CO₂ significantly increased tiller number, root dry weight, shoot dry weight, and total plant dry weight, even in the presence of cadmium stress (Jia et al., 2011). These findings align with our results, where elevated CO₂ increased root and shoot FW across the tested species.

CO₂ enrichment has also been shown to enhance plant height, with studies reporting increases of 4%–8% and 8%–29% during the flowering and podding stages, respectively (Lamichaney et al., 2021). Our study agreed with these findings and found that elevated CO₂ promoted plant height. However, in contrast to previous research, we observed a reduction in root length under CO₂ enrichment across the tested species, except for *G. globosa*, where root length remained unchanged. Additionally, our findings suggest that *G. globosa* exhibited enhanced growth under elevated CO₂

TABLE 4 The plant's root FW (g) under varying CO₂ concentrations.

Treatment group	Root fresh weight		
	<i>Mentha × piperita</i>	<i>Capsicum annuum</i>	<i>Gomphrena globosa</i>
Group I	3.3 ± 0.6 a	2.3 ± 0.6 a	2.0 ± 0.2 a
Group II	5.3 ± 0.6 a	3.3 ± 0.6 a	3.7 ± 0.8 a
Group III	6.3 ± 0.6 a	4.6 ± 0.7 a	5.2 ± 0.8 a
Two-way repeated-measures ANOVA			
Plant species	<0.001***		
Treatment	<0.001***		
Plant × Treatment × Time	<0.001***		

Significance levels: $p < 0.05$ (*), $p < 0.01$ (**), $p < 0.001$ (***); "ns" indicates non-significant differences ($p > 0.05$). Different letters (a, b) denote significant differences according to Tukey's HSD test at $p \leq 0.05$. Abbreviation: ANOVA, analysis of variance. CO₂ treatments: Group I (Control, ~400 ppm CO₂), Group II (Elevated CO₂, ~2,900 ppm CO₂), and Group III (Highly Elevated CO₂, ~5,400 ppm CO₂).



conditions, aligning with the results reported by [Silva et al. \(2024\)](#). Previous research has shown that elevated atmospheric CO₂ enhances photosynthesis and growth in amaranth. However, grain yield remains unaffected, protein content declines, and there is an increase in secondary metabolites and stress tolerance ([Silva et al., 2024](#)). These findings reinforce that CO₂ enrichment can stimulate plant growth, but the overall physiological and metabolic responses

TABLE 5 Plant FW (g) in *Mentha × piperita*, *C. annuum*, and *G. globosa*.

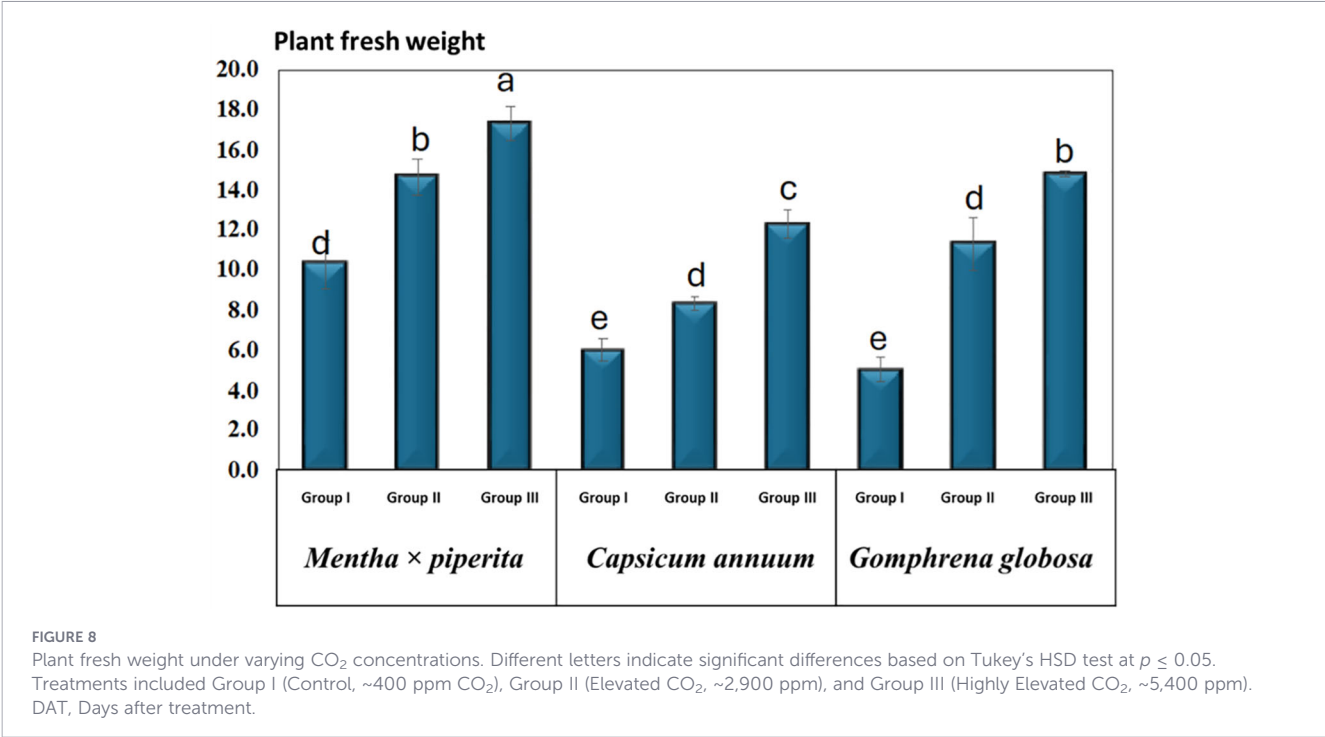
Treatment group	Plant fresh weight		
	<i>Mentha × piperita</i>	<i>Capsicum annuum</i>	<i>Gomphrena globosa</i>
Group I	10.3 ± 2.1 c	6.0 ± 1.0 c	5.0 ± 1.0 c
Group II	14.7 ± 1.5 b	8.3 ± 0.6 b	11.3 ± 2.3 b
Group III	17.3 ± 1.5 a	12.3 ± 1.2 a	14.8 ± 0.3 a
Two-way repeated-measures ANOVA			
Plant species	<0.001***		
Treatment	<0.001***		
Plant × Treatment × Time	<0.001***		

Significance levels: $p < 0.05$ (*), $p < 0.01$ (**), $p < 0.001$ (***); "ns" indicates non-significant differences ($p > 0.05$). Different letters (a, b) denote significant differences according to Tukey's HSD test at $p \leq 0.05$. Abbreviations: ANOVA, analysis of variance; Plant FW, plant fresh weight. CO₂ treatments: Group I (Control, ~400 ppm CO₂), Group II (Elevated CO₂, ~2,900 ppm CO₂), and Group III (Highly Elevated CO₂, ~5,400 ppm CO₂).

depend on species-specific traits. Both our study and that of [Pereyda-González et al. \(2022\)](#) indicate that CO₂ enrichment can enhance plant growth, although its effects are temperature-dependent ([Pereyda-González et al., 2022](#)). In our study, *C. annuum* exhibited the highest growth and chlorophyll content at 5,400 ppm CO₂, suggesting a strong positive response to elevated CO₂ levels. Similarly, previous research has demonstrated that *C. annuum* experiences improved seedling growth at moderate temperatures (30 °C). However, at higher temperatures (40 °C), CO₂ enrichment fails to mitigate heat stress, impairing fruiting and development ([Pereyda-González et al., 2022](#)).

Our findings also align with previous research on *M. piperita*, where CO₂ enrichment (620 ppm) increased biomass production by 48% and enhanced essential oil yield. Although our study tested higher CO₂ concentrations (2,900 and 5,400 ppm), the observed growth stimulation supports the idea that *M. piperita* is exceptionally responsive to CO₂ enrichment ([Al Jaouni et al., 2018](#)).

Similarly, a study on sweet pepper (*C. annuum*) found that short-term exposure to elevated CO₂ (800 mmol mol⁻¹) increased photosynthetic rates and biomass accumulation. However, long-term exposure led to acclimation, reducing photosynthetic efficiency after 135 days ([Porras et al., 2017](#)). [Kumari et al. \(2016\)](#) also reported that CO₂ enrichment (550 ppm) combined with high temperatures improved yield attributes but reduced photosynthetic efficiency due to acclimation ([Kumari et al., 2016](#)). However, our study suggests that photosynthetic efficiency remains enhanced even at higher CO₂ levels as indicated by



increased chlorophyll content and plant biomass. This discrepancy may be attributed to differences in experimental conditions, such as controlled environments versus field studies (Karami et al., 2023). A notable trade-off observed in multiple studies is the dilution effect on nitrogen content (Dong et al., 2018; Yang et al., 2023).

TABLE 6 The shoot/root ratio (g.g⁻¹) in plants.

Treatment group	Shoot/root ratio		
	<i>Mentha × piperita</i>	<i>Capsicum annuum</i>	<i>Gomphrena globosa</i>
Group I	2.2 ± 1.1 a	1.6 ± 0.3 a	1.5 ± 0.5 a
Group II	1.8 ± 0.2 a	1.6 ± 0.5 a	2.1 ± 0.2 a
Group III	1.7 ± 0.1 a	1.7 ± 0.2 a	1.9 ± 0.4 a
Two-way repeated-measures ANOVA			
Plant species	>0.05 ns		
Treatment	>0.05 ns		
Plant × Treatment × Time	>0.05 ns		

Statistical analysis of plant growth parameters under different CO₂ treatments. “ns” denotes non-significant differences ($p > 0.05$). Different letters (a, b) indicate statistically significant differences according to Tukey’s HSD test at $p \leq 0.05$. Abbreviation: ANOVA, analysis of variance. CO₂ treatments: Group I (Control, ~400 ppm CO₂), Group II (Elevated CO₂, ~2,900 ppm CO₂), and Group III (Highly Elevated CO₂, ~5,400 ppm CO₂).

While CO₂ elevation enhances biomass accumulation, it can also lead to a decline in total leaf nitrogen content, a key factor influencing photosynthetic enzyme activity. This suggests that although elevated CO₂ initially boosts growth, long-term plant responses may be constrained by nitrogen availability, potentially affecting overall nutritional quality (Andrews et al., 2019). This effect is particularly evident in crop species, where acclimation effects and eventual nitrogen limitations follow early increases in photosynthetic efficiency (Porrás et al., 2017). Additionally, CO₂ enrichment may reduce the concentration of essential nutrients, impacting the nutritional quality of crops. These findings underscore the importance of adopting a holistic approach when evaluating the impacts of elevated CO₂ on agriculture and food security (Dong et al., 2018; Yang et al., 2023). This discussion highlights that while CO₂ enrichment influences plant growth, its effects must be assessed within the broader context of environmental interactions, ensuring a comprehensive understanding of future agricultural challenges (Gonzalez-Meler et al., 2004; Jeong et al., 2018; Boretti and Florentine, 2019).

Future studies evaluating protein-to-carbohydrate ratios and carbon–nitrogen balance would provide deeper insight into metabolic adjustments under elevated CO₂ conditions.

It should be noted that the present experiment evaluated short-term responses (14 DAT) under controlled conditions. Longer-term studies extending to full developmental stages and harvest would be necessary to determine the sustained effects of elevated CO₂ on yield and reproductive performance. Future studies should include intermediate CO₂ concentrations (e.g., 600–1,200 ppm) that better represent projected atmospheric scenarios, in order to assess gradual physiological and yield-related responses under more realistic climate conditions.

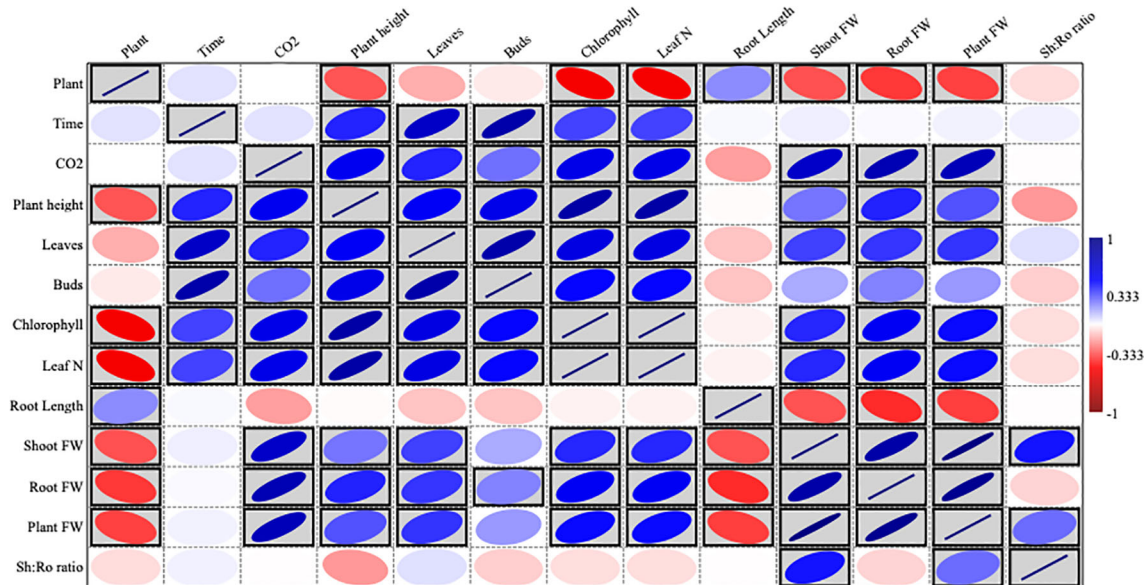


FIGURE 9

Blue–red heatmap illustrating the interrelationships between study variables. Blue indicates a positive correlation, red represents a negative (inverse) correlation, and white denotes no correlation. Gray boxes highlight statistically significant correlations.

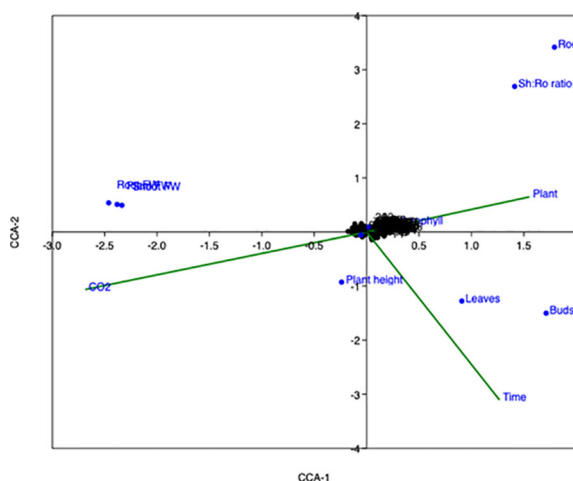


FIGURE 10

Canonical correspondence analysis (CCA) illustrating the interactions among study variables. Arrows represent the relationships between dependent and independent variables, with direction and length indicating the strength and significance of the associations.

Despite the overall positive impact of elevated CO₂, variations in photosynthetic efficiency, biomass allocation, and nutrient assimilation underscore the need for further investigations. Future research should focus on understanding the long-term physiological and biochemical adaptations of plants under CO₂ enrichment, particularly in the context of climate change and global food security. Additionally, exploring the interactions between elevated CO₂, nutrient availability, and abiotic stress factors will be crucial for developing sustainable agricultural strategies to mitigate potential challenges associated with rising atmospheric CO₂ concentrations.

Data availability statement

The original contributions presented in the study are included in the article/[Supplementary Material](#). Further inquiries can be directed to the corresponding author.

Author contributions

AA: Conceptualization, Investigation, Methodology, Resources, Supervision, Validation, Visualization, Writing – original draft, Writing – review & editing. EA: Investigation, Resources, Writing – review & editing. RA: Investigation, Resources, Writing – review & editing. AA: Investigation, Resources, Writing – review & editing. AA: Investigation, Resources, Writing – review & editing. RA: Investigation, Resources, Writing – review & editing. SA: Conceptualization, Investigation, Methodology, Resources, Supervision, Validation, Visualization, Writing – original draft, Writing – review & editing.

5 Conclusion

This study demonstrates that elevated CO₂ levels significantly enhance plant growth and photosynthetic efficiency in *C. annuum*, *G. globosa*, and *M. piperita*. The observed growth stimulation can be attributed to improved water-use efficiency, enhanced RuBisCO activity, and increased photosynthetic capacity. However, the magnitude of these effects varied among species, highlighting the species-specific nature of plant responses to CO₂ enrichment.

Funding

The author(s) declared that financial support was received for this work and/or its publication. The United Arab Emirates University provided support for the publication of this article.

Conflict of interest

The author(s) declared that this work was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The author(s) declared that generative AI was not used in the creation of this manuscript.

Any alternative text (alt text) provided alongside figures in this article has been generated by Frontiers with the support of artificial

intelligence and reasonable efforts have been made to ensure accuracy, including review by the authors wherever possible. If you identify any issues, please contact us.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpls.2026.1772837/full#supplementary-material>

References

- Ainsworth, E. A., and Long, S. P. (2021). 30 years of free-air carbon dioxide enrichment (FACE): What have we learned about future crop productivity and its potential for adaptation? *Glob. Change Biol.* 27, 27–49. doi: 10.1111/gcb.15375
- Akhlaq, M., Chuan, Z., Haofang, Y., Shaowei, L., Ni, Y., Zhou, J., et al. (2023). Exploring adequate CO₂ elevation for optimum tomato growth and yield under protected cultivation. *J. Plant Physiol.* 289, 154093. doi: 10.1016/j.jplph.2023.154093
- Al Jaouni, S., Saleh, A. M., Wadaan, M. A. M., Hozzein, W. N., Selim, S., and Abdelgawad, H. (2018). Elevated CO₂ induces a global metabolic change in basil (*Ocimum basilicum* L.) and peppermint (*Mentha piperita* L.) and improves their biological activity. *J. Plant Physiol.* 224–225, 121–131. doi: 10.1016/j.jplph.2018.03.016
- Almasri, R. S., Bedir, A. S., and Raish, S. M. A. (2025). Comprehensive ethnopharmacological analysis of medicinal plants in the UAE: *Lawsonia inermis*, *Nigella sativa*, *Ziziphus spina-christi*, *Allium cepa*, *Allium sativum*, *Cymbopogon schoenanthus*, *Matricaria aurea*, *Phoenix dactylifera*, *Portulaca oleracea*, *Reichardia tingitana*, *Salvadora persica*, *Solanum lycopersicum*, *Trigonella foenum-graecum*, *Withania somnifera*, and *Ziziphus lotus*. *Nutrients* 17, 411. doi: 10.3390/nu17030411
- Al Raish, S. M. A. (2026). The efficacy of plant extracts against key food-borne pathogens: A mechanistic, applications, and advances. *Microorganisms* 14. doi: 10.3390/microorganisms14030621
- Al Raish, S. M. A., Almasri, R. S., Bedir, A. S., and Elkahwagy, A. A. (2025). Phytochemical composition, bioactive compounds, and antidiabetic potential of four medicinal plants native to the UAE: *Capparis spinosa*, *Citrullus colocynthis*, *Morus alba*, and *Rhazya stricta*. *Biology* 14, 1146. doi: 10.3390/biology14091146
- Al Raish, S. M., Alsheriafi, H. N., AlKuwaiti, A. A., Safi, S. K., and Safi, A. S. (2026). Insights into knowledge, attitudes, and practices of medicinal plant use in the United Arab Emirates: a cross-sectional study. *Front. Nutr.* 13, 1755440. doi: 10.3389/fnut.2026.1755440
- Ancin, M., Gámez, A. L., Jauregui, I., Galmes, J., Sharwood, R. E., Erice, G., et al. (2024). Does the response of Rubisco and photosynthesis to elevated [CO₂] change with unfavourable environmental conditions? *J. Exp. Bot.* 75, 7351–7364. doi: 10.1093/jxb/erae379
- Andrews, M., Condon, L. M., Kemp, P. D., Topping, J. F., Lindsey, K., Hodge, S., et al. (2019). Elevated CO₂ effects on nitrogen assimilation and growth of C₃ vascular plants are similar regardless of N-form assimilated. *J. Exp. Bot.* 70, 683–690. doi: 10.1093/jxb/ery371
- Baslam, M., and Sanz-Saez, A. (2023). Editorial: Photosynthesis in a changing global climate: A matter of scale, volume II. *Front. Plant Sci.* 14. doi: 10.3389/fpls.2023.1158816
- Bedir, A. S., Almasri, R. S., and Al Raish, S. M. (2025). Therapeutic efficacy of *Nigella sativa* and *Ziziphus lotus*: sustainable strategies for diabetes, antimicrobial resistance, and health treatment. *Front. Nutr.* 12. doi: 10.3389/fnut.2025.1592423
- Boretti, A., and Florentine, S. (2019). Atmospheric CO₂ concentration and other limiting factors in the growth of C₃ and C₄ plants. *Plants* 8, 92. doi: 10.3390/plants8040092
- Bravdo, B.-A., and Canvin, D. (1979). Effect of carbon dioxide on photorespiration 1. *Plant Physiol.* 63, 399–401. doi: 10.1104/pp.63.2.399
- Busch, F. A. (2020). Photorespiration in the context of Rubisco biochemistry, CO₂ diffusion and metabolism. *Plant J. Cell. Mol. Biol.* 101, 919–939. doi: 10.1111/tpj.14674
- Cai, Y., Miao, Y., Wu, H., and Wang, D. (2021). Hyperspectral estimation models of winter wheat chlorophyll content under elevated CO₂. *Front. Plant Sci.* 12. doi: 10.3389/fpls.2021.642917
- Cao, Q., Li, G., and Liu, F. (2022). Elevated CO₂ enhanced water use efficiency of wheat to progressive drought stress but not on maize. *Front. Plant Sci.* 13. doi: 10.3389/fpls.2022.953712
- Cassia, R., Nocioni, M., Correa-Aragunde, N., and Lamattina, L. (2018). Climate change and the impact of greenhouse gases: CO₂ and NO, friends and foes of plant oxidative stress. *Front. Plant Sci.* 9. doi: 10.3389/fpls.2018.00273
- Chen, S., Zhao, W., Zhang, R., Sun, X., Zhou, Y., and Liu, L. (2023). Higher sensitivity of NIRvRad in detecting net primary productivity of C₄ than that of C₃: Evidence from ground measurements of wheat and maize. *Remote Sens.* 15, 1133. doi: 10.3390/rs15041133
- Cheng, Y., and He, D. (2019). A photosynthesis continuous monitoring system for CAM plants. *Int. J. Agric. Biol. Eng.* 12, 141–146. doi: 10.25165/j.ijabe.20191203.4885
- Dong, J., Gruda, N., Lam, S. K., Li, X., and Duan, Z. (2018). Effects of elevated CO₂ on nutritional quality of vegetables: A review. *Front. Plant Sci.* 9. doi: 10.3389/fpls.2018.00924
- Ellsworth, D. S., Anderson, I. C., Crous, K. Y., Cooke, J., Drake, J. E., Gherlenda, A. N., et al. (2017). Elevated CO₂ does not increase eucalypt forest productivity on a low-phosphorus soil. *Nat. Clim. Change* 7, 279–282. doi: 10.1038/nclimate3235
- Engineer, C. B., Hashimoto-Sugimoto, M., Negi, J., Israelsson-Nordström, M., Azoulay-Shemer, T., Rappel, W.-J., et al. (2016). CO₂ sensing and CO₂ regulation of stomatal conductance: Advances and open questions. *Trends Plant Sci.* 21, 16–30. doi: 10.1016/j.tplants.2015.08.014
- F U S., and Ferris, H. (2006). Plant species, atmospheric CO₂ and soil N interactively or additively control C allocation within plant-soil systems. *Sci. China C. Life.* 49, 603–612. doi: 10.1007/s11427-006-2026-x
- Gonzalez-Meler, M. A., Taneva, L., and Trueman, R. J. (2004). Plant respiration and elevated atmospheric CO₂ concentration: Cellular responses and global significance. *Ann. Bot.* 94, 647–656. doi: 10.1093/aob/mch189
- Haghpahan, M., Hashemipetroudi, S., Arzani, A., and Araniti, F. (2024). Drought tolerance in plants: Physiological and molecular responses. *Plants* 13. doi: 10.3390/plants13212962
- Hatfield, J. L., and Dold, C. (2019). Water-use efficiency: Advances and challenges in a changing climate. *Front. Plant Sci.* 10. doi: 10.3389/fpls.2019.00103
- Idso, S. B., Kimball, B. A., Anderson, M. G., and Mauney, J. R. (1987). Effects of atmospheric CO₂ enrichment on plant growth: The interactive role of air temperature. *Agric. Ecosyst. Environ.* 20 (1), 1–10. doi: 10.1016/0167-8809(87)90023-5

- Jeong, H.-M., Kim, H.-R., Hong, S., and You, Y.-H. (2018). Effects of elevated CO₂ concentration and increased temperature on leaf quality responses of rare and endangered plants. *J. Ecol. Environ.* 42, 1. doi: 10.1186/s41610-017-0061-0
- Jia, Y., Tang, S., Ju, X., Shu, L., Tu, S., Feng, R., et al. (2011). Effects of elevated CO₂ levels on root morphological traits and Cd uptakes of two *Lolium* species under Cd stress. *J. Zhejiang Univ.-Sci. B* 12, 313–325. doi: 10.1631/jzus.B1000181
- Jiang, M., Caldararu, S., Zhang, H., Fleischer, K., Crous, K. Y., Yang, J., et al. (2020). Low phosphorus supply constrains plant responses to elevated CO₂: A meta-analysis. *Glob. Change Biol.* 26, 5856–5873. doi: 10.1111/gcb.15277
- Jiang, J., Shi, S., and Raftery, A. E. (2025). Mitigation efforts to reduce carbon dioxide emissions and meet the Paris Agreement have been offset by economic growth. *Commun. Earth Environ.* 6, 823. doi: 10.1038/s43247-025-02743-x
- Karami, S., Shiran, B., Ravash, R., and Fallahi, H. (2023). A comprehensive analysis of transcriptomic data for comparison of plants with different photosynthetic pathways in response to drought stress. *PLoS One* 18, e0287761. doi: 10.1371/journal.pone.0287761
- Kaur, H., Kumar, A., Choudhary, A., Sharma, S., Choudhary, D. R., and Mehta, S. (2023). “Effect of elevated CO₂ on plant growth, active constituents, and production,” in *Plants and Their Interaction to Environmental Pollution* (India: Elsevier), 61–77. doi: 10.1016/B978-0-323-99978-6.00016-9
- Kaur, N., Thenveetil, N., Sehgal, A., Bheemanahalli, R., Reddy, K. N., Gao, W., et al. (2025). Effect of elevated carbon dioxide on the growth, development, and nutrient composition of C3 and C4 functional groups. *Front. Earth Sci.* doi: 10.1007/s11707-025-1178-6
- Keenan, T. F., Luo, X., De Kauwe, M. G., Medlyn, B. E., Prentice, I. C., Stocker, B. D., et al. (2022). Retraction note: A constraint on historic growth in global photosynthesis due to increasing CO₂. *Nature* 606, 420–420. doi: 10.1038/s41586-022-04869-w
- Kumari, M., Verma, S. C., Bhardwaj, S. K., Thakur, A. K., Gupta, R. K., and Sharma, R. (2016). Effect of elevated CO₂ and temperature on growth parameters of pea (*Pisum sativum* L.) crop. *JANS* 8, 1941–1946. doi: 10.31018/jans.v8i4.1067
- Lamichaney, A., Tewari, K., Basu, P. S., Katiyar, P. K., and Singh, N. P. (2021). Effect of elevated carbon-dioxide on plant growth, physiology, yield and seed quality of chickpea (*Cicer arietinum* L.) in Indo-Gangetic plains. *Physiol. Mol. Biol. Plants* 27, 251–263. doi: 10.1007/s12298-021-00928-0
- Laza, H., Baker, J. T., Yates, C., Mahan, J. R., Burrow, M. D., Puppala, N., et al. (2021). Effect of elevated CO₂ on peanut performance in a semi-arid production region (Accessed February 25, 2026).
- Li, F., He, C., Chang, Z., Ma, C., Yu, J., Liu, L., et al. (2023). Effects of elevated carbon dioxide on plant growth and leaf photosynthesis of annual ryegrass along a phosphorus deficiency gradient. *Front. Plant Sci.* 14. doi: 10.3389/fpls.2023.1271262
- Mndela, M., Tjelele, J. T., Madakadze, I. C., Mangwane, M., Samuels, I. M., Muller, F., et al. 2022 A global meta-analysis of woody plant responses to elevated CO₂: implications on biomass, growth, leaf N content, photosynthesis and water relations. Available online at: <https://agris.fao.org/search/ar/records/67053efcb1dfe472e1468056> (Accessed December 10, 2025).
- Mukundan, N. S., Satyamoorthy, K., and Sankar Babu, V. (2024). Investigating photosynthetic evolution and the feasibility of inducing C4 syndrome in C3 plants. *Plant Biotechnol. Rep.* 18, 449–463. doi: 10.1007/s11816-024-00908-2
- Pereyda-González, J. M., De-la-Peña, C., Tezara, W., Zamora-Bustillos, R., Andueza-Noh, R. H., Noh-Kú, J. G., et al. (2022). High temperature and elevated CO₂ modify phenology and growth in pepper plants. *Agronomy* 12. doi: 10.3390/agronomy12081836
- Porras, M. E., Lorenzo, P., Medrano, E., Sánchez-González, M. J., Otálora-Alcón, G., Piñero, M. C., et al. (2017). Photosynthetic acclimation to elevated CO₂ concentration in a sweet pepper (*Capsicum annuum*) crop under Mediterranean greenhouse conditions: Influence of the nitrogen source and salinity. *Funct. Plant Biol. FPB* 44, 573–586. doi: 10.1071/FP16362
- Raines, C. A. (2011). Increasing photosynthetic carbon assimilation in C3 plants to improve crop yield: Current and future strategies. *Plant Physiol.* 155, 36–42. doi: 10.1104/pp.110.168559
- Reich, P. B., Hobbie, S. E., Lee, T. D., and Pastore, M. A. (2018). Unexpected reversal of C3 versus C4 grass response to elevated CO₂ during a 20-year field experiment. *Science* 360, 317–320. doi: 10.1126/science.aas9313
- Sage, R. F. (1994). Acclimation of photosynthesis to increasing atmospheric CO₂: The gas exchange perspective. *Photosynth. Res.* 39, 351–368. doi: 10.1007/BF00014591
- Sendall, K., Muñoz, C., Ritter, A., Rich, R., Noyce, G., and Megonigal, P. (2024). Effects of warming and elevated CO₂ on stomatal conductance and chlorophyll fluorescence of C3 and C4 coastal wetland species. *Wetlands* 44. doi: 10.1007/s13157-024-01780-0
- Shishegaran, A., Shishegaran, A., Najari, M., Ghotbi, A., and Nazem Boushehri, A. (2020). Effect of plants on an environment with high carbon dioxide concentration. *Clean. Eng. Technol.* 1, 100002. doi: 10.1016/j.clet.2020.100002
- Silva, B. E. P., Pires, S. N., Teixeira, S. B., Lucho, S. R., Fagundes, N. S., Centeno, L. H., et al. (2024). Physiological and biochemical responses of pseudocereals with C3 and C4 photosynthetic metabolism in an environment with elevated CO₂. *Plants* 13. doi: 10.3390/plants13233453
- Tang, X., Zhao, J., Zhou, J., Zhu, Q., Sheng, X., and Yue, C. (2024). Elevated CO₂ shifts photosynthetic constraint from stomatal to biochemical limitations during induction in *Populus tomentosa* and *Eucalyptus robusta*. *Plants* 14. doi: 10.3390/plants14010047
- Thompson, M., Gamage, D., Hirotsu, N., Martin, A., and Seneweera, S. (2017). Effects of elevated carbon dioxide on photosynthesis and carbon partitioning: A perspective on root sugar sensing and hormonal crosstalk. *Front. Physiol.* 8. doi: 10.3389/fphys.2017.00578
- Verhage, L. (2021). Model behavior: finding out how to increase photosynthesis in C4 crops. *Plant J.* doi: 10.1111/tpj.15408
- Wang, J., Li, L., Lam, S. K., Shi, X., and Pan, G. (2023). Changes in plant nutrient status following combined elevated [CO₂] and canopy warming in winter wheat. *Front. Plant Sci.* 14. doi: 10.3389/fpls.2023.1132414
- Wang, A., Lv, J., Wang, J., and Shi, K. (2022). CO₂ enrichment in greenhouse production: towards a sustainable approach. *Front. Plant Sci.* 13. doi: 10.3389/fpls.2022.1029901
- Wei, Z., Abdelhakim, L. O. A., Fang, L., Peng, X., Liu, J., and Liu, F. (2022). Elevated CO₂ effect on the response of stomatal control and water use efficiency in amaranth and maize plants to progressive drought stress. *Agric. Water Manage.* 266. doi: 10.1016/j.agwat.2022.107609
- Xu, Z., Jiang, Y., Jia, B., and Zhou, G. (2016). Elevated-CO₂ response of stomata and its dependence on environmental factors. *Front. Plant Sci.* 7. doi: 10.3389/fpls.2016.00657
- Yadav, A., Bhatia, A., Yadav, S., Kumar, V., and Singh, B. (2019). The effects of elevated CO₂ and elevated O₃ exposure on plant growth, yield and quality of grains of two wheat cultivars grown in north India. *Heliyon* 5, e02317. doi: 10.1016/j.heliyon.2019.e02317
- Yang, Q., Li, P., Zhang, D., Lin, W., Hao, X., and Zong, Y. (2023). Effects of elevated CO₂ on the photosynthesis, chlorophyll fluorescence and yield of two wheat cultivars (*Triticum aestivum* L.) under persistent drought stress. *Sustainability* 15. doi: 10.3390/su15021593
- Zhang, N., Berman, S. R., van den Berg, T., Chen, Y., Marcelis, L. F. M., and Kaiser, E. (2024). Biochemical versus stomatal acclimation of dynamic photosynthetic gas exchange to elevated CO₂ in three horticultural species with contrasting stomatal morphology. *Plant Cell Environ.* 47, 4516–4529. doi: 10.1111/pce.15043
- Zhang, Y., Chen, Y., Guo, Y., Ma, Y., Yang, M., Fu, R., et al. (2022). Elevated CO₂ delayed yellowing by maintaining chlorophyll biosynthesis and inhibiting chlorophyll degradation and carotenoid accumulation of postharvest broccoli. *Postharvest Biol. Technol.* 194. doi: 10.1016/j.postharvbio.2022.112089
- Zheng, Y., Li, F., Hao, L., Yu, J., Guo, L., Zhou, H., et al. (2019). Elevated CO₂ concentration induces photosynthetic down-regulation with changes in leaf structure, non-structural carbohydrates and nitrogen content of soybean. *BMC Plant Biol.* 19, 255. doi: 10.1186/s12870-019-1788-9