

RESEARCH

Open Access



Monotropein attenuates apoptosis and pyroptosis in chondrocytes and alleviates osteoarthritis progression in mice

Zhen Li^{1†}, Zhenyue Chen^{2†}, Jiayi Chen^{3†}, Zhutong Liu¹, Zehui Li¹, He Sun¹, Xiaochao Wang¹, Jinqiang Wei¹, Xuewei Cao^{1,4*} and Decai Zheng^{1,5*}

Abstract

Background Osteoarthritis (OA) is a chronic degenerative joint disease characterized by loss of joint function, which seriously reduces the quality of life of the elderly and imposes a heavy socioeconomic burden worldwide. Monotropein (MON), the main active ingredient of *Morinda officinalis* F.C. How, has exhibited therapeutic effects in different disease models. However, its potential effects on chondrocytes in an arthritic model remain unclear. This study aimed to evaluate the effects of MON in chondrocytes and a mouse model of OA, and explore the potential mechanisms.

Materials and methods Murine primary chondrocytes were pretreated with 10 ng/ml interleukin (IL)-1 β for 24 h to establish an in vitro model of OA, and then treated with different concentrations of MON (0, 25, 50 and 100 μ M) for 24 h. The proliferation of the chondrocytes was assayed using ethynyl-deoxyuridine (EdU) staining. Immunofluorescence staining, western blotting and TUNEL staining were performed to assess the effects of MON on cartilage matrix degradation, apoptosis and pyroptosis. The mouse model of OA was constructed by surgical destabilization of the medial meniscus (DMM), and the animals were randomly divided into the sham-operated, OA and OA + MON groups. Following OA induction, the mice were given intraarticular injection of 100 μ M MON or equal volume of normal saline twice a week for 8 weeks. The effects of MON on cartilage matrix degradation, apoptosis and pyroptosis were assessed as indicated.

Results MON significantly accelerated the proliferation of chondrocytes, and inhibited cartilage matrix degradation, apoptosis and pyroptosis in the IL-1 β -stimulated cells by blocking the nuclear factor-kappa B (NF- κ B) signaling pathway. In the mouse model as well, MON treatment alleviated OA progression and promoted cartilage repair by inhibiting cartilage matrix degradation, and chondrocyte apoptosis and pyroptosis through the inactivation of the NF- κ B signaling pathway. Furthermore, the MON-treated arthritic mice exhibited better articular tissue morphology and lower OARSI scores.

Conclusions MON alleviated OA progression by inhibiting cartilage matrix degradation, and the apoptosis and pyroptosis of chondrocytes via NF- κ B pathway inactivation, and is a promising alternative for the treatment of OA.

Keywords Monotropein, Osteoarthritis, Cartilage matrix degradation, Apoptosis, Pyroptosis

[†]Zhen Li, Zhenyue Chen and Jiayi Chen contributed equally to this study

*Correspondence:

Xuewei Cao
caoxuewei2021@163.com
Decai Zheng
sandzheng@163.com

Full list of author information is available at the end of the article



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Introduction

Osteoarthritis (OA) is a chronic degenerative joint disease with pathological features including articular cartilage degeneration, synovial inflammation, and secondary bone hyperplasia [2, 9]. It is characterized by a prolonged disease course, high morbidity and disability rates, and joint pain, dysfunction and deformity, which severely affects quality of life and imposes a heavy socioeconomic burden worldwide [12, 27]. OA is currently managed by basic treatment, pharmacological intervention, restorative treatments and reconstruction [19, 36], depending on the degree of arthritic progression [17]. However, these therapies only mitigate the symptoms, and cannot prevent or reverse the disease course [28]. Therefore, novel treatment strategies are needed to inhibit the development and progression of OA.

Monotropein (MON), a natural iridoid glycoside compound, is the main active ingredient of *Morinda officinalis* F. C. How, which is widely used for treating OA [47]. The plant name has been checked with <http://www.worldfloraonline.org/> (2022. 11. 28). Pharmacological studies have shown that MON exerts anti-inflammatory, antioxidant, anti-apoptotic and autophagic effects. It alleviated inflammation, oxidative stress and apoptosis in senescent endothelial cells and mice with acute kidney injury by inhibiting the nuclear factor-kappa B (NF- κ B) signaling pathway [15, 45]. In addition, Chen et al. reported that MON alleviated secondary liver injury in a mouse model of chronic colitis by inactivating the NF- κ B pathway and NLR Family Pyrin Domain Containing 3 (NLRP3) inflammasome [4]. Furthermore, MON significantly attenuated H₂O₂-induced oxidative stress in osteoblasts by enhancing autophagy [31], and protected endothelial progenitor cells against apoptosis and autophagy [33].

Although MON has shown therapeutic effects in different diseases models, its possible effects on chondrocytes in an arthritic model remain unclear. Therefore, the aim of this study was to evaluate the therapeutic potential of MON in in vitro and in vivo models of OA, and explore the potential mechanisms.

Materials and methods

Primary culture of chondrocytes

Primary chondrocytes were extracted from neonate C57BL/6 mice as previously described [41, 42]. Briefly, the cartilage tissues were isolated from the knee joints under sterile conditions by removing the bone and connective tissues, washed thrice with PBS, and cut into small pieces. The minced cartilage tissues were then homogenized with 0.25% trypsin (Gibco) for 30 min at 37 °C. After removing the supernatant, the tissues were rinsed with PBS and subsequently digested with 0.2% collagenase type II (Sigma, Beijing, China) for 12 h at

37 °C on a shaker. The macerated tissue suspensions were passed through a nylon mesh to remove the residue, and then clarified by centrifuging at 1000 rpm for 5 min. The primary chondrocytes were resuspended in Dulbecco's Modified Eagle Medium (DMEM, Gibco) containing 10% fetal bovine serum (FBS; Gibco), 1% penicillin and streptomycin (Gibco), and seeded into tissue culture flasks. The cells were cultured at 37 °C under 5% CO₂ and saturated humidity, and the medium was changed every 2 days. The second and third passages of chondrocytes were used for in vitro experiments at 80% confluency.

Cell viability assay

MON ($\geq 98\%$ purity) was purchased from Sigma-Aldrich Chemical Corporation (St Louis, MO, USA) and dissolved in 0.9% normal saline. Cell viability was determined using the Cell Counting Kit-8 (CCK-8, KeyGEN, China) [23]. Briefly, primary chondrocytes were seeded into 96-well plates at the density of 5000 cells per well and incubated with different concentrations of MON (0–1000 μ M) for 24 h. According to previous studies on drug concentration and concentration screening in this experiment, the optimal experimental concentration was confirmed [33, 34, 47]. In another experiment, the cells were first stimulated with 10 ng/ml IL-1 β for 24 h, and then treated with MON (0, 25, 50 and 100 μ M) for 24 and 48 h. At the indicated time points, 10 μ l CCK-8 solution was added to the respective wells, and the cells were incubated for 2 h. The absorbance at 450 nm (OD₄₅₀) was measured using a multifunctional microplate reader (Bio-Rad).

Ethynyl-deoxyuridine (EdU) staining

Primary chondrocytes were seeded in 96-well plates, pre-treated with 10 ng/ml IL-1 β for 24 h, and then incubated with MON (0, 25, 50 and 100 μ M) for 24 h. Cell proliferation was assayed using BeyoClick™ EdU-488 Cell Proliferation Kit (Beyotime, Beijing, China) [38, 40]. The EdU-labeled cells were observed under a fluorescence microscope (Olympus IX73), and the proportion of proliferating cells were calculated using Image J.

Establishment of OA model and treatment

Sixty-three 10-weeks-old male specific pathogen-free (SPF) C57BL/6 mice (20–25 g) were supplied by the Experimental Animal Center of Guangzhou University of Chinese Medicine. Fifteen mice were used in the preliminary test and forty-eight were used in the formal test. All animal experiments were conducted in accordance with the internationally accepted principles for laboratory animal use and care as found in the European Community guidelines (EEC Directive of 1986; 86/609/EEC). The OA model was constructed by surgical destabilization

of the medial meniscus (DMM) as previously described [3]. Briefly, after the mice were anesthetized, their right knee was depilated and a 0.5 cm long incision was made along the medial capsule of the right knee. The joint was then exposed and holding the patella firmly, the tissues surrounding the medial meniscus were bluntly dissected. The medial meniscus ligament was then dissected and displaced medially, thereby destabilizing the tissue. After suturing and sterilizing the wounds, the mice were placed under an electric blanket till they regained consciousness.

The animals were randomly divided into the sham-operated, OA and OA + MON groups. Consistent with *in vitro* experiments, three concentration doses (25, 50 and 100 μ M) were used to perform the animal pretest. According to the pre-experimental results, we found that the concentration of 100 μ M MON showed significant statistical difference, so we used the single-dose concentration to illustrate the effect of MON. The mice in the OA + MON group were given intra-articular injections of 100 μ M MON twice a week for 8 weeks after OA induction, and 10 μ L of the solution was slowly injected into right knees. Animals in the sham-operated and OA groups received equal volume of normal saline. After 8 weeks of MON treatment, all mice were sacrificed and the right knee joint was excised for further evaluation.

Histological assessment

After eight weeks of MON treatment, the animals were euthanized and the right knee tissue was fixed with 4% PFA and then decalcified with ethylenediaminetetraacetic acid (EDTA). After decalcification, the tissues were dehydrated through an ethanol gradient (70%, 80%, 95–100%), cleared with xylene, and embedded in paraffin. The wax blocks were sliced into 5 μ m-thick sections that were baked overnight in an oven at 37 °C. The sections were dewaxed and stained 0.2% Safranin O solution for 15 min, rinsed with distilled water, counterstained with 0.2% Fast Green solution for 5 min (Sigma-Aldrich, USA), and sealed. The structural changes in the cartilage tissues were evaluated in a blinded manner according to the Osteoarthritis Research Society International (OARSI) scoring system [6]. Hematoxylin–eosin (HE) staining was performed as per standard protocols, and the inflammation around the lesions was scored. The slides were observed using the Olympus IX73 microscope.

Western blotting

The cartilage tissues were sonicated in the radio immunoprecipitation assay (RIPA) lysis buffer (Gibco, Grand Island, NY, USA), and centrifuged twice at 12000 g for 10 min at 4 °C. The protein concentration in the supernatants was measured using a bicinchoninic acid (BCA) kit (Bio-Rad Laboratories, CA, USA). Twenty micrograms

protein per sample was diluted in the loading buffer and denatured at 99 °C for 10 min. The protein samples were separated by 10% SDS-PAGE, and then transferred to polyvinylidene difluoride (PVDF) membranes. After blocking with 5% skimmed milk, the blots were incubated overnight with primary antibodies targeting matrix metalloproteinase 3 (MMP3, Proteintech, 66338-1-Ig), MMP13 (Proteintech, 18165-1-AP), collagen II (Abcam, ab34712), SOX9 (Proteintech, 67439-1-Ig), Bcl-2 (CST, 15071), Bax (CST, 41162), cleaved caspase-3 (CST, 9661), NLRP3 (Affinity, DF7438), apoptosis-associated speck-like protein containing a CARD (ASC, CST, 67824), cleaved caspase-1 (CST, 89332), gasdermin D (GSDMD, Affinity, AF4012), phosphorylated I κ B α (p-I κ B α , Affinity, AF2002), I κ B α (Affinity, AF5002), p-p65 (CST, 3033), p65 (CST, 8242) and GAPDH (CST, 5174) at 4 °C. All primary antibodies were used at 1:1000 dilution. The membranes were incubated with the secondary antibody (Beyotime, 1:1000, A0208 anti-rabbit or A0216 anti-mouse) for 90 min at room temperature (RT). The positive bands were visualized using a ChemiDoc™ MP Imaging System (Bio-Rad), and the relative intensities of each band were determined using Image J (National Institutes of Health, Bethesda, MD).

Immunofluorescence staining

Immunofluorescence was performed as per standard protocols. The suitably treated chondrocytes were fixed with 4% PFA for 30 min and permeabilized in 0.3% Triton X-100 at RT for 30 min. Tissue sections were treated with xylene and alcohol gradient, and heated in citric acid for antigen retrieval. After incubating with 10% serum for 1 h at RT to block non-specific binding, the cells or tissue sections were incubated overnight with primary antibodies against collagen II (1:200, Abcam, ab34712), MMP13 (1:200, Proteintech, 18165-1-AP), NLRP3 (1:200, Affinity, DF7438), GSDMD (1:100, Affinity, AF4012), p-I κ B α (1:200, Affinity, AF2002) and p-p65 (1:1000, CST, 3033) at 4 °C. The following day, the cells or tissue sections were incubated with fluorochrome-conjugated rabbit or mouse secondary antibodies (1:300, Invitrogen, A-11008 anti-rabbit or A-11001 anti-mouse), and mounted with DAPI medium (Solarbio). The samples were observed under a fluorescence microscope (Olympus IX73), and the fluorescence intensity of the positively stained cells or tissue area was measured using Image J.

TUNEL assay

TUNEL staining was performed using the Apoptosis Detection Kit (Yeasen Biotech, Shanghai, China) [10]. Briefly, cells or tissue sections were fixed for 25 min and then permeabilized with proteinase K (20 μ g/ml) for 20 min. After incubating with TdT for 1 h, the cells

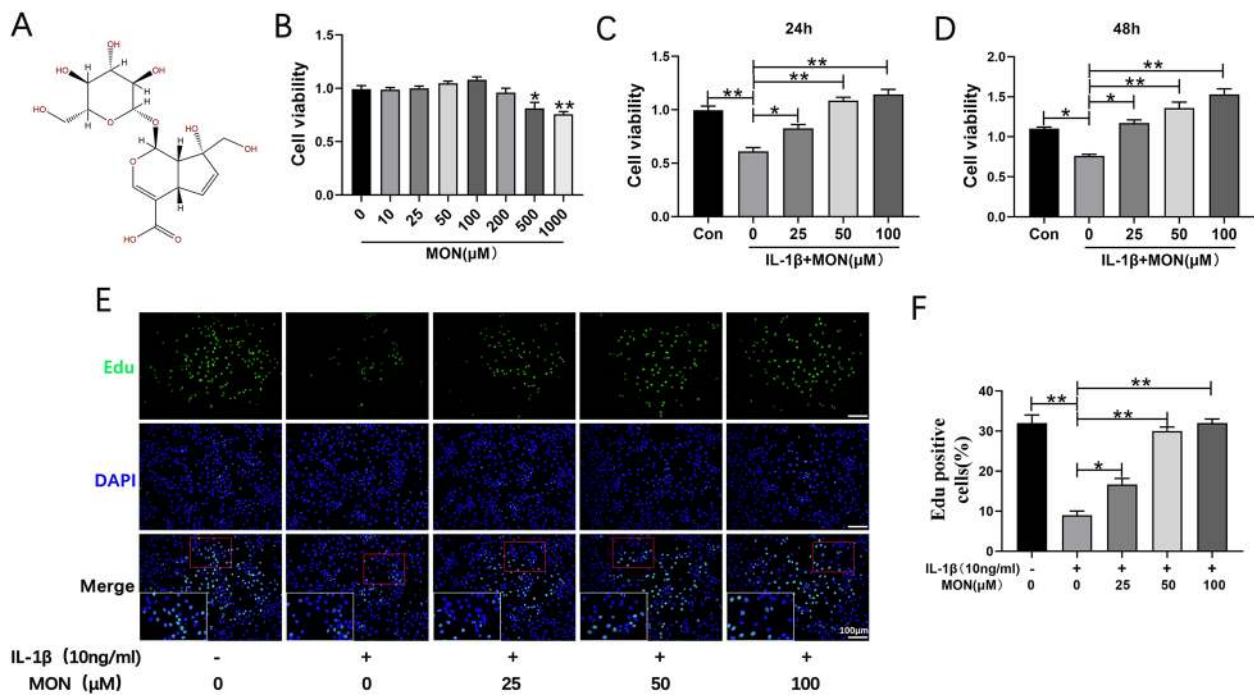


Fig. 1 MON enhanced the survival of IL-1 β -treated chondrocytes. **A** The molecular structure of MON. **B** The cell counting kit-8 assay showing the cell viability in different concentration of MON (0–1000 μ M) for 24 h. **C, D** The cell counting kit-8 assay showing the cell viability of chondrocytes pretreatment with or without 10 ng/ml IL-1 β for 24 h, followed by treatment with different concentration of MON (0, 25, 50 and 100 μ M) for 24 h or 48 h. **E** Edu staining showing the proliferation of chondrocytes pretreated with or without 10 ng/ml IL-1 β for 24 h, followed by treatment with different concentration of MON (0, 25, 50 and 100 μ M) for another 24 h. **F** Quantification of Edu positive cells. All experiments were performed in triplicated and data were presented as the mean $\pm$ SD, $n = 3$ per group. * $P < 0.05$, ** $P < 0.01$

or tissue sections were counterstained with DAPI. The number and proportion of apoptotic cells were evaluated under a fluorescence microscope (Olympus IX73).

Statistical analysis

The SPSS 20.0 software (SPSS, Chicago, IL, USA) was used for statistical analyses. Data are presented as mean $\pm$ standard deviation (SD). One-way analysis of variance was used to analyse multiple groups, and unpaired Student's t -test was used to analyse differences between two groups.

$P < 0.05$ was considered as statistically significant.

Results

MON enhanced the survival of IL-1 β -treated chondrocytes

To determine the effects of MON (chemical structure is shown in Fig. 1) on chondrocyte viability in vitro, primary mouse chondrocytes were treated with the different concentrations of MON with or without prior IL-1 β stimulation. As shown in Fig. 1B, MON was non-toxic to the mouse chondrocytes at concentrations lower than 200 μ M. Accordingly, 25, 50 and 100 μ M doses were used in subsequent experiments. IL-1 β stimulation markedly reduced the viability of chondrocytes, which was reversed by MON in a dose- and time-dependent manner (Fig. 1C, D). Consistent with this, Edu incorporation assay showed that MON significantly increased the proportion of the proliferating IL-1 β -stimulated

(See figure on next page.)

Fig. 2 MON inhibited IL-1 β -stimulated cartilage matrix degradation and apoptosis of chondrocytes. **A** Co-immunofluorescence staining of Collagen II (green)/MMP13 (red) and relative quantification of Collagen II or MMP13 positive cells in IL-1 β -induced chondrocytes after MON treatment. **B** Western blot analysis and relative quantification showing the expression levels of cartilage matrix degradation related proteins (MMP3, MMP13, Collagen II and SOX9) in IL-1 β -induced chondrocytes after MON treatment. **C** TUNEL staining showing the chondrocyte apoptosis and quantitative estimation of the number of TUNEL positive cells in IL-1 β -induced chondrocytes after MON treatment. **D** Western blot analysis and relative quantification showing the expression levels of apoptosis related proteins (Bcl-2, Bax and Cleaved caspase-3) in IL-1 β -induced chondrocytes after MON treatment. All experiments were performed in triplicated and data were presented as the mean $\pm$ SD, $n = 3$ per group. ## $P < 0.01$ versus control group; * $P < 0.05$, ** $P < 0.01$ versus IL-1 β group

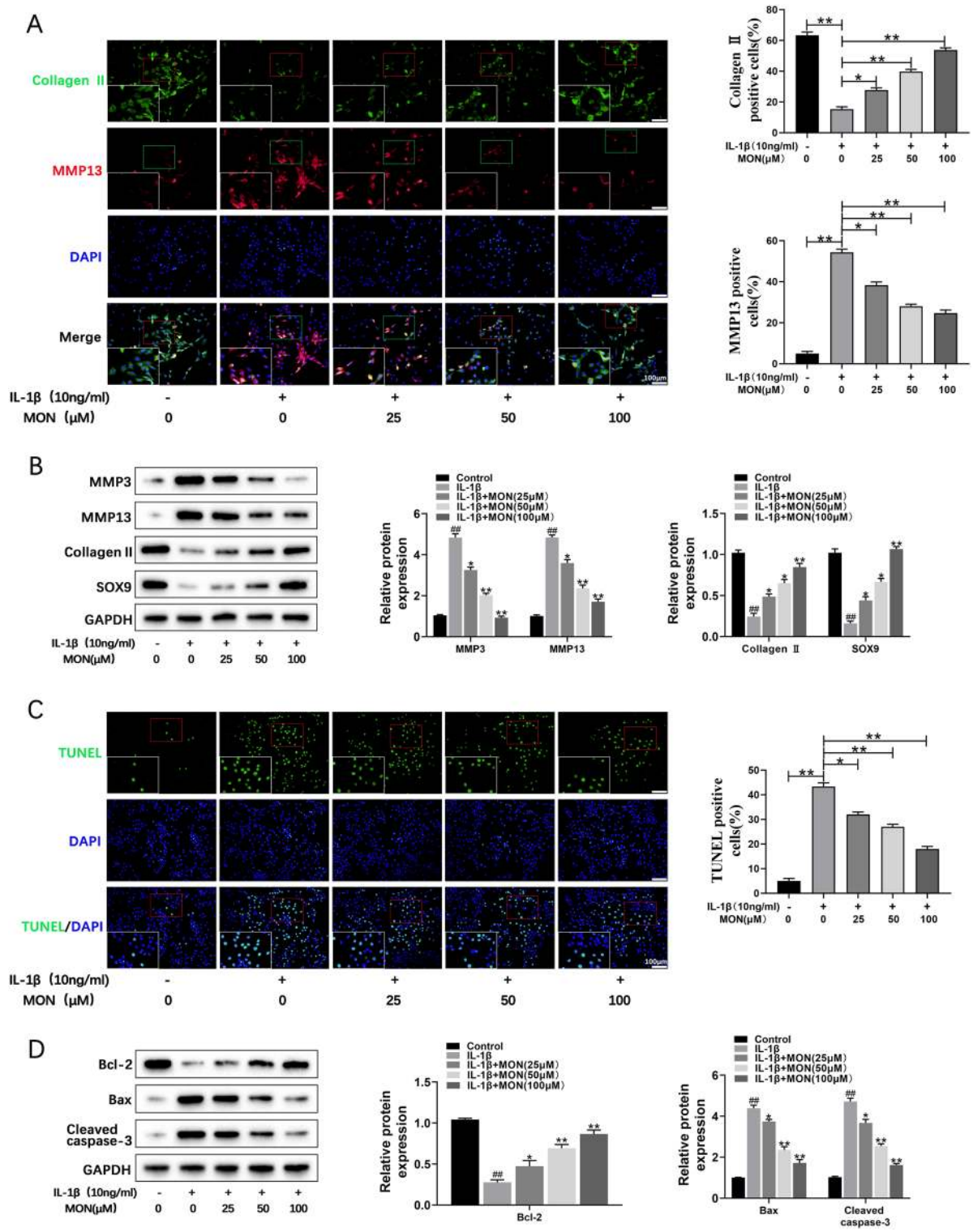


Fig. 2 (See legend on previous page.)

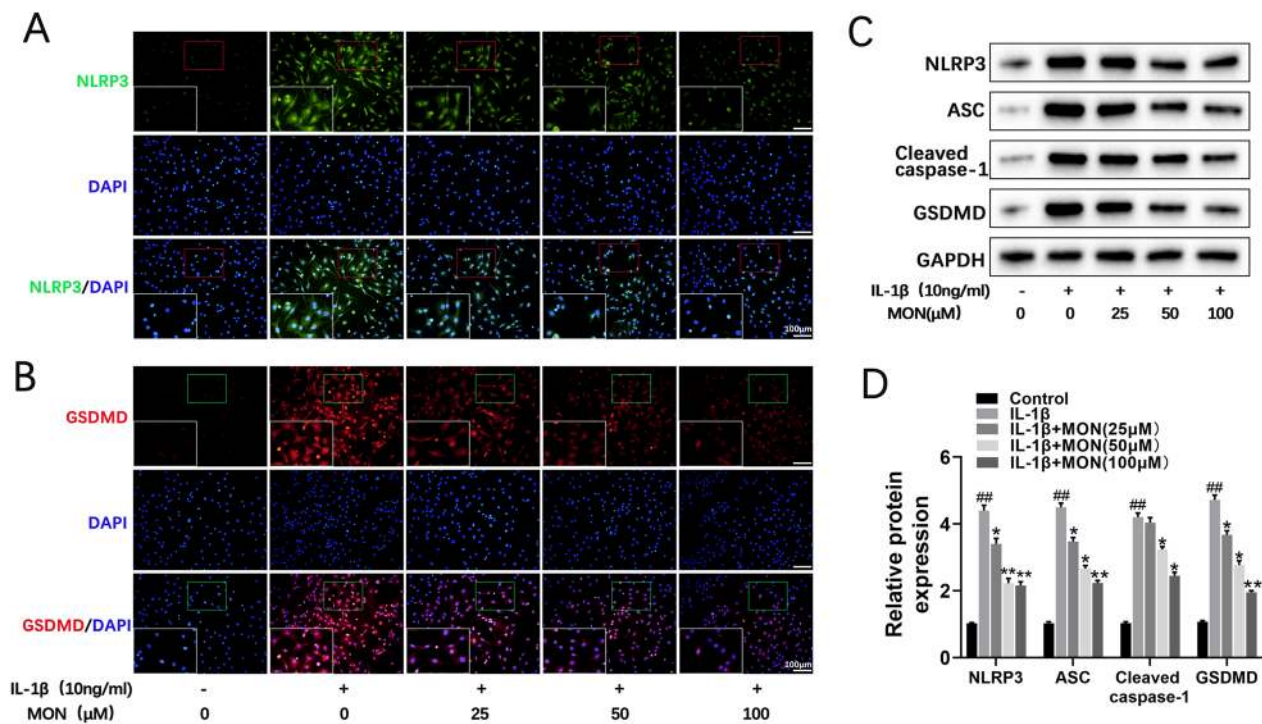


Fig. 3 MON inhibited IL-1 β -induced pyroptosis in chondrocytes. **A, B** Immunofluorescence staining of NLRP3 (green) and GSDMD (red) were performed respectively in IL-1 β -induced chondrocytes after MON treatment. **C, D** Western blot analysis and relative quantification showing the expression levels of pyroptosis related proteins (NLRP3, ASC, Cleaved caspase-1 and GSDMD) in IL-1 β -induced chondrocytes after MON treatment. All experiments were performed in triplicated and data were presented as the mean $\pm$ SD, $n = 3$ per group. $^{##}P < 0.01$ versus control group; $^{*}P < 0.05$, $^{**}P < 0.01$ versus IL-1 β group

chondrocytes in a dose-dependent manner (Fig. 1E, F). In summary, MON treatment protected the chondrocytes against IL-1 β , which is the major effector of the NLRP3 inflammasome.

MON inhibited IL-1 β -stimulated cartilage matrix degradation and apoptosis of chondrocytes

To further evaluate the protective effects of MON, we treated the IL-1 β -stimulated primary chondrocytes with 0, 25, 50 and 100 μ M MON for 24 h. As shown in Fig. 2A, IL-1 β treatment decreased the in situ expression of collagen II and increased that of MMP13 in the primary chondrocytes. However, MON restored the levels of both matrix proteins in a dose-dependent manner. Analysis of the total protein fraction from the IL-1 β -stimulated chondrocytes further showed that MON increased the levels of collagen II and SOX9, and decreased that of MMP3 and MMP13 (Fig. 2B). These findings suggested that MON can effectively reverse cartilage matrix degradation during chronic inflammation. TUNEL staining further showed that the number of apoptotic cells increased markedly after IL-1 β treatment, which was mitigated by MON in a dose-dependent manner compared to the controls (Fig. 2C). Consistent with this, the

anti-apoptotic Bcl-2 protein was significantly down-regulated, whereas the pro-apoptotic Bax and cleaved caspase-3 proteins were upregulated following IL-1 β exposure. MON treatment not only restored the expression levels of Bcl-2 but also downregulated Bax and cleaved caspase-3 (Fig. 2D). Taken together, MON inhibited IL-1 β -stimulated degradation of the cartilage matrix and apoptosis of chondrocytes in vitro.

MON inhibited IL-1 β -induced pyroptosis in chondrocytes

Pyroptosis is a pro-inflammatory type of programmed cell death mediated by the gasdermin (GSDMD) protein [41, 42]. The binding of NLRP3 to the adapter protein ASC cleaves caspase-1, and activates IL-1 β and IL-18. Furthermore, GSDMD cleavage results in the formation of membrane pores that release the inflammatory factors, eventually leading to pyroptosis [20, 30]. We found that IL-1 β significantly increased the expression of NLRP3 and GSDMD in the primary chondrocytes, whereas MON treatment led to their downregulation in a dose-dependent manner (Fig. 3A, B). Western blotting further showed that IL-1 β treatment markedly increased the expression of NLRP3, ASC, cleaved caspase-1 and GSDMD, which was reversed by MON in a

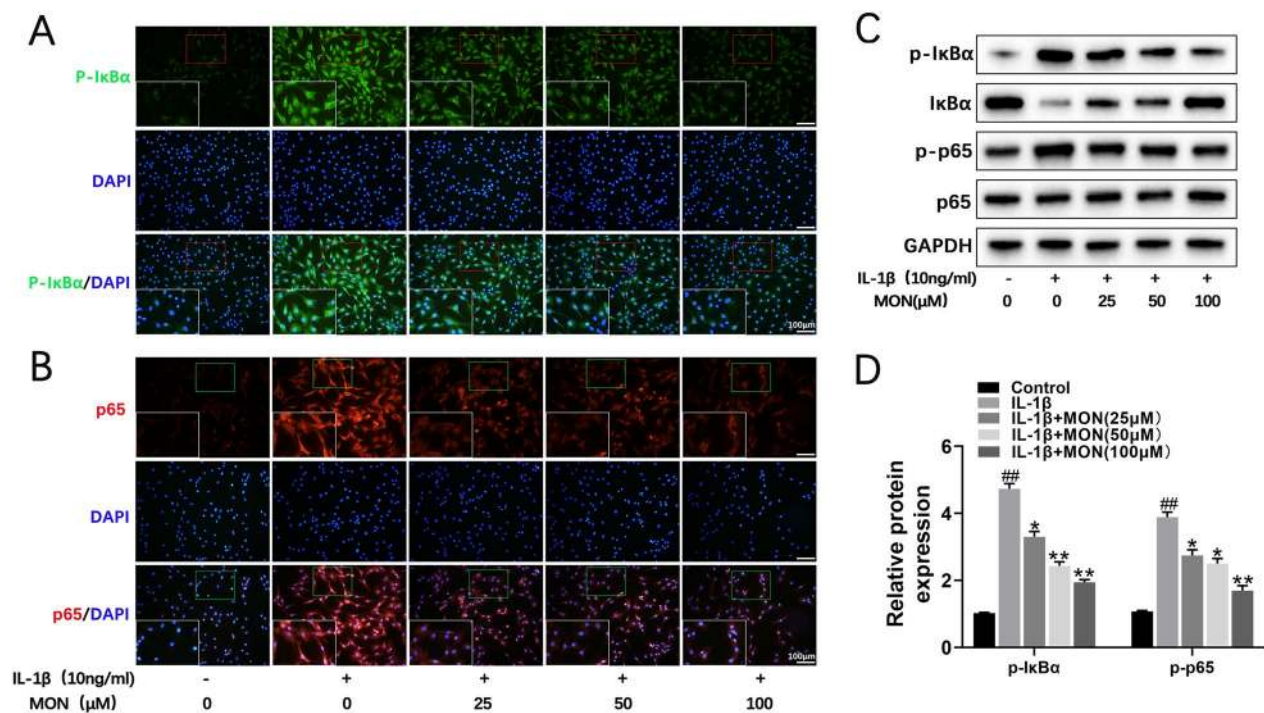


Fig. 4 MON inhibited the NF- κ B signaling pathway in chondrocytes. **A, B** Immunofluorescence staining showing the phosphorylation of I κ B α (green) and the nuclear translocation of p65 (red) in IL-1 β -induced chondrocytes after MON treatment. **C, D** Western blot analysis and relative quantification showing the expression levels of NF- κ B signaling pathway related proteins (p-I κ B α , I κ B α , p-p65 and p65) in IL-1 β -induced chondrocytes after MON treatment. All experiments were performed in triplicated and data were presented as the mean $\pm$ SD, $n = 3$ per group. ## $P < 0.01$ versus control group; * $P < 0.05$, ** $P < 0.01$ versus IL-1 β group

dose-dependent manner (Fig. 3C, D). Collectively, these findings indicated that MON treatment can attenuate OA progression by inhibiting IL-1 β -induced pyroptosis in chondrocytes.

MON inhibited the NF- κ B signaling pathway in chondrocytes

Activation of the IKK complex phosphorylates the NF- κ B inhibitor protein I κ B α , which leads to its proteasomal degradation and the translocation of NF- κ B dimers to the nucleus to drive transcription of target genes [21, 44, 46]. IL-1 β treatment not only increased the expression of p-I κ B α (Fig. 4A) but also promoted nuclear translocation of p65 (Fig. 4B) in the primary chondrocytes, whereas MON reversed these effects of IL-1 β in a dose-dependent manner (Fig. 4A, B). Furthermore, IL-1 β also upregulated p-I κ B α and p-p65 and activated the NF- κ B pathway, while MON decreased the phosphorylation of I κ B α and p65 without affecting the total p65 levels (Fig. 4C, D). Taken together, MON can inactivate the NF- κ B pathway in IL-1 β -stimulated chondrocytes.

The protective effects of MON are mediated through the inactivation of NF- κ B signaling

To next determine whether the NF- κ B pathway is critical to the effects of MON on the cartilage matrix proteins and chondrocytes, we stimulated the primary chondrocytes with 10 ng/ml IL-1 β and 20 μ M recombinant NF- κ B (R-NF- κ B, Cloud-Clone Corp) before treating them with 100 μ M MON. Recombinant NF- κ B reversed the effects of MON on MMP3, MMP13, collagen II and SOX9 (Fig. 5A), as well as on the expression levels of apoptosis markers (Fig. 5B) in the IL-1 β -stimulated cells. Furthermore, recombinant NF- κ B also enhanced the expression of pyroptosis proteins including NLRP3, ASC, cleaved caspase-1 and GSDMD despite MON treatment (Fig. 5C). Collectively, these findings demonstrated that MON inhibited IL-1 β -induced cartilage matrix degradation, and the apoptosis and pyroptosis of chondrocytes by targeting the NF- κ B pathway.

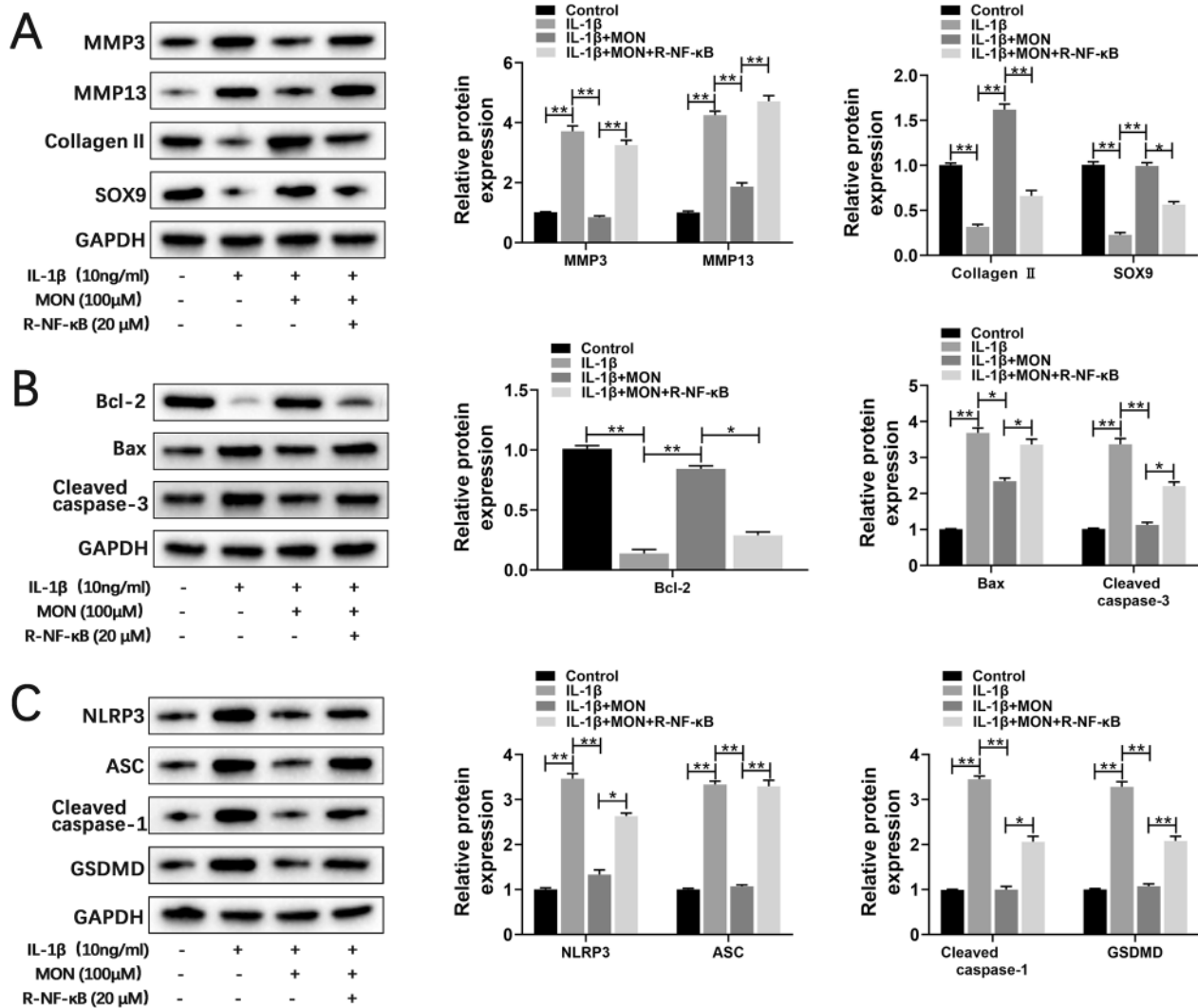


Fig. 5 The protective effects of MON are mediated through the inactivation of NF- κ B signaling. **A–C** Western blot analysis and relative quantification showing the expression levels of cartilage matrix degradation related proteins (MMP3, MMP13, Collagen II and SOX9), apoptosis related proteins (Bcl-2, Bax and Cleaved caspase-3) and pyroptosis related proteins (NLRP3, ASC, Cleaved caspase-1 and GSDMD) in IL-1 β -induced chondrocytes after MON and recombinant NF- κ B protein treatment. All experiments were performed in triplicated and data were presented as the mean $\pm$ SD, $n = 3$ per group. * $P < 0.05$, ** $P < 0.01$

MON alleviated arthritic progression and promoted cartilage repair in the DMM model by targeting the NF- κ B pathway

The pre-experimental results suggested that the concentration of 100 μ M MON showed significant statistical difference, the single-dose concentration was used in subsequent animal experiments (Fig. 6A–D). To determine the effects of MON on OA progression in vivo, the mice were injected with 100 μ M MON twice a week into the operated knee for 8 weeks after OA induction (Fig. 6E). Safranin O-Fast Green and HE staining of the knee joint sections of arthritic mice indicated that DMM eroded the articular cartilage, which was attenuated

by MON treatment (Fig. 6F). Furthermore, the OARSI scores also decreased significantly in the MON-treated mice compared to the untreated controls, which confirmed that MON reversed the cartilage damage associated with OA (Fig. 6G). Consistent with the gross findings, collagen II deposition decreased significantly and the number of MMP13-positive cells increased in the OA mice. However, MON treatment increased collagen II levels and decreased that of MMP13 (Fig. 7A–D). Analysis of the total protein levels of the cartilage tissues also indicated that MON treatment reversed the increase in MMP3 and MMP13, and the decrease in collagen II and SOX9 in the arthritic mice (Fig. 7E–G), which was

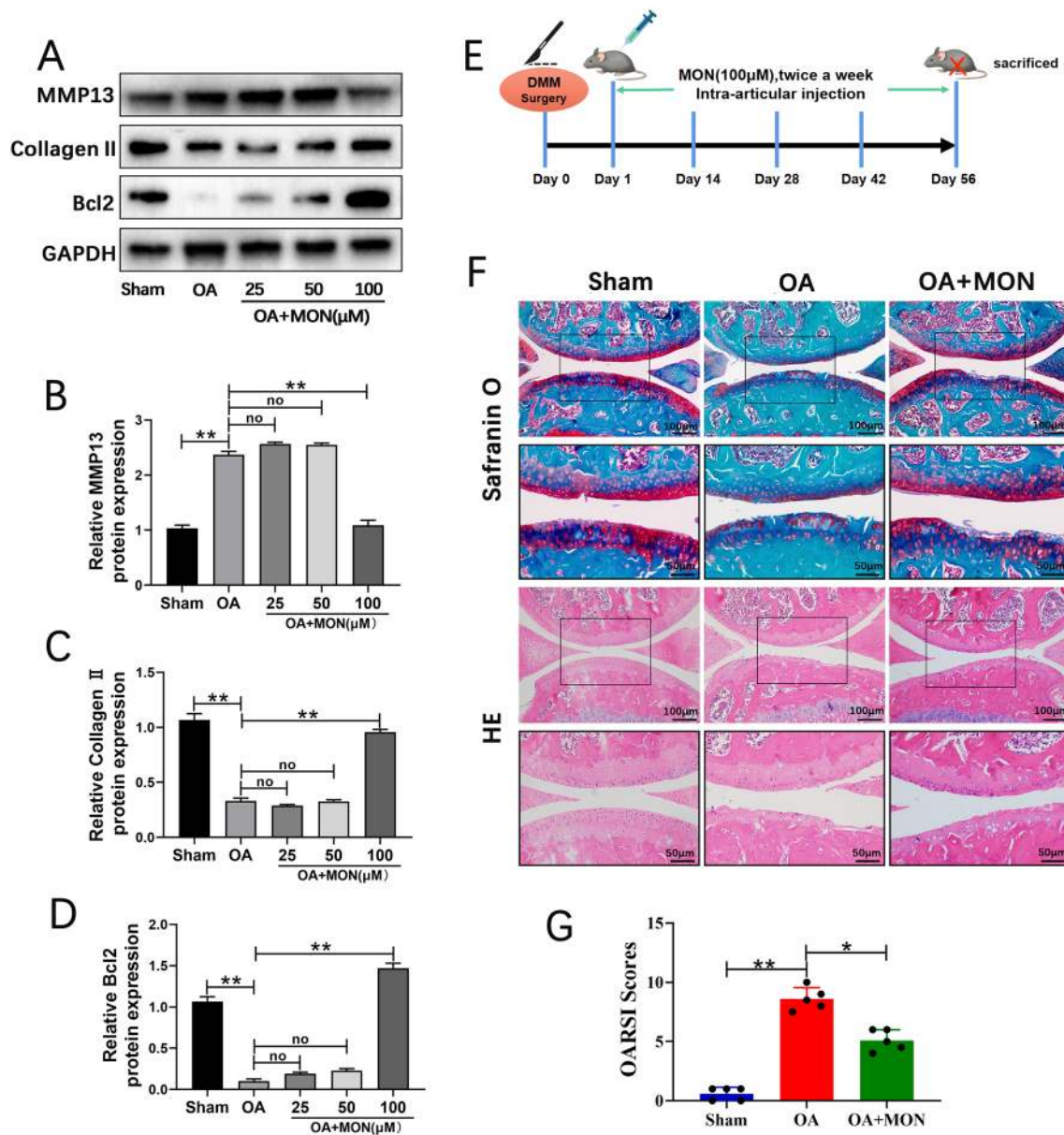


Fig. 6 MON alleviated OA progression and promoted cartilage repair in DMM mice. **A–D** The pre-experimental results suggested that the concentration of 100 μ M MON showed significant statistical difference. **E** DMM mice model and MON treatment protocol. **F** Safranin O-Fast Green and HE staining of mice knee joints were performed to assess the histological morphology of the different groups at 56 days after MON treatment. **G** The OARSI scores of the different groups were calculated respectively at 56 days after MON treatment. All experiments were performed in triplicated and data were presented as the mean $\pm$ SD, $n = 3$ per group. * $P < 0.05$, ** $P < 0.01$

consistent with the in vitro results. TUNEL staining of the cartilage sections further showed that a significant increase in the proportion of apoptotic chondrocytes was observed in the OA group, which was accompanied by elevated Bax and cleaved caspase-3 levels, and the down-regulation of Bcl2. Intra-articular injection of MON alleviated chondrocyte apoptosis and restored the levels of the related markers (Fig. 8A–E). In line with the in vitro

results, the pyroptosis-related markers including NLRP3, ASC, cleaved caspase-1 and GSDMD were elevated significantly in the OA mice compared to the sham-operated controls, and were reversed by MON treatment (Fig. 9A–G). Taken together, MON treatment alleviated the structural damage to the cartilage due to DMM and promoted chondrocyte survival, thereby halting arthritic progression.

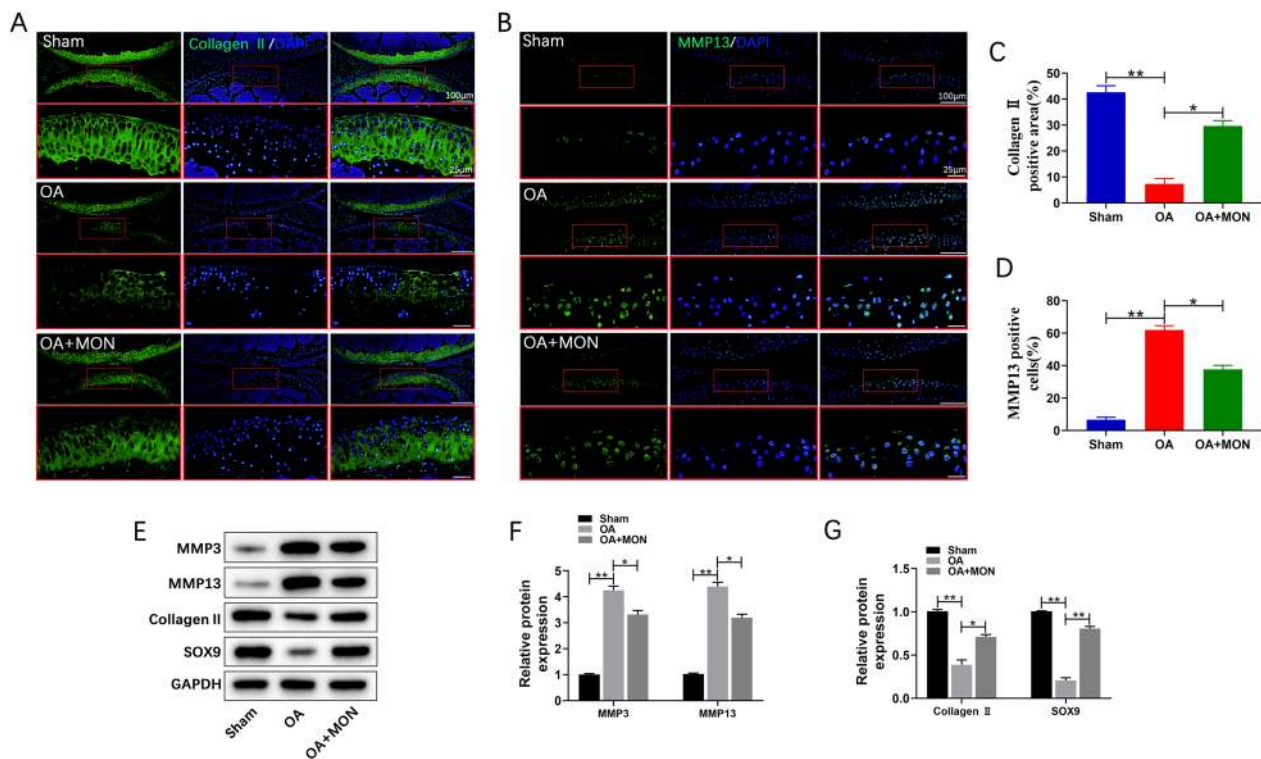


Fig. 7 MON inhibited cartilage matrix degradation in DMM mice. **A, B** Immunofluorescence staining of Collagen II and MMP13 were performed respectively in mice knee joints at 56 days after MON treatment. **C, D** The relative quantification analysis of Collagen II positive area and MMP13 positive cells. **E–G** Western blot analysis and relative quantification showing the expression levels of cartilage matrix degradation related proteins (MMP3, MMP13, Collagen II and SOX9) in DMM mice at 56 days after MON treatment. All experiments were performed in triplicated and data were presented as the mean $\pm$ SD, $n = 3$ per group. * $P < 0.05$, ** $P < 0.01$

To ascertain the molecular basis of the effects of MON in vivo, we analyzed I κ B α and p65 activation in the articular cartilage of the arthritic mice. As expected, DMM significantly increased the levels of p-I κ B α and p-p65, as well as the nuclear translocation of p65 compared to the sham-operated group, which was reversed by MON treatment (Fig. 10A, B). Furthermore, MON treatment decreased the expression of p-I κ B α and p-p65 without affecting total p65 levels in the knee joints of arthritic mice (Fig. 10C, D). In summary, MON protected against cartilage erosion after DMM by targeting the NF- κ B signaling pathway. Moreover, A schematic diagram illustrating the theory in the current study can be found in Fig. 11.

Discussion

Osteoarthritis (OA) is a chronic inflammatory disease of the joints that is characterized by the destruction of articular cartilage [37]. Extracellular matrix degradation, chondrocyte apoptosis and pyroptosis form the pathological basis of OA progression. The current treatment strategies for OA can only provide transient relief in the symptoms but cannot effectively prevent disease

occurrence or progression [18, 28]. Hence, it is essential to develop therapeutic strategies that can enhance cartilage repair and block OA progression.

MON, a bioactive compound of the medicinal herb *Morinda officinalis*, has documented anti-inflammatory, anti-oxidant, anti-apoptotic and autophagic effects. MON alleviated inflammation, oxidative stress and apoptosis associated with acute kidney injury by inhibiting NF- κ B signaling [45]. In addition, MON attenuated liver injury in a mouse model via inactivation of the NF- κ B pathway and NLRP3 inflammasome [4], and also decreased H₂O₂-induced oxidative stress by enhancing autophagy in osteoblasts [31]. Furthermore, previous studies have proved that MON could exert protective effects against IL-1 β -induced apoptosis and catabolic responses in chondrocytes [34]. In the present study, we found that MON inhibited cartilage matrix degradation and promoted the survival of chondrocytes in the in vitro and in vivo models of OA by targeting the NF- κ B signaling pathway.

Excessive degradation of articular cartilage plays a crucial part in the pathogenesis of OA [25, 48]. The imbalance between extracellular matrix synthesis

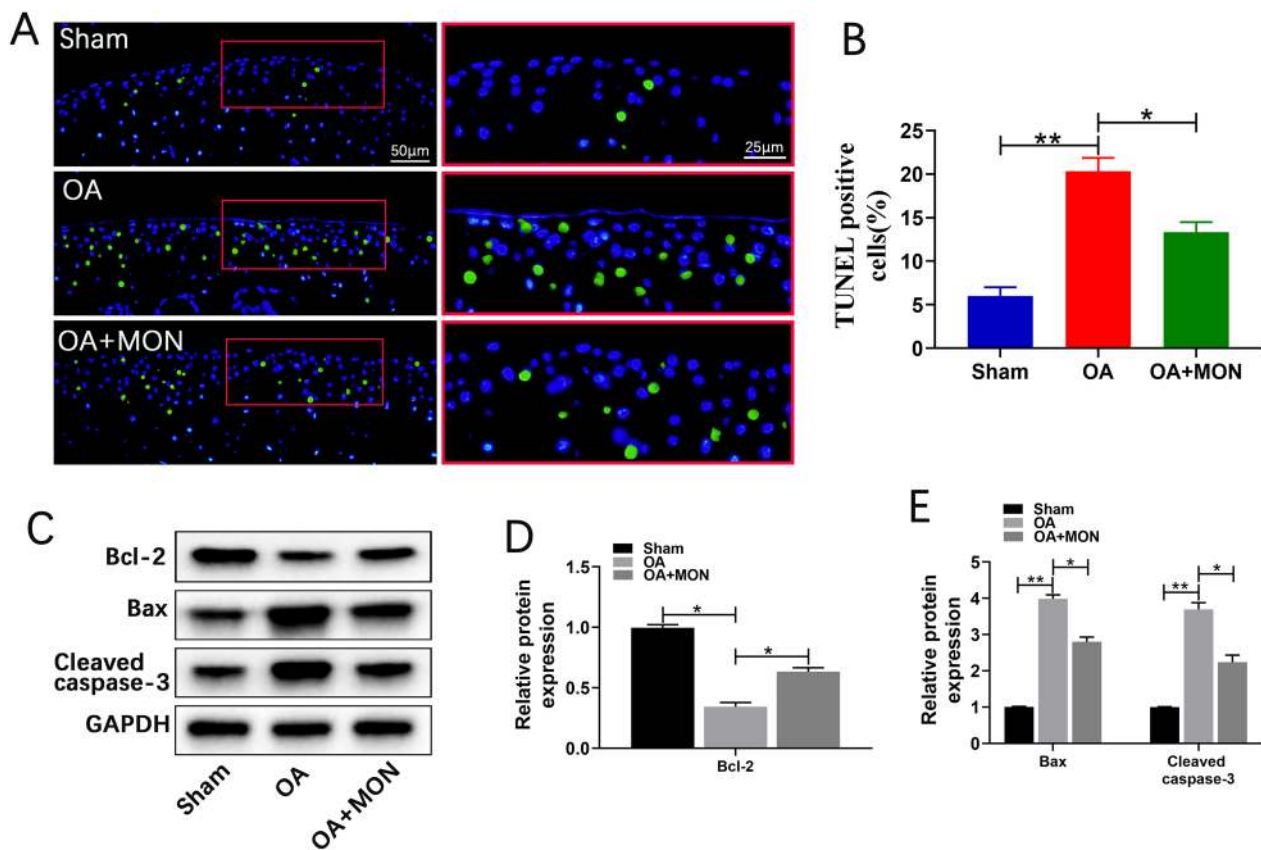


Fig. 8 MON inhibited chondrocyte apoptosis in DMM mice. **A, B** TUNEL staining showing the chondrocyte apoptosis and quantitative estimation of the number of TUNEL positive cells in mice knee joints at 56 days after MON treatment. **C–E** Western blot analysis and relative quantification showing the expression levels of apoptosis related proteins (Bcl-2, Bax and Cleaved caspase-3) in DMM mice at 56 days after MON treatment. All experiments were performed in triplicated and data were presented as the mean $\pm$ SD, $n = 3$ per group. * $P < 0.05$, ** $P < 0.01$

and degradation is the direct cause of articular cartilage erosion [8, 29]. OA is characterized by progressive destruction of cartilage, culminating in complete loss of chondrocytes. Moreover, collagen II and MMPs are thought to be the major contributors to cartilage degeneration during OA pathogenesis [25, 26]. Therefore, inhibiting cartilage matrix degradation and restoring its synthesis may ameliorate OA progression. We found that MON treatment upregulated the matrix proteins collagen II and SOX9, and down-regulated MMP3 and MMP13 in the IL-1 β -stimulated chondrocytes as well as in the articular cartilage of OA mice. Thus, MON can mitigate OA progression by inhibiting cartilage matrix degradation. These findings are in accordance with previous studies indicating that MON upregulated COL2A1 and downregulated MMPs in the chondrocytes [34].

The loss of chondrocytes in the arthritic joints is attributed to apoptosis, which also results in the destruction of the cartilage homeostasis and the degeneration of articular cartilage [13, 32]. Therefore, promoting chondrocyte

survival and inhibiting apoptosis can inhibit cartilage degeneration. MON has been previously demonstrated to attenuate apoptosis in IL-1 β -stimulated chondrocytes [34]. Similar results were obtained in our study as well, and the anti-apoptotic effects of MON were accompanied by an increase in the expression of Bcl-2, and decrease in Bax and cleaved caspase-3 levels in the IL-1 β -induced chondrocytes and mouse model. Thus, MON alleviated OA progression by inhibiting chondrocyte apoptosis. This is consistent with a previous finding that MON inhibited apoptosis in senescent endothelial cells and an acute kidney injury mice model by targeting NF- κ B signaling [15, 45].

Pyroptosis, a pro-inflammatory type of programmed cell death mediated by the NLRP3 inflammasome, has been implicated in the pathological progression of OA [24, 39]. Activation of NLRP3 cleaves the caspase-1 precursor to accelerate the release of inflammatory factors into the extracellular space [1]. Pro-inflammatory factors produced during pyroptosis of chondrocytes or synovial macrophages can directly lead to synovial

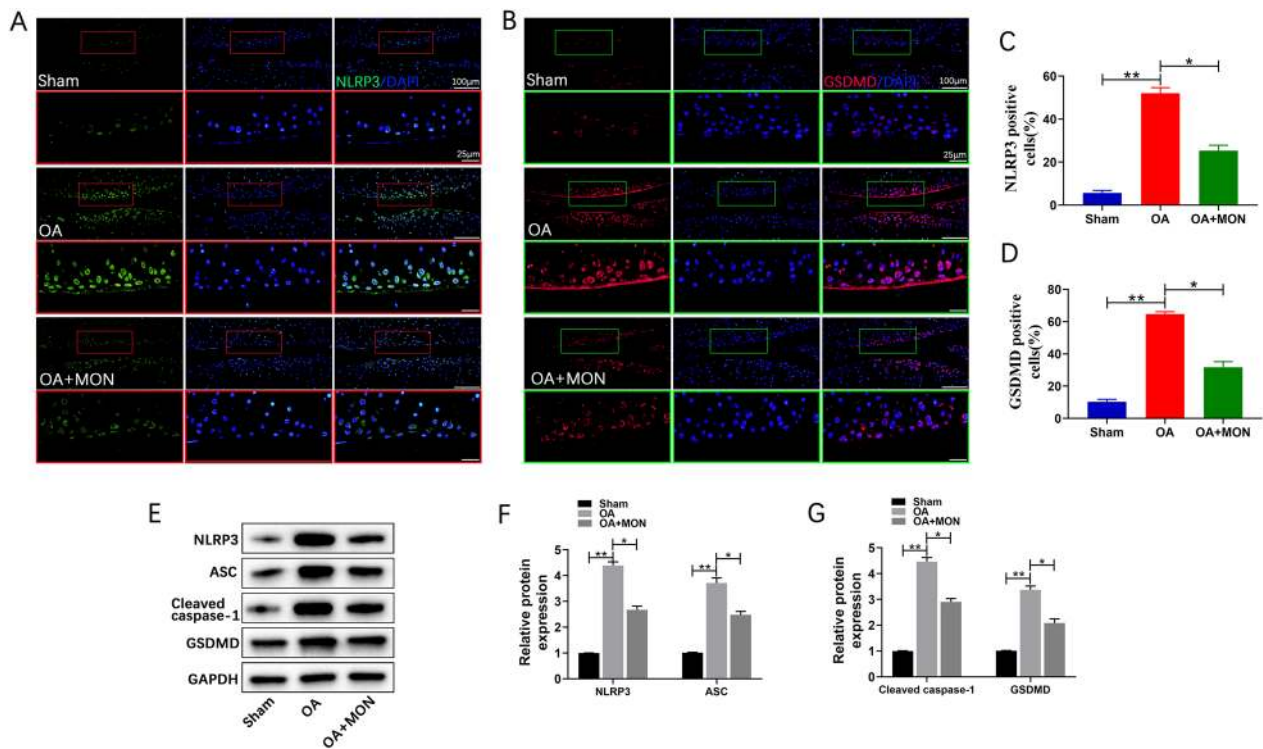


Fig. 9 MON inhibited chondrocyte pyroptosis in DMM mice. **A, B** Immunofluorescence staining of NLRP3 (green) and GSDMD (red) were performed respectively in mice knee joints at 56 days after MON treatment. **C, D** The relative quantification analysis of NLRP3 and GSDMD positive cells. **E–G** Western blot analysis and relative quantification showing the expression levels of pyroptosis related proteins (NLRP3, ASC, Cleaved caspase-1 and GSDMD) in DMM mice at 56 days after MON treatment. All experiments were performed in triplicated and data were presented as the mean $\pm$ SD, $n = 3$ per group. * $P < 0.05$, ** $P < 0.01$

inflammation and the degradation of cartilage matrix, and thus promote OA progression [1, 43]. MON treatment dramatically down-regulated the pyroptosis-related proteins in IL-1 β -stimulated chondrocytes and DMM-induced mice, suggesting that inhibition of chondrocyte pyroptosis may be a potential mechanism by which MON alleviates OA progression. Consistent with these results, MON attenuated liver injury by down-regulating the activity of NLRP3 inflammasome [4]. Furthermore, Zhang et al. also reported that the NLRP3 inhibitor CY-09 protected chondrocytes against inflammation and alleviated OA progression by blocking NLRP3 inflammasome-mediated pyroptosis [44, 46]. Another study showed that Icaritin alleviated LPS-induced chondrocyte pyroptosis by inhibiting NLRP3/caspase-1 signaling pathway, and mitigated the symptoms of OA [49]. Therefore, the NLRP3 inflammasome is a promising therapeutic target for OA.

Apoptosis is a process of programmed cell death that does not cause any inflammatory response, in contrast, pyroptosis is the programmed inflammatory cell death mediated by the NLRP3 inflammasome and releases plenty of pro-inflammatory factors [1, 13, 39]. It has been

proved that apoptosis and pyroptosis of chondrocytes mediate inflammation of cartilage or synovium, aggravate cartilage matrix degradation, break the dynamic balance of extracellular matrix synthesis and degradation, and aggravate the development of OA [13, 39]. Thus, chondrocyte apoptosis and pyroptosis as well as cartilage matrix degradation are crucial pathological changes in OA.

Numerous previous studies have demonstrated that the NF- κ B pathway is a classical inflammatory pathway involved in OA development [5]. Activation of the NF- κ B pathway culminates in the production of pro-inflammatory cytokines such as IL-1 β , TNF α , etc., which induces chondrocyte apoptosis and pyroptosis, promotes cartilage matrix degradation, and aggravates OA progression [16]. Therefore, inhibiting NF- κ B pathway activation and reducing inflammatory response can aid in protecting cartilage tissues and alleviating OA progression. In our study as well, the NF- κ B pathway was activated in both IL-1 β -stimulated chondrocytes and DMM-induced mice, and MON treatment blocked the pathway via inhibiting the phosphorylation of I κ B α and nuclear translocation of p65. Furthermore, the recombinant NF- κ B reversed the

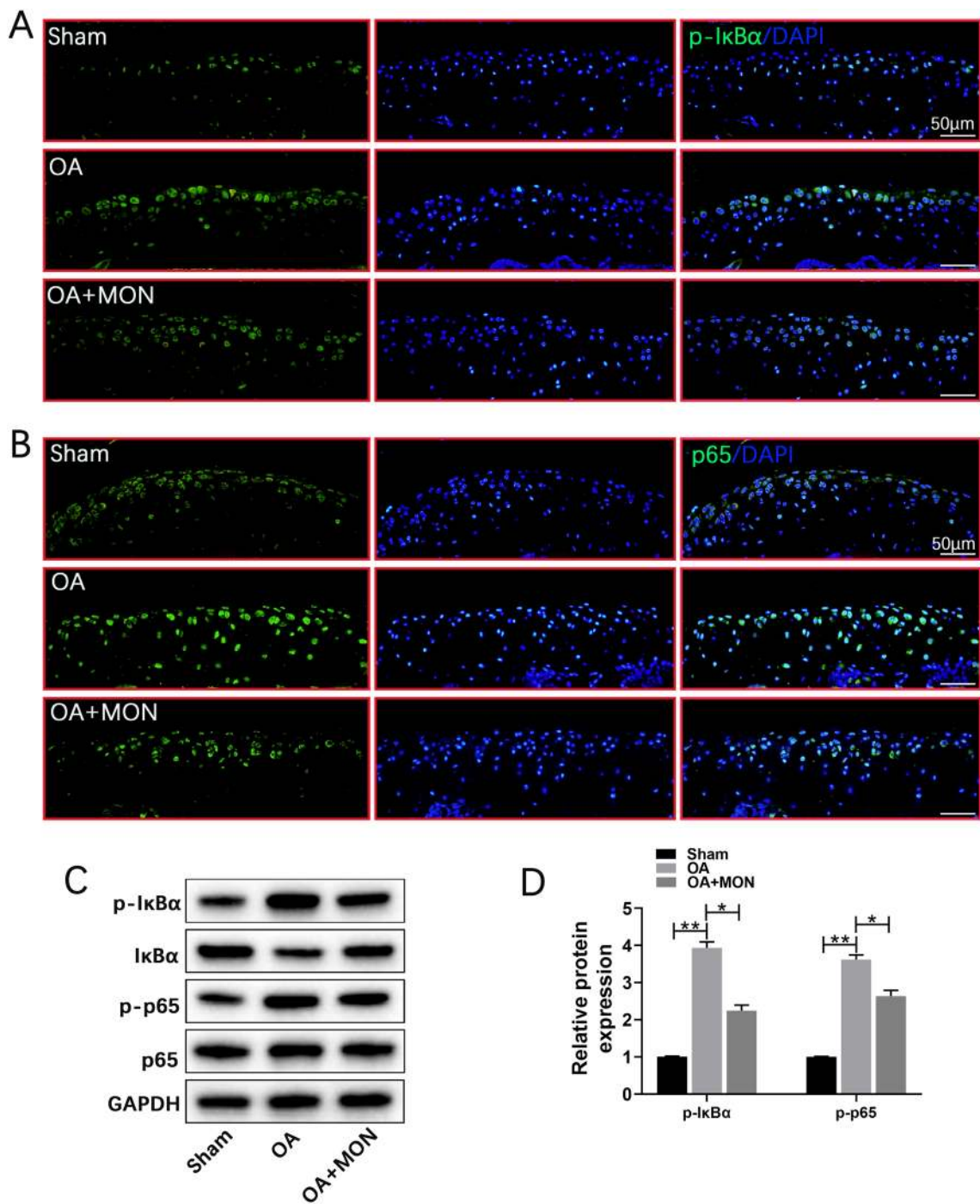


Fig. 10 MON inhibited the activation of NF-κB signaling pathway in DMM mice. **A, B** Immunofluorescence staining showing the phosphorylation of IκBα (green) and the nuclear translocation of p65 (green) in mice knee joints at 56 days after MON treatment. **C, D** Western blot analysis and relative quantification showing the expression levels of NF-κB signaling pathway related proteins (p-IκBα, IκBα, p-p65 and p65) in DMM mice at 56 days after MON treatment. All experiments were performed in triplicated and data were presented as the mean ± SD, *n* = 3 per group. **P* < 0. 05, ***P* < 0. 01

therapeutic effects of MON, indicating that the NF-κB signaling pathway is critical to the role of MON in the chondrocytes. Previous studies have also shown that MON inactivated the NF-κB pathway in different disease models, such as chronic colitis and acute kidney injury [4, 45]. Jiang et al. [15] reported that MON alleviated

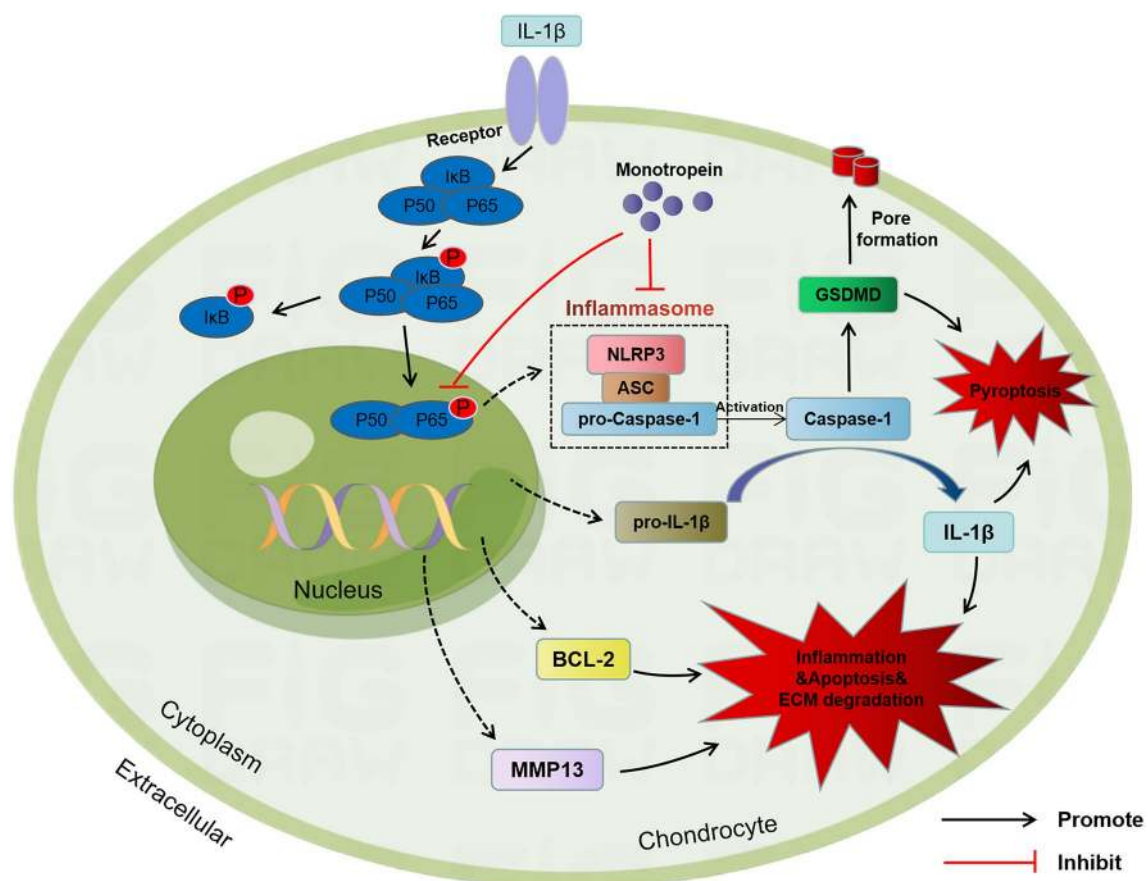


Fig. 11 A schematic diagram illustrating the theory in the current study. MON inhibited cartilage matrix degradation and alleviated apoptosis and pyroptosis in the in vitro and in vivo models of OA by targeting the NF-κB signaling pathway

inflammation, oxidative stress and apoptosis in senescent endothelial cells by inhibiting NF-κB signaling.

The regulatory relationship between NF-κB signaling pathway and pyroptosis has been elucidated in recent years [22, 41, 42]. NF-κB lies upstream of NLRP3 and induces its transcription [7, 22, 35]. Activation of the NF-κB pathway dissociates NF-κB from IκB, and the free NF-κB translocates to the nucleus and transcriptionally activates the NLRP3 inflammasome, thus promoting the release of IL-1β and IL-18, which leads to chondrocyte pyroptosis [14, 38, 40]. Previous studies have shown that Moroniside alleviated chondrocyte pyroptosis in OA mice by reducing the levels of NLRP3 inflammasome and caspase-1 via NF-κB inhibition [41, 42]. Another study showed that Loganin significantly inhibited the NF-κB pathway and decreased the levels of cryopyrin and caspase-1 in OA mice, which prevented chondrocyte pyroptosis [11]. Therefore, targeted inhibition of the NF-κB signaling pathway may protect chondrocytes from pyroptosis and thus alleviate OA progression.

Limitations

However, our current study also has some limitations. First of all, we used positive control drug in the preliminary experiment, however, there were no positive drug for OA mouse model in the formal experiment. Furthermore, micro-CT were not used for the evaluation of the bone situation especially cartilage erosion due to lack of funding. Further studies will be performed by us in the future to refine these limitations.

Conclusion

In summary, MON inhibited cartilage matrix degradation and alleviated apoptosis and pyroptosis in the in vitro and in vivo models of OA by targeting the NF-κB signaling pathway. Therefore, MON is a promising drug for promoting cartilage repair and stalling OA progression, and warrants clinical validation.

Abbreviations

ASC	Apoptosis-associated speck-like protein containing a CARD
BCA	Bicinchoninic acid
CCK-8	Cell counting kit-8
DMEM	Dulbecco's modified eagle medium
DMM	Destabilization of the medial meniscus
EDTA	Ethylenediaminetetraacetic acid
EdU	Ethynyl-deoxyuridine
FBS	Fetal bovine serum
GSDMD	Gasdermin D
HE	Hematoxylin–eosin
MMP13	Matrix metalloproteinase 13
MMP3	Matrix metalloproteinase 3
MON	Monotropein
NLRP3	NLR family pyrin domain containing 3
OA	Osteoarthritis
OARSI	Osteoarthritis Research Society International
PBS	Phosphate-buffered saline
p-IkBα	Phosphorylated IκBα
PVDF	Polyvinylidene difluoride
RIPA	Radio Immunoprecipitation Assay
R-NF-κB	Recombinant NF-κB
RT	Room temperature
SDs	Standard deviations
SPF	Specific pathogen-free
TBST	Tris buffer solution
TLR ₄	Toll-like receptor 4

Acknowledgements

Not applicable.

Author contributions

ZL and ZC performed the experiments and wrote the paper draft. ZL and JC analyzed the data and prepared the figures. ZL, HS, XW and JW performed the experiments. DZ and XC supervised the experimenters and corrected the paper draft. All authors read and approved the final manuscript.

Funding

This study was supported by a grant from the Construction project of the inheritance studio of Xianzhang Huang, a famous Chinese medicine doctor in Guangdong Province (NO. 20200741, 20212148, 20220037) and Guangdong Provincial Hospital of Traditional Chinese Medicine and the School of Biomedicine, Chinese University of Hong Kong School of Medicine, Basic Clinical Collaborative Innovation Project (NO. YN2018HK04).

Availability of data and materials

The data used to support the findings of this study are available from the corresponding author upon request.

Declarations

Ethics approval and consent to participate

All animal experiments were approved by the Animal Ethics Committee of Guangzhou University of Chinese Medicine (NO. 44005800012467) and performed according to the guidelines of the Chinese National Institutes of Health.

Consent for publication

We declare that the publisher has the author's permission to publish the relevant contribution.

Competing interests

The authors declare that there is no competing interest regarding the publication of this paper.

Author details

¹The Second Clinical College of Guangzhou, University of Chinese Medicine, The Second Affiliated Hospital of Guangzhou University of Chinese Medicine, Guangdong Provincial Hospital of Chinese Medicine, Guangzhou 510120, Guangdong, China. ²The First Clinical College of Guangzhou, University

of Chinese Medicine, Guangzhou 510405, Guangdong, China. ³Zhongshan Hospital of Traditional Chinese Medicine Affiliated to Guangzhou University of Traditional Chinese Medicine, Zhongshan 528401, Guangdong, China.

⁴Department of Orthopaedic Surgery, Guangdong Provincial Hospital of Chinese Medicine, 111 Dade Road, Yuexiu District, Guangzhou 510120, Guangdong, China. ⁵Department of Rehabilitation, Guangdong Provincial Hospital of Chinese Medicine, 261 Datong Road, Yuexiu District, Guangzhou 510105, Guangdong, China.

Received: 26 December 2022 Accepted: 7 April 2023

Published online: 19 April 2023

References

- An S, Hu H, Li Y, Hu Y. Pyroptosis plays a role in osteoarthritis. *Aging Dis*. 2020;11:1146–57.
- Arden N, Nevitt MC. Osteoarthritis: epidemiology. *Best Pract Res Clin Rheumatol*. 2006;20:3–25.
- Cao Y, Tang S, Nie X, Zhou Z, Ruan G, Han W, Zhu Z, Ding C. Decreased miR-214-3p activates NF-κB pathway and aggravates osteoarthritis progression. *EBioMedicine*. 2021;65:103283.
- Chen Y, Lu Y, Pei C, Liang J, Ding P, Chen S, Hou SZ. Monotropein alleviates secondary liver injury in chronic colitis by regulating TLR4/NF-κB signaling and NLRP3 inflammasome. *Eur J Pharmacol*. 2020;883:173358.
- Choi MC, Jo J, Park J, Kang HK, Park Y. NF-κB signaling pathways in osteoarthritic cartilage destruction. *Cells-Basel*. 2019. <https://doi.org/10.3390/cells8070734>.
- Glasson SS, Chambers MG, Van Den Berg WB, Little CB. The OARSI histopathology initiative - recommendations for histological assessments of osteoarthritis in the mouse. *Osteoarthritis Cartilage*. 2010;18(Suppl 3):S17–23.
- Guo H, Callaway JB, Ting JP. Inflammasomes: mechanism of action, role in disease, and therapeutics. *Nat Med*. 2015;21:677–87.
- Guo Q, Chen X, Chen J, Zheng G, Xie C, Wu H, Miao Z, Lin Y, Wang X, Gao W, Zheng X, Pan Z, Zhou Y, Wu Y, Zhang X. STING promotes senescence, apoptosis, and extracellular matrix degradation in osteoarthritis via the NF-κB signaling pathway. *Cell Death Dis*. 2021;12:13.
- Han D, Fang Y, Tan X, Jiang H, Gong X, Wang X, Hong W, Tu J, Wei W. The emerging role of fibroblast-like synoviocytes-mediated synovitis in osteoarthritis: an update. *J Cell Mol Med*. 2020;24:9518–32.
- Hou Y, Luan J, Huang T, Deng T, Li X, Xiao Z, Zhan J, Luo D, Hou Y, Xu L, Lin D. Tauroursodeoxycholic acid alleviates secondary injury in spinal cord injury mice by reducing oxidative stress, apoptosis, and inflammatory response. *J Neuroinflammation*. 2021;18:216.
- Hu J, Zhou J, Wu J, Chen Q, Du W, Fu F, Yu H, Yao S, Jin H, Tong P, Chen D, Wu C, Ruan H. Loganin ameliorates cartilage degeneration and osteoarthritis development in an osteoarthritis mouse model through inhibition of NF-κB activity and pyroptosis in chondrocytes. *J Ethnopharmacol*. 2020;247:112261.
- Hunter DJ, March L, Chew M. Osteoarthritis in 2020 and beyond: a lancet commission. *Lancet*. 2020;396:1711–2.
- Hwang HS, Kim HA. Chondrocyte apoptosis in the pathogenesis of osteoarthritis. *Int J Mol Sci*. 2015;16:26035–54.
- Jia S, Yang Y, Bai Y, Wei Y, Zhang H, Tian Y, Liu J, Bai L. Mechanical stimulation protects against chondrocyte pyroptosis through irisinduced suppression of PI3K/Akt/NF-κB signaling pathway in osteoarthritis. *Front Cell Dev Biol*. 2022;10:797855.
- Jiang F, Xu XR, Li WM, Xia K, Wang LF, Yang XC. Monotropein alleviates H₂O₂ induced inflammation, oxidative stress and apoptosis via NFκB/AP1 signaling. *MOL MED REP*. 2020;22:4828–36.
- Jimi E, Fei H, Nakatomi C. 2019. NF-κB Signaling regulates physiological and pathological chondrogenesis. *INT J MOL SCI* 20.
- Jones IA, Togashi R, Wilson ML, Heckmann N, Vangsness CJ. Intra-articular treatment options for knee osteoarthritis. *NAT REV RHEUMATOL*. 2019;15:77–90.
- Katz JN, Arant KR, Loeser RF. Diagnosis and treatment of hip and knee osteoarthritis: a review. *JAMA*. 2021;325:568–78.
- Kloppenburg M, Berenbaum F. Osteoarthritis year in review 2019: epidemiology and therapy. *Osteoarthritis Cartilage*. 2020;28:242–8.

20. Kovacs SB, Miao EA. Gasdermins: effectors of pyroptosis. *Trends Cell Biol.* 2017;27:673–84.
21. Lawrence T. The nuclear factor NF-kappaB pathway in inflammation. *Cold Spring Harb Perspect Biol.* 2009;1: a1651.
22. Li Z, Huang Z, Zhang H, Lu J, Tian Y, Wei Y, Yang Y, Bai L. P2X7 receptor induces pyroptotic inflammation and cartilage degradation in osteoarthritis via NF-kappaB/NLRP3 crosstalk. *Oxid Med Cell Longev.* 2021;2021:8868361.
23. Luo D, Li X, Hou Y, Hou Y, Luan J, Weng J, Zhan J, Lin D. Sodium tanshinone IIA sulfonate promotes spinal cord injury repair by inhibiting blood spinal cord barrier disruption in vitro and in vivo. *Drug Dev Res.* 2022;83:669–79.
24. McAllister MJ, Chemaly M, Eakin AJ, Gibson DS, McGilligan VE. NLRP3 as a potentially novel biomarker for the management of osteoarthritis. *Osteoarthritis Cartilage.* 2018;26:612–9.
25. Mehana EE, Khafaga AF, El-Blehi SS. The role of matrix metalloproteinases in osteoarthritis pathogenesis: an updated review. *Life Sci.* 2019;234: 116786.
26. Poole AR, Kobayashi M, Yasuda T, Lavery S, Mwale F, Kojima T, Sakai T, Wahl C, El-Maadawy S, Webb G, Tchetina E, Wu W. Type II collagen degradation and its regulation in articular cartilage in osteoarthritis. *Ann Rheum Dis.* 2002;61(Suppl 2):i78–81.
27. Prieto-Alhambra D, Judge A, Javaid MK, Cooper C, Diez-Perez A, Arden NK. Incidence and risk factors for clinically diagnosed knee, hip and hand osteoarthritis: influences of age, gender and osteoarthritis affecting other joints. *Ann Rheum Dis.* 2014;73:1659–64.
28. Qvist P, Bay-Jensen AC, Christiansen C, Dam EB, Pastoureau P, Karsdal MA. The disease modifying osteoarthritis drug (DMOAD): is it in the horizon? *Pharmacol Res.* 2008;58:1–7.
29. Rahmati M, Nalesso G, Mobasheri A, Mozafari M. Aging and osteoarthritis: central role of the extracellular matrix. *Ageing Res Rev.* 2017;40:20–30.
30. Shi J, Gao W, Shao F. Pyroptosis: gasdermin-mediated programmed necrotic cell death. *Trends Biochem Sci.* 2017;42:245–54.
31. Shi Y, Liu XY, Jiang YP, Zhang JB, Zhang QY, Wang NN, Xin HL. Monotropein attenuates oxidative stress via Akt/mTOR-mediated autophagy in osteoblast cells. *Biomed Pharmacother.* 2020;121:109566.
32. Wang BW, Jiang Y, Yao ZL, Chen PS, Yu B, Wang SN. Aucubin protects chondrocytes against il-1beta-induced apoptosis in vitro and inhibits osteoarthritis in mice model. *Drug Des Devel Ther.* 2019;13:3529–38.
33. Wang C, Mao C, Lou Y, Xu J, Wang Q, Zhang Z, Tang Q, Zhang X, Xu H, Feng Y. Monotropein promotes angiogenesis and inhibits oxidative stress-induced autophagy in endothelial progenitor cells to accelerate wound healing. *J Cell Mol Med.* 2018;22:1583–600.
34. Wang F, Wu L, Li L, Chen S. Monotropein exerts protective effects against IL-1beta-induced apoptosis and catabolic responses on osteoarthritis chondrocytes. *Int Immunopharmacol.* 2014;23:575–80.
35. Wang Q, Ou Y, Hu G, Wen C, Yue S, Chen C, Xu L, Xie J, Dai H, Xiao H, Zhang Y, Qi R. Naringenin attenuates non-alcoholic fatty liver disease by down-regulating the NLRP3/NF-kappaB pathway in mice. *Br J Pharmacol.* 2020;177:1806–21.
36. Whittaker JL, Truong LK, Dhiman K, Beck C. Osteoarthritis year in review 2020: rehabilitation and outcomes. *Osteoarthritis Cartilage.* 2021;29:190–207.
37. Won Y, Shin Y, Chun CH, Cho Y, Ha CW, Kim JH, Chun JS. Pleiotropic roles of metallothioneins as regulators of chondrocyte apoptosis and catabolic and anabolic pathways during osteoarthritis pathogenesis. *Ann Rheum Dis.* 2016;75:2045–52.
38. Yang G, Wang Y, Chen Y, Huang R. UFL1 attenuates IL-1beta-induced inflammatory response in human osteoarthritis chondrocytes. *Int Immunopharmacol.* 2020;81:106278.
39. Yang J, Hu S, Bian Y, Yao J, Wang D, Liu X, Guo Z, Zhang S, Peng L. Targeting cell death: pyroptosis, ferroptosis, apoptosis and necroptosis in osteoarthritis. *Front Cell Dev Biol.* 2021;9:789948.
40. Yang W, Zhang S, Ou T, Jiang H, Jia D, Qi Z, Zou Y, Qian J, Sun A, Ge J. Interleukin-11 regulates the fate of adipose-derived mesenchymal stem cells via STAT3 signalling pathways. *Cell Prolif.* 2020;53:e12771.
41. Yu H, Yao S, Zhou C, Fu F, Luo H, Du W, Jin H, Tong P, Chen D, Wu C, Ruan H. Morroniside attenuates apoptosis and pyroptosis of chondrocytes and ameliorates osteoarthritic development by inhibiting NF-kappaB signaling. *J Ethnopharmacol.* 2021;266:113447.
42. Yu P, Zhang X, Liu N, Tang L, Peng C, Chen X. Pyroptosis: mechanisms and diseases. *Signal Transduct Target Ther.* 2021;6:128.
43. Zhang L, Xing R, Huang Z, Zhang N, Zhang L, Li X, Wang P. Inhibition of synovial macrophage pyroptosis alleviates synovitis and fibrosis in knee osteoarthritis. *Mediators Inflamm.* 2019;2019:2165918.
44. Zhang T, Ma C, Zhang Z, Zhang H, Hu H. NF-kappaB signaling in inflammation and cancer. *MedComm.* 2021;2020(2):618–53.
45. Zhang Y, Chen Y, Li B, Ding P, Jin D, Hou S, Cai X, Sheng X. The effect of monotropein on alleviating cisplatin-induced acute kidney injury by inhibiting oxidative damage, inflammation and apoptosis. *Biomed Pharmacother.* 2020;129: 110408.
46. Zhang Y, Lin Z, Chen D, He Y. CY-09 attenuates the progression of osteoarthritis via inhibiting NLRP3 inflammasome-mediated pyroptosis. *Biochem Biophys Res Commun.* 2021;553:119–25.
47. Zhang Z, Zhang Q, Yang H, Liu W, Zhang N, Qin L, Xin H. Monotropein isolated from the roots of *Morinda officinalis* increases osteoblastic bone formation and prevents bone loss in ovariectomized mice. *Fitoterapia.* 2016;110:166–72.
48. Zheng L, Zhang Z, Sheng P, Mobasheri A. The role of metabolism in chondrocyte dysfunction and the progression of osteoarthritis. *Ageing Res Rev.* 2021;66: 101249.
49. Zu Y, Mu Y, Li Q, Zhang ST, Yan HJ. Icariin alleviates osteoarthritis by inhibiting NLRP3-mediated pyroptosis. *J Orthop Surg Res.* 2019;14:307.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

